Fiber_Comparison

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```
Load in ParathaakSGB table
parathaa_ksgb_assignments <- read.table("~/Repos/Parathaa2_OP3/Datasets/fiber_amplicons_true/taxonomic_</pre>
parathaa_abundance_table <- read.table("~/Repos/Parathaa2_0P3/Datasets/fiber_amplicons_true/taxonomic_a
colnames(parathaa_abundance_table)[1] <- "query.name"</pre>
parathaa_abundance_table[,-1] <- sweep(parathaa_abundance_table[,-1], 2, colSums(parathaa_abundance_tab
parathaa_abundance_table_kSGB <- full_join(parathaa_ksgb_assignments, parathaa_abundance_table, by="que
#remove control samples
parathaa_abundance_table_kSGB <- parathaa_abundance_table_kSGB[,-c(256:263, 10)]</pre>
Load in MetaPhlAn4 table
m4_assignments <- read.table("~/Repos/Parathaa2_0P3/Datasets/fiber_amplicons_true/taxonomic_assignments
colnames(m4_assignments) <- gsub("_.*", "", colnames(m4_assignments))</pre>
indexs <- match(colnames(parathaa_abundance_table), colnames(m4_assignments))</pre>
##ignore NAs for the moment..
indexs <- indexs[!is.na(indexs)]</pre>
m4_filt <- m4_assignments[,c(1,indexs)]</pre>
m4_filt <- m4_filt[-which(rowSums(m4_filt[,-1])==0),]</pre>
spec_index <- grep("s__", m4_filt$X..taxonomy)</pre>
SGB_index <- grep("t__", m4_filt$X..taxonomy)</pre>
only_spec <- setdiff(spec_index, SGB_index)</pre>
m4_filt$Species <- gsub(".*s__", "", m4_filt$X..taxonomy)</pre>
m4_filt$Species <- gsub("\\|.*", "", m4_filt$Species)</pre>
m4_filt$Genus <- gsub(".*g__", "", m4_filt$X..taxonomy)</pre>
m4_filt$Genus <- gsub("\\|.*", "", m4_filt$Genus)</pre>
m4_filt$Family <- gsub(".*f__", "", m4_filt$X..taxonomy)</pre>
```

m4_filt\$Family <- gsub("\\|.*", "", m4_filt\$Family)</pre>

```
m4_filt$Order <- gsub(".*o__", "", m4_filt$X..taxonomy)</pre>
m4_filt$Order <- gsub("\\|.*", "", m4_filt$Order)</pre>
m4_filt$Class <- gsub(".*c__", "", m4_filt$X..taxonomy)</pre>
m4_filt$Class <- gsub("\\|.*", "", m4_filt$Class)</pre>
m4_filt$Phylum <- gsub(".*p__", "", m4_filt$X..taxonomy)</pre>
m4_filt$Phylum <- gsub("\\|.*", "", m4_filt$Phylum)</pre>
m4_filt_spec <- m4_filt[c(3,only_spec),]</pre>
Load in Parathaa SILVA.SEED
parathaa_seed_assignments <- read.table("~/Repos/Parathaa2_OP3/SILVA_run_V3V4/Fiber_assignments/taxonom
parathaa_abundance_table_seed <- full_join(parathaa_seed_assignments, parathaa_abundance_table, by="que
parathaa_abundance_table_seed <- parathaa_abundance_table_seed[,-c(1, 256:263)]
Load in Dada2 assignments
DADA2_assignments <- read.table("~/Repos/Parathaa2_OP3/Datasets/fiber_amplicons_true/taxonomic_assignments
colnames(DADA2_assignments)[8] <- "query.name"</pre>
dada2_abundance_table <- full_join(DADA2_assignments, parathaa_abundance_table, by="query.name")
dada2_abundance_table <- dada2_abundance_table[,-c(1, 256:263)]</pre>
Load in Dada2 seed assignments
DADA2_assignments_seed <- read.table("~/Repos/Parathaa2_0P3/Datasets/fiber_amplicons_true/taxonomic_ass
colnames(DADA2_assignments_seed)[8] <- "query.name"</pre>
dada2_seed_abundance_table <- full_join(DADA2_assignments_seed, parathaa_abundance_table, by="query.nam"
dada2_seed_abundance_table <- dada2_seed_abundance_table[,-c(1,256:263)]</pre>
```

Number of assignments

Functions

