



Review Paper

Past, present, and future perspectives of environmental DNA (eDNA) metabarcoding: A systematic review in methods, monitoring, and applications of global eDNA

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ABSTRACT

Environmental DNA (eDNA) metabarcoding is a novel method of assessing biodiversity wherein samples are taken from the environment via water, sediment or air from which DNA is extracted, and then amplified using general or universal primers in polymerase chain reaction and sequenced using next-generation sequencing to generate thousands to millions of reads. From this data, species presence can be determined, and overall biodiversity assessed. It is an interdisciplinary method that brings together traditional field-based ecology with in-depth molecular methods and advanced computational tools. As an emerging monitoring method, there are many pitfalls and roadblocks to be considered and avoided, but the method may still have the ability to revolutionize modern biodiversity surveys for the molecular era. In this paper, we review the basic methodology, benefits, and concerns of eDNA metabarcoding, and systematically cover the applications of the method in global ecology thus far, including biodiversity monitoring across all habitats and taxonomic groups, ancient ecosystem reconstruction, plant-pollinator interactions, diet analysis, invasive species detection, pollution responses, and air quality monitoring. We also discuss the future applications of the method as well as expected technological advances and how they may impact the way that eDNA metabarcoding may be used in the future. eDNA metabarcoding is a unique method still in development and will likely remain in flux for some time as technology advances and procedures become standardized. However, as metabarcoding is optimized and its use becomes more widespread, it is likely to become an essential tool for ecological monitoring and global conservation study.

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1. Introduction

Environmental DNA or eDNA describes the genetic material present in environmental samples such as sediment, water, and air, including whole cells, extracellular DNA and potentially whole organisms (Ficetola et al., 2008; Barnes and Turner,

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2016). eDNA can be captured from environmental samples and preserved, extracted, amplified, sequenced, and categorized based on its sequence (Deiner et al., 2015). From this information, detection and classification of species is possible. eDNA may come from skin, mucous, saliva, sperm, secretions, eggs, feces, urine, blood, roots, leaves, fruit, pollen, and rotting bodies of larger organisms, while microorganisms may be obtained in their entirety (Taberlet et al., 2012a; Bohmann et al., 2014; Barnes and Turner, 2016). eDNA production is dependent on biomass, age and feeding activity of the organism as well as physiology, life history, and space use (Barnes and Turner, 2016; Goldberg et al., 2016; Hering et al., 2018). Despite being a relatively new method of surveying, eDNA has already proven to have enormous potential in biological monitoring. Conventional methods for surveying richness and abundance are limited by taxonomic identification, may cause disturbance or destruction of habitat, and may rely on methods in which it is difficult to detect small or elusive species, thus making estimates for entire communities impossible. eDNA can complement these methods by targeting different species, sampling greater diversity, and increasing taxonomic resolution (Deiner et al., 2017). Additionally, eDNA is capable of detecting rare species, but not of determining population quality information such as sex ratios and body conditions, so it is ideal for supplementing traditional studies (Goldberg et al., 2016; Deiner et al., 2017). Regardless, it has useful applications in detecting the first occurrences of invasive species, the continued presence of native species thought to be extinct or otherwise threatened, and other elusive species occurring in low densities that would be difficult to detect by traditional means.

Degradation of eDNA in the environment limits the scope of eDNA studies, as often only small segments of genetic material remain, particularly in warm, tropical regions. Additionally, the varying lengths of time to degradation based on environmental conditions and the potential of DNA to travel throughout media such as water can affect inference of fine-scale spatiotemporal trends of species and communities (Coissac et al., 2012; Taberlet et al., 2012a; Eichmiller et al., 2016; Goldberg et al., 2016; Deiner et al., 2017; Hering et al., 2018). Despite these drawbacks, eDNA still has the potential to determine relative or rank abundance as some studies have found it to correspond with biomass, though the variation inherent in environmental samples makes it difficult to quantify (Bohmann et al., 2014; Hering et al., 2018). While eDNA has numerous applications in conservation, monitoring, and ecosystem assessment, as well as others yet to be described, the highly variable concentrations of eDNA and potential heterogeneity through the water body makes it essential that the procedure is optimized, ideally with a pilot study for each new application to ensure that the sampling design is appropriate to detect the target (Carew et al., 2013; Goldberg et al., 2016; Deiner et al., 2017).

While the definition of eDNA seems straightforward, the lines between different forms of DNA become blurred, particularly in comparison to community DNA, which is described as bulk organismal samples (Deiner et al., 2017). A question arises regarding whole microorganisms captured in eDNA samples: do these organisms alter the classification of the sample to a community DNA sample? Additionally, the classification of genetic material from feces is problematic and often referred to as eDNA (Deiner et al., 2017). Differentiation between the two is important as community DNA indicates organismal presence at a particular time and place, while eDNA may have come from a different location, from predator feces, or from past presence, however this differentiation is often impossible (Creer et al., 2016; Deiner et al., 2017). For the sake of simplicity, this paper will consider eDNA loosely classified as including many sectors of DNA biodiversity research, including fecal analysis and bulk samples when they are applicable to biodiversity research and ecosystem analysis.

Methods in eDNA research have recently expanded to be able to assess whole communities from a single sample. This process involves metabarcoding, which can be precisely defined as the use of general or universal polymerase chain reaction (PCR) primers on mixed DNA samples from any origin followed by high-throughput next-generation sequencing (NGS) to determine the species composition of the sample. This method has been common in microbiology for years, but is only just finding its footing in assessment of macroorganisms (Coissac et al., 2012; Creer et al., 2016; Deiner et al., 2017). Ecosystem-wide applications of eDNA metabarcoding have the potential to not only describe communities and biodiversity, but also to detect interactions and functional ecology over large spatial scales, though it may be limited by false readings due to contamination or other errors (Bohmann et al., 2014; Ficetola et al., 2016; Creer et al., 2016; Hering et al., 2018). Altogether, eDNA metabarcoding increases speed, accuracy, and identification over traditional barcoding and decreases cost, but needs to be standardized and unified, integrating taxonomy and molecular methods for full ecological study (Coissac et al., 2012; Yu et al., 2012; Cristescu, 2014; Gibson et al., 2015; Hering et al., 2018).

This review is intended to provide an overview of methodological and bioinformatics considerations for eDNA metabarcoding studies, as well as to summarize published eDNA metabarcoding studies to date, describing some of the numerous applications therein. Additionally, critical considerations, areas needing improvement, and future directions will be discussed.

2. Methods and technical considerations

Within published eDNA metabarcoding studies, the methodologies often differ substantially, making cross-study comparisons impossible, but the uniqueness of each study site and target taxa likely make standardization a challenging goal (Hering et al., 2018). Regardless, a coordinated global initiative for biodiversity research and evaluation via metabarcoding has the potential to standardize methods to some degree (Cristescu, 2014; Hering et al., 2018). In this section, we will briefly cover the process of eDNA metabarcoding from sample collection to data analysis.

As eDNA can be applied to numerous ecosystems, collection methods must correspond with these different sample types. Microbes and pollen can be easily collected from the air, biofilms can be swabbed or scraped, water can be precipitated or filtered, and sediments can be processed (Creer et al., 2016). For water samples, large volumes of water and multiple field

replicates should be used; however, the choice between precipitation and filtration (and subsequently filter size) is reliant on the target taxa, as different taxa can be isolated more efficiently in different ways (Deiner et al., 2015; Creer et al., 2016; Goldberg et al., 2016; Hering et al., 2018). For soil and sediment samples, larger volumes of sample over larger spatial scales are required from larger size classes of organisms, and samples should be extracted from multiple locations to avoid heterogeneity of samples and to better describe the biodiversity of the area (Taberlet et al., 2012b; Creer et al., 2016). In all studies, field equipment, supplies, and personnel must be separated from areas where PCR, tissue, and organisms are processed or handled due to the high potential for contamination, and decontamination must occur between samples to maintain independence (Goldberg et al., 2016). Negative field controls are also essential to ensuring valid sampling and to identify contamination; this can include periodically filtering clean water as field blanks and processing the filters with the same protocol as the samples (Goldberg et al., 2016; Hering et al., 2018). Preservation methods are straightforward among sample types: drying, freezing at -20°C , or preserving in 100% ethanol or a cell lysis buffer are all adequate methods for sample preservation (Creer et al., 2016; Goldberg et al., 2016). Immediate preservation of samples is essential to avoid degradation, which can be affected by environmental variables such as temperature, pH, and exposure to light, as well as target species (Barnes and Turner, 2016; Strickler et al., 2015; Tsuji et al., 2017).

Laboratory procedures tend to be more homogenous among sample types but differ between studies. eDNA should be processed in a clean lab with equipment and supplies that cannot be removed; additionally, decontamination of personnel and supplies must occur before entering the clean lab (Goldberg et al., 2016). Processing areas in the lab should be cleaned with bleach and UV periodically, and during all procedures, filter pipet tips and clean gloves should be used (Champlot et al., 2010; Goldberg et al., 2016). In addition to the field blanks discussed previously, it is important to have a lab procedure that includes positive controls, negative controls, and replicates at all steps (Ficetola et al., 2016; Deiner et al., 2017). Kit-based extraction procedures are probably the most common, and are generally effective; however different kits may be more effective depending on the target taxa, particularly between eukaryotes and prokaryotes (Deiner et al., 2015; Creer et al., 2016). Phenol chloroform extractions are also effective, and may isolate more genetic material than other methods in certain scenarios (Creer et al., 2016; Deiner et al., 2017).

Perhaps one of the most important considerations in eDNA metabarcoding studies is PCR primer design. Different primers and regions differ in coverage, resolution, and bias between taxa. Cytochrome oxidase-I (COI) for metazoans and Ribulose biphosphate carboxylase large chain (*rcbL*) for plants are the standards established by the barcode of life, but other regions such as 12S or 16S ribosomal RNA may be more appropriate for different taxa (Epp et al., 2012; Taberlet et al., 2012b; Gibson et al., 2015; Creer et al., 2016; Deiner et al., 2017; Hering et al., 2018). Primers for eDNA metabarcoding need to be short enough to amplify degraded samples, identical within but variable between species, and flanked by highly conserved regions to amplify a variety of species without sacrificing specificity of the target group (Epp et al., 2012). In addition to these specifications, primer choice has the potential to bias results by preferentially amplifying some target sequences more than others, as well as amplifying non-target groups (Cristescu, 2014). One potential solution to this issue is the use of multiple primer sets, particularly evolutionarily independent primer sets coinciding with standardized barcodes for the target taxonomic groups (Drummond et al., 2015). Although it can reduce primer bias and increase taxonomic coverage, this method suffers from being more costly and time-consuming (Cristescu, 2014; Creer et al., 2016; Alberdi et al., 2018). Another important factor in PCR and primer design is in replicates; multiple PCR replicates increase species detection and decrease the likelihood of false negatives, but the number of replicates used often differs between studies and depends on detection probabilities, research objectives, sequencing depth, primer choice, cost constraints, and sequencing platform (Ficetola et al., 2015; Alberdi et al., 2018). It is also worth noting that the use of PCR replicates is not an adequate substitute for biological replicates; rather, both are necessary for comprehensive results (Goldberg et al., 2016). In running PCR for next generation sequencing (NGS), primers can be tagged with short nucleotide sequences to uniquely identify their source in a process commonly referred to as multiplexing. These tags, while useful, also have the potential to bias results, particularly when located on the 5' end, so they also require rigorous testing before implementation (Binladen et al., 2007; Berry et al., 2011). In the sequencing step a number of platforms are available, but Illumina sequencing currently outperforms other NGS platforms in terms of depth and accuracy (Deiner et al., 2017). It is also important to note that diversity has been found to increase with sequencing depth before reaching a plateau, so theoretically a maximum sequencing depth could be determined for individual studies (Alberdi et al., 2018).

The final technical step to be discussed is bioinformatics pathways. Unique bioinformatics pathways must be developed for eDNA metabarcoding and must be adapted for each study, previously requiring consultation with a computational specialist (Coissac et al., 2012). As technology has advanced, however, programs have been developed specifically for analysis of information from NGS data, making the method more accessible and user-friendly (Alberdi et al., 2018). NGS results in millions of reads which describe the genetic code of each strand of DNA that has been sequenced. These reads are typically grouped into operational taxonomic units (OTUs) which are not standard across sites, studies, or species, and cannot be universally applicable between them (Cristescu, 2014; Hering et al., 2018). OTUs are used to differentiate species or taxa based on similarity of genetic code as determined by sequencing, but they also need to be linked with ecological and physiological traits of species for proper identification. Various programs exist to help, but intragenetic variation hampers this process (Coissac et al., 2012; Cristescu, 2014; Deiner et al., 2017). OTU clustering is based on similarity to a specific sequence and subsequent grouping under similarity cutoffs, with 97–99% similarity being a common cutoff range; however ranges for studies depend on intraspecific diversity estimated from existing reference databases, such as GenBank (Deiner et al., 2017). Species identification from OTUs is often based on similarity (BLAST), phylogeny (maxlike), character (diagnostic regions),

classification algorithms, and coalescence (evaluation of quality using likelihood/Bayesian), or even species delimitation where sequences are partitioned based on differences and not similarity to a known sequence, but these should be combined with other ecological characteristics (Coissac et al., 2012; Deiner et al., 2017). Unassigned reads are another concern and their impact on analyses needs to be limited, but as so many species have yet to be described, analyses may wrongly exclude target species; poor species representation in databases may lead to incorrect identification and affect assessments of ecological quality (Deiner et al., 2017; Hering et al., 2018). Errors in false positives and negatives can be best avoided by combining metabarcoding data with direct observation and generalized occupancy models. It is recommended that occupancy models be run to evaluate detection probability, rate of false presences, and whether the level of replication is appropriate to control for false negatives (Ficetola et al., 2015, 2016; Deiner et al., 2017). Calculation of diversity indices is often used to influence diversity modelling and interpretation of results (Deiner et al., 2017). Ficetola et al. (2016) developed formulas to calculate the number of positives needed to be confident that they are not false positives:

$$\Pr(x \geq i | \text{site not occupied}) = \sum_{j=i}^k \binom{k}{j} \times p_{10}^j \times (1 - p_{10})^{k-j}$$

where p_{10} is the false positive rate, k is the number of replicated analyses on one sample, and i is the number of detections in PCR replicates from the same sample, and

$$\Pr(\text{site occupied} | x) = \frac{\psi p_{11}^x (1 - p_{11})^{k-x}}{\psi p_{11}^x (1 - p_{11})^{k-x} + (1 - \psi) p_{10}^x (1 - p_{10})^{k-x}}$$

where x is the number of detections, ψ is the estimated probabilities of occupancy, p_{11} is true detection, and p_{10} is false detection.

Quantitative analysis of eDNA metabarcoding data is questionable, and while there is some data that suggests a relationship between biomass and number of copies, there are many confounding variables, so multiple sampling occasions and replicates would be necessary as well as in-depth testing and optimization. Additionally, the scope of the study must be considered as general estimates may be appropriate for some studies (Goldberg et al., 2016; Deiner et al., 2017; Hering et al., 2018). There are some core programming skills required for analysis of metabarcoding data: the ability to operate from the command line, the capacity to use/manipulate/write programs in Perl or Python for analyzing large volumes of data, and familiarity with R (Creer et al., 2016). Some of the software often used for analyzing these datasets includes USEARCH, VSEARCH, CROP, swarm, UNIX, QIIME, mothur, OBITools, and phyloseq (Coissac et al., 2012; Creer et al., 2016). Most importantly, data from eDNA metabarcoding studies must be shared and accessible in its raw format with standardized metadata, ideally in cloud-based storage or common-storage databases to be used by future researchers (Creer et al., 2016; Deiner et al., 2017). Altogether, workflows are often determined by personal preference, available equipment, and cost (Deiner et al., 2015). Specificity and suggestions for bioinformatics pathways are also reviewed by Deiner et al. (2017). It is important to note that even if all methods in an eDNA metabarcoding study are executed flawlessly with primers that bind perfectly, some error should be expected. As eDNA is composed of small fragments heterogeneously distributed in the environment, there is always a chance that these fragments will not be picked up (Pilliod et al. 2013; Goldberg et al., 2016). This supports the need for lab-based testing before entering the field and replicates across sampling areas, but also the idea that absence cannot be confirmed; even if a species is not detected, that does not mean it is not there. Additionally, a positive eDNA result can be introduced through confounding sources (Darling and Mahon, 2011; Goldberg et al., 2016). Many issues with eDNA procedures may be resolved over time as the process matures, but the goal is to be focused on reducing falsehoods rather than perfect detection. Regardless, the data able to be gleaned from eDNA metabarcoding is invaluable and can be applied to a myriad of projects in a variety of ways.

3. Ecosystem and biodiversity monitoring

3.1. Advantages and disadvantages

Perhaps the most exciting and thus far the most explored avenue of eDNA metabarcoding research is the application for ecosystem and biodiversity monitoring, either for specific species groups, or potentially for entire ecosystem studies. Biodiversity is known to be declining worldwide, but it is difficult to monitor using traditional means (Thomsen and Willerslev, 2015). Additionally, some traditional survey methods have relied on destructive techniques and often require extensive taxonomic expertise, making them imperfect methods of detection (Thomsen and Willerslev, 2015; Holdaway et al., 2017). eDNA can remain viable for variable periods of time from weeks to hundreds of thousands of years, allowing for applications in molecular biology, ecology, palaeontology, and environmental sciences, particularly when coupled with next-generation sequencing (Thomsen and Willerslev, 2015). eDNA metabarcoding can even be automated to further streamline the procedure (Holdaway et al., 2017). Taking automation even further, supervised machine learning has been used to build predictive models for biodiversity monitoring without requiring taxonomic assignment, removing the need for human

interference in development of reference databases and subsequent species assignment, an innovation particularly exciting for monitoring of extremely diverse and sensitive groups of bioindicators that would require extensive effort to fully describe (Cordier et al., 2017). eDNA has superior species detectability, requires lower effort, causes no ecosystem disturbance, allows detection without a priori knowledge of species, and can be implemented in areas where traditional surveys are impossible, making it extremely promising for the future of biodiversity monitoring (Valentini et al., 2016).

There are concerns regarding methodology with eDNA metabarcoding, as would be expected with any emerging science. Every step of the procedure has the potential to influence the results, and when these results are in turn used to influence important conservation decisions, it is essential that the procedures be optimized and standardized. Many of these concerns are discussed in the previous section, but some specifically relevant to biodiversity monitoring include: optimization of sample collection and extraction; ensuring samples are free of contamination; designing primers to have minimal bias without sacrificing resolution or spread; development of extensive reference databases; using proper bioinformatics pathways; lack of standardization in classification; and difficulty in estimating DNA degradation rates, thus allowing for the misrepresentation of species presence in areas which they may have vacated (Deiner et al., 2015; Thomsen and Willerslev, 2015; Valentini et al., 2016; Holdaway et al., 2017). There is information on populations that simply cannot be obtained via metabarcoding. Population structure and size, sex and age ratios, and individual conditions cannot be obtained by using standard eDNA metabarcoding alone and are often critical to the development of conservation plans (Valentini et al., 2016; Evans et al., 2016a; Holdaway et al., 2017). Despite this, there are ways to accurately measure biodiversity with the integration of eDNA metabarcoding. The most common solution to this lack of data is to integrate metabarcoding into the traditional monitoring framework (Holdaway et al., 2017). While eDNA metabarcoding cannot ascertain information on population structure, it can detect rare, cryptic, or elusive species from a single sample, including species that may be considered pests, diseases, or species of conservation concern (Holdaway et al., 2017). An integration of the two methods can allow for the benefits of both without being hindered by the drawbacks of using only one; as such, this integrated biodiversity monitoring approach has been recommended to be used with government monitoring projects in New Zealand, including in biosecurity and agriculture (Holdaway et al., 2017). Fully removing the need for traditional sampling in the determination of ecological networks, it may be possible to describe interactions between species and their subsequent influence on ecosystem function from metabarcoding alone. Evans et al. (2016a,b) developed a methodology employing the use of nested tags allowing analysis of samples as individuals, enabling the construction of ecological networks from environmental samples alone. Traditional methods of describing ecological networks rely on indicator species or bipartite networks, but DNA-based networks have been found to be faster, more efficient, and more comprehensive than traditional methods, particularly as the complexity of the region increases or in poorly known ecosystems (Evans et al., 2016a). Overall, a combination of methods yields the most resolved networks, and has been suggested for use in conservation, restoration, and assessment of how human-induced climate change affects ecosystem function (Evans et al., 2016a). Altogether, an integration of conventional and novel techniques appears to be the next step in acquiring information for land management and biodiversity monitoring. The following discusses some of the studies in which eDNA metabarcoding has already been applied for monitoring across marine, estuarine, freshwater, and terrestrial ecosystems (Fig. 1).

