BSG-MDS Practical 2 Statistical Genetics

Biel Caballero and Gerard Gomez

07/11/2023, submission deadline 14/11/2023

```
## Loading required package: combinat
##
## Attaching package: 'combinat'
## The following object is masked from 'package:utils':
##
       combn
## Loading required package: gdata
##
## Attaching package: 'gdata'
## The following objects are masked from 'package:data.table':
##
       first, last
##
  The following object is masked from 'package:stats':
##
##
##
##
  The following object is masked from 'package:utils':
##
##
       object.size
## The following object is masked from 'package:base':
##
##
       startsWith
## Loading required package: gtools
## Loading required package: MASS
## Loading required package: mvtnorm
##
## NOTE: THIS PACKAGE IS NOW OBSOLETE.
##
##
     The R-Genetics project has developed an set of enhanced genetics
     packages to replace 'genetics'. Please visit the project homepage
##
     at http://rgenetics.org for informtion.
##
##
##
## Attaching package: 'genetics'
```

```
## The following objects are masked from 'package:base':
##
##
       %in%, as.factor, order
## Loading required package: mice
##
## Attaching package: 'mice'
##
  The following object is masked from 'package:stats':
##
##
       filter
## The following objects are masked from 'package:base':
##
       cbind, rbind
##
## Loading required package: Rsolnp
## Loading required package: nnet
#Hardy Weinberg Equilibrium
##Create dataset
data<-fread("TSIChr22v4.raw", header = TRUE)</pre>
geneticData <- as.data.frame(data[,c(-1:-6)])</pre>
```

1. How many variants are there in this database? What percentage of the data is missing?

```
print(paste0("There are ",ncol(geneticData), " varaints"))

## [1] "There are 1102156 varaints"

sum(is.na(data))

## [1] 0

print("No data is missing")

## [1] "No data is missing"
```

2. Calculate the percentage of monomorphic variants. Exclude all monomorphics from the database for all posterior computations of the practical. How many variants do remain in your database?

```
monomorphicVariants <- c()
for(i in 1:ncol(geneticData)){
   if(length(unique(geneticData[,i])) == 1){
      monomorphicVariants <- append(monomorphicVariants,i)
   }
}
print(paste0("The parcentage of monomorphic variants are ", length(monomorphicVariants)/ncol(geneticDat
## [1] "The parcentage of monomorphic variants are 0.810304530393157"
noMonomorphic <- geneticData[,-monomorphicVariants]</pre>
```

3. Extract polymorphism rs587756191_T from the data, and determine its genotype counts. Apply a chi-square test for Hardy-Weinberg equilibrium, with and without continuity correction. Also try an exact test, and a permutation test. You can use thee functions HWChisq, HWExact and HWPerm for this purpose. Do you think this variant is in equilibrium? Argue your answer.

```
library(HardyWeinberg)
rs587756191_T <- noMonomorphic[,"rs587756191_T"]
counts <- table(rs587756191_T)</pre>
HWChisq(c(AA = 106, AB = 1, BB = 0))
## Warning in HWChisq(c(AA = 106, AB = 1, BB = 0)): Expected counts below 5: chi-
## square approximation may be incorrect
## Chi-square test with continuity correction for Hardy-Weinberg equilibrium (autosomal)
## Chi2 = 106.2512 DF = 1 p-value = 6.495738e-25 D = 0.002336449 f = -0.004694836
HWExact(c(AA = 106, AB = 1, BB = 0))
## Haldane Exact test for Hardy-Weinberg equilibrium (autosomal)
## using SELOME p-value
## sample counts: nAA = 106 nAB = 1 nBB = 0
## HO: HWE (D==0), H1: D <> 0
## D = 0.002336449 \text{ p-value} = 1
HWPerm(c(AA = 106, AB = 1, BB = 0))
## Permutation test for Hardy-Weinberg equilibrium
## Observed statistic: 0.002358439
                                      17000 permutations. p-value: 1
All tests give a p-value of 1, hence they are not in Hardy-Weiberg Equilibrium
```

4. Determine the genotype counts for all polymorphic variants, and store them in a p \times 3 matrix.

```
polymorphic <- matrix(nrow = ncol(noMonomorphic),ncol = 3)
for(i in 1:ncol(noMonomorphic)){
  tab = table(noMonomorphic[,i])
  na = names(tab)
  for(n in na){
    polymorphic[i,as.numeric(n)+1] <- tab[n]
  }
}</pre>
```

5. Apply an exact test for Hardy-Weinberg equilibrium to each SNP. You can use function HWExactStats for fast computation. What is the percentage of significant SNPs (use alpha = 0.05)? Is this the number of markers that you would expect to be out of equilibrium by the effect of chance alone?

```
hwe<-HWExactStats(polymorphic, x.linked = FALSE)
hwe_signSNP<-which(hwe<0.05)
print(paste0("The parcentage of significant SNPs are ", round((length(hwe_signSNP)*100)/length(hwe),1),</pre>
```