```
tax_levels <- c("Species", "Genus", "Family", "Order", "Class", "Phylum")
calculate_assignments <- function(tax_assignments){

ret_frame <- data.frame(matrix(nrow=12, ncol=1))
    multi_names <- paste("multi", tax_levels, sep=" ")
    rownames(ret_frame) <- c(tax_levels, multi_names)
    colnames(ret_frame) <- "Number of Assignments"

for(i in 1:length(tax_levels)){
    ret_frame[tax_levels[i],1] <- length(which(!is.na(tax_assignments[,tax_levels[i]])))
    ret_frame[multi_names[i],1] <- length(which(grepl(";", tax_assignments[,tax_levels[i]])))
    }
    return(ret_frame)
}</pre>
```

	Number of Assignments
Species	285
Genus	1437
Family	2630
Order	3021
Class	3486
Phylum	3628
multi Species	89
multi Genus	116
multi Family	107
multi Order	74
multi Class	72
multi Phylum	35

	Number of Assignments
Species	193
Genus	1704
Family	2929
Order	3498
Class	3727
Phylum	3885
multi Species	58
multi Genus	64
multi Family	49
multi Order	17
multi Class	3
multi Phylum	2

Parathaa kSGB

Assignment_levels_kSGB <- calculate_assignments(parathaa_ksgb_assignments)
kable_styling(kable((Assignment_levels_kSGB)))</pre>

Parathaa SEED

Assignment_levels_SEED <- calculate_assignments(parathaa_seed_assignments)
kable_styling(kable(Assignment_levels_SEED))</pre>

Dada2

Assignment_levels_DADA2 <- calculate_assignments(DADA2_assignments)
kable_styling(kable(Assignment_levels_DADA2))</pre>

Dada2 seed

Assignment_levels_DADA2_seed <- calculate_assignments(DADA2_assignments_seed)
kable_styling(kable(Assignment_levels_DADA2_seed))</pre>

Yikes these are a bit concerning...

NT 1 CA .
Number of Assignments
176
2835
3664
3824
3930
3961
0
0
0
0
0
0

	Number of Assignments		
Species	48		
Genus	2467		
Family	3243		
Order	3494		
Class	3728		
Phylum	3872		
multi Species	0		
multi Genus	0		
multi Family	0		
multi Order	0		
multi Class	0		
multi Phylum	0		

	# of Assign. kSGB	# of Assign. SEED	# of Assign. DADA2	# of Assign. DADA2.Seed
Species	285	193	176	48
Genus	1437	1704	2835	2467
Family	2630	2929	3664	3243
Order	3021	3498	3824	3494
Class	3486	3727	3930	3728
Phylum	3628	3885	3961	3872
multi Species	89	58	0	0
multi Genus	116	64	0	0
multi Family	107	49	0	0
multi Order	74	17	0	0
multi Class	72	3	0	0
multi Phylum	35	2	0	0

Combined table

Composition Analysis

```
generate_tax_level_table <- function(table, level, prop=1){</pre>
  res_list <- list()</pre>
  Other <- c()
  for(i in unique(table[,level])){
    tmp_df <- table[which(table[,level]==i),]</pre>
    tmp_df <- tmp_df %>% select_if(is.numeric)
    tmp <- colSums(tmp_df)</pre>
    if(mean(tmp) < prop){</pre>
      if(length(Other)==0){
        Other <- tmp
      }else{
      Other <- Other + tmp
    }else
      res_list[[i]] <- tmp</pre>
  res_list[["Other"]] <- Other</pre>
  res_df <- data.frame(do.call(rbind, res_list))</pre>
  if("maxDist" %in% colnames(res df)){
      res_df <- res_df[,-which(colnames(res_df) == "maxDist")]</pre>
  res_df$Taxon <- rownames(res_df)</pre>
  return(res_df)
```

}

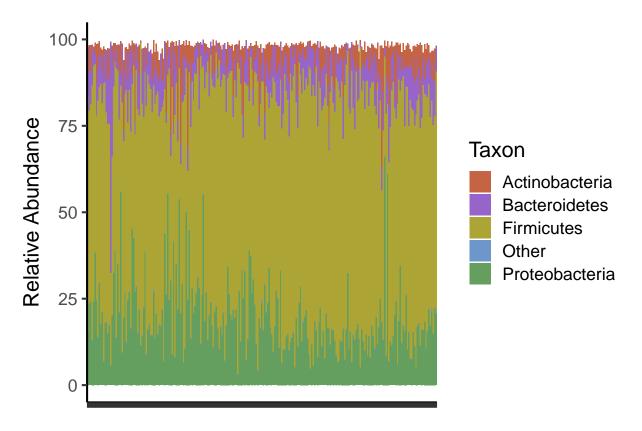
Phylum