3.2. Marine monitoring

3.2.1. Sediments

eDNA metabarcoding has been applied to marine environments by use of several collection procedures targeting many different groups of organisms. Marine sediments in particular are extremely species-rich, have key ecological roles and ecosystem services, and are sensitive to anthropogenic disturbance, but they have been poorly studied using traditional means and are not well understood because taxonomic work to describe the species found in marine sediments is difficult and rarely performed (Guardiola et al., 2015). eDNA metabarcoding of sediment has been used in attempts to describe marine eukaryotic communities, though it has suffered from a lack of taxonomic knowledge. Pawlowski et al. (2011) used metabarcoding on marine sediments to examine deep-sea eukaryotic richness and detected 942–1756 taxa per sample, dominated by dinoflagellates, cercozoans, ciliates, and euglenozoans; even photosynthetic taxa were present. Between 31 and 71 metazoan taxa were identified per sample despite small sample volumes, indicating free, extracellular, remnant, or resting DNA/organisms in the sample (Pawlowski et al., 2011). They determined the deep-sea floor to be a global DNA repository, describing organisms living both in the sediment as well as in the water column, making it useful for monitoring environmental changes. Interpretation of DNA data from the deep-sea floor was difficult due to lack of direct observational data, but still offers important insights into living marine organisms and the potential to study ancient marine life by analysis of deep soil cores (Pawlowski et al., 2011). A similar study by Guardiola et al. (2015) using metabarcoding on deep sea sediments identified over 1600 taxa dominated by Metazoa, Alveolata, Stramenopiles, and Rhizaria, with noticeable differences on a small scale, such as within the same core sample, between core samples, within localities, and between geographic zones (Guardiola et al., 2015). While composition was found to change between sites, overall biodiversity was similar throughout the dataset, with some differentiation due to differences in metazoan lifestyles (Guardiola et al., 2015). Guardiola et al. (2016) also published a similar study the following year, finding over 5500 taxa dominated by Metazoa, Alveolata, Rhizaria, Nemata, Arthropoda, and Annelida and discovering significant differences between layers of sediment, zone (slope or canyon),

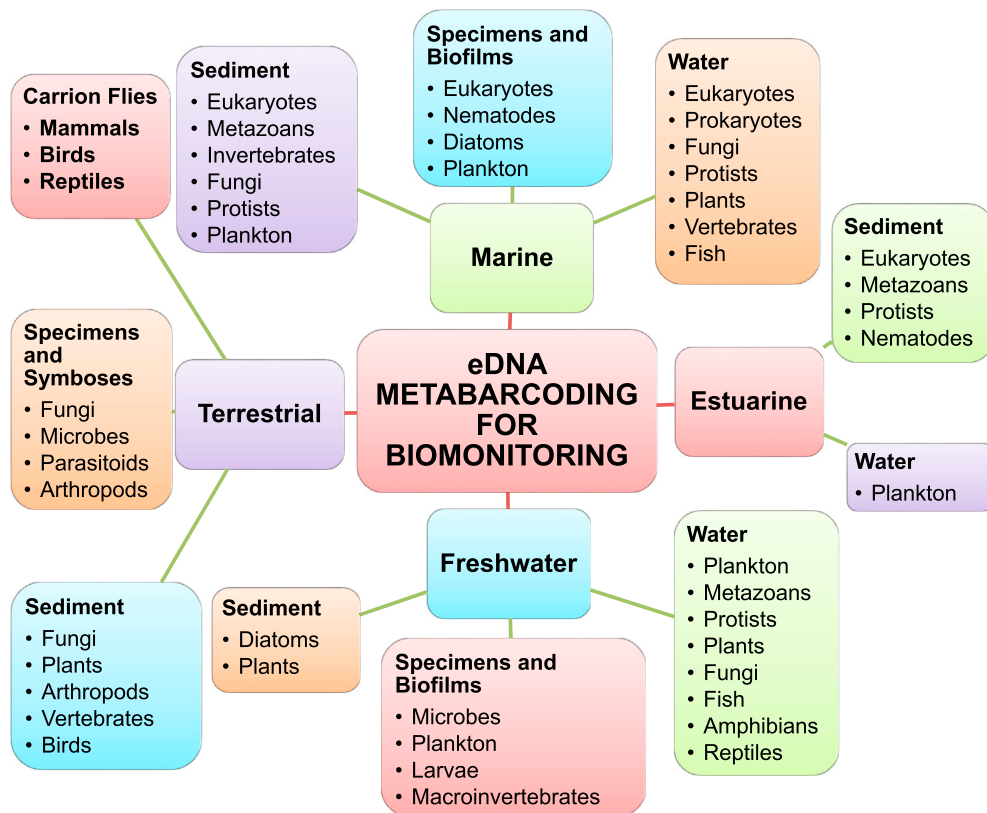


Fig. 1. Schematic diagram of global ecosystem and biodiversity monitoring with environmental DNA metabarcoding.

depth, and season. They determined that despite the use of a hypervariable region, diversity estimates were likely conservative due to the huge diversity of eukaryotes and that a lack of extensive databases limits the completeness of the study; as such, differences in taxonomic units may only be variations within taxa rather than different taxa entirely. In the same year, Sinniger et al. (2016) published a study in which they used eDNA metabarcoding of sediments to assess deep-sea benthic metazoan diversity at varying depths and locations worldwide. They found their data to be dominated by meiobenthic taxa, and while all phyla were identified, diversity within phyla remains largely unknown due to lack of studies exploring benthic metazoans and therefore a reference database that lacked specificity. Finally, Lanzén et al. (2017) tested the use of metabarcoding genomic DNA obtained from soil samples for determining diversity and community structure of meio- and microbenthic eukaryote communities, finding that sequencing depth and extraction replicates are important for optimization of diversity estimates. However, the precise number of replicates or sequencing depth will vary based on the aim of the study.

Looking at marine metazoan biodiversity, Fonseca et al. (2010) used metabarcoding to analyze eDNA from sediment in Scotland. From this study, they aimed to determine taxonomic richness based on identifiable taxa, to describe the breadth of unidentified microbial metazoans, and to assess the ecological importance of various groups. They argued that biodiversity assessments are biased towards large organisms, thus leading to the lack of descriptions for millions of microscopic metazoans, a group estimated to comprise up to 60% of the animal phyla that perform important tasks in the marine ecosystem. Their study confirmed this immense lack of knowledge, for which they suggest further exploration of microbial metazoan phylogeny by use of metabarcoding in order to better understand this important and highly diverse group (Fonseca et al., 2010).

eDNA metabarcoding of marine sediment has also been used to identify invertebrate communities. Cowart et al. (2015) tested the reliability of metabarcoding by comparing it to morphological methods of identifying invertebrates associated with seagrass meadows. They determined that metabarcoding enabled them to identify greater diversity with similar spatial differences when compared to morphological surveys; however, 36% of the species identified with metabarcoding were not present in the database, indicating the need for optimization of the library and procedures before future studies are employed (Cowart et al., 2015). Aylagas et al. (2016) compared taxonomic inferences of benthic macroinvertebrates of known taxonomy between different DNA sources, different COI barcodes, and different amplification conditions. They determined that metabarcoding was comparable to morphologic surveys but was able to consistently identify more benthic macroinvertebrate taxa. eDNA metabarcoding of marine sediments has also been used to investigate the distribution of fungi diversity and potential factors influencing said distribution (Li et al., 2016). This study identified 816 taxa, 130 known genera, 36 orders, 14 classes, and 5 phyla, with Ascomycota making up 72–79% of all units, and found that species compositions varied across

regions, with temperature determined to be the most important factor in fungal community distribution (Li et al., 2016). eDNA metabarcoding of sediments has the potential to quickly and accurately assess community structure, benthic health, and response to changes in environment, making it a promising methodology for biodiversity monitoring. However, it currently continues to require integration or comparison with traditional methods until it is fully optimized (Coward et al., 2015; Aylagas et al., 2016; Sinniger et al., 2016).

3.2.2. Bulk specimens and biofilms

In addition to sediment, marine eDNA metabarcoding can be applied directly to collected specimens, particularly of microscopic organisms that would otherwise require high levels of taxonomic expertise to identify. This technique has been applied to plankton biodiversity studies to determine the applicability of metabarcoding for evaluation of large scale trends (Chain et al., 2016). From bulk samples of zooplankton, Chain et al. (2016) discovered approximately 400 metazoan families in ports, including over 30 orders not previously reported. Zooplankton communities were also found to vary temporally and to be distinct throughout habitats (Chain et al., 2016). More recently, Djurhuus et al. (2018) compared metabarcoding of plankton samples against morphological identification, as well as the potential differences between direct seawater filtering, prefiltering, and plankton tissue metabarcoding. They found that different metabarcoding techniques revealed significant differences in taxonomic composition, with prefiltering detecting less than other methods. Deagle et al. (2018) also published a study comparing plankton metabarcoding to morphological identification; however, their study relied on the use of continuous plankton recorders which have been used to characterize plankton diversity for decades. They found that metabarcoding increased the number of species identified, increased the taxonomic resolution, and produced more detections overall compared to microscopy. Collectively, these results indicate that the potential use of metabarcoding in addition to continuous plankton recorders already in place has great potential, though it would require a shift in perspective regarding biodiversity surveys to integrate molecular methods in what is traditionally a morphological field.

Metabarcoding of specimens has also been applied to eukaryotes as a general group. Leray and Knowlton (2015) used eDNA metabarcoding of biofilms to describe the diversity and structuring of animals living on oyster reefs. They identified over 2000 taxonomic units, but only 11% could be matched to reference databases and only 8% to the species level in part due to broad but nonspecific taxonomic coverage. Despite issues in coverage, they were able to determine that smaller organisms were more diverse and that spatial overlap between sites was minimal, illustrating the usefulness of the method for presence/absence surveys as well as relative abundances and comprehensive community-level diversity analyses (Leray and Knowlton, 2015). Wangensteen et al. (2018a,b) developed a DNA metabarcoding protocol for biodiversity characterization of marine hard-bottom communities using two molecular markers. They determined that the analyzed communities contained a large amount of undescribed eukaryotic biodiversity, and that while gaps in reference databases were common, they typically applied to “rare” taxa present on a low number of sequencing reads (Wangensteen et al., 2018a,b). Specimen metabarcoding has also been applied to focal groups of nematodes and diatoms. Dell’Anno et al. (2015) compared eDNA metabarcoding to morphological data of nematodes and found that the analyses only matched at the family level due to poor resolution and incomplete reference databases for the metabarcoding methodology. As such, they called for further research on the subject before widespread application, but acknowledged its potential for studying the hyper-diverse nematodes in the deep-sea benthos (Dell’Anno et al., 2015). For diatoms, Rivera et al. (2018a,b) explored the use of metabarcoding biofilms from the shells of sea turtles to infer turtle behavior. They compared metabarcoding and morphological approaches and determined that each approach resulted in different diatom assemblages. Metabarcoding was able to distinguish between individual sea turtles based on diatom communities, but was limited by reference databases, while microscopy provided more ecological information on sea turtles, making a combination of both methods ideal for using biofilms to study sea turtle behavior (Rivera et al., 2018a). This study illustrates the widespread potential of eDNA metabarcoding of ecosystems, as it can be applied not only to ecosystem monitoring but also to movements of larger vertebrates. Altogether, metabarcoding is powerful and sensitive for large scale biodiversity studies, and as reference databases are expanded, the method will be further improved. It is a robust, fast, objective, and affordable method for characterizing complex communities of any size of organism with the potential to be further expanded with the use of multiple primer sets for optimal coverage (Chain et al., 2016; Pawlowski et al., 2018; Wangensteen et al., 2018a,b).

3.2.3. Water

Metabarcoding of water samples has been integral to the development of biomonitoring methodologies. Marine micro-organismal diversity was assessed by Brannock et al. (2016) in which metabarcoding was used to assess micro-eukaryotic diversity over time at various locations throughout the Alabama continental shelf. They found that community composition correlated with salinity, temperature, and dissolved silicate, and that it also differed seasonally. This information on diversity and community variance is important to the study of marine ecosystems, but without the use of advanced molecular techniques, it would be nearly impossible to gain a comprehensive understanding (Brannock et al., 2016). Bridging the gap between micro- and macro- organismal metabarcoding studies, Stat et al. (2017) compared shotgun sequencing to tree-of-life metabarcoding or “the use of multiple metabarcoding assays to survey a wide array of biotic diversity,” in determining biodiversity in a tropical marine environment. They found shotgun sequencing to be very inefficient due to the high number of reads required to obtain a clear picture of biodiversity, but the tree-of-life methodology was able to detect 287 taxa across the major eukaryotic divisions (animals, fungi, protozoa, plantae, chromista, bacteria, archaea) (Stat et al., 2017). However, multiple sets of “universal” primers were needed to obtain the full scope of biodiversity (Stat et al., 2017). In addition, they

were able to isolate haplotypes from a specific fish genus from the same water samples, further expanding the biodiversity applications of eDNA metabarcoding (Stat et al., 2017).

Metabarcoding studies focusing solely on the identification of vertebrates from marine water samples are numerous. A study by Thomsen et al. (2012) investigated the potential of metabarcoding eDNA from seawater to determine marine fish biodiversity and compared the data against traditional fish surveys. They successfully obtained DNA from 15 species of fish, including species important for human use and species rarely recorded traditionally, as well as 4 bird species, determining that eDNA metabarcoding identified the same or more species than traditional sampling. Additionally, they found that DNA fragments degraded within days in seawater, indicating that positive eDNA signals indicate recent species presence. Kelly et al. (2014) tested the ability of eDNA metabarcoding to detect species of bony fish in a large mesocosm aquarium, discovering a correlation between the rank abundance of eDNA and species biomass. They were also able to detect non-native species used as food in the aquarium. Miya et al. (2015) focused on the development of a truly universal primer set for fish identification to the species level via metabarcoding, testing their primers against over 800 different species and calling their creation “MiFish”. These primers were tested at an aquarium with known species, detecting 93.3% of all species present (123 genera and 59 families), and around natural coral reefs detecting 232 species (70 families and 152 genera), supporting their effectiveness and universal application. These primers have since been used in numerous fish eDNA metabarcoding studies (Miya et al., 2015). One such study was published in 2017 by Yamamoto et al. (2017) in which the ability of eDNA metabarcoding to reveal fish community structure in coastal waters at 47 spatially distinct stations was evaluated and compared to 14 years of data from visual surveys. The metabarcoding study detected 128 species, 62.5% of which were seen by visual surveys, and 20 species that were expected to occur but had never before been documented, indicating that the method has potential to enhance marine ecosystem analyses and fish community surveys, particularly as it was determined to have a higher detection rate than other methods (Yamamoto et al., 2017).

eDNA metabarcoding of water samples has also been used to describe entire vertebrate communities. Port et al. (2016) tested eDNA metabarcoding for marine vertebrates along a transect gradient of habitats and compared the results to simultaneous visual surveys by divers, finding spatial correspondence between metabarcoding and visual surveys, and significant differences between habitats indicating local community compositions. Additionally, metabarcoding was found to be able to reliably detect cryptic species missed by visual surveys, though not all species were consistently detected with both methods, suggesting that the current applications of eDNA metabarcoding are best suited to augment traditional data for the monitoring of community dynamics over space and time (Port et al., 2016). Andruszkiewicz et al. (2017) published a study in which they used eDNA metabarcoding to characterize marine vertebrate communities at 10 locations by collecting water samples in replicates at multiple depths per station. They were able to identify 7 families, 3 subfamilies, 10 genera, and 72 species from 33 unique marine vertebrate taxa with all but one family already known to inhabit the area. They also found that communities differed based on sampling depth and between sampling locations, indicating that eDNA metabarcoding is able to provide an accurate census of vertebrate communities and thus has enormous potential for biomonitoring at fine scale over space and time (Andruszkiewicz et al., 2017).

The effectiveness of marine eDNA metabarcoding has also been compared between sample types. Massana et al. (2015) analyzed the taxonomic diversity of both planktonic and benthic protist communities over space and time. They found different species compositions between water and sediment samples and that community composition varied between sites. They also determined that seasonality was important to consider, and that many samples are needed per site to adequately estimate diversity and community composition (Massana et al., 2015). Altogether, the use of sediment and water each yielded different results in community composition and spatiotemporal differences, further indicating the need to ensure that studies are optimized for the desired species groups. Overall, marine eDNA metabarcoding is non-invasive, efficient, cost-effective, and sensitive, with potential to complement other methods of biodiversity monitoring for ecological study and management on large spatial and temporal scales. Although it requires optimization in replicates, marker selection, avoidance of false negatives/positives, and reference database development, it has enormous potential across sample types and target taxa (Miya et al., 2015; Port et al., 2016; Pawlowski et al., 2016; Andruszkiewicz et al., 2017).

3.3. Estuarine monitoring

3.3.1. Sediment

Estuarine biodiversity monitoring is often included with marine monitoring due to their similarities; however, estuaries are a unique ecosystem overlapping with both marine and freshwater ecosystems and as such some eDNA metabarcoding studies have focused specifically on estuarine biodiversity. The majority of these studies have been concerned with the identification of benthic communities from sediment samples which often contain taxa such as nematodes that are commonly considered to be indicators of environmental health (Lallias et al., 2015; Avó et al., 2017). Morphology-based profiling of benthic communities has so far been low-throughput and ambiguous, and metabarcoding has been poorly applied to estuarine macrobenthos communities. Additionally, models to predict estuarine ecosystem diversity are limited for eukaryotes, which are a huge resource for assessing ecosystem health (Lallias et al., 2015; Lobo et al., 2017). As such, future studies are needed to explore estuarine sedimentary diversity via metabarcoding; however, some exploratory studies have been published. Chariton et al. (2010) applied metabarcoding to estuarine sediments to identify eukaryotes in human-

impacted ecosystems, successfully obtaining sequencing data for thousands of eukaryotes from 54 phyla ranging from microscopic non-metazoans to large metazoans, with a large proportion of richness stemming from micro- and meiofauna that are often overlooked in traditional surveys. Chariton et al. (2015) also published a study in which metabarcoding was used to assess benthic eukaryotic composition of five estuaries of varying conditions. They found that eukaryotic community composition reflected environmental condition and was strongly correlated with nutrients, turbidity, and pH, with the highest biotic richness occurring in the most disturbed environment. In the same year, Lallias et al. (2015) published a study in which metabarcoding was used to assess environmental impacts on microbial metazoan and protist diversity in two distinct estuaries with different histories of use by using longer primers focused for live individuals, they aimed to offer an immediate picture of the microbial community. They found that taxa were responding to hydrodynamics, salinity, and granulometry based on their life history, and that they exhibited estuary-specific similarity within salinity ranges. In development and assessment of metabarcoding methodology, Lobo et al. (2017) created several test communities to assess the viability of metabarcoding, and then compared metabarcoding to morphological identification in field tests at four distinct estuaries. They found that field morphological methods underestimated species richness, identifying only 23 species as compared to the 61 species identified using metabarcoding. They concluded that metabarcoding was able to provide many high-quality species identifications, and sensitive enough to determine differences in species compositions between communities. Avó et al. (2017) also tested the ability of metabarcoding to accurately describe communities by comparing metabarcoding of sediments from an estuary that had been previously characterized using traditional methodology. Metabarcoding generated 25–26 of the traditionally identified genera and identified 34–43 species in total, including 7–16 new genera for the databases, while morphological analysis matched approximately 90% of the metabarcoding genera (Avó et al., 2017). Between the two methods, they found strong temporal differences, but not spatial differences in species composition, with metabarcoding appearing to be a promising alternative for the labor-intensive and time-consuming traditional monitoring methods (Avó et al., 2017).

3.3.2. Bulk specimens

eDNA metabarcoding has also been applied to studies of the plankton community. Abad et al. (2016) evaluated the capacity of metabarcoding for estuarine plankton monitoring by comparing it with traditional microscopic taxonomic analysis. They found similar spatial and temporal trends of taxonomic diversity for zooplankton but not phytoplankton, possibly due to a lack of an extensive phytoplankton database. Additionally, there was high correspondence between abundances in the two methodologies, suggesting that there is potential for using metabarcoding to estimate species abundances as well as invasive species detection, as two non-indigenous species were discerned (Abad et al., 2016).

Limitations of eDNA metabarcoding in estuaries are similar to those found in other ecosystems and applications, though the lack of studies assessing estuaries using metabarcoding results in a greater need for exploratory studies. Overall, eDNA metabarcoding of estuaries is fast, scalable, objective comprehensive, extremely sensitive, has potential for high taxonomic resolution, is able to identify species of any size, and is cost- and time-effective with the ability to enhance traditional monitoring techniques and inform environmental policies (Lallias et al., 2015; Chariton et al., 2015; Abad et al., 2016).

3.4. Freshwater monitoring

Biodiversity monitoring studies in freshwater using eDNA metabarcoding are quite similar to those in marine and estuarine environments, with the main areas of focus being in sediment, biofilms, bulk specimens, and water samples. Freshwater fauna are sensitive to environmental change, making them excellent indicators of ecosystem health, but traditional surveys are labor intensive, require physical capture which may injure individuals, and often result in low detection. eDNA metabarcoding has potential to improve aquatic biodiversity surveys; however further research of species characteristics and life history traits that may affect detection and refinement of quantification is needed (Evans et al., 2016b).

3.4.1. Sediments

Metabarcoding has been used on freshwater sediments to analyze plant communities. Alsos et al. (2018) analyzed the composition of lake vegetation and vascular plant eDNA at 11 lakes by using both metabarcoding and traditional vegetation sampling to determine the degree to which modern taxa are found in sediment eDNA samples. They recorded 489 taxa by traditional surveys, of which 17–49% were detected using eDNA, while 47 plant taxa were identified using eDNA, of which 73% matched those recorded in vegetation surveys. They concluded that the taxa which were present in eDNA but not surveys were likely missed during the surveys or grew outside the survey area, and those missed in eDNA may have been found with increased sampling effort. Additionally, they determined that detection rates varied among plant families, and terrestrial plant DNA was dependent on sampling distance (Alsos et al., 2018). For aquatic vegetation, eDNA may be superior to traditional surveys; however, further optimization is needed before the procedure is ready for widespread application. Altogether, metabarcoding of sediments appears to be a promising method for biodiversity monitoring, though it requires standardization and expansion of databases before it is able to reach its full potential as a staple in biomonitoring methodology (Alsos et al., 2018).

3.4.2. Bulk specimens and biofilms

Similar to marine environments, specimen collection for metabarcoding is a popular choice for describing freshwater biodiversity, with many of these studies focusing on plankton. Liu et al. (2017) assessed the effect of prefiltering on detection

of microeukaryotic plankton communities and found that approximately 25% of reads were for metazoans in both groups. Additionally, they found no significant differences in richness, diversity, and spatiotemporal patterns, suggesting that pre-filtering does nothing to differentiate size classes of eukaryotic plankton in freshwater environments. In the same year, Wurzbacher et al. (2017) published a study in which they tested eDNA metabarcoding for total plankton community dynamics over time, including eukaryotes and prokaryotes in their analysis. They discovered changes in community composition over time, following the same patterns as detected by morphological methods, but morphological methods missed the bloom of a rare dominant bacterium that was impacting the planktonic community. These results highlight the usefulness of microbial and macrobial community metabarcoding studies for understanding whole ecosystems and species interactions and community dynamics. Banerji et al. (2018) published a similar study in which they ran metabarcoding analysis on samples collected from multiple sites in a reservoir over 4 months, from which they detected over 1300 taxonomic units, including metazoans, protists, chlorophytes and fungi. They found that diversity estimates differed between sites and between taxonomic groups, suggesting that different groups of plankton respond to different environmental factors. Previous to these studies, Sohlberg et al. (2015) explored planktonic fungal communities in groundwater associated with deep bedrock fractures, finding fungi at all depths sampled at higher diversity than expected, dominated by Ascomycota. Fungi were found to be active in the fracture zones, indicating an ability to maintain cellular activity potentially by adaptation to the environment. Some of the fungal sequences isolated may represent novel species that may play an important role in subsurface nutrient cycling (Sohlberg et al., 2015). Yang et al. (2017) used metabarcoding to characterize zooplankton communities by comparing results to traditional morphological methods and found agreement between the two methods regarding species composition, spatial distribution, differences between water body type, and abundance based on biomass, indicating that metabarcoding is valid for use on a large scale for biomonitoring of ecosystems to influence conservation decisions. Additionally, they found metabarcoding to identify more species than morphology-based identification, though it was limited by biases commonly associated with metabarcoding studies such as technical biases, bioinformatics differences, lack of comprehensive databases, and an inability to detect life stage and health of individuals (Yang et al., 2017). Another study applying metabarcoding to freshwater specimens was published the same year, in which a new method for analysis of larval fish communities using metabarcoding of bulk ichthyoplankton samples was developed and compared to individual larval sequencing (Maggia et al., 2017). They found that both methods were comparable in results, indicating the potential of metabarcoding to inform understanding of fish recruitment and river preservation, particularly in tropical regions where fish larval ecology is poorly studied due to difficulty in identification (Maggia et al., 2017). Metabarcoding of plankton communities has the potential to provide better taxonomic coverage and resolution than morphological methods, and comparison to conventional data revealed similar abundance and diversity data. However, each method included groups not seen in the other, suggesting the use of the two methods simultaneously is the best way to achieve optimum coverage and the most accurate estimates of diversity (Banerji et al., 2018).