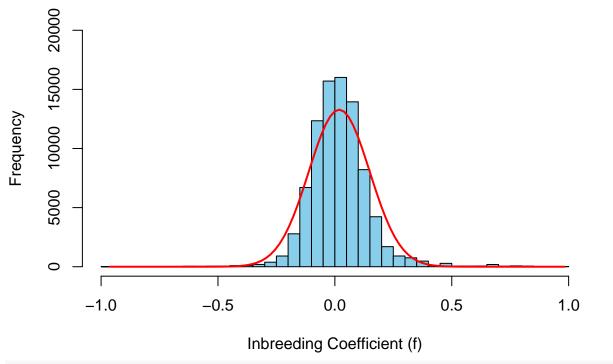
[1] "The parcentage of significant SNPs are 2.5%"

```
print("The number of SNPs we would expect to be out of equilibrium is around 60%")
## [1] "The number of SNPs we would expect to be out of equilibrium is around 60%"
6. Which SNP is most significant according to the exact test results? Give its genotype counts.
In which sense is this genotypic composition unusual?
ms_signSNP<-polymorphic[which.min(hwe),c(1:3)]</pre>
print(paste0("The genotype counts are: AA = ",ms_signSNP[1], " AB = ",ms_signSNP[2], " and BB = ",ms_si
## [1] "The genotype counts are: AA = 56 AB = 1 and BB = 50"
print("These genotypic composition is unusual because the predominant genotypes are the monoallelic one
## [1] "These genotypic composition is unusual because the predominant genotypes are the monoallelic on
7. Compute the inbreeding coefficient (f) for each SNP, and make a histogram of f. You can
use function HWf for this purpose. Give descriptive statistics (mean, standard deviation, etc)
of f calculated over the set of SNPs. What distribution do you expect f to follow theoretically?
Use a probability plot to confirm your idea.
ic<-HWf(polymorphic)</pre>
summary(ic)
##
      Min. 1st Qu. Median
                              Mean 3rd Qu.
                                               Max.
                                                       NA's
##
     -0.96
             -0.06
                              0.02
                      0.01
                                      0.08
                                               0.98
                                                    122389
print(paste0("The descriptive statistiucs are:"))
## [1] "The descriptive statistiucs are:"
                  - Mean: ",summary(ic)[4]))
print(paste0("
## [1] "
            - Mean: 0.0187182056859644"
print(paste0("
                - Standard deviation: ",sd(ic, na.rm = TRUE)))
## [1] "
            - Standard deviation: 0.130155015109863"
print(paste0("
                  - Min: ",summary(ic)[1]))
            - Min: -0.962616822429907"
print(paste0("
                - Max: ",summary(ic)[6]))
            - Max: 0.981249452378866"
print(paste0("
                - Number of NAs: ", summary(ic)[7]))
           - Number of NAs: 122389"
hist_data <- hist(ic, breaks = 30, col = "skyblue", xlab = "Inbreeding Coefficient (f)",
```

ylab = "Frequency", main = "Histogram of Inbreeding Coefficients", plot = FALSE)

```
ylab = "Frequency", main = "Histogram of Inbreeding Coefficients", ylim = c(0,20000))
print("The distribution we expect f to follow theoretically is a normal distribution")
## [1] "The distribution we expect f to follow theoretically is a normal distribution"
# Fit a normal distribution curve to the histogram
mu <- mean(ic, na.rm = TRUE)
sigma <- sd(ic, na.rm = TRUE)
x <- seq(min(ic, na.rm = TRUE), max(ic, na.rm = TRUE), length.out = 100)
y <- dnorm(x, mean = mu, sd = sigma) * length(ic[is.finite(ic)]) * diff(breaks[1:2])
lines(x, y, col = "red", lwd = 2)</pre>
```

Histogram of Inbreeding Coefficients



print("The distribution we expect f to follow theoretically is a normal distribution")

[1] "The distribution we expect f to follow theoretically is a normal distribution"

8. Apply the exact test for HWE to each SNP, using different significant levels. Report the number and percentage of significant variants using an exac test for HWE with alpha = 0.10, 0.05, 0.01 and 0.001. State your conclusions.

```
hwe<-HWExactStats(polymorphic, x.linked = FALSE)
hwe_signSNP<-which(hwe<0.1)
print(paste0("The number of significant SNPs (alpha = 0.1) parcentage of significant SNPs (alpha = 0.1)</pre>
```

[1] "The number of significant SNPs (alpha = 0.1) parcentage of significant SNPs (alpha = 0.1) are 4
hwe_signSNP<-which(hwe<0.05)
print(paste0("The number of significant SNPs (alpha = 0.05) parcentage of significant SNPs (alpha = 0.0</pre>

[1] "The number of significant SNPs (alpha = 0.05) parcentage of significant SNPs (alpha = 0.05) are

```
hwe_signSNP<-which(hwe<0.01)
print(paste0("The number of significant SNPs (alpha = 0.01) parcentage of significant SNPs (alpha = 0.0
## [1] "The number of significant SNPs (alpha = 0.01) parcentage of significant SNPs (alpha = 0.01) are
hwe_signSNP<-which(hwe<0.001)
print(paste0("The number of significant SNPs (alpha = 0.001) parcentage of significant SNPs (alpha = 0.001)
```

[1] "The number of significant SNPs (alpha = 0.001) parcentage of significant SNPs (alpha = 0.001) a