```
parathaa_kSGB_Phylum <- generate_tax_level_table(parathaa_abundance_table_kSGB, "Phylum")</pre>
parathaa_seed_Phylum <- generate_tax_level_table(parathaa_abundance_table_seed, "Phylum")</pre>
dada2_full_Phylum <- generate_tax_level_table(dada2_abundance_table, "Phylum")</pre>
dada2_seed_Phylum <- generate_tax_level_table(dada2_seed_abundance_table, "Phylum")
m4_Phylum <- generate_tax_level_table(m4_filt_spec, "Phylum")</pre>
unique(c(parathaa kSGB Phylum$Taxon, parathaa seed Phylum$Taxon, dada2 full Phylum$Taxon,
       dada2_seed_Phylum$Taxon, m4_Phylum$Taxon))
                           "Proteobacteria"
                                                                   "Actinobacteria"
## [1] "Firmicutes"
                                               "Bacteroidetes"
## [5] "Other"
                           "Actinobacteriota" "Fusobacteriota"
                                                                   "UNCLASSIFIED"
parathaa_kSGB_Phylum_melt <- melt(parathaa_kSGB_Phylum)</pre>
## Using Taxon as id variables
parathaa_seed_Phylum_melt <- melt(parathaa_seed_Phylum)</pre>
## Using Taxon as id variables
dada2_full_Phylum_melt <- melt(dada2_full_Phylum)</pre>
## Using Taxon as id variables
dada2_seed_Phylum_melt <- melt(dada2_seed_Phylum)</pre>
## Using Taxon as id variables
m4_Phylum_melt <- melt(m4_Phylum)</pre>
## Using Taxon as id variables
Phyla_colors <- c('Actinobacteria'="#c56445", "Bacteroidetes"="#9764c9", 'Firmicutes'="#aca535",
                   'Other'="#6d96ca", "Proteobacteria"="#659f60", "UNCLASSIFIED"="#c65c8a",
                   'Actinobacteriota'="#c56445", 'Bacteroidota'="#9764c9", "Fusobacteriota"="cyan")
```

Parathaa kSGB

```
parathaa_kSGB_phyla_plot <- parathaa_kSGB_Phylum_melt %>%
   ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
   theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
   xlab("") + ylab("Relative Abundance") +
   scale_fill_manual(values=Phyla_colors)

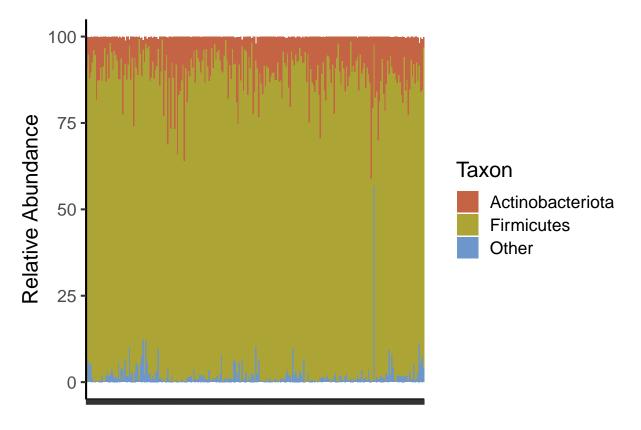
parathaa_kSGB_phyla_plot
```



Parathaa SEED

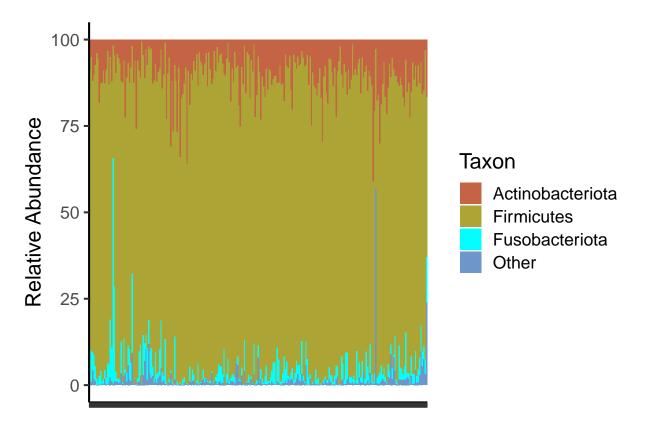
```
parathaa_seed_phyla_plot <- parathaa_seed_Phylum_melt %>%
   ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
   theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
   xlab("") + ylab("Relative Abundance") +
   scale_fill_manual(values=Phyla_colors)