In addition to plankton, metabarcoding of collected freshwater specimens has been applied to bulk macroinvertebrate samples to describe biodiversity. Carew et al. (2013) tested whether metabarcoding could be used to identify species of Chironomidae from field specimens, an important indicator group of aquatic ecosystem health. They found that with one gene region (COI) metabarcoding could identify 96% of species, but when using two (COI and CytB), 99% of species could be identified. They also found a strong relationship between number of sequences and number of individuals, indicating the possibility of estimating abundance using metabarcoding. Metabarcoding has been compared to morphological macroinvertebrate metrics across a gradient linked to watershed features where the two study types were found to consistently indicate the same differences within the stream gradient, again suggesting the usefulness of macroinvertebrate DNA metabarcoding for large-scale biomonitoring (Emilsson et al., 2017).

A unique study published by Hajibabaei et al., in 2012 investigated the use of preservative ethanol as a nondestructive source for DNA from macroinvertebrates to determine biodiversity and compared this data to both the morphological identification of species in the ethanol as well as metabarcoding of all the specimens homogenized together, as is typically done with specimen metabarcoding studies. They found that ethanol-based metabarcoding identified 87% of taxa while tissue metabarcoding identified 89%, with the missing taxa for both being those of lowest abundances in the mixture. This methodology presents a solution to the need for both sample and voucher specimens by not destroying samples in order to obtain data, thus taking advantage of existing preservation standards to run molecular biodiversity surveys. It has the potential to be applied to target pests, pathogens, vectors, or other organisms associated with those that have been preserved in ethanol in addition to the larger macroinvertebrates (Hajibabaei et al., 2012). Altogether, metabarcoding of specimens has great potential in the realm of freshwater biodiversity monitoring as it has been found to reduce the cost and time of biomonitoring, though as with all metabarcoding studies thus far, it requires refinement and further development of reference databases (Maggia et al., 2017; Yang et al., 2017; Emilsson et al., 2017).

The testing of biofilms for diatom diversity has been applied several times in freshwater diversity; diatoms are often a focus of freshwater monitoring due to their status as bioindicators (Kermarrec et al., 2014; Rivera et al., 2018a,b). Metabarcoding of biofilm diatoms has been compared to morphological surveys several times with similar results throughout. In a mock community, Kermarrec et al. (2014) found that species richness was always higher with microscopy but noted that some species detections from microscopy may have been from dead cells, which cannot always be differentiated from living organisms morphologically. Several studies have used natural freshwater communities that often function as indicators of water quality to assess the effectiveness of metabarcoding. Zimmermann et al. (2015) found that metabarcoding consistently identified more taxa and more knowledge on taxonomic diversity than morphological methods, while Vasselon et al. (2017)

determined that while the community structures of the two methods were highly correlated and the ecological quality estimates were similar, only 13% of species were shared by both approaches, likely due to incompleteness of databases, limits in taxonomic knowledge, and biased abundance estimates. Similarly, [Rivera et al. \(2018a,b\)](#) found similar assemblages with both approaches structured by the same environmental factors; but the ecological assessment for each approach differed, perhaps due to the lack of a comprehensive DNA reference library, an issue that will likely resolve itself over time. Despite this limitation, the authors suggest an integrated approach of taxonomy, autecology, life forms, cell sizes, and ecological guilds from morphology, molecular data, and ecological aspects for determination of freshwater diatom biodiversity; metabarcoding offers the ability to obtain diversity data that can then be applied on different spatial, temporal, and taxonomic levels for large scale biomonitoring and management, including functional and ecological community relations and water quality assessment ([Kerमारrec et al., 2014](#), [Zimmermann et al., 2015](#), [Rivera et al., 2018a,b](#)). In short, metabarcoding has great potential for the augmentation of traditional diatom surveys rather than the complete replacement.

Freshwater biofilms have also been used for the metabarcoding of microbes in general. A study by [Besemer et al. \(2013\)](#) used eDNA metabarcoding to test fluvial biofilms to determine the distribution of diversity of ecologically critical benthic microbes. They were able to determine from metabarcoding data alone that microbial diversity decreased downstream, particularly at confluences, suggesting that environmental and biotic factors may influence biofilm connectivity and diversity. Additionally, they found a high degree of variability in species composition between streams not explained by geographic distance, suggesting that fluvial networks constrain microbial distribution ([Besemer et al., 2013](#)). Patterns of diversity discovered in this metabarcoding study indicate the importance of headwater communities for regional diversity and conservation as well as management of ecosystem function ([Besemer et al., 2013](#)).

3.4.3. Water

Water sampling is perhaps the most common method of collecting eDNA for metabarcoding in freshwater environments, largely dominated by studies focusing on fish and other aquatic vertebrates. One of the earliest studies in freshwater vertebrate metabarcoding compared eDNA metabarcoding to electrofishing to characterize stream fish communities ([Olds et al., 2016](#)). They found electrofishing to yield 12 species and an estimated richness of 16 species, while eDNA sampling detected 16 species, including 3 species that had never been detected in the water body using electrofishing ([Olds et al., 2016](#)). They determined that eDNA metabarcoding has the potential to enhance fish surveys without disturbing fish or habitat in order to better inform management decisions despite being unable to provide specific data on population quality ([Olds et al., 2016](#)). The same year, [Civade et al. \(2016\)](#) used eDNA metabarcoding to assess fish biodiversity in a freshwater ecosystem and investigated signal variability by testing organization of fish communities along a longitudinal gradient. They found eDNA metabarcoding to be more efficient and more likely to detect species presence than traditional sampling, and that fish species identified with metabarcoding were comparable to those identified by traditional sampling, thus giving an in-depth description of local fish biodiversity along a longitudinal system ([Civade et al., 2016](#)). [Hanfling et al. \(2016\)](#) compared eDNA metabarcoding of a lake fish community to long-term traditional data in an attempt to define the qualitative and quantitative accuracy of eDNA metabarcoding. They found that eDNA detected 14 of the 16 known species in the lake from one sampling session, while the most recent traditional sampling effort had detected only 4, making eDNA much more sensitive; the two species missed by eDNA were unlikely to be in the lake at the time of sampling. They also found that the number of sequence reads corresponded with both site occupancy and abundance estimated from previous surveys, indicating that eDNA metabarcoding could potentially be used quantitatively for describing fish communities ([Hanfling et al., 2016](#)). [Shaw et al. \(2016\)](#) also compared eDNA metabarcoding to traditional methods, focusing on the comparison with netting surveys. They found that water samples detected 100% of netted species (including species of conservation concern, rare, and invasive species), and that five 1 L water samples per site were more effective for characterizing fish communities than smaller water samples or sediment samples. The following year, [Evans et al. \(2017\)](#) tested eDNA metabarcoding estimates of species richness by comparing them to traditional methods to define how spatial distribution and bioinformatics affect efficiency. They discovered that eDNA metabarcoding could detect all species found with traditional sampling plus 11 more when considering all markers separately. Despite this success with a variety of markers, they determined that when species needed to be detected by all markers, only 7 known, traditionally captured species were found. Additionally, they determined that eDNA was relatively evenly distributed throughout the reservoir but predicted that this would not be the case with moving water bodies such as streams and rivers. [Nakagawa et al. \(2018\)](#) assessed the performance of eDNA metabarcoding in a moving water body, focusing on stream fish species and comparing metabarcoding data against existing observational data; additionally, they tested for detection bias between upstream and downstream sampling locations. They found upstream eDNA data to be most similar to observational data, detecting 86.4% of reported species as well as two additional species, and that eDNA results were consistent with turnover and assemblage patterns of some species ([Nakagawa et al., 2018](#)). [Sato et al. \(2017\)](#) explored the differences in freshwater fish biodiversity assessments resulting from pooling water for eDNA metabarcoding. They found more species in the individual samples, including moderate spatial correlation, indicating that the pooling strategy is not suitable for assessing richness, but may be useful for comparing fish communities within sites. Additionally, they found that increased replicates lead to increased sensitivity with water volumes and adequate distance between sampling points being necessary, and thus suggested small volumes from many locations for maximum detectability ([Sato et al., 2017](#)).

Expanding the scope from fish alone, [Evans et al. \(2017\)](#) created aquatic mesocosms to test the ability of eDNA metabarcoding to detect both fish and amphibian species in different densities and relative abundances. They found that

metabarcoding was able to detect all species in all mesocosms, and were able to identify a small positive relationship between species abundance and read abundance, illustrating the potential of eDNA metabarcoding use as an index of species abundance and quantification of diversity. [Valentini et al. \(2016\)](#) took this a step further by designing fish and amphibian primer sets and testing them in both lab and field conditions for applicability in ecological studies and aquatic biodiversity monitoring. They tested their primer designs against 53 amphibian species, 64 freshwater fish, and 22 marine fish, initially tested with individual tissue to construct a reference database, next with pooled tissue, then with spiked water, and finally with field sampling in areas with known species compositions. Following these tests, occupancy data was compared to historical and traditional survey data, finding that the use of eDNA metabarcoding with these primers could be an applicable methodology for freshwater vertebrate biodiversity studies ([Valentini et al., 2016](#)). [Lim et al. \(2016\)](#) also assessed species diversity across taxa, using universal metazoan primers on water samples from two reservoirs with known species diversity. They found signatures for over 500 species, of which approximately 100 could be identified to genus or species level, including an invasive snail and a rare fish.

Studies focusing only on amphibians have also been published. A study by [Lopes et al. \(2017\)](#) tested eDNA metabarcoding for anurans in tropical streams by comparing metabarcoding data to that from traditional visual and audio surveys. They successfully detected all species known to inhabit the streams with one eDNA visit, with a greater number of reads coming from more common species that are more closely tied to the streams. They also found that detection was maximized by filtering large quantities of water and sampling multiple sites along streams, but quantification was not found to be possible. eDNA detection was higher per sampling event than traditional field surveys, was able to detect various life stages that may be missed by traditional surveys, could be applied outside of the species reproductive peak, is noninvasive, and could be used on a wide taxonomic scale and large spatial scale in relatively short time periods, making it useful for anuran surveys in tropical environments ([Lopes et al., 2017](#)). [Lacoursière-Roussel et al. \(2016\)](#) also focused on herpetofauna and found metabarcoding to be effective in assessing species distributions. Most recently, [Harper et al. \(2018\)](#) compared the sensitivity of eDNA metabarcoding to eDNA quantitative PCR (qPCR) detection for great crested newts. They found qPCR to provide greater detection for this species, but when thresholds were removed from metabarcoding, it performed as well as qPCR suggesting that a good potential methodology would be initial community surveys using metabarcoding followed by species-specific surveys using qPCR for species of concern as indicated by the metabarcoding studies ([Harper et al., 2018](#)). eDNA metabarcoding appears to be an extremely promising avenue in freshwater biodiversity studies with the capability to collect monumental amounts of data in short periods of time without disrupting ecosystems ([Shaw et al., 2016](#); [Nakagawa et al., 2018](#)). Although it has the expected downsides discussed in previous sections, as technology advances, metabarcoding can be expected to decrease in cost and increase in effectiveness. As in other ecosystems, it is probably best suited as a complementary method to traditional surveys ([Valentini et al., 2016](#); [Hänfling et al., 2016](#)).

3.5. Terrestrial monitoring

Terrestrial eDNA metabarcoding for biodiversity is not an intuitive application of the methodology, but there are several important sources of terrestrial eDNA that can be traced, such as soil, saltlicks, water sources, and bulk specimens. It seems improbable that soil samples alone would allow reliable detection of non-soil dwelling species. Indeed, the majority of terrestrial eDNA metabarcoding studies focus on soil samples for invertebrates in frequent, direct contact with the soil, but other associations have been detected and studied ([Calvignac-Spencer et al., 2013](#); [Schubert et al., 2015](#); [Lee et al., 2016](#)). These creative workarounds are unique and certainly seem impossible without the advent of metabarcoding. Rapid survey approaches for monitoring biodiversity could help to assess and understand threats to ecosystems worldwide, and metabarcoding provides new opportunities for large-scale biodiversity studies as well as being an efficient and economical approach to studying ecosystem structure ([Yoccoz et al., 2012](#)).

3.5.1. Soil

Terrestrial metabarcoding began with soil sampling, with a study by [Buée et al., in 2009](#) in which universal fungal primers were used on six distinct forest soil samples, discovering over 1000 separate taxonomic units, primarily of Ascomycota and Basidiomycota, with Agaricomycetes as the dominant taxa. From this data, diversity analyses suggested strong spatial heterogeneity and unexpectedly high richness. Additionally, they found that analyzed sequences corresponded 73% of the time with only 26 identified taxa, thus highlighting the urgent need for more extensive sequence databases. Soil fungi also play an important ecological role but little is known about the structure, richness, and distribution of different fungal communities and ecological groups, making metabarcoding an exciting prospect for soil fungal studies in the future ([Buée et al., 2009](#)).

Other studies have focused on determining plant diversity from soil sample metabarcoding. Plant community composition can help to define the ecological quality of sites, but aboveground visual surveys are not all-encompassing; eDNA has the potential to identify active and dormant seeds, pollen, and detritus of species simultaneous with metabarcoding, thus providing a more comprehensive view of plant diversity ([Fahner et al., 2016](#)). [Yoccoz et al. \(2012\)](#) used metabarcoding of a short DNA fragment using plant-specific primers to estimate taxonomic diversity on 8 plant communities from two vegetation types and assessed DNA persistence by metabarcoding formerly cultivated soils. They found that plant diversity from metabarcoding was consistent with estimates from conventional surveys but revealed taxa not detectable through traditional surveys. In addition, soil samples were spatially distinct in species composition and diversity, with crop DNA sequence detection from the formerly cultivated fields dependent on the amount of time since cultivation, dropping off almost

completely after approximately 50 years (Yoccoz et al., 2012). Fahner et al. (2016) also assessed soil metabarcoding for measuring plant diversity, evaluating sequence recovery, annotation, and resolution for 4 plant DNA markers first with database sequences, then with soil samples from a remote wetland. They determined that internal transcribed spacer (ITS)-2 and ribulose biphosphate carboxylase (rbc)-L were the most effective regions for metabarcoding vascular plants for generalized databases, and that full-length DNA barcode regions could outperform the shorter sequences in taxonomic resolution and detection. This finding is especially noteworthy as Yoccoz et al. (2012) noted the limitations of the short barcoding region in taxonomic resolution (Fahner et al., 2016).

Like in aquatic biodiversity studies, metabarcoding of soil and sediment has been applied to the characterization of eukaryotic communities. Drummond et al. (2015) compared a broad suite of markers against traditional biodiversity monitoring tools for terrestrial eukaryotes, examining the 16S, 18S, trnL, ITS, and COI regions to see if eDNA could be a proxy for traditional biodiversity sampling, which combinations of markers are best, and how sensitive the measures are to markers/parameters. They found that alpha and gamma diversities were sensitive to number of reads, while beta diversity was stable after a couple thousand reads. Additionally, they determined that the COI region had the strongest correlation with traditional sampling data, especially for vegetation and invertebrates, but that sampling efforts required at least two markers to cover all phyla (Drummond et al., 2015). Invertebrate-species metabarcoding has also been explored. Yang et al. (2014) compared the results of soil and leaf litter metabarcoding to traditional surveys for arthropods and found that the two methodologies yielded different species compositions. However, they described similar ecological information in regards to relative richness and diversity, with metabarcoding being perhaps more sensitive to habitat differences than traditional surveys (Yang et al., 2014). Soil metabarcoding can also be applied to the assessment of eukaryotic communities in response to environmental changes. Štursová et al. (2014) used metabarcoding to analyze the response of fungal soil and leaf litter communities during a bark beetle-induced tree dieback, finding that the biomass of the fungal communities decreased, and that community composition changed with a loss of root-associated fungi and an increase in saprotrophic species (Štursová et al., 2014).

Andersen et al. (2012) used metabarcoding to attempt to characterize vertebrate taxonomic richness and structural diversity by use of soil samples from areas with known species compositions such as zoological gardens, safari parks, and farms. They found that DNA from the soil reflects overall taxonomic richness and relative biomass but may not be sensitive to recent changes in communities. Moreover, they found no impact of soil pH on DNA amplification, but that high organic content did inhibit amplification, and that DNA was capable of traveling through strata depending on soil texture and structure (Andersen et al., 2012). DNA was not uniformly distributed throughout the area, with higher concentrations found around latrine or den areas, so they concluded that spatial replicates are necessary to optimize detection. Despite this, they determined metabarcoding to be a quick and reliable alternative to traditional surveys (Andersen et al., 2012). Perhaps the most comprehensive study on terrestrial biodiversity from soil samples was published by Epp et al. (2012) in which they developed metabarcodes for fungi, bryophytes, enchytraeids, beetles and birds, and evaluated *in silico* PCR and *in vitro* PCR of surface soil and frozen sediment. They found that *in silico* primer design was effective in creating working primers for the field and that the most promising universal metabarcoding regions were located on 12s and 16s which resulted in higher coverage than other regions. They also found that there was high variation between the modern and ancient species compositions (Epp et al., 2012).

3.5.2. Root associations

In a similar environment to soil samples, sampling of communities associated with roots has been used for metabarcoding biodiversity studies. Blaaid et al. (2012) investigated changes in fungal communities associated with the root systems of *Bistorta vivipara*, a perennial herbaceous flowering plant, along a primary succession gradient using metabarcoding to detect variation in fungal richness and community composition as the ecosystems change. They identified a total of 470 taxonomic units found to correspond with between 8 and 41 fungal sequences dominated by basidiomycetes. Additionally, total taxa increased as the ecosystem developed, but average per root system did not change, though there was a high degree of patchiness in fungal distribution across root systems (Blaaid et al., 2012). Martínez-García et al. (2015) also looked at root-associated fungal communities by using metabarcoding. They looked at a long-term soil sequence that included ecosystem progression and retrogression to determine the importance of host plant identity as a factor driving fungal community composition. They also combined metabarcoding methods (for determining community per site) with terminal fragment length polymorphism (T-RFLP) methods (for determining community within sites) to acquire a full view of community composition, diversity, and change over time. They found fungal communities to be highly structured by ecosystem age with strong niche differentiation. They also found that diversity decreased as the ecosystem aged, and 39% of fungal variance could be attributed to the host plant, suggesting that fungal communities are tightly linked to plant communities (Martínez-García et al., 2015).

3.5.3. Bulk specimens

Bulk specimen metabarcoding has also been explored in terrestrial environments. Conventional ecosystem assessments based on morphology or barcodes are costly and time-consuming and thus less than ideal for large numbers of samples, hence the future applications of metabarcoding (Gibson et al., 2014). Yu et al. (2012) presented protocols for determining taxonomy of bulk arthropod samples via metabarcoding by constructing seven communities of arthropods and comparing community composition and diversity between the two datasets. They found that it is possible to precisely estimate community differences and diversities by metabarcoding despite the loss of some taxonomic information, and that metabarcoding of bulk

samples is effective for measuring biodiversity. In the following year, [Ji et al. \(2013\)](#) validated metabarcoding of bulk arthropod samples by testing it against 3 high quality standard datasets from Malaysia, China and the United Kingdom. For all locations, metabarcoding returned correlated estimates of species richness, identified treatments and responses, and returned similar statistical models in the same taxon. [Gibson et al. \(2014\)](#) published a similar study in which they used multiple universal primers on bulk arthropod samples to test the ability of metabarcoding to describe diversity. They found that metabarcoding was able to identify 91% of the arthropods as well as detect microbes associated with the arthropods. Additionally, they determined that metabarcoding was more effective than other methods and greatly reduced the time and cost of biodiversity studies, making it ideal for a variety of further applications in ecology, including interactions between the macro- and micro-biome ([Gibson et al., 2014](#)).

Metabarcoding of bulk specimens may also have applications in the assessment of host-parasitoid and community interactions within biodiversity studies, an area of study essential to biodiversity monitoring, but difficult to discern using traditional methods. [Sigut et al. \(2017\)](#) used mock samples of insect larvae and parasitoids to test the potential of metabarcoding for identification and assessed the completeness of the barcode database by comparing it to a known host-parasitoid database for the study area. In this study, metabarcoding successfully identified 92.8% of all taxa present in mock samples and was able to identify taxa consistently across all taxonomic levels. Additionally, they found that 39.4% parasitoid and 90.7% host taxa were able to be identified based on the reference database, indicating a definite need for expansion of the parasitoid database. Metabarcoding data in this study suggests higher parasitoid diversity than traditional surveys, identifying the potential of metabarcoding to achieve more accurate identifications from reliable libraries and to better characterize diversity ([Sigut et al., 2017](#)). [Miller et al. \(2016\)](#) established a protocol for metabarcoding fungal communities directly from the DNA extracted from homogenized beetles and found that metabarcoding produced viable information on fungal species abundance associated with beetles particularly when large numbers of PCR replicates are used.

A unique application of metabarcoding bulk arthropod samples lies in the use of carrion fly samples to estimate vertebrate diversity. Carrion flies feed ubiquitously on dead and decaying animals and have not been found to have any distinct host preferences. This lack of preference makes them an excellent potential model for determining vertebrate diversity, assuming death rates between species are relatively constant ([Calvignac-Spencer et al., 2013](#)). Applications of this method may even be able to be applied to monitoring efforts of mortality dynamics or identification and monitoring of individual populations ([Calvignac-Spencer et al., 2013](#)). One of the earliest studies employing this creative methodology for monitoring terrestrial biodiversity was by [Calvignac-Spencer et al. \(2013\)](#) in which they used metabarcoding on 201 pooled carrion flies collected in Cote d'Ivoire and Madagascar, retrieving DNA from 22 to 4 species respectively, from different forest strata and different size ranges, including very rare species. Though a full spectrum of mammalian diversity was not established in this study, they estimated further sampling effort could achieve a more complete estimation of biodiversity. In addition, amphibians and birds were detected, suggesting further applications in biodiversity monitoring ([Calvignac-Spencer et al., 2013](#)). [Schubert et al. \(2015\)](#) screened carrion flies with nonspecific and specific primers to determine the differences in detection rates between the two primers. They found that while specific primers increased the detection of target species, it was still possible to detect these species using general primers. They also determined that the DNA collected from carrion flies was enough to attempt genotyping individuals in addition to the intended use to estimate population abundance and assess overall mammalian diversity ([Schubert et al., 2015](#)). [Lee et al. \(2016\)](#) attempted to calibrate the use of metabarcoding of carrion flies by comparing the metabarcoding-derived biodiversity estimates against traditional methods including cage trapping, mist netting, hair trapping, and scat collection in two forests, one with known megafauna and one without. They found metabarcoding to consistently detect more species than other methods, and predicted that as study times progressed, it would continue to outperform traditional methods ([Lee et al., 2016](#)). Finally, [Rodgers et al. \(2017\)](#) conducted vertebrate-specific metabarcoding on carrion flies in an area with a well-known mammal community to test the efficiency of the methodology for vertebrate detection and monitoring by comparing metabarcoding results against diurnal transect counts and camera trapping. All surveys were conducted over the same time period (including fly collection), and metabarcoding was found to detect more species than other methods over the same time period, but less than those documented over the past 7 years of surveys ([Rodgers et al., 2017](#)). Their data suggests that metabarcoding is powerful and efficient for biodiversity studies, particularly when used in conjunction with current monitoring methods. However, more complete reference databases, PCR optimization, multiple markers, greater sampling effort, and greater sequencing depth are needed, sentiments shared by other studies ([Lee et al., 2016](#); [Rodgers et al., 2017](#)). This unique application of metabarcoding may be a rapid, unbiased, and cost- and time-efficient wildlife monitoring methodology with capabilities to detect species and individuals, and the potential to join the standard mammal monitoring field methods. Moreover, further investigation is also needed into carrion fly ecology before it can be fully integrated into terrestrial biodiversity monitoring efforts.

3.5.4. Water

Though they do not reside in the water, terrestrial animals still require water and other nutrient sources to survive. As such, eDNA can be collected from the sources and analyzed to determine species use. Assessing species presence using eDNA in forest habitats, which are often difficult to survey, particularly when containing elusive or rare species, could be a viable alternative or augmentation to traditional study methods. A study in 2017 used metabarcoding to characterize the mammalian community utilizing forest ponds in Japan, from which they were able to detect several common mammals from the area. Not all common mammals were detected, however, as pond use is dependent on individual need and likely varies greatly over space and time ([Ushio et al., 2017](#)). A similar study using universal bird primers has also been published, however

variation in species and a lack of reference sequences reduced detection (Ushio et al., 2018). Another terrestrial location that may be visited more frequently is saltlicks or other nutrient centers. Environmental DNA metabarcoding has been applied to saltlicks in Borneo for detection of mammals; in this study, several endangered species were detected, but many smaller species were missed due to lack of reference sequences (Ishige et al., 2017). Regardless, this is an area of research that shows great promise with the improvement of techniques and reference databases.

4. Further applications

While the prospects for ecosystem and biodiversity monitoring using eDNA metabarcoding are thrilling and in need of further research, it is not the only area in which the method can be applied. Metabarcoding can be used to reconstruct ancient ecosystems from DNA found throughout the world, to explore the interconnectness between plants and pollinators, to assess diet without need for feeding observation or stomach flushing, to detect invasive species before they could possibly be detected via traditional means, to determine community responses to pollution, and even to assess air quality and its implications for human health. Though these are the topics covered in this review paper, more applications of eDNA metabarcoding are surely yet to come (Fig. 2).

4.1. Paleology and ancient ecosystems

Environmental metabarcoding is not only applicable for monitoring the health and status of modern ecosystems but has also been used to reconstruct ancient ecosystems from carbon-dated deep sediment samples that have been carbon dated (Pedersen et al., 2013). As traditional methods of ancient ecosystem reconstruction are based on pollen and microfossil identification, they are often time-consuming and prone to human error; metabarcoding studies when used in conjunction with traditional data may be able to give a more complete view of ancient ecosystems (Sønstebo et al., 2010; Jørgensen et al., 2012). These reconstructed ecosystems can in turn be used for future conservation planning, including backwards testing of climate change models, invasive species emergence tracking, and assessment of anthropogenic influences to past biodiversity and landscape (Sønstebo et al., 2010; Jørgensen et al., 2012; Bellemain et al., 2013; Giguët-Covex et al., 2014).

One of the first studies using both metabarcoding and next-generation sequencing on ancient DNA was published in 2010, wherein Sønstebo et al. (2010) constructed a library of 842 widespread/important arctic flora species and identified chloroplast region P6 as a successful metabarcoding region for these plants. They found that using metabarcoding primers with NGS resulted in greater taxonomic resolution than traditional methods. However, using both methods together was the most successful in identifying the greatest number of taxa to the smallest resolution (Sønstebo et al., 2010). Also studying arctic plant composition, Jørgensen et al. (2012) compared sedimentary DNA retrieved from nunataks to data from detailed surveys of the past 130 years. Based on traditional surveys, biodiversity had drastically decreased since 1967 from 60 species to just

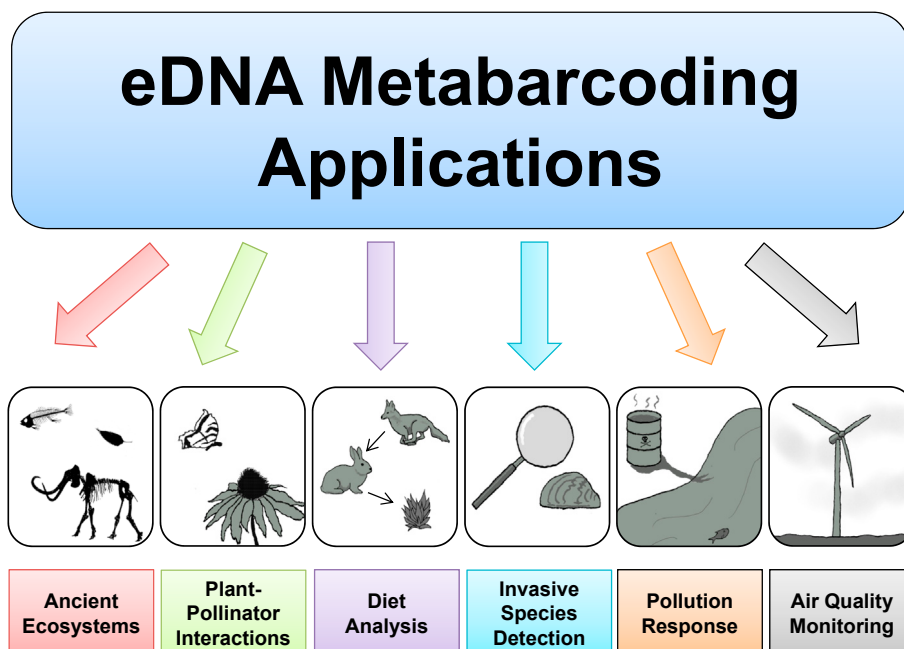


Fig. 2. Applications of environmental DNA metabarcoding in aquatic and terrestrial ecosystems.

eight in 2009. They determined that metabarcoding plant DNA was more successful than traditional surveys, as it was able to identify twice as many species as the 2009 survey, including 3 taxa that had never been documented on the nunatak (Jørgensen et al., 2012). As there is no animal life on these nunataks, changes in biodiversity can be assumed to directly correlate with climate, making data from this location especially valuable for climate change research (Jørgensen et al., 2012). Pedersen et al. (2013) compared metabarcoding of sediment from a lake in Greenland to a previously determined biodiversity record to identify ancient plant biodiversity. Sedimentary DNA was able to identify two previously undiscovered families at the study site; however, others were missed due to the incomplete nature of genetic libraries. As such, they determined that DNA metabarcoding has the potential to reconstruct local vegetation in greater detail than traditional means, but that further work is needed on resources such as GenBank to ensure that no species are missed during the process. Despite increased rates of degradation due to climatic differences, even tropical regions can yield useful metabarcoding data from sediment. Boessenkool et al. (2014) were able to identify species from tropical sedimentary DNA to a greater taxonomic resolution than was found with pollen sampling, but more species were identified with pollen. This discrepancy could be due to DNA degradation, or it could be a product of the transient nature of pollen; where sedimentary DNA is relatively local, pollen is able to spread before landing in sediment, and thus may not give an accurate representation of species composition in a specific location (Boessenkool et al., 2014).

To achieve a more complete look at ancient ecosystems, Willerslev et al. (2014) built upon a previous study from 2003 by studying not only plant diversity from sediment DNA, but also nematode diversity as a proxy for vegetation cover and the diets of ancient herbaceous megafauna as a method of confirming plant species presence and ecosystem-level interactions (Willerslev et al., 2003). By combining all results of ancient eDNA sequencing, the study was able to determine the circumpolar vegetation cover and the changes therein over time. Additionally, metabarcoding of stomach contents of ancient megafauna reveals information regarding habitat choice, behavior, and biology of species and their influence on the plant communities (Willerslev et al., 2014). Wood et al. (2012) also used metabarcoding to analyze the diet of extinct herbivores, focusing on the moa of New Zealand. They were able to identify at least 67 plant taxa, revealing a generalist feeding ecology across available habitats. Additionally, intact seeds indicate the potential importance of moa for plant dispersal (Wood et al., 2012). Microorganisms such as fungi also play an important role in ancient ecosystems and can even be used to determine plant and animal diversity based on associated fungal presence (Bellemain et al., 2013). Metabarcoding of permafrost has been successfully used to identify fungal communities with almost double the number of taxonomic units identified than with Sanger sequencing (Bellemain et al., 2013). While this is undoubtedly useful, differences in stability during cryopreservation and bias in PCR amplification and sequencing make it impossible to fully describe ancient fungal communities, though overall changes in diversity over time may be observed (Bellemain et al., 2013). Ancient DNA metabarcoding can also be used to determine anthropogenic effects on the landscape, as explored by Giguët-Covex et al. (2014). Lake sediments were used to reconstruct plant cover and livestock farming history to determine human causes of erosive events as compared to naturally predicted erosion. High amounts of cattle and sheep DNA in the sediment indicated intense grazing and a disappearance of tree species from deforestation would reduce soil stability and contribute to erosion (Giguët-Covex et al., 2014). Pansu et al. (2015) also assessed ancient human impacts on plant communities by metabarcoding lake sediments; they were able to identify anthropogenic changes to vegetation communities indicating the establishment of an agropastoral landscape.

The majority of ancient metabarcoding studies have used sediment to determine biodiversity, but bone fragments can be sequenced to determine their species of origin (Haouchar et al., 2014; Grealy et al., 2015). Bone fragments that are unidentifiable based on morphology can be combined and sequenced together to determine the species at a certain site where bones were found. In particular, bulk bone metabarcoding “has applications for the detection of cryptic or morphologically undiscoverable taxa, historical faunal turnover and population fragmentation” which may provide a powerful tool for species conservation as well as “identifying taxonomic assemblages, estimating genetic diversity, and assessing general ancient DNA preservation” (Grealy et al., 2015). Bulk bone metabarcoding can be combined with sediment analysis as indicated by Haouchar et al. (2014). They used a variety of general and taxa-specific primers on bone and sediment samples to assess the genetic biodiversity of a cave in Australia, from which at least 15 plant and animal taxa were identified, providing valuable insight to the ancient biodiversity of the area.

The possible applications of ancient DNA metabarcoding are staggering, but it is important to acknowledge potential flaws and limitations of current technology. Limitations of genetic libraries, differing rates of degradation between species and geographic location, and PCR bias are all major concerns (Bellemain et al., 2013; Pedersen et al., 2013). Another major concern is contamination; the presence of a modern species replicated by the primers can not only give false positives, but also false negatives by overwhelming the concentration of taxa to where low-concentration DNA is not able to replicate (Boessenkool et al., 2012). One of the contaminants most difficult to avoid is human DNA, which can cause problems when using mammal primers (Boessenkool et al., 2012). To combat this, Boessenkool et al. (2014) developed human-blocking primers to reduce the amplification of human DNA and found that species diversity increased, and new taxa were evident when the human DNA was suppressed. Other concerns include climatic/temporal limitations, biological origin/state, stratigraphic reliability, and sterile sampling procedure (Rawlence et al., 2014). In particular, ancient DNA is limited by the climate of the region; most studies so far have employed the technique in arctic or temperate climates; tropical regions are typically both warmer and more humid, which increases the rate of DNA degradation over time. As such, microclimates such as caves may be essential to the application of ancient eDNA metabarcoding in more tropical regions (Gutiérrez-García et al., 2014).

4.2. Plant-pollinator interactions

Environmental DNA metabarcoding can be used to characterize interactions between organisms and the environment. One of the most promising areas where this is being applied is in analyzing pollinator interactions with plants. Pollinators are essential to human survival and to general ecosystem health, but many populations are declining. The worldwide decline of bee species and colony collapse has the potential to be detrimental to global food production. In determining plant species most used by pollinators, conservation plans can be constructed to ensure these species are available for pollinators to use. Previously, the methods for determining the plants visited by pollinators involved either visually observing which plants individuals visited or collecting pollen to be identified under a microscope. Both methods are time-consuming and subject to bias, with the latter requiring a taxonomic pollen expert to do the identification. [Valentini et al. \(2010\)](#) used universal barcodes and NGS to determine the geographical origin of honey based on the plant DNA present in the honey samples. They found the method to be simple, fast, robust, and successful in identifying plant presence to pinpoint the origin of the samples. Additionally, the nature of honey prevents DNA degradation, so multiple, longer primer sets could be used if certain regions are indistinguishable with the universal primers ([Valentini et al., 2010](#)). [Hawkins et al. \(2015\)](#) tested the effectiveness of the metabarcoding method as compared to traditional, microscopic identification of pollen grains in honey. They found that the two methods had a correspondence of 92% when only high abundance taxa were considered, but that when all taxa were considered, the correspondence dropped to below 50%, with both methods finding taxa that were missed by the other. While metabarcoding is highly sensitive and able to provide greater taxonomic resolution, it is less likely to detect pollen present in low levels than traditional methods due to inherent bias in amplification and sequencing. [de Vere et al. \(2017\)](#) applied metabarcoding of honey samples directly to determination of bee plant preference. By testing honey in a highly diverse botanical garden, they were able to determine that bees preferentially used the same plant taxa composed of native or near-native plants found in woodlands and hedgerows, suggesting that maintenance of these natural environments free from invasive species is essential for bees to thrive ([de Vere et al., 2017](#)).

In addition to honey samples, pollen can be taken directly from pollinators for identification. [Keller et al. \(2015\)](#) compared metabarcoding to microscopy for mixed pollen samples collected from pollinators, testing whether primers were actually universal while being able to differentiate species. They found that at 85% match of sequencing results to genome libraries, 98.9% of samples were classified to genus and 61.6% were classified to species, with metabarcoding identifying 13 taxa missed by microscopy. Microscopy found 4 taxa missed by metabarcoding, but overall richness and abundance estimates were similar ([Keller et al., 2015](#)). [Richardson et al. \(2015a,b\)](#) published two studies on metabarcoding pollen from pollinators. The first tested a multi-locus approach to metabarcoding to characterize pollen from honey bees. This study found that using multiple loci was more reliable for identifying pollen as it helps to eliminate false positives and to avoid primer amplification bias. They also compared traditional pollen analysis to metabarcoding of pollen samples from honey bees. The results from this study indicate that metabarcoding is sensitive enough for identification of plants and is able to identify taxa missed by traditional analysis. However, it cannot quantify pollen analysis due to replication bias, and they therefore recommended that both methods be used together when quantification is necessary ([Richardson et al., 2015b, 2015a](#)). In the same year, [Sickel et al. \(2015\)](#) published a dual-index multiplexing method adapted from bacterial studies and applied it to the sequencing of pollen samples from multiple bee species simultaneously. They were able to adapt the method so that labelling and PCR were completed in one step and determined that 2000 to 3000 quality reads were needed in order to identify all taxa in the sample. In addition to general plant species use, metabarcoding of pollen from pollinators can be applied to look at landscape-level plant diversity of taxa most used by pollinators ([Cornman et al., 2015](#)). Networking and quantifying plant-pollinator interactions can also be accomplished by metabarcoding. [Pornon et al. \(2016\)](#) tested whether metabarcoding could circumvent the limits of conventional sampling in detecting and quantifying plant-pollinator interactions and found that metabarcoding of insect pollen loads revealed 2.5x more plant-pollinator interactions. Additionally, there was correlation between number of sequences with both pollen-carrying load and number of visits per plant species ([Pornon et al., 2016](#)). These interactions can be used to determine pollen transport networks and plant species choice among groups, as studied by Lucas et al., in 2018. They used metabarcoding of pollen from hoverflies to determine that there appears to be individual seasonal specialization in plant choice, while within groups there was redundancy ([Lucas et al., 2018](#)). Using eDNA metabarcoding on the pollen from migratory pollinators can reveal migratory distances and geographic ranges of migratory species ([Liu et al., 2017](#); [Suchan et al., 2018](#); [Chang et al., 2018](#)). [Suchan et al. \(2018\)](#) used this method on a model butterfly species along the European Mediterranean coast, finding pollen from taxa native to Africa, suggesting the possibility of insect-mediated transcontinental pollination.

Pollen metabarcoding is faster and more effective than direct observation for interactions and allows the study of plant-pollinator networks across ecological gradients ([Pornon et al., 2016](#)). Additionally, it is able to detect numerous interactions that would otherwise be missed by traditional methods, thus better describing pollinator behavior and plant choice ([Pornon et al., 2017](#)). [Bell et al. \(2017\)](#) tested the effectiveness of metabarcoding for creating networks by sampling pollen from 38 bee species across varying environments. From this data, they were able to match bee species to pollen presence and number of interacting individuals to quantify the interactions. Though metabarcoding of pollen to determine plant-pollinator interactions has benefits including faster turnaround time, hands-off work during which researchers can focus on other activities, the ability to include all pollen grains, more standardized and known techniques, greater taxonomic sensitivity, identification of novel taxa, and more predictable costs that may sometimes be lower than traditional pollen ID, there are

some downsides to the technique; namely, lack of complete libraries, potential for contamination and misidentification, and potential reduced amplification of rare species due to primer bias (Bell et al., 2017; Pornon et al., 2017).

4.3. Diet analysis

Perhaps one of the earliest applications of metabarcoding was in non-invasive diet analysis from fecal matter or stomach contents. Traditional diet analysis is labor intensive and involves expert morphological identification of partially or fully digested food material which is particularly difficult for species that are rare, small, aquatic, nocturnal, fast-moving, elusive, or that consume primarily soft-bodied organisms (Boyer et al., 2013; Berry et al., 2017). As such, a new method was needed to fully characterize the diets of species in the wild. Metabarcoding has shown to be a viable alternative and has been found to result in more in-depth characterization of diet than other methods for both herbivores and predators, particularly when standard forward and reverse primers are combined with a blocking primer to reduce amplification of the host species (Vestheim and Jarman, 2008; Deagle et al., 2009, 2010; McInnes et al., 2017a; Guillerault et al., 2017). Metabarcoding of feces and stomach contents has further applications beyond diet characterization. It has been used to study resource partitioning, dietary overlap and competition, impacts to forestry and fisheries, comparative conservation strategies, range management, habitat use, species interactions, large-scale monitoring, ecological impacts of changing populations, and human dimensions (Valentini et al., 2009; Kowalczyk et al., 2011; Shehzad et al., 2012a, 2012b; Baamrane et al., 2012; Quéméré et al., 2013; Hibert et al., 2013; Soininen et al., 2013, 2014; Jakubavičiūtė et al., 2017). The metabarcoding approach to diet analysis is fast, simple, robust, widely applicable, typically cheaper than traditional sampling and identification, often has high taxonomic resolution, and is able to detect DNA from digested materials even for soft-bodied or gelatinous prey (Valentini et al., 2009; Shehzad et al., 2012a; Boyer et al., 2013; Buglione et al., 2018). It has even been described as “one of the most accurate approaches available for increasing the understanding on the feeding habits of animals” (Buglione et al., 2018). Despite these definite advantages, there are still drawbacks to using metabarcoding for diet analysis. Poor database quality and imprecise taxonomic resolution are issues yet to be overcome. Despite this, it can be expected that as technology advances, metabarcoding will make these issues merely a problem of the past (Hibert et al., 2013; Yoon et al., 2017).

One of the most important specific methodologies for characterization of herbivorous diet using metabarcoding was developed by Valentini et al., in 2009: the *trnL* approach. This approach has been found to be able to identify about 50% of plant taxa to species from degraded DNA from feces by amplifying a 100–150 base pair region of the P6 loop of the chloroplast *trnL* intron, and can be applied to herbivorous birds, mammals, insects, and molluscs (Valentini et al., 2009). This method has been applied to numerous herbivore diet analysis studies since its development, including an experiment by Kowalczyk et al. (2011) studying European bison restored after extinction in the wild. In this study, the *trnL* approach of metabarcoding was used on feces from bison of different feeding groups (intensively fed, less intensively fed, non-fed in forest, and non-fed in agriculture surrounding forest), and found that when fed, bison consumed more herbs, grasses, and sedges, but when not fed they consumed more woody materials (Kowalczyk et al., 2011). As supplementary feeding has a negative impact on bison ecology and health, a conservation plan can be developed based on their dietary choices when human-provided food is not available; notably, the species of trees consumed could be identified and were determined to be insignificant to the forestry industry, further supporting the change to a non-feeding management style (Kowalczyk et al., 2011). Also using the *trnL* approach, Baamrane et al. (2012) evaluated food habits of the endangered Moroccan dorcas gazelle by metabarcoding fecal pellets and comparing results against a previously compiled database of plant species found on the study site. Plant preferences were able to be identified and data was compared to the results of a microhistological study in the 1980s. The modern study found less taxonomic richness, but a much more diversified diet, potentially due to changes in nutritional quality of plants over time (Baamrane et al., 2012). Hibert et al. (2013) used the *trnL* approach to study the diet of tapir, an elusive large rainforest herbivore. They were able to achieve taxonomic resolution to genus, and found great correlation with plants already known to be consumed by tapirs in the region, but new families of plants were also identified. Additionally, seasonality of diet could be observed by their use of metabarcoding tapir feces throughout the year (Hibert et al., 2013). In the same year, Quéméré et al. (2013) studied the dietary plasticity of the golden-crowned sifaka lemur in Madagascar by using the *trnL* approach on fecal material compared to a large DNA reference database. They discovered remarkable dietary diversity including over 130 identified species from varying habitats, suggesting a high flexibility of foraging strategies, including consumption of cultivated and naturalized plants, and thus confirming the species' ability to adapt to human-induced landscape changes.