parathaa_seed_phyla_plot
```



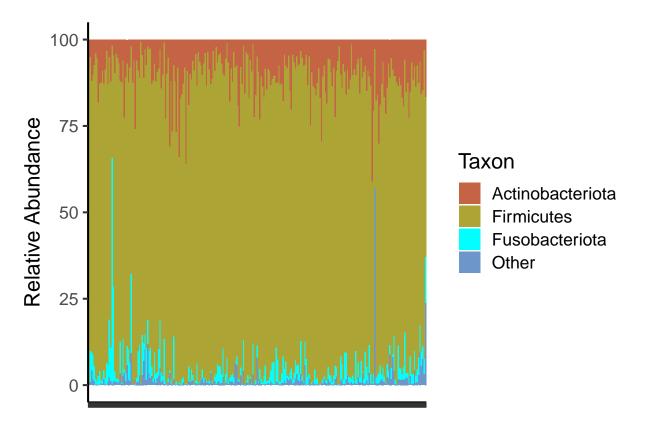
Dada2 Full

```
dada2_full_phyla_plot <- dada2_full_Phylum_melt %>%
   ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
   theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
   xlab("") + ylab("Relative Abundance") +
   scale_fill_manual(values=Phyla_colors)
dada2_full_phyla_plot
```



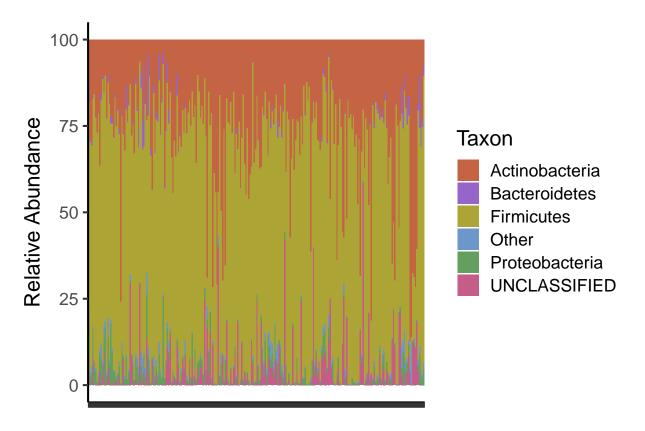
Dada2 SEED

```
dada2_seed_phyla_plot <- dada2_seed_Phylum_melt %>%
   ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
   theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
   xlab("") + ylab("Relative Abundance") +
   scale_fill_manual(values=Phyla_colors)
dada2_seed_phyla_plot
```

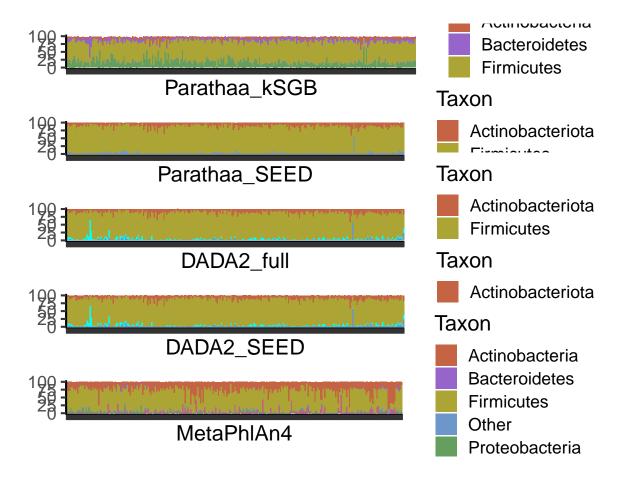


MetaPhlAn4