Metabarcoding has also thoroughly characterized the diet of lemmings. Through metabarcoding of stomach contents, Soininen et al. (2013) found that lemmings primarily ingested grass, but that vascular plant consumption differed between habitat types, suggesting regional variation in lemming diet. Metabarcoding of stomach contents of several species of lemmings has revealed differences in composition and temporal variation (Soininen et al., 2014). Metabarcoding of lemming feces has been used to assess diet overlap and potential for competition in sympatric lemming species. The species were found to utilize the same plant as their primary food source in proportion with resource availability but differed in other dietary items, indicating a high potential for competition in populations that were not exhibiting any actual competition (Soininen et al., 2015). Kartzin et al. (2015) used metabarcoding and next-generation sequencing of fecal material from 7 large mammalian herbivore species in Kenya to determine resource partitioning between species. They found diet to be similar within species and strongly divergent between species, even when considering similar groups (i.e. grazing mammals may eat a similar percentage of grass, but different species will consume different grasses), suggesting that food choice is more closely

linked to plant species than type. Most recently, metabarcoding was used to characterize the diet and seasonal variations therein of the vulnerable Italian hare from only 22 fecal pellets, detecting 99 taxa of plants (Buglione et al., 2018). Applications of metabarcoding of herbivorous diets can be combined with those used for carnivorous diets to characterize the wide variety of food items consumed by omnivores, such as brown bears, which consume plants, invertebrates and vertebrates (De Barba et al., 2014).

Metabarcoding of feces and stomach contents can also be used to characterize the diets of carnivorous invertebrates. A study published in 2013 used metabarcoding of feces to analyze the diet of a carnivorous and critically endangered land snail, finding that the snails mostly consume earthworms that either live in the leaf litter or come to the surface to feed on the leaf litter at night, suggesting that the snail will eat any earthworm it encounters in the leaf litter (Boyer et al., 2013). This knowledge influences conservation decisions without having to harm or disturb the snails in their natural environment. In the same year, Piñol et al. (2014) published a study which investigated the diet of a generalist arthropod predator (linyphiid spider) using metabarcoding without a predator-blocking oligonucleotide. Of this data, only 6% of usable reads were from prey, but this amounted to over 61,000 reads and was enough to identify many prey taxa and to reach the conclusion that the spider exploits a large range of prey. Additionally, this study reached the conclusion that it is possible to characterize diet without developing a blocking oligonucleotide, which would increase the time, testing volume, and cost.

Metabarcoding of vertebrate predator feces emerged around the same time as did that of herbivores. Deagle et al. (2009) used 4 metabarcoding primer sets and a blocking oligonucleotide on feces of Australian fur seals in order to compare the process to traditional diet analysis. They identified 62 species in the seals' diet (including cartilaginous prey missed by traditional identification due to degradation), and were able to identify the most important prey species to the seals as well as location-based differences in prey composition. Also studying aquatic mammalian predators, Berry et al. (2017) applied metabarcoding to Australian sea lion scats to find 6 classes from 3 phyla for over 8 taxa altogether, some of which confirmed the consumption known dietary items while others had not previously been recorded as prey for these sea lions. These results indicate that the Australian sea lion is an opportunistic predator primarily consuming demersal and benthic fauna of different species composition between study sites (Berry et al., 2017). Metabarcoding of feces has also been used to compare diets of sympatric mammalian predators, such as the study by Bohmann et al. (2011) with African free-tailed bats. They found a large diversity of prey in the samples that were identified to species level at times and were able to determine the differences between the two species' diets. A similar study was recently published by Arrizabalaga-Escudero et al. (2018) in which they used metabarcoding on the feces of two Mediterranean bats in order to determine dietary differences; similar to Bohmann et al. (2011), they were able to characterize the diet of each species and determine the overlap therein (Arrizabalaga-Escudero et al., 2018). Shehzad et al. (2012a,b) developed a methodology for studying mammalian carnivore diet via metabarcoding of feces using the leopard cat as a model species, and then applied the same methodology to snow leopards. To do this, they used a vertebrate primer set on feces samples and tested the methodology both with and without a predator blocking oligonucleotide. In the leopard cat, they found 18 prey taxa including mammals, birds, amphibians, and fish, with most taxa and reads coming from rodents, confirming previous assumptions that the cat has an eclectic diet focused on rodents. The methodology was adapted for use with snow leopard feces where they observed 5 large ungulate prey items including domestic livestock. This knowledge of snow leopard diet can help to influence conservation strategies regarding livestock predation and retaliatory killing (Shehzad et al., 2012a).

Deagle et al. (2010) ran another metabarcoding study in which they pyrosequenced markers from feces of captive little penguins to see if sequences reflected proportions of species consumed. They found that the same species that were fed were detected, but the proportions were much different, and discovered that DNA can persist in feces for at least 4 days based on identification of species ingested by the penguins before they were taken into captivity for testing (Deagle et al., 2010). McInnes et al. (2017a,b) also studied seabird diet via metabarcoding by developing scat collection protocols for detection of food DNA in vertebrate scat by using shy albatross as a case study for methods that could be potentially applied to numerous species. They found that fresh scat yielded highest food sequences while collection from dirt reduced food sequences and increased contamination, that food DNA detection varied throughout breeding season, and that older individuals produced scat with more food DNA. They then proceeded to apply this procedure to assess albatross consumption of gelatinous zooplankton, finding that while fish were the primary dietary item, cnidarians were also prevalent in samples and detectable at all breeding and life stages (McInnes et al., 2017b). They compared this data with fishery statistics regarding jellyfish catch to discover a relationship that may indicate that jellyfish consumption in albatross is selective and contributes a significant factor to diet. This study would have been near-impossible before metabarcoding, as gelatinous zooplankton dissolve quickly once consumed (McInnes et al., 2017b).

Finally, metabarcoding has been used to characterize the diets of fish species including catfish, stickleback, and toothfish. Using the feces of a non-native catfish, Guillerault et al. (2017) compared metabarcoding to traditional prey identification finding 11 species with DNA and five with stomach content analysis for a total of 14 species, with two species identified using both methods. Even more amazingly, they were able to detect prey indirectly via parasite presence: while eels were not detected, an eel-specific parasite was, indicating that the catfish do in fact consume eels. Jakubaviciūtė et al. (2017) were able to identify 120 prey taxa consumed by the three-spined stickleback, a fish whose population is currently steeply increasing. From this unusually high prey diversity they were able to predict the potential impacts of the sticklebacks' increasing populations (Jakubaviciūtė et al., 2017). Applying metabarcoding to the stomach contents of Antarctic toothfish, Yoon et al. (2017) were able to identify a total of 19 species and to determine the most important prey species for the toothfish.

Additionally, they successfully obtained genotypes of prey items via next-generation sequencing and identified 17 more haplotypes, further expanding knowledge of toothfish diet. The scope of dietary metabarcoding studies is already wide and can only be expected to grow as technology improves and techniques are refined.

4.4. Detection of invasive species

Invasive and non-native species are some of the greatest threats to ecosystems as our planet becomes more connected. In particular, aquatic vessels have incredible potential to unknowingly transport dangerously prolific species into new habitats where they are capable of outcompeting native species. Current methodologies for aquatic microbiota inventory are diverse and rely on labor-intensive sampling and expert taxonomic identification (Borrell et al., 2017). Additionally, many marine pests cannot be identified morphologically at the larval/juvenile level, so they may go unnoticed at the early stages of infestation when they have the highest prospects of successful management (Pochon et al., 2013). Many factors can influence the transmission of species via vessels, such as activities performed on the boats, types of boats, and the season: motorboats have a higher rate of species translocation, and propagation is more likely during warmer months (Fletcher et al., 2017; Pochon et al., 2017). Invertebrates are able to survive in bilge and ballast water for at least 16 days, with protozoan species in particular able to multiply during that time and emerge from the pumps unharmed, possibly long distances away from their native environment (Zaiko et al., 2015a; Ardura et al., 2015; Fletcher et al., 2017). Metabarcoding offers a potential method to identify these species early and implement management plans, even for benthic organisms at the planktonic larval stage (Zaiko et al., 2015a). It is extremely sensitive, able to detect DNA at concentrations from 0.02 to 0.64% of the total biomass analyzed, able to detect a variety of taxa simultaneously, and able to be applied to both freshwater and marine environments (Pochon et al., 2013; Hatzenbuehler et al., 2017).

The practice of using metabarcoding to detect nonnative species was first used in marine environments. Ardura et al. (2015) sampled from ballast waters in a ship across its journey from the North Sea to the tropics and found that mollusk species could still be detected, including the invasive European mud snail. In the same year, Zaiko et al. (2015a,b) published two studies comparing results of metabarcoding to traditional identification. In ballast waters, metabarcoding found several noteworthy species that had been missed by traditional methods. In a study with a focus on plankton communities not associated with a vessel, they identified five different non-indigenous species, four of which were not found with traditional monitoring. In 2016, Zaiko et al. (2016) explored the scope of marine eukaryotic metabarcoding by sampling from settlement plates in a busy port. They found more than 400 taxa (several of which were non-indigenous), but even at poor resolution, data was consistent with the 25 taxa found by morphological analysis. In a study testing whether a minimum sampling effort for metabarcoding could compare with previously compiled nonnative invertebrate species, Borrell et al. (2017) found three nonnative species matching the results of traditional studies from only three 1 L bottles of water per port at eight ports. Testing metabarcoding on 30 bilge water samples, Fletcher et al. (2017) were able to identify five different non-indigenous species in 23 water samples, all at high taxonomic resolution. Pochon et al. (2017) took bilge water sampling beyond the status quo, using both eDNA and eRNA metabarcoding to test the potential for invasive identification. They found that eDNA and enhancer RNA (eRNA) gave different results in terms of species presence, with eDNA showing mostly fungi while eRNA showed mostly ciliates and metazoans. They hypothesized that this was due to the increased level of cell activity of ciliates and metazoans, and therefore increased eRNA output (Pochon et al., 2017).

As previously mentioned, metabarcoding is applicable to freshwater ecosystems as well as marine. In 2016, Brown et al. sampled zooplankton in Canadian ports to attempt to identify non-indigenous species via metabarcoding and successfully identified 25 non-indigenous species, 11 of which had never previously been reported in their locations. Blackman et al. (2017) used metabarcoding of freshwater samples from rivers and found it possible to reliably differentiate between native and invasive gammarid species, and that the method could be applied to the assessment of geographic distribution of invasive species. In the following year, Klymus et al. (2017a,b) developed two metabarcoding primers to use with eDNA from the Great Lakes, focusing on the detection of invasive gastropod and bivalve species. Their dual primer design allowed for inference of relative abundance of species, as well as detection of species missed by morphological survey. Most recently, Borrell et al. (2018) used eDNA metabarcoding in two estuaries to test detection of non-native species, and despite limited sampling, one invasive was identified and later confirmed by classic DNA barcoding studies of the estuaries.

Metabarcoding is a powerful tool, and is a more comprehensive method of monitoring and checking for invasive presence than traditional identification (Zaiko et al., 2015b; Brown et al., 2016). Additionally, the cost of metabarcoding is currently comparable to other methods and is expected to decrease as it becomes more common (Borrell et al., 2017). As with other applications, it does have drawbacks. For one, it may bias biodiversity assessments by capturing signals from dead organisms, extracellular DNA, or false readings due to taxonomic selectivity and restricted sensitivity of primers (Fletcher et al., 2017). False readings, either positive or negative, can be problematic in invasive species detection; false positives could trigger an unwarranted and potentially expensive response, while false negatives can result in an invasion that could have potentially be prevented (Darling and Mahon, 2011; Ficetola et al., 2015). Another drawback to this method is the current lack of comprehensive genetic databases, and the unavoidability of primer and amplification bias make quantification impossible and require careful analysis of results (Brown et al., 2016; Hatzenbuehler et al., 2017). Regardless, metabarcoding is a unique tool that can only be expected to improve over time from its already wholly applicable and useful state.

4.5. Pollution response

Uses of eDNA metabarcoding extend to the ability to track ecosystem responses to pollution. While many pollutants do not have DNA and thus cannot be detected, they often cause changes in microbiological communities affected by them. Studies can be done to determine which species are most likely to be present or absent in the case of a specific pollutant to help determine the severity of the pollution. Until that point, responses to pollution can be monitored in areas with known pollutants or in controlled situations to determine differences in community composition. Perhaps some of the more high-profile studies of pollution response are those which aim to determine differences in communities surrounding offshore oil drilling platforms or in response to oil pollution events. [Smith et al. \(2015\)](#) used statistical analysis of microbial communities to identify environmental contaminants at a nuclear waste site, and then applied the approach to hydrocarbon contamination from the Deepwater Horizon oil spill. They found that even after the contaminants had been fully degraded, the bacterial communities affected were altered based on the interaction, indicating that bacterial communities have the potential to be used as environmental sensors to evaluate human impacts. [Coelho et al. \(2016\)](#) used microbial activity profiling to evaluate responses to ocean acidification and oil hydrocarbon contamination in lab conditions, determining that the resulting lower relative abundance of sulphate-reducing bacteria may be coupled to changes in sediment archaeal communities. Additionally, they found increased abundance of hydrocarbonoclastic fungi and changes to the sediment chemistry due to the changes in the microbial community, all of which impair the ecosystem's ability to recover from oil contamination events. In terms of community responses surrounding offshore drilling sites, [Lanzén et al. \(2016\)](#) found changes in sediment communities surrounding oil platforms, with metazoans appearing to be the most sensitive eukaryotic taxa to the contaminant effects of drilling, while [Laroche et al. \(2018\)](#) determined that bacteria were highly responsive to drilling activity as well, noting the presence of hydrocarbon-degrading taxa at the petroleum impacted stations, indicating presence of taxa highly specific to site conditions. Both studies identified potential pollution indicator species. Another important sphere of pollution to be considered is that of marine fisheries. Products from these facilities have the potential to affect the marine microbiome, but traditional monitoring of these species is time-consuming and requires extensive specialization. Environmental metabarcoding can be used to estimate community composition, and to identify indicator species. Foraminiferal responses have been studied multiple times, and candidate bioindicator species identified therein. [Pawlowski et al. \(2014\)](#) found high variation between communities collected near the fish farms versus at a distance, with species richness decreasing nearer to the farms, while [Pochon et al. \(2015\)](#) found clear shifts in community structures between farm sites and control sites and differences in structure between high and low flow environments. Bacterial communities may also be important for assessing the impact of fisheries on ecosystems. [Dowle et al. \(2015\)](#) tested the impact of organic sediment enrichment from a salmon farm on bacterial communities within the sediment along a gradient of distances from the farm. They found stark differences between bacterial communities near the farm as compared to control sites, and identified a strong relationship between bacterial communities and total organic matter, suggesting that metabarcoding bacterial communities may be a useful tool in assessing impacts of aquaculture ([Dowle et al., 2015](#)). [Stoeck et al.](#) published two studies in 2018 regarding microbial response to fish farming, one assessing benthic bacterial communities and the other testing protist presence. They found that deltaproteobacteria were predominant in immediate vicinity of cages, while gammaproteobacteria dominated less impacted sites, indicating changes in community structure due to salmon farming. Additionally, alpha and beta diversity correlated with macrofauna diversity metrics, indicating similar responses to farming, thus making bacterial eDNA metabarcoding a potential alternative bio-monitoring method. For protist response, they found that ciliate communities were the most powerful indicators of fisheries pollution when compared to traditional macrofaunal surveys, and that the use of ciliates can allow for an even finer scale resolution ([Stoeck et al., 2018a,b](#)).

Aquatic communities may be subject to pollution and contamination beyond that of oil drilling and commercial fisheries, further extending the potential applications of environmental metabarcoding. [Xie et al. \(2017\)](#) studied eukaryotic community response to different land use types and chemical contaminants, finding that structures of eukaryotic communities were significantly different between land-use types and identifying key contaminants such as pyrethroids, Mn, Zn, Pb, Cr and polycyclic aromatic hydrocarbons (PAHs). Negative associations of eukaryotic classes with these contaminants suggest that there is some degree of toxicity. Other studies focused on specific contaminant effects. [Chariton et al. \(2014\)](#) examined the responses of benthic communities exposed to elevated concentrations of triclosan, a common antimicrobial agent, and found a decline of metazoan diversity, especially in the taxa Chromadorea, Bivalvia, and Kinorhyncha. [Frontalini et al. \(2018\)](#) tested the response of benthic foraminifera communities exposed to different concentrations of Hg, finding that Hg pollution is detrimental to foraminifera with diversity decreasing as Hg levels increase. Potential bioindicators of Hg pollution were also identified ([Frontalini et al., 2018](#)).

This technique has been used in terrestrial as well as aquatic environments. [Bell et al. \(2014\)](#) compared fungal and bacterial communities of 66 soil samples from the rhizospheres of planted willows and six unplanted control samples at the site of a former petrochemical plant by metabarcoding, finding that fungal communities changed drastically between sites, and bacterial communities changed to a lesser degree between sites with a decrease in diversity in hydrocarbon contaminated sites. Additionally, they found that willow phylogeny influenced fungal diversity in these sites. Heavy metals are also a concern in polluted soils. [Azarbad et al. \(2015\)](#) tested microbial communities associated with forests in metal (Zn and Pb) polluted soils in Poland and found that the metal pollution level impacted bacterial community structure, but not taxon richness or community composition; rather, the relative abundance of specific bacterial taxa

changed. In the same year, Zappellini et al. (2015) reported bacterial and fungal communities from both chlor-alkali residue (high in Hg and As) and nearby undisturbed soil to determine community composition. They found a dominance of gammaproteobacteria and *Pseudomonas* sp. at the study site, and minor differences between genera of fungal communities. Foulon et al. (2016) compared microbial community composition at two sites with varying environmental conditions, but both contaminated with Zn, Cd, and Pb: Pierrelaye, which was a sandier soil type enriched with P, C, and total N, and Leforest, which was a more silt/clay soil type with a lower pH and higher amounts of Cd and Zn (Foulon et al., 2016). They found that Ascomycota dominated overall poplar soil, root samples at Pierrelaye, and unplanted soil, while Basidiomycota was more dominant in root samples from Leforest (Foulon et al., 2016). The dominance of an ectomycorrhizal community at Leforest was potentially favored due to strong resistance to environmental stress, while at Pierrelaye where soils were more fertile and resources more available, other species are able to thrive (Foulon et al., 2016). Most recently, fungal communities at an Hg-contaminated phytomanagement site of poplar trees were characterized using environmental metabarcoding (Durand et al., 2017). Communities were found to be negatively affected by Hg concentration, and aboveground and belowground fungi communities were found to be almost completely isolated from one another (Durand et al., 2017).

Even areas specifically denoted to contain pollution are capable of having an effect on wildlife. In 2017, Klymus et al. tested eDNA metabarcoding as a survey tool to examine potential use of uranium mine containment ponds as water sources by wildlife by testing ponds near mines and one mine containment pond using two markers for vertebrates. They detected two species missed by traditional surveys, including the presence of tiger salamanders in the containment pond. Finally, while perhaps not a response to a pollutant in the traditional sense, environmental metabarcoding can be used to test the effects of invasive species on native communities. Wangenstein et al. (2018a,b) used multigene metabarcoding to observe littoral community responses to three invasive, canopy-forming seaweeds. They found that the presence of invasive seaweed had a significant effect on the understory community for two of the three species. Overall, pollution response studies have found metabarcoding to be more sensitive in revealing community changes than traditional surveys; however, it is still subject to amplification bias, interpretation uncertainties, and other roadblocks previously discussed (Klymus et al., 2017b; Frontalini et al., 2018).

4.6. Air quality monitoring

DNA metabarcoding can also have applications in human health; determining the microbiome in the air can influence health decisions regarding allergens, pathogenic organisms, and air pollution. Current methods in studying airborne allergens are microscope-based, labor intensive, and cannot identify to the genus level in all relevant groups (Kraaijeveld et al., 2015). By collecting samples from the air and applying metabarcoding and next-generation sequencing technology, it is possible to determine the composition of the airborne microbiome and its potential implications for human and ecosystem health.

Metabarcoding of pollen was found in 2014 to be more efficient and more sensitive than current pollen monitoring techniques; however, a simple pollen count is needed in addition to metabarcoding data in order to properly quantify airborne pollen (Kraaijeveld et al., 2015). Leontidou et al. (2018) created a standardized procedure for sample preparation and DNA extraction for airborne pollen metabarcoding by comparing metabarcoding results to morphological identification. They found that even in complex samples with more than 46 families, plants can be identified efficiently by metabarcoding with greater taxonomic resolution than by morphological identification.

Airborne fungal diversity is another important facet of the aerial microbiome. Presence and abundance of taxa in fungal airborne samples can be variable over time and impacted by climatic conditions, and because many mycelium fragments and spores have the potential to be toxic or cause allergies, it is important to describe airborne fungal diversity (Banchi et al., 2018). Previous studies have relied on a few indicator species and taxa of key importance, but with metabarcoding all taxa can be analyzed. Oh et al. (2014) used metagenomics to study airborne fungal diversity and found over 1000 unique OTUs, with a set of 40 potentially allergy-inducing genera identified accounting for approximately 20% of the airborne fungal diversity. Banchi et al. (2018) specifically compared metabarcoding to traditional airborne fungal analysis and found that metabarcoding provided a 10x more comprehensive determination of taxa compared to microscopy; even so, some taxa were only identified in one technique or the other, making both methods necessary for comprehensive characterization of the fungal community.

While the most frequent use of environmental metabarcoding is in natural environments, the methodology can be applied to indoor locations as well. In particular, airborne microorganisms are a critical concern in hospitals, where some have the potential to cause hospital-associated infections. Tong et al. (2017) compared results of metabarcoding air samples and traditional culture-based methods to describe the microbial diversity. In this study, few bacteria were grown after cultivation, but sequencing revealed the relative abundance of fungi, archaea, bacteria, and viruses. Knowing the species present can inform infection control plans in the case of an outbreak, as sampling different areas of the hospital can offer a better idea of what control methods are necessary (Tong et al., 2017). Cao et al. (2014) applied a similar method of characterizing the full microbiome; however, their study was outdoors in Beijing during a severe smog event. While particulate matter air pollution is well-understood, inhalable microorganisms have not been well studied, and most studies have been purposed for taxonomic identification rather than pathogenic potential (Cao et al., 2014). This study used metabarcoding to characterize the airborne microbial community and was able to identify most taxa to the species level with the results. In addition, they discovered the presence of some respiratory microbial allergens and pathogens (Cao et al., 2014).

5. Future directions

For eDNA metabarcoding to truly take off, current assessments of ecological quality would need to be adapted to the eDNA metabarcoding framework. These changes must be feasible on a large scale, particularly when considering thresholds between countries and the differences between traditional and molecular methods. To calibrate, molecular methods would need to be applied simultaneously with existing systems in key environmental gradients which would likely be accomplished during development and testing of molecular methods (Hering et al., 2018). As such, a comprehensive eDNA monitoring program could be possible in just a few years across a variety of environments and taxa, thus providing a global framework for ecosystem modelling and function to inform management decisions. To handle this shift, reference databases would need to be expanded to include identifiable sequences for all target biodiversity, and advanced facilities will need to be created to house the growing data libraries. Wide-spread implementation would benefit from optimized bioinformatics pathways with user-friendly interfaces, as well as general improvements to the eDNA collection and extraction methodologies, particularly in terms of accounting for degradation. Metabarcoding has the potential to complement higher cost biodiversity measures and even replace them in some instances, especially as it becomes more widespread and the cost goes down (Bohmann et al., 2014; Creer et al., 2016; Deiner et al., 2017). Ecological indicators are essential for conservation science, but full biodiversity assessments that are rapid, cost-effective, and noninvasive are also important to conservation biology (Ji et al., 2013; Deiner et al., 2017). While eDNA metabarcoding cannot give information as to the health or demographics of a population, it has the potential to assess long-term changes in ecosystems and local extinctions over time, including for modeling the impacts of climate change on populations (Deiner et al., 2017).

eDNA also has applications in citizen science by engagement of citizen collection of eDNA using commercially available sampling kits, involving the public in biodiversity sampling in a way that could be complementary to already established methods. However it is limited by the difficulty in visualizing and explaining data to laypeople. More user-friendly methods of data exploration are needed (Bohmann et al., 2014; Deiner et al., 2017; Harper et al., 2018). It may even be possible to automate eDNA sampling in the future, but further research is needed into the temporal longevity and spatial dispersal of eDNA in all ecosystems for studies of abundance and richness to be considered fully valid (Barnes and Turner, 2016; Bohmann et al., 2014; Deiner et al., 2017). Metagenomics, or shotgun sequencing of environmental samples without relying on a PCR amplification step, avoids primer bias and could theoretically be used for biodiversity studies. However, because it requires a huge increase in sequencing power to acquire adequate depth, the cost is often prohibitive. As technology advances, this may become a commonplace method of biodiversity monitoring (Taberlet et al., 2012a; Yu et al., 2012; Cristescu, 2014; Creer et al., 2016; Deiner et al., 2017; Hering et al., 2018). There is a definite need to let methods mature as eDNA metabarcoding is a developing field, and thus apply the evolution of the method to reliable indices and ensure both forward and backward compatibility (Hering et al., 2018).

6. Conclusion

In conclusion, the potential of eDNA metabarcoding in biological research seems almost limitless, but the technique requires scientific collaboration and coordination. As technology advances and procedures are optimized, metabarcoding will likely emerge as an essential tool for a number of scientific tasks not only in biodiversity monitoring but also in the numerous areas discussed above. eDNA metabarcoding brings together several fields for the assessment of communities for purposes ranging from ecosystem restoration to human health, making it extremely versatile and vital to the future of molecular research.

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References

- Abad, D., Albaina, A., Aguirre, M., Laza-Martínez, A., Uriarte, I., Iriarte, A., Villate, F., Estonba, A., 2016. Is metabarcoding suitable for estuarine plankton monitoring? A comparative study with microscopy. *Mar. Biol.* 163, 149.
- Alberdi, A., Aizpurua, O., Gilbert, M.T.P., Bohmann, K., 2018. Scrutinizing key steps for reliable metabarcoding of environmental samples. *Meth. Ecol. Evolut.* 9, 134–147.
- Alsos, I.G., Lammers, Y., Yoccoz, N.G., Jørgensen, T., Sjögren, P., Gielly, L., Edwards, M.E., 2018. Plant DNA metabarcoding of lake sediments: how does it represent the contemporary vegetation. *PLoS One* 13, e0195403.
- Andersen, K., Bird, K.L., Rasmussen, M., Haile, J., Breuning-Madsen, H., Kjær, K.H., Orlando, L., Gilbert, M.T.P., Willerslev, E., 2012. Meta-barcoding of “dirt” DNA from soil reflects vertebrate biodiversity. *Mol. Ecol.* 21, 1966–1979.
- Andruszkiewicz, E.A., Starks, H.A., Chavez, F.P., Sassoubre, L.M., Block, B.A., Boehm, A.B., 2017. Biomonitoring of marine vertebrates in Monterey Bay using eDNA metabarcoding. *PLoS One* 12, e0176343.
- Ardura, A., Zaiko, A., Martinez, J.L., Samuiloviene, A., Borrell, Y., Garcia-Vazquez, E., 2015. Environmental DNA evidence of transfer of North Sea molluscs across tropical waters through ballast water. *J. Molluscan Stud.* 81, 495–501.

- Arrizabalaga-Escudero, A., Clare, E.L., Salsamendi, E., Alberdi, A., Garin, I., Aihartza, J., Goiti, U., 2018. Assessing niche partitioning of co-occurring sibling bat species by DNA metabarcoding. *Mol. Ecol.* 27, 1273–1283.
- Avó, A.P., Daniell, T.J., Neilson, R., Oliveira, S., Branco, J., Adão, H., 2017. DNA barcoding and morphological identification of benthic nematodes assemblages of estuarine intertidal sediments: advances in molecular tools for biodiversity assessment. *Front. Mar. Sci.* 4.
- Aylagas, E., Borja, Á., Irigoien, X., Rodríguez-Espeleta, N., 2016. Benchmarking DNA metabarcoding for biodiversity-based monitoring and assessment. *Front. Mar. Sci.* 3.
- Azarbad, H., Niklińska, M., Laskowski, R., van Straalen, N.M., van Gestel, C.A.M., Zhou, J., He, Z., Wen, C., Röling, W.F.M., 2015. Microbial community composition and functions are resilient to metal pollution along two forest soil gradients. *FEMS (Fed. Eur. Microbiol. Soc.) Microbiol. Ecol.* 91, 1–11.
- Baamrane, M.A.A., Shehzad, W., Ouhammou, A., Abbad, A., Naimi, M., Coissac, E., Taberlet, P., Znari, M., 2012. Assessment of the food habits of the Moroccan dorcas gazelle in M'Sabih Talaa, west Central Morocco, using the *trnL* approach. *PLoS One* 7, e35643.
- Banchi, E., Ametrano, C.G., Stanković, D., Verardo, P., Moretti, O., Gabrielli, F., Lazzarin, S., Borney, M.F., Tassan, F., Tretiach, M., Pallavicini, A., Muggia, L., 2018. DNA metabarcoding uncovers fungal diversity of mixed airborne samples in Italy. *PLoS One* 13, e0194489.
- Banerji, A., Bagley, M., Elk, M., Pilgrim, E., Martinson, J., Domingo, J.S., 2018. Spatial and temporal dynamics of a freshwater eukaryotic plankton community revealed via 18S rRNA gene metabarcoding. *Hydrobiologia* 818, 71–86.
- Barnes, M.A., Turner, C.R., 2016. The ecology of environmental DNA and implications for conservation genetics. *Conserv. Genet.* 17, 1–17.
- Bell, K.L., Fowler, J., Burgess, K.S., Dobbs, E.K., Gruenewald, D., Lawley, B., Morozumi, C., Brosi, B.J., 2017. Applying pollen DNA metabarcoding to the study of plant–pollinator interactions. *Appl. Plant Sci.* 5, 1600124.
- Bell, T.H., Hassan, S.E., Lauron-Moreau, A., Al-Otaibi, F., Hijri, M., Yergeau, E., St-Arnaud, M., 2014. Linkage between bacterial and fungal rhizosphere communities in hydrocarbon-contaminated soils is related to plant phylogeny. *ISME J.* 8, 331–343.
- Bellemain, E., Davey, M.L., Kausrud, H., Epp, L.S., Boessenkool, S., Coissac, E., Geml, J., Edwards, M., Willerslev, E., Gussarova, G., Taberlet, P., Brochmann, C., 2013. Fungal palaeodiversity revealed using high-throughput metabarcoding of ancient DNA from arctic permafrost. *Environ. Microbiol.* 15, 1176–1189.
- Berry, D., Mahfoudh, K.B., Wagner, M., Loy, A., 2011. Barcoded primers used in multiplex amplicon pyrosequencing bias amplification. *Appl. Environ. Microbiol.* 77, 7846–7849.
- Berry, T.E., Osterrieder, S.K., Murray, D.C., Coghlan, M.L., Richardson, A.J., Grealy, A.K., Stat, M., Bejder, L., Bunce, M., 2017. DNA metabarcoding for diet analysis and biodiversity: a case study using the endangered Australian sea lion (*Neophoca cinerea*). *Ecol. Evol.* 7, 5435–5453.
- Besemer, K., Singer, G., Quince, C., Bertuzzo, E., Sloan, W., Battin, T.J., 2013. Headwaters are critical reservoirs of microbial diversity for fluvial networks. *Proc. R. Soc. B* 280, 20131760.
- Binladen, J., Gilbert, M.T.P., Bollback, J.P., Panitz, F., Bendixen, C., Nielsen, R., Willerslev, E., 2007. The use of coded PCR primers enables high-throughput sequencing of multiple homolog amplification products by 454 parallel sequencing. *PLoS One* 2, e197.
- Blaalid, R., Carlsen, T., Kumar, S., Halvorsen, R., Ugland, K.I., Fontana, G., Kausrud, H., 2012. Changes in the root-associated fungal communities along a primary succession gradient analysed by 454 pyrosequencing. *Mol. Ecol.* 21, 1897–1908.
- Blackman, R.C., Constable, D., Hahn, C., Sheard, A.M., Durkota, J., Hänfling, B., Handley, L.L., 2017. Detection of a new non-native freshwater species by DNA metabarcoding of environmental samples – first record of *Gammarus fossarum* in the UK. *Aquat. Invasions* 12, 177–189.
- Boessenkool, S., Epp, L.S., Haile, J., Bellemain, E., Edwards, M., Coissac, E., Willerslev, E., Brochmann, C., 2012. Blocking human contaminant DNA during PCR allows amplification of rare mammal species from sedimentary ancient DNA. *Mol. Ecol.* 21, 1806–1815.
- Boessenkool, S., McGlynn, G., Epp, L.S., Taylor, D., Pimentel, M., Gizaw, A., Nemomissa, S., Brochmann, C., Popp, M., 2014. Use of ancient sedimentary DNA as a novel conservation tool for high-altitude tropical biodiversity. *Conserv. Biol.* 28, 446–455.
- Bohmann, K., Evans, A., Gilbert, M.T.P., Carvalho, G.R., Creer, S., Knapp, M., Yu, D.W., de Bruyn, M., 2014. Environmental DNA for wildlife biology and biodiversity monitoring. *Trends Ecol. Evol.* 29, 358–367.
- Bohmann, K., Monadjem, A., Noer, C.L., Rasmussen, M., Zeale, M.R.K., Clare, E., Jones, G., Willerslev, E., Gilbert, M.T.P., 2011. Molecular diet analysis of two African free-tailed bats (*Molossidae*) using high throughput sequencing. *PLoS One* 6, e21441.
- Borrell, Y.J., Miralles, L., Huu, H.D., Mohammed-Geba, K., Garcia-Vazquez, E., 2017. DNA in a bottle—rapid metabarcoding survey for early alerts of invasive species in ports. *PLoS One* 12, e0183347.
- Borrell, Y.J., Miralles, L., Martínez-Marqués, A., Semeraro, A., Arias, A., Carleos, C.E., García-Vázquez, E., 2018. Metabarcoding and post-sampling strategies to discover non-indigenous species: a case study in the estuaries of the central south Bay of Biscay. *J. Nat. Conserv.* 42, 67–74.
- Boyer, S., Wratten, S.D., Holyoake, A., Abdelkrim, J., Cruickshank, R.H., 2013. Using next-generation sequencing to analyse the diet of a highly endangered land snail (*Powelliphanta augusta*) feeding on endemic earthworms. *PLoS One* 8, e75962.
- Brannock, P.M., Ortmann, A.C., Moss, A.G., Halanich, K.M., 2016. Metabarcoding reveals environmental factors influencing spatio-temporal variation in pelagic micro-eukaryotes. *Mol. Ecol.* 25, 3593–3604.
- Brown, E.A., Chain, F.J.J., Zhan, A., MacIsaac, H.J., Cristescu, M.E., 2016. Early detection of aquatic invaders using metabarcoding reveals a high number of non-indigenous species in Canadian ports. *Divers. Distrib.* 22, 1045–1059.
- Buée, M., Reich, M., Murat, C., Morin, E., Nilsson, R.H., Uroz, S., Martin, F., 2009. 454 Pyrosequencing analyses of forest soils reveal an unexpectedly high fungal diversity. *New Phytol.* 184, 449–456.
- Buglione, M., Maselli, V., Ripa, D., de Filippo, G., Trapanese, M., Fulgione, D., 2018. A pilot study on the application of DNA metabarcoding for non-invasive diet analysis in the Italian hare. *Mamm. Biol.* 88, 31–42.
- Calvignac-Spencer, S., Merkel, K., Kutzner, N., Köhl, H., Boesch, C., Kappeler, P.M., Metzger, S., Schubert, G., Leendertz, F.H., 2013. Carrion fly-derived DNA as a tool for comprehensive and cost-effective assessment of mammalian biodiversity. *Mol. Ecol.* 22, 915–924.
- Cao, C., Jiang, W., Wang, B., Fang, J., Lang, J., Tian, G., Jiang, J., Zhu, T.F., 2014. Inhalable microorganisms in Beijing's PM2.5 and PM10 pollutants during a severe smog event. *Environ. Sci. Technol.* 48, 1499–1507.
- Carew, M.E., Pettigrove, V.J., Metzeling, L., Hoffmann, A.A., 2013. Environmental monitoring using next generation sequencing: rapid identification of macroinvertebrate bioindicator species. *Front. Zool.* 10, 45.
- Chain, F.J.J., Brown, E.A., MacIsaac, H.J., Cristescu, M.E., 2016. Metabarcoding reveals strong spatial structure and temporal turnover of zooplankton communities among marine and freshwater ports. *Divers. Distrib.* 22, 493–504.
- Champlot, S., Berthelot, C., Pruvost, M., Bennett, E.A., Grange, T., Geigl, E.-M., 2010. An efficient multistrategy DNA decontamination procedure of PCR reagents for hypersensitive PCR applications. *PLoS One* 5, e13042.
- Chang, H., Guo, J., Fu, X., Liu, Y., Wyckhuys, K.A.G., Hou, Y., Wu, K., 2018. Molecular-assisted pollen grain analysis reveals spatiotemporal origin of long-distance migrants of a noctuid moth. *Int. J. Mol. Sci.* Basel 19, 567.
- Chariton, A.A., Court, L.N., Hartley, D.M., Colloff, M.J., Hardy, C.M., 2010. Ecological assessment of estuarine sediments by pyrosequencing eukaryotic ribosomal DNA. *Front. Ecol. Environ.* 8, 233–238.
- Chariton, A.A., Ho, K.T., Proestou, D., Bik, H., Simpson, S.L., Portis, L.M., Cantwell, M.G., Baguley, J.G., Burgess, R.M., Pelletier, M.M., Perron, M., Gansch, C., Matthews, R.A., 2014. A molecular-based approach for examining responses of eukaryotes in microcosms to contaminant-spiked estuarine sediments. *Environ. Toxicol. Chem.* 33, 359–369.
- Chariton, A.A., Stephenson, S., Morgan, M.J., Steven, A.D.L., Colloff, M.J., Court, L.N., Hardy, C.M., 2015. Metabarcoding of benthic eukaryote communities predicts the ecological condition of estuaries. *Environ. Pollut.* 203, 165–174.
- Civade, R., Dejean, T., Valentini, A., Roset, N., Raymond, J., Bonin, A., Taberlet, P., Pont, D., 2016. Spatial representativeness of environmental DNA metabarcoding signal for fish biodiversity assessment in a natural freshwater system. *PLoS One* 11, e0157366.
- Coelho, F.J.R.C., Cleary, D.F.R., Costa, R., Ferreira, M., Polónia, A.R.M., Silva, A.M.S., Simões, M.M.Q., Oliveira, V., Gomes, N.C.M., 2016. Multitaxon activity profiling reveals differential microbial response to reduced seawater pH and oil pollution. *Mol. Ecol.* 25, 4645–4659.
- Coissac, E., Riaz, T., Puillandre, N., 2012. Bioinformatic challenges for DNA metabarcoding of plants and animals. *Mol. Ecol.* 21, 1834–1847.

- Cordier, T., Esling, P., Lejzerowicz, F., Visco, J., Ouadahi, A., Martins, C., Cedhagen, T., Pawlowski, J., 2017. Predicting the ecological quality status of marine environments from eDNA metabarcoding data using supervised machine learning. *Environ. Sci. Technol.* 51, 9118–9126.
- Cornman, R.S., Otto, C.R.V., Iwanowicz, D., Pettis, J.S., 2015. Taxonomic characterization of honey bee (*Apis mellifera*) pollen foraging based on non-overlapping paired-end sequencing of nuclear ribosomal loci. *PLoS One* 10, e0145365.
- Cowart, D.A., Pinheiro, M., Mouchel, O., Maguer, M., Grall, J., Miné, J., Arnaud-Haond, S., 2015. Metabarcoding is powerful yet still blind: a comparative analysis of morphological and molecular surveys of seagrass communities. *PLoS One* 10, e0117562.
- Creer, S., Deiner, K., Frey, S., Porazinska, D., Taberlet, P., Thomas, W.K., Potter, C., Bik, H.M., 2016. The ecologist's field guide to sequence-based identification of biodiversity. *Meth. Ecol. Evolut.* 7, 1008–1018.
- Cristescu, M.E., 2014. From barcoding single individuals to metabarcoding biological communities: towards an integrative approach to the study of global biodiversity. *Trends Ecol. Evol.* 29, 566–571.
- Darling, J.A., Mahon, A.R., 2011. From molecules to management: adopting DNA-based methods for monitoring biological invasions in aquatic environments. *Environ. Res.* 111, 978–988.
- Deagle, B.E., Chiaradia, A., McInnes, J., Jarman, S.N., 2010. Pyrosequencing faecal DNA to determine diet of little penguins: is what goes in what comes out? *Conserv. Genet.* 11, 2039–2048.
- Deagle, B.E., Clarke, L.J., Kitchener, J.A., Polanowski, A.M., Davidson, A.T., 2018. Genetic monitoring of open ocean biodiversity: an evaluation of DNA metabarcoding for processing continuous plankton recorder samples. *Mol. Ecol. Res.* 18, 391–406.
- Deagle, B.E., Kirkwood, R., Jarman, S.N., 2009. Analysis of Australian Fur seal diet by pyrosequencing prey DNA in faeces. *Mol. Ecol.* 18, 2022–2038.
- De Barba, M.D., Miquel, C., Boyer, F., Mercier, C., Rioux, D., Coissac, E., Taberlet, P., 2014. DNA metabarcoding multiplexing and validation of data accuracy for diet assessment: application to omnivorous diet. *Mol. Ecol. Res.* 14, 306–323.
- Deiner, K., Bik, H.M., Mächler, E., Seymour, M., Lacoursière-Roussel, A., Altermatt, F., Creer, S., Bista, I., Lodge, D.M., de Vere, N., Pfrender, M.E., Bernatchez, L., 2017. Environmental DNA metabarcoding: Transforming how we survey animal and plant communities. *Mol. Ecol.* 26, 5872–5895.
- Deiner, K., Walser, J., Mächler, E., Altermatt, F., 2015. Choice of capture and extraction methods affect detection of freshwater biodiversity from environmental DNA. *Biol. Conserv.* 183, 53–63.
- Dell'Anno, A., Carugati, L., Corinaldesi, C., Riccioni, G., Danovaro, R., 2015. Unveiling the biodiversity of deep-sea nematodes through metabarcoding: are we ready to bypass the classical taxonomy? *PLoS One* 10, e0144928.
- de Vere, N., Jones, L.E., Gilmore, T., Moscrop, J., Lowe, A., Smith, D., Hegarty, M.J., Creer, S., Ford, C.R., 2017. Using DNA metabarcoding to investigate honey bee foraging reveals limited flower use despite high floral availability. *Sci. Rep.* 7, 42838.
- Djurhuus, A., Pitz, K., Sawaya, N.A., Rojas-Márquez, J., Michaud, B., Montes, E., Müller-Karger, F., Breitbart, M., 2018. Evaluation of marine zooplankton community structure through environmental DNA metabarcoding. *Limnol. Oceanogr. Methods* 16, 209–221.
- Dowle, E., Pochon, X., Keeley, N., Wood, S.A., 2015. Assessing the effects of salmon farming seabed enrichment using bacterial community diversity and high-throughput sequencing. *FEMS (Fed. Eur. Microbiol. Soc.) Microbiol. Ecol.* 91.
- Drummond, A.J., Newcomb, R.D., Buckley, T.R., Xie, D., Dopheide, A., Potter, B.C., Heled, J., Ross, H.A., Tooman, L., Grosser, S., Park, D., Demetras, N.J., Stevens, M.I., Russell, J.C., Anderson, S.H., Carter, A., Nelson, N., 2015. Evaluating a multigene environmental DNA approach for biodiversity assessment. *GigaScience* 4.
- Durand, A., Maillard, F., Foulon, J., Gweon, H.S., Valot, B., Chalot, M., 2017. Environmental metabarcoding reveals contrasting belowground and aboveground fungal communities from poplar at a Hg phytomanagement site. *Microb. Ecol.* 74, 795–809.
- Eichmiller, J.J., Best, S.E., Sorensen, P.W., January 28, 2016. Effects of temperature and trophic state on degradation of environmental DNA in lake water. *Environ. Sci. Technol.* 50, 1859–1867.
- Emilson, C.E., Thompson, D.G., Venier, L.A., Porter, T.M., Swystun, T., Chartrand, D., Capell, S., Hajibabaei, M., 2017. DNA metabarcoding and morphological macroinvertebrate metrics reveal the same changes in boreal watersheds across an environmental gradient. *Sci. Rep.* 7, 12777.
- Epp, L.S., Boessenkool, S., Bellemain, E.P., Haile, J., Esposito, A., Riaz, T., Erősz, C., Gusarov, V.I., Edwards, M.E., Johnsen, A., Stenøien, H.K., Hassel, K., Kausrud, H., Yoccoz, N.G., Bråthen, K.A., Willerslev, E., Taberlet, P., Coissac, E., Brochmann, C., 2012. New environmental metabarcodes for analysing soil DNA: potential for studying past and present ecosystems. *Mol. Ecol.* 21, 1821–1833.
- Evans, D.M., Kitson, J.J.N., Lunt, D.H., Straw, N.A., Pocock, M.J.O., 2016a. Merging DNA metabarcoding and ecological network analysis to understand and build resilient terrestrial ecosystems. *Funct. Ecol.* 30, 1904–1916.
- Evans, N.T., Li, Y., Renshaw, M.A., Olds, B.P., Deiner, K., Turner, C.R., Jerde, C.L., Lodge, D.M., Lamberti, G.A., Pfrender, M.E., 2017. Fish community assessment with eDNA metabarcoding: effects of sampling design and bioinformatic filtering. *Can. J. Fish. Aquat. Sci.* 74, 1362–1374.
- Evans, N.T., Olds, B.P., Renshaw, M.A., Turner, C.R., Li, Y., Jerde, C.L., Mahon, A.R., Pfrender, M.E., Lamberti, G.A., Lodge, D.M., 2016b. Quantification of mesocosm fish and amphibian species diversity via environmental DNA metabarcoding. *Mol. Ecol. Res.* 16, 29–41.
- Fahner, N.A., Shokralla, S., Baird, D.J., Hajibabaei, M., 2016. Large-scale monitoring of plants through environmental DNA metabarcoding of soil: recovery, resolution, and annotation of four DNA markers. *PLoS One* 11, e0157505.
- Ficetola, G.F., Miaud, C., Pompanon, F., Taberlet, P., 2008. Species detection using environmental DNA from water samples. *Biol. Lett.* 4, 423–425.
- Ficetola, G.F., Pansu, J., Bonin, A., Coissac, E., Giguët-Covex, C., Barba, M.D., Gielly, L., Lopes, C.M., Boyer, F., Pompanon, F., Rayé, G., Taberlet, P., 2015. Replication levels, false presences and the estimation of the presence/absence from eDNA metabarcoding data. *Mol. Ecol. Res.* 15, 543–556.
- Ficetola, G.F., Taberlet, P., Coissac, E., 2016. How to limit false positives in environmental DNA and metabarcoding? *Mol. Ecol. Res.* 16, 604–607.
- Fletcher, L.M., Zaiko, A., Atalah, J., Richter, I., Dufour, C.M., Pochon, X., Wood, S.A., Hopkins, G.A., 2017. Bilge water as a vector for the spread of marine pests: a morphological, metabarcoding and experimental assessment. *Biol. Invasions* 19, 2851–2867.
- Fonseca, V.G., Carvalho, G.R., Sung, W., Johnson, H.F., Power, D.M., Neill, S.P., Packer, M., Blaxter, M.L., Lambshead, P.J.D., Thomas, W.K., Creer, S., 2010. Second-generation environmental sequencing unmasks marine metazoan biodiversity. *Nat. Commun.* 1, 98.
- Foulon, J., Zappellini, C., Durand, A., Valot, B., Girardclos, O., Blaudez, D., Chalot, M., 2016. Environmental metabarcoding reveals contrasting microbial communities at two poplar phytomanagement sites. *Sci. Total Environ.* 571, 1230–1240.
- Frontalini, F., Greco, M., Di Bella, L., Lejzerowicz, F., Reo, E., Caruso, A., Cosentino, C., Maccotta, A., Scopelliti, G., Nardelli, M.P., Losada, M.T., Armynot du Châtelet, E., Coccioni, R., Pawlowski, J., 2018. Assessing the effect of mercury pollution on cultured benthic foraminifera community using morphological and eDNA metabarcoding approaches. *Mar. Pollut. Bull.* 129, 512–524.
- Gibson, J.F., Shokralla, S., Curry, C., Baird, D.J., Monk, W.A., King, I., Hajibabaei, M., 2015. Large-scale biomonitoring of remote and threatened ecosystems via high-throughput sequencing. *PLoS One* 10, e0138432.
- Gibson, J., Shokralla, S., Porter, T.M., King, I., van Konyenburg, S., Janzen, D.H., Hallwachs, W., Hajibabaei, M., 2014. Simultaneous assessment of the macrobiome and microbiome in a bulk sample of tropical arthropods through DNA metasyntematics. *Proc. Natl. Acad. Sci. Unit. States Am.* 111, 8007–8012.
- Giguët-Covex, C., Pansu, J., Arnaud, F., Rey, P., Griggo, C., Gielly, L., Domaizon, I., Coissac, E., David, F., Choler, P., Poulenard, J., Taberlet, P., 2014. Long livestock farming history and human landscape shaping revealed by lake sediment DNA. *Nat. Commun.* 5, 3211.
- Goldberg, C.S., Turner, C.R., Deiner, K., Klymus, K.E., Thomsen, P.F., Murphy, D.A., Spear, S.F., McKee, A., Oyler-McCance, S.J., Cornman, R.S., Laramie, M.B., Mahon, A.R., Lance, R.F., Pilliod, D.S., Strickler, K.M., Waits, L.P., Fremier, A.K., Takahara, T., Herder, J.E., Taberlet, P., 2016. Critical considerations for the application of environmental DNA methods to detect aquatic species. *Meth. Ecol. Evolut.* 7, 1299–1307.
- Grealy, A.C., McDowell, M.C., Scofield, P., Murray, D.C., Fusco, D.A., Haile, J., Prideaux, G.J., Bunce, M., 2015. A critical evaluation of how ancient DNA bulk bone metabarcoding complements traditional morphological analysis of fossil assemblages. *Quat. Sci. Rev.* 128, 37–47.
- Guardiola, M., Uriz, M.J., Taberlet, P., Coissac, E., Wangenstein, O.S., Turon, X., 2015. Deep-sea, deep-sequencing: metabarcoding extracellular DNA from sediments of marine canyons. *PLoS One* 10, e0139633.

- Guardiola, M., Wangenstein, O.S., Taberlet, P., Coissac, E., Uriz, M.J., Turon, X., 2016. Spatio-temporal monitoring of deep-sea communities using metabarcoding of sediment DNA and RNA. *PeerJ* 4 e2807.
- Guillerault, N., Bouletreau, S., Iribar, A., Valentini, A., Santoul, F., 2017. Application of DNA metabarcoding on faeces to identify European catfish *Silurus glanis* diet. *J. Fish. Biol.* 90, 2214–2219.
- Gutiérrez-García, T.A., Vázquez-Domínguez, E., Arroyo-Cabrales, J., Kuch, M., Enk, J., King, C., Poinar, H.N., 2014. Ancient DNA and the tropics: a rodent's tale. *Biol. Lett.* 10.
- Hajibabaei, M., Spall, J.L., Shokralla, S., van Konynenburg, S., 2012. Assessing biodiversity of a freshwater benthic macroinvertebrate community through non-destructive environmental barcoding of DNA from preservative ethanol. *BMC Ecol.* 12, 28.
- Hänfling, B., Handley, L.L., Read, D.S., Hahn, C., Li, J., Nichols, P., Blackman, R.C., Oliver, A., Winfield, I.J., 2016. Environmental DNA metabarcoding of lake fish communities reflects long-term data from established survey methods. *Mol. Ecol.* 25, 3101–3119.
- Haouchar, D., Haile, J., McDowell, M.C., Murray, D.C., White, N.E., Allcock, R.J.N., Phillips, M.J., Prideaux, G.J., Bunce, M., 2014. Thorough assessment of DNA preservation from fossil bone and sediments excavated from a late Pleistocene–Holocene cave deposit on Kangaroo Island, South Australia. *Quat. Sci. Rev.* 84, 56–64.
- Harper, L.R., Handley, L.L., Hahn, C., Boonham, N., Rees, H.C., Gough, K.C., Lewis, E., Adams, I.P., Brotherton, P., Phillips, S., Hänfling, B., 2018. Needle in a haystack? A comparison of eDNA metabarcoding and targeted qPCR for detection of the great crested newt (*Triturus cristatus*). *Ecol. Evolut.* 8, 6330–6341.
- Hatzenbuehler, C., Kelly, J.R., Martinson, J., Okum, S., Pilgrim, E., 2017. Sensitivity and accuracy of high-throughput metabarcoding methods for early detection of invasive fish species. *Sci. Rep.* 7, 46393.
- Hawkins, J., de Vere, N., Griffith, A., Ford, C.R., Allainguillaume, J., Hegarty, M.J., Baillie, L., Adams-Groom, B., 2015. Using DNA metabarcoding to identify the floral composition of honey: a new tool for investigating honey bee foraging preferences. *PLoS One* 10, e0134735.
- Hering, D., Borja, A., Jones, J.L., Pont, D., Boets, P., Bouchez, A., Bruce, K., Drakare, S., Hänfling, B., Kahlert, M., Leese, F., Meissner, K., Mergen, P., Reyjol, Y., Segurado, P., Vogler, A., Kelly, M., 2018. Implementation options for DNA-based identification into ecological status assessment under the European Water Framework Directive. *Water Res.* 138, 192–205.
- Hiibert, F., Taberlet, P., Chave, J., Scotti-Saintagne, C., Sabatier, D., Richard-Hansen, C., 2013. Unveiling the diet of elusive rainforest herbivores in next generation sequencing era? The tapir as a case study. *PLoS One* 8, e60799.
- Holdaway, R., Wood, J., Landcare Research, Dickie, I., Orwin, K., Bellingham, P., Richardson, S., Lyver, P., Timoti, P., Buckley, T., 2017. Using DNA metabarcoding to assess New Zealand's terrestrial biodiversity. *N. Z. J. Ecol.* 41.
- Ishige, T., Miya, M., Ushio, M., Sado, T., Ushioda, M., Maebashi, K., Yonechi, R., Lagan, P., Matsubayashi, H., 2017. Tropical-forest mammals as detected by environmental DNA at natural saltlicks in Borneo. *Biol. Conserv.* 210, 281–285.
- Jakubaviciūtė, E., Bergström, U., Eklöf, J.S., Haenel, Q., Bourlat, S.J., 2017. DNA metabarcoding reveals diverse diet of the three-spined stickleback in a coastal ecosystem. *PLoS One* 12, e0186929.
- Ji, Y., Ashton, L., Pedley, S.M., Edwards, D.P., Tang, Y., Nakamura, A., Kitching, R., Dolman, P.M., Woodcock, P., Edwards, F.A., Larsen, T.H., Hsu, W.W., Benedick, S., Hamer, K.C., Wilcove, D.S., Bruce, C., Wang, X., Levi, T., Lott, M., Emerson, B.C., Yu, D.W., 2013. Reliable, verifiable and efficient monitoring of biodiversity via metabarcoding. *Ecol. Lett.* 16, 1245–1257.
- Jørgensen, T., Kjær, K.H., Haile, J., Rasmussen, M., Boessenkool, S., Andersen, K., Coissac, E., Taberlet, P., Brochmann, C., Orlando, L., Gilbert, M.T.P., Willerslev, E., 2012. Islands in the ice: detecting past vegetation on Greenlandic nunataks using historical records and sedimentary ancient DNA Metabarcoding. *Mol. Ecol.* 21, 1980–1988.
- Kartzinel, T.R., Chen, P.A., Coverdale, T.C., Erickson, D.L., Kress, W.J., Kuzmina, M.L., Rubenstein, D.I., Wang, W., Pringle, R.M., 2015. DNA metabarcoding illuminates dietary niche partitioning by African large herbivores. *Proc. Natl. Acad. Sci. Unit. States Am.* 112, 8019–8024.
- Keller, A., Danner, N., Grimmer, G., Ankenbrand, M., von der Ohe, K., von der Ohe, W., Rost, S., Härtel, S., Steffan-Dewenter, I., 2015. Evaluating multiplexed next-generation sequencing as a method in palynology for mixed pollen samples. *Plant Biol.* 17, 558–566.
- Kelly, R.P., Port, J.A., Yamahara, K.M., Crowder, L.B., 2014. Using environmental DNA to census marine fishes in a large mesocosm. *PLoS One* 9, e86175.
- Kermarrec, L., Franc, A., Rimet, F., Chaumeil, P., Frigerio, J., Humbert, J., Bouchez, A., 2014. A next-generation sequencing approach to river biomonitoring using benthic diatoms. *Freshw. Sci.* 33, 349–363.
- Klymus, K.E., Marshall, N.T., Stepien, C.A., 2017a. Environmental DNA (eDNA) metabarcoding assays to detect invasive invertebrate species in the Great Lakes. *PLoS One* 12, e0177643.
- Klymus, K.E., Richter, C.A., Thompson, N., Hinck, J.E., 2017b. Metabarcoding of environmental DNA samples to explore the use of uranium mine containment ponds as a water source for wildlife. *Diversity* 9, 54.
- Kowalczyk, R., Taberlet, P., Coissac, E., Valentini, A., Miquel, C., Kamiński, T., Wójcik, J.M., 2011. Influence of management practices on large herbivore diet—case of European bison in Białowieża primeval forest (Poland). *For. Ecol. Manag.* 261, 821–828.
- Kraaijeveld, K., de Weger, L.A., García, M.V., Buermans, H., Frank, J., Hiemstra, P.S., den Dunnen, J.T., 2015. Efficient and sensitive identification and quantification of airborne pollen using next-generation DNA sequencing. *Mol. Ecol. Res.* 15, 8–16.
- Lacoursière-Roussel, A., Dubois, Y., Normandeau, E., Bernatchez, L., 2016. Improving herpetological surveys in eastern North America using the environmental DNA method. *Genome* 59, 991–1007.
- Lallias, D., Hiddink, J.G., Fonseca, V.G., Gaspar, J.M., Sung, W., Neill, S.P., Barnes, N., Ferrero, T., Hall, N., Lamshead, P.J.D., Packer, M., Thomas, W.K., Creer, S., 2015. Environmental metabarcoding reveals heterogeneous drivers of microbial eukaryote diversity in contrasting estuarine ecosystems. *ISME J.* 9, 1208–1221.
- Lanzén, A., Lekang, K., Jonassen, I., Thompson, E.M., Troedsson, C., 2016. High-throughput metabarcoding of eukaryotic diversity for environmental monitoring of offshore oil-drilling activities. *Mol. Ecol.* 25, 4392–4406.
- Lanzén, A., Lekang, K., Jonassen, I., Thompson, E.M., Troedsson, C., 2017. DNA extraction replicates improve diversity and compositional dissimilarity in metabarcoding of eukaryotes in marine sediments. *PLoS One* 12, e0179443.
- Laroche, O., Wood, S.A., Tremblay, L.A., Ellis, J.L., Lear, G., Pochon, X., 2018. A cross-taxa study using environmental DNA/RNA metabarcoding to measure biological impacts of offshore oil and gas drilling and production operations. *Mar. Pollut. Bull.* 127, 97–107.
- Lee, P.-S., Gan, H.M., Clements, G.R., Wilson, J.-J., 2016. Field calibration of blowfly-derived DNA against traditional methods for assessing mammal diversity in tropical forests. *Genome* 59, 1008–1022.
- Leontidou, K., Vernesi, C., Groeve, J.D., Cristofolini, F., Vokou, D., Cristofori, A., 2018. DNA metabarcoding of airborne pollen: new protocols for improved taxonomic identification of environmental samples. *Aerobiologia* 34, 63–74.
- Leray, M., Knowlton, N., 2015. DNA barcoding and metabarcoding of standardized samples reveal patterns of marine benthic diversity. *Proc. Natl. Acad. Sci. U. S. A.* 112, 2076–2081.
- Li, W., Wang, M.M., Wang, X.G., Cheng, X.L., Guo, J.J., Bian, X.M., Cai, L., 2016. Fungal communities in sediments of subtropical Chinese seas as estimated by DNA metabarcoding. *Sci. Rep.* 6, 26528.
- Lim, N.K.M., Tay, Y.C., Srivathsan, A., Tan, J.W.T., Kwik, J.T.B., Baloğlu, B., Meier, R., Yeo, D.C.J., 2016. Next-generation freshwater bioassessment: eDNA metabarcoding with a conserved metazoan primer reveals species-rich and reservoir-specific communities. *R. Soc. Open Sci.* 3, 160635.
- Liu, Y., Fu, X., Mao, L., Xing, Z., Wu, K., 2017. Identification of host plant use of adults of a long-distance migratory insect, *Mythimna separata*. *PLoS One* 12, e0184116.
- Liu, L., Liu, M., Wilkinson, D.M., Chen, H., Yu, X., Yang, J., 2017. DNA metabarcoding reveals that 200-µm-size-fractionated filtering is unable to discriminate between planktonic microbial and large eukaryotes. *Mol. Ecol. Res.* 17, 991–1002.
- Lobo, J., Shokralla, S., Costa, M.H., Hajibabaei, M., Costa, F.O., 2017. DNA metabarcoding for high-throughput monitoring of estuarine macrobenthic communities. *Sci. Rep.* 7.

- Lopes, C.M., Sasso, T., Valentini, A., Dejean, T., Martins, M., Zamudio, K.R., Haddad, C.F.B., 2017. eDNA metabarcoding: a promising method for anuran surveys in highly diverse tropical forests. *Mol. Ecol. Res.* 17, 904–914.
- Lucas, A., Bodger, O., Brosi, B.J., Ford, C.R., Forman, D.W., Greig, C., Hegarty, M., Neyland, P.J., de Vere, N., 2018. Generalisation and specialisation in hoverfly (Syrphidae) grassland pollen transport networks revealed by DNA metabarcoding. *J. Anim. Ecol.* 87, 1008–1021.
- Maggia, M.E., Vigouroux, Y., Renno, J.F., Duponchelle, F., Desmarais, E., Nunez, J., García-Dávila, C., Carvajal-Vallejos, F.M., Paradis, E., Martin, J.F., Mariac, C., 2017. DNA metabarcoding of Amazonian ichthyoplankton swarms. *PLoS One* 12, e0170009.
- Martínez-García, L.B., Richardson, S.J., Tylianakis, J.M., Peltzer, D.A., Dickie, I.A., 2015. Host identity is a dominant driver of mycorrhizal fungal community composition during ecosystem development. *New Phytol.* 205, 1565–1576.
- Massana, R., Gobet, A., Audic, S., Bass, D., Bittner, L., Boutte, C., Chambouvet, A., Christen, R., Claverie, J.-M., Decelle, J., Dolan, J.R., Dunthorn, M., Edvardsen, B., Forn, I., Forster, D., Guillou, L., Jaillon, O., Kooistra, W.H.C.F., Logares, R., Mahé, F., Not, F., Ogata, H., Pawlowski, J., Pernice, M.C., Probert, I., Romain, S., Richards, T., Santini, S., Shalchian-Tabrizi, K., Siano, R., Simon, N., Stoeck, T., Vaulot, D., Zingone, A., de Vargas, C., 2015. Marine protist diversity in European coastal waters and sediments as revealed by high-throughput sequencing. *Environ. Microbiol.* 17, 4035–4049.
- McInnes, J.C., Alderman, R., Deagle, B.E., Lea, M.-A., Raymond, B., Jarman, S.N., 2017a. Optimised scat collection protocols for dietary DNA metabarcoding in vertebrates. *Meth. Ecol. Evolut.* 8, 192–202.
- McInnes, J.C., Alderman, R., Lea, M.-A., Raymond, B., Deagle, B.E., Phillips, R.A., Stanworth, A., Thompson, D.R., Catry, P., Weimerskirch, H., Suazo, C.G., Gras, M., Jarman, S.N., 2017b. High occurrence of jellyfish predation by black-browed and Campbell albatross identified by DNA metabarcoding. *Mol. Ecol.* 26, 4831–4845.
- Miller, K.E., Hopkins, K., Inward, D.J.G., Vogler, A.P., 2016. Metabarcoding of fungal communities associated with bark beetles. *Ecol. Evolut.* 6, 1590–1600.
- Miya, M., Sato, Y., Fukunaga, T., Sado, T., Poulsen, J.Y., Sato, K., Minamoto, T., Yamamoto, S., Yamanaka, H., Araki, H., Kondoh, M., Iwasaki, W., 2015. MiFish, a set of universal PCR primers for metabarcoding environmental DNA from fishes: detection of more than 230 subtropical marine species. *R. Soc. Open Sci.* 2, 150088.
- Nakagawa, H., Yamamoto, S., Sato, Y., Sado, T., Minamoto, T., Miya, M., 2018. Comparing local- and regional-scale estimations of the diversity of stream fish using eDNA metabarcoding and conventional observation methods. *Freshw. Biol.* 63, 569–580.
- Oh, S., Fong, J.J., Park, M.S., Chang, L., Lim, Y.W., 2014. Identifying airborne fungi in Seoul, Korea using metagenomics. *J. Microbiol.* 52, 465–472.
- Olds, B.P., Jerde, C.L., Renshaw, M.A., Li, Y., Evans, N.T., Turner, C.R., Deiner, K., Mahon, A.R., Brueske, M.A., Shirey, P.D., Pfrender, M.E., Lodge, D.M., Lamberti, G.A., 2016. Estimating species richness using environmental DNA. *Ecol. Evolut.* 6, 4214–4226.
- Pansu, J., Giguet-Coxe, C., Ficetola, G.F., Gielly, L., Boyer, F., Zinger, L., Arnaud, F., Poulenard, J., Taberlet, P., Choler, P., 2015. Reconstructing long-term human impacts on plant communities: an ecological approach based on lake sediment DNA. *Mol. Ecol.* 24, 1485–1498.
- Pawlowski, J., Christen, R., Lecroq, B., Bachar, D., Shahbazkia, H.R., Amaral-Zettler, L., Guillou, L., 2011. Eukaryotic richness in the abyss: insights from pyrotag sequencing. *PLoS One* 6, e18169.
- Pawlowski, J., Esling, P., Lejzerowicz, F., Cedhagen, T., Wilding, T.A., 2014. Environmental monitoring through protist next-generation sequencing metabarcoding: assessing the impact of fish farming on benthic foraminifera communities. *Mol. Ecol. Res.* 14, 1129–1140.
- Pawlowski, J., Kelly-Quinn, M., Altermatt, F., Apotheloz-Perret-Gentil, L., Beja, P., Boggero, A., Borja, A., Bouchez, A., Cordier, T., Domaizon, I., Feio, M.J., Filipe, A.F., Fornari, R., Graf, W., Herder, J., van der Hoorn, B., Iwan Jones, J., Sagova-Marekova, M., Moritz, C., Barquin, J., Piggott, J.J., Pinna, M., Rimet, F., Rinkevich, B., Sousa-Santos, C., Specchia, V., Trobajo, R., Vasselton, V., Vitecek, S., Zimmerman, J., Weigand, A., Leese, F., Kahlert, M., 2018. The future of biotic indices in the ecogenomic era: integrating (e)DNA metabarcoding in biological assessment of aquatic ecosystems. *Sci. Total Environ.* 637–638, 1295–1310.
- Pawlowski, J., Lejzerowicz, F., Apotheloz-Perret-Gentil, L., Visco, J., Esling, P., 2016. Protist metabarcoding and environmental biomonitoring: time for change. *Eur. J. Protistol.* 55, 12–25.
- Pedersen, M.W., Ginolhac, A., Orlando, L., Olsen, J., Andersen, K., Holm, J., Funder, S., Willerslev, E., Kjær, K.H., 2013. A comparative study of ancient environmental DNA to pollen and macrofossils from lake sediments reveals taxonomic overlap and additional plant taxa. *Quat. Sci. Rev.* 75, 161–168.
- Pilliod, D.S., Goldberg, C.S., Arkle, R.S., Waits, L.P., 2013. Estimating occupancy and abundance of stream amphibians using environmental DNA from filtered water samples. *Can. J. Fish. Aquat. Sci.* 70, 1123–1130.
- Piñol, J., Andrés, V.S., Clare, E.L., Mir, G., Symondson, W.O.C., 2014. A pragmatic approach to the analysis of diets of generalist predators: the use of next-generation sequencing with no blocking probes. *Mol. Ecol. Res.* 14, 18–26.
- Pochon, X., Bott, N.J., Smith, K.F., Wood, S.A., 2013. Evaluating detection limits of next-generation sequencing for the surveillance and monitoring of international marine pests. *PLoS One* 8, e73935.
- Pochon, X., Wood, S.A., Keeley, N.B., Lejzerowicz, F., Esling, P., Drew, J., Pawlowski, J., 2015. Accurate assessment of the impact of salmon farming on benthic sediment enrichment using foraminiferal metabarcoding. *Mar. Pollut. Bull.* 100, 370–382.
- Pochon, X., Zaiko, A., Fletcher, L.M., Laroche, O., Wood, S.A., 2017. Wanted dead or alive? Using metabarcoding of environmental DNA and RNA to distinguish living assemblages for biosecurity applications. *PLoS One* 12, e0187636.
- Pornon, A., Andalo, C., Burrus, M., Escaravage, N., 2017. DNA metabarcoding data unveils invisible pollination networks. *Sci. Rep.* 7, 16828.
- Pornon, A., Escaravage, N., Burrus, M., Holota, H., Khimoun, A., Mariette, J., Pellizzari, C., Iribar, A., Etienne, R., Taberlet, P., Vidal, M., Winterton, P., Zinger, L., Andalo, C., 2016. Using metabarcoding to reveal and quantify plant-pollinator interactions. *Sci. Rep.* 6, 27282.
- Port, J.A., O'Donnell, J.L., Romero-Maraccini, O.C., Leary, P.R., Litvin, S.Y., Nickols, K.J., Yamahara, K.M., Kelly, R.P., 2016. Assessing vertebrate biodiversity in a kelp forest ecosystem using environmental DNA. *Mol. Ecol.* 25, 527–541.
- Quéméré, E., Hibert, F., Miquel, C., Lhuillier, E., Rasolondraibe, E., Champeau, J., Rabarivola, C., Nusbaumer, L., Chatelain, C., Gautier, L., Ranirison, P., Crouau-Roy, B., Taberlet, P., Chikhi, L., 2013. A DNA metabarcoding study of a primate dietary diversity and plasticity across its entire fragmented range. *PLoS One* 8, e58971.
- Rawlence, N.J., Lowe, D.J., Wood, J.R., Young, J.M., Churchman, G.J., Huang, Y., Cooper, A., 2014. Using palaeoenvironmental DNA to reconstruct past environments: progress and prospects. *J. Quat. Sci.* 29, 610–626.
- Richardson, R.T., Lin, C., Quijia, J.O., Riusech, N.S., Goodell, K., Johnson, R.M., 2015a. Rank-based characterization of pollen assemblages collected by honey bees using a multi-locus metabarcoding approach. *Appl. Plant Sci.* 3.
- Richardson, R.T., Lin, C., Sponsler, D.B., Quijia, J.O., Goodell, K., Johnson, R.M., 2015b. Application of ITS2 metabarcoding to determine the provenance of pollen collected by honey bees in an agroecosystem. *Appl. Plant Sci.* 3, 1400066.
- Rivera, S.F., Vasselton, V., Ballorain, K., Carpentier, A., Wetzel, C.E., Ector, L., Bouchez, A., Rimet, F., 2018a. DNA metabarcoding and microscopic analyses of sea turtles biofilms: complementary to understand turtle behavior. *PLoS One* 13, e0195770.
- Rivera, S.F., Vasselton, V., Jacquet, S., Bouchez, A., Ariztegui, D., Rimet, F., 2018b. Metabarcoding of lake benthic diatoms: from structure assemblages to ecological assessment. *Hydrobiologia* 807, 37–51.
- Rodgers, T.W., Xu, C.C.Y., Giacalone, J., Kapheim, K.M., Saltonstall, K., Vargas, M., Yu, D.W., Somervuo, P., McMillan, W.O., Jansen, P.A., 2017. Carrion fly-derived DNA metabarcoding is an effective tool for mammal surveys: evidence from a known tropical mammal community. *Mol. Ecol. Res.* 17, e133–e145.
- Sato, H., Sogo, Y., Doi, H., Yamanaka, H., 2017. Usefulness and limitations of sample pooling for environmental DNA metabarcoding of freshwater fish communities. *Sci. Rep.* 7, 14860.
- Schubert, G., Stockhausen, M., Hoffmann, C., Merkel, K., Vigilant, L., Leendertz, F.H., Calvignac-Spencer, S., 2015. Targeted detection of mammalian species using carrion fly-derived DNA. *Mol. Ecol. Res.* 15, 285–294.
- Shaw, J.L.A., Clarke, L.J., Wedderburn, S.D., Barnes, T.C., Weyrich, L.S., Cooper, A., 2016. Comparison of environmental DNA metabarcoding and conventional fish survey methods in a river system. *Biol. Conserv.* 197, 131–138.
- Shehzad, W., McCarthy, T.M., Pompanon, F., Purejav, L., Coissac, E., Riaz, T., Taberlet, P., 2012a. Prey preference of snow leopard (*Panthera uncia*) in south gobi, Mongolia. *PLoS One* 7, e32104.

- Shehzad, W., Riaz, T., Nawaz, M.A., Miquel, C., Poillot, C., Shah, S.A., Pompanon, F., Coissac, E., Taberlet, P., 2012b. Carnivore diet analysis based on next-generation sequencing: application to the leopard cat (*Prionailurus bengalensis*) in Pakistan. *Mol. Ecol.* 21, 1951–1965.
- Sickel, W., Ankenbrand, M.J., Grimmer, G., Holzschuh, A., Härtel, S., Lanzen, J., Steffan-Dewenter, I., Keller, A., 2015. Increased efficiency in identifying mixed pollen samples by meta-barcoding with a dual-indexing approach. *BMC Ecol.* 15, 20.
- Sigut, M., Kostovčík, M., Šigutová, H., Hulcr, J., Drozd, P., Hřeček, J., 2017. Performance of DNA metabarcoding, standard barcoding, and morphological approach in the identification of host–parasitoid interactions. *PLoS One* 12, e0187803.
- Sinniger, F., Pawlowski, J., Harii, S., Gooday, A.J., Yamamoto, H., Chevaldonné, P., Cedhagen, T., Carvalho, G., Creer, S., 2016. Worldwide analysis of sedimentary DNA reveals major gaps in taxonomic knowledge of deep-sea benthos. *Front. Mar. Sci.* 3.
- Smith, M.B., Rocha, A.M., Smillie, C.S., Olesen, S.W., Paradis, C., Wu, L., Campbell, J.H., Fortney, J.L., Mehlhorn, T.L., Lowe, K.A., Earles, J.E., Phillips, J., Techtman, S.M., Joyner, D.C., Elias, D.A., Bailey, K.L., Hurt, R.A., Preheim, S.P., Sanders, M.C., Yang, J., Mueller, M.A., Brooks, S., Watson, D.B., Zhang, P., He, Z., Dubinsky, E.A., Adams, P.D., Arkin, A.P., Fields, M.W., Zhou, J., Alm, E.J., Hazen, T.C., 2015. Natural bacterial communities serve as quantitative geochemical biosensors. *mBio* 6 e00326–15.
- Sohlberg, E., Bomberg, M., Miettinen, H., Nyyssönen, M., Salavirta, H., Vikman, M., Itävaara, M., 2015. Revealing the unexplored fungal communities in deep groundwater of crystalline bedrock fracture zones in Olkiluoto, Finland. *Front. Microbiol.* 6.
- Soininen, E.M., Ehrlich, N., Lecomte, N., Yoccoz, N.G., Tarroux, A., Berteaux, D., Gauthier, G., Gielly, L., Brochmann, C., Gussarova, G., Ims, R.A., 2014. Sources of variation in small rodent trophic niche: new insights from DNA metabarcoding and stable isotope analysis. *Isot. Environ. Health Stud.* 50, 361–381.
- Soininen, E.M., Gauthier, G., Bilodeau, F., Berteaux, D., Gielly, L., Taberlet, P., Gussarova, G., Bellemain, E., Hassel, K., Stenøien, H.K., Epp, L., Schröder-Nielsen, A., Brochmann, C., Yoccoz, N.G., 2015. Highly overlapping winter diet in two sympatric lemming species revealed by DNA metabarcoding. *PLoS One* 10, e0115335.
- Soininen, E.M., Zinger, L., Gielly, L., Bellemain, E., Bräthen, K.A., Brochmann, C., Epp, L.S., Gussarova, G., Hassel, K., Henden, J., Killengreen, S.T., Rämä, T., Stenøien, H.K., Yoccoz, N.G., Ims, R.A., 2013. Shedding new light on the diet of Norwegian lemmings: DNA metabarcoding of stomach content. *Polar Biol.* 36, 1069–1076.
- Sønstebo, J.H., Gielly, L., Brysting, A.K., Elven, R., Edwards, M., Haile, J., Willerslev, E., Coissac, E., Rioux, D., Sannier, J., Taberlet, P., Brochmann, C., 2010. Using next-generation sequencing for molecular reconstruction of past Arctic vegetation and climate. *Mol. Ecol. Res.* 10, 1009–1018.
- Stat, M., Huggett, M.J., Bernasconi, R., DiBattista, J.D., Berry, T.E., Newman, J.P., Harvey, E.S., Bunce, M., 2017. Ecosystem biomonitoring with eDNA: metabarcoding across the tree of life in a tropical marine environment. *Sci. Rep.* 7, 12240.
- Stoeck, T., Frühe, L., Forster, D., Cordier, T., Martins, C.L.M., Pawlowski, J., 2018a. Environmental DNA metabarcoding of benthic bacterial communities indicates the benthic footprint of salmon aquaculture. *Mar. Pollut. Bull.* 127, 139–149.
- Stoeck, T., Kochems, R., Forster, D., Lejzerowicz, F., Pawlowski, J., 2018b. Metabarcoding of benthic ciliate communities shows high potential for environmental monitoring in salmon aquaculture. *Ecol. Indic.* 85, 153–164.
- Strickler, K.M., Fremier, A.K., Goldberg, C.S., 2015. Quantifying effects of UV-B, temperature, and pH on eDNA degradation in aquatic microcosms. *Biol. Conserv.* 183, 85–92.
- Štursová, M., Šnajdr, J., Cajthaml, T., Bárta, J., Šantrůčková, H., Baldrian, P., 2014. When the forest dies: the response of forest soil fungi to a bark beetle-induced tree dieback. *ISME J.* 8, 1920–1931.
- Suchan, T., Talavera, G., Sáez, L., Ronikier, M., Vila, R., 2018. Pollen metabarcoding as a tool for tracking long-distance insect migrations. *Mol. Ecol. Res.* 0, 2045–2050.
- Taberlet, P., Coissac, E., Pompanon, F., Brochmann, C., Willerslev, E., 2012a. Towards next-generation biodiversity assessment using DNA metabarcoding. *Mol. Ecol.* 21, 2045–2050.
- Taberlet, P., Prud'homme, S.M., Campione, E., Roy, J., Miquel, C., Shehzad, W., Gielly, L., Rioux, D., Choler, P., Clément, J., Melodelima, C., Pompanon, F., Coissac, E., 2012b. Soil sampling and isolation of extracellular DNA from large amount of starting material suitable for metabarcoding studies. *Mol. Ecol.* 21, 1816–1820.
- Thomsen, P.F., Kielgast, J., Iversen, L.L., Møller, P.R., Rasmussen, M., Willerslev, E., 2012. Detection of a diverse marine fish fauna using environmental DNA from seawater samples. *PLoS One* 7, e41732.
- Thomsen, P.F., Willerslev, E., 2015. Environmental DNA – an emerging tool in conservation for monitoring past and present biodiversity. *Biol. Conserv.* 183, 4–18.
- Tong, X., Xu, H., Zou, L., Cai, M., Xu, X., Zhao, Z., Xiao, F., Li, Y., 2017. High diversity of airborne fungi in the hospital environment as revealed by meta-sequencing-based microbiome analysis. *Sci. Rep.* 7, 39606.
- Tsuji, S., Ushio, M., Sakurai, S., Minamoto, T., Yamanaka, H., 2017. Water temperature-dependent degradation of environmental DNA and its relation to bacterial abundance. *PLoS One* 12, e0176608.
- Ushio, M., Fukuda, H., Inoue, T., Makoto, K., Kishida, O., Sato, K., Murata, K., Nikaido, M., Sado, T., Sato, Y., Takeshita, M., Iwasaki, W., Yamanaka, H., Kondoh, M., Miya, M., 2017. Environmental DNA enables detection of terrestrial mammals from forest pond water. *Mol. Ecol. Resour.* 17, e63–e75.
- Ushio, M., Murata, K., Sado, T., Nishiumi, I., Takeshita, M., wasaki, W., Miya, M., 2018. Demonstration of the potential of environmental DNA as a tool for the detection of avian species. *Sci. Rep.* 8, 4493.
- Valentini, A., Miquel, C., Nawaz, M.A., Bellemain, E., Coissac, E., Pompanon, F., Gielly, L., Cruaud, C., Nascetti, G., Wincker, P., Swenson, J.E., Taberlet, P., 2009. New perspectives in diet analysis based on DNA barcoding and parallel pyrosequencing: the *tmL* approach. *Mol. Ecol. Res.* 9, 51–60.
- Valentini, A., Miquel, C., Taberlet, P., 2010. DNA Barcoding for honey biodiversity. *Diversity* 2, 610–617.
- Valentini, A., Taberlet, P., Miaud, C., Civate, R., Herder, J., Thomsen, P.F., Bellemain, E., Besnard, A., Coissac, E., Boyer, F., Gaboriaud, C., Jean, P., Poulet, N., Roset, N., Copp, G.H., Geniez, P., Pont, D., Argillier, C., Baudoin, J., Peroux, T., Crivelli, A.J., Olivier, A., Acqueberge, M., Brun, M.L., Møller, P.R., Willerslev, E., Dejean, T., 2016. Next-generation monitoring of aquatic biodiversity using environmental DNA metabarcoding. *Mol. Ecol.* 25, 929–942.
- Vasselon, V., Rimet, F., Tapolczai, K., Bouchez, A., 2017. Assessing ecological status with diatoms DNA metabarcoding: Scaling-up on a WFD monitoring network (Mayotte island, France). *Ecol. Indic.* 82, 1–12.
- Vestheim, H., Jarman, S.N., 2008. Blocking primers to enhance PCR amplification of rare sequences in mixed samples – a case study on prey DNA in Antarctic krill stomachs. *Front. Zool.* 5, 12.
- Wangensteen, O.S., Cebrian, E., Palacín, C., Turon, X., 2018a. Under the canopy: community-wide effects of invasive algae in Marine Protected Areas revealed by metabarcoding. *Mar. Pollut. Bull.* 127, 54–66.
- Wangensteen, O.S., Palacín, C., Guardiola, M., Turon, X., 2018b. DNA metabarcoding of littoral hard-bottom communities: high diversity and database gaps revealed by two molecular markers. *PeerJ* 6 e4705.
- Willerslev, E., Davison, J., Moora, M., Zobel, M., Coissac, E., Edwards, M.E., Lorenzen, E.D., Vestergård, M., Gussarova, G., Haile, J., Craine, J., Gielly, L., Boessenkool, S., Epp, L.S., Pearman, P.B., Cheddadi, R., Murray, D., Bräthen, K.A., Yoccoz, N., Binney, H., Cruaud, C., Wincker, P., Goslar, T., Alsos, I.G., Bellemain, E., Brysting, A.K., Elven, R., Sønstebo, J.H., Murton, J., Sher, A., Rasmussen, M., Rønn, R., Mourier, T., Cooper, A., Austin, J., Möller, P., Froese, D., Zazula, G., Pompanon, F., Rioux, D., Niderkorn, V., Tikhonov, A., Savvinov, G., Roberts, R.G., MacPhee, R.D.E., Gilbert, M.T.P., Kjær, K.H., Orlando, L., Brochmann, C., Taberlet, P., 2014. Fifty thousand years of Arctic vegetation and megafaunal diet. *Nature* 506, 47–51.
- Willerslev, E., Hansen, A.J., Binladen, J., Brand, T.B., Gilbert, M.T.P., Shapiro, B., Bunce, M., Wiuf, C., Gilichinsky, D.A., Cooper, A., 2003. Diverse plant and animal genetic records from Holocene and Pleistocene sediments. *Science* 300, 791–795.
- Wood, J.R., Wilmshurst, J.M., Wagstaff, S.J., Worthy, T.H., Rawlence, N.J., Cooper, A., 2012. High-resolution coproecology: using coprolites to reconstruct the habits and habitats of New Zealand's extinct Upland Moa (*Megapteryx didinus*). *PLoS One* 7, e40025.
- Wurzbacher, C., Attermeyer, K., Kettner, M.T., Flintrop, C., Warthmann, N., Hilt, S., Grossart, H., Monaghan, M.T., 2017. DNA metabarcoding of unfractionated water samples relates phyto-, zoo- and bacterioplankton dynamics and reveals a single-taxon bacterial bloom. *Environ. Microbiol. Rep.* 9, 383–388.
- Xie, Y., Wang, J., Yang, J., Giesy, J.P., Yu, H., Zhang, X., 2017. Environmental DNA metabarcoding reveals primary chemical contaminants in freshwater sediments from different land-use types. *Chemosphere* 172, 201–209.

- Yamamoto, S., Masuda, R., Sato, Y., Sado, T., Araki, H., Kondoh, M., Minamoto, T., Miya, M., 2017. Environmental DNA metabarcoding reveals local fish communities in a species-rich coastal sea. *Sci. Rep.* 7, 40368.
- Yang, C., Wang, X., Miller, J.A., de Blécourt, M., Ji, Y., Yang, C., Harrison, R.D., Yu, D.W., 2014. Using metabarcoding to ask if easily collected soil and leaf-litter samples can be used as a general biodiversity indicator. *Ecol. Indic.* 46, 379–389.
- Yang, J., Zhang, X., Xie, Y., Song, C., Zhang, Y., Yu, H., Burton, G.A., 2017. Zooplankton community profiling in a eutrophic freshwater ecosystem-lake Tai basin by DNA metabarcoding. *Sci. Rep.* 7, 1773.
- Yoccoz, N.G., Bråthen, K.A., Gielly, L., Haile, J., Edwards, M.E., Goslar, T., Stedingk, H.V., Brysting, A.K., Coissac, E., Pompanon, F., Sønstebo, J.H., Miquel, C., Valentini, A., Bello, F.D., Chave, J., Thuiller, W., Wincker, P., Cruaud, C., Gavory, F., Rasmussen, M., Gilbert, M.T.P., Orlando, L., Brochmann, C., Willerslev, E., Taberlet, P., 2012. DNA from soil mirrors plant taxonomic and growth form diversity. *Mol. Ecol.* 21, 3647–3655.
- Yoon, T., Kang, H., Lee, S.R., Lee, J., Baeck, G.W., Park, H., Kim, H., 2017. Metabarcoding analysis of the stomach contents of the antarctic toothfish (*Dissostichus mawsoni*) collected in the antarctic ocean. *PeerJ* 5.
- Yu, D.W., Ji, Y., Emerson, B.C., Wang, X., Ye, C., Yang, C., Ding, Z., 2012. Biodiversity soup: metabarcoding of arthropods for rapid biodiversity assessment and biomonitoring. *Meth. Ecol. Evolut.* 3, 613–623.
- Zaiko, A., Martinez, J.L., Schmidt-Petersen, J., Ribicic, D., Samuiloviene, A., Garcia-Vazquez, E., 2015a. Metabarcoding approach for the ballast water surveillance – an advantageous solution or an awkward challenge? *Mar. Pollut. Bull.* 92, 25–34.
- Zaiko, A., Samuiloviene, A., Ardura, A., Garcia-Vazquez, E., 2015b. Metabarcoding approach for nonindigenous species surveillance in marine coastal waters. *Mar. Pollut. Bull.* 100, 53–59.
- Zaiko, A., Schimanski, K., Pochon, X., Hopkins, G.A., Goldstien, S., Floerl, O., Wood, S.A., 2016. Metabarcoding improves detection of eukaryotes from early biofouling communities: implications for pest monitoring and pathway management. *Biofouling* 32, 671–684.
- Zappellini, C., Karimi, B., Foulon, J., Lacercat-Didier, L., Maillard, F., Valot, B., Blaudez, D., Cazaux, D., Gilbert, D., Yergeau, E., Greer, C., Chalot, M., 2015. Diversity and complexity of microbial communities from a chlor-alkali tailings dump. *Soil Biol. Biochem.* 90, 101–110.
- Zimmermann, J., Glöckner, G., Jahn, R., Enke, N., Gemeinholzer, B., 2015. Metabarcoding vs. morphological identification to assess diatom diversity in environmental studies. *Mol. Ecol. Res.* 15, 526–542.