```
m4_Phyla_plot <- m4_Phylum_melt %>%
    ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
    theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
    xlab("") + ylab("Relative Abundance") +
    scale_fill_manual(values=Phyla_colors)
m4_Phyla_plot
```



Combined



Genus Comparison

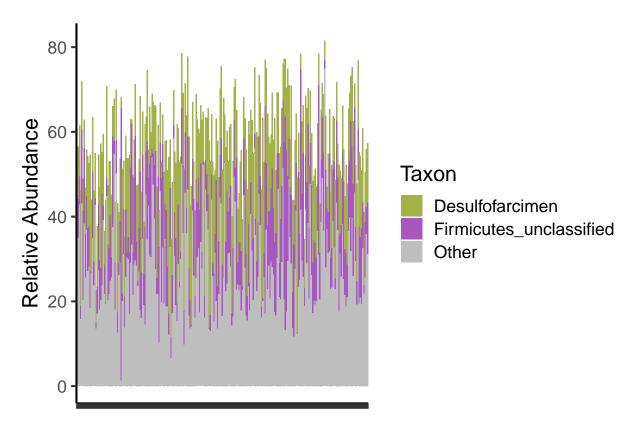
```
parathaa_kSGB_Genus <- generate_tax_level_table(parathaa_abundance_table_kSGB, "Genus", prop=5)
parathaa_seed_Genus <- generate_tax_level_table(parathaa_abundance_table_seed, "Genus", prop=5)
dada2_full_Genus <- generate_tax_level_table(dada2_abundance_table, "Genus", prop=5)
dada2_seed_Genus <- generate_tax_level_table(dada2_seed_abundance_table, "Genus", prop=5)
m4_genus <- generate_tax_level_table(m4_filt_spec, "Genus", prop=5)</pre>
unique(c(parathaa_kSGB_Genus$Taxon, parathaa_seed_Genus$Taxon, dada2_full_Genus$Taxon, dada2_seed_Genus
    [1] "Desulfofarcimen"
                                   "Firmicutes_unclassified"
    [3] "Other"
                                   "Bacillus"
##
                                   "Peptoclostridium"
##
    [5] "Clavibacter"
       "Turicibacter"
                                   "Romboutsia"
##
    [7]
   [9] "Blautia"
                                   "Bifidobacterium"
## [11] "Terrisporobacter"
                                   "Paraclostridium"
   [13] "GGB51725"
                                   "GGB47957"
##
  [15] "GGB77090"
                                   "Collinsella"
parathaa_kSGB_Genus_melt <- melt(parathaa_kSGB_Genus)</pre>
## Using Taxon as id variables
parathaa_seed_Genus_melt <- melt(parathaa_seed_Genus)</pre>
```

```
## Using Taxon as id variables
dada2_full_Genus_melt <- melt(dada2_full_Genus)</pre>
## Using Taxon as id variables
dada2_seed_Genus_melt <- melt(dada2_seed_Genus)</pre>
## Using Taxon as id variables
m4 genus melt <- melt(m4 genus)
## Using Taxon as id variables
Genera colors 5 <- c("Desulfofarcimen"="#a2b148",
                     "Firmicutes_unclassified"="#a756c3",
                     "Other"="grey",
                     "Bacillus"="#5bb94e",
                     "Clavibacter"="#6b6dc6",
                     "Peptoclostridium"="#d39a3b",
                     "Turicibacter"="#6999d4",
                     "Romboutsia"="#ca542a",
                     "Blautia"="#54c0a9",
                     "Bifidobacterium"="#d3425f",
                     "GGB51725"="#49874d",
                     "GGB47957"="#da73b6",
                     "GGB77090"="#83732f",
                     "GGB77090"="#a34d72",
                     "Collinsella"="#cc7d62")
```

Parathaa kSGB

```
parathaa_kSGB_genus_plot <- parathaa_kSGB_Genus_melt %>%
   ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
   theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
   xlab("") + ylab("Relative Abundance") +
   scale_fill_manual(values=Genera_colors_5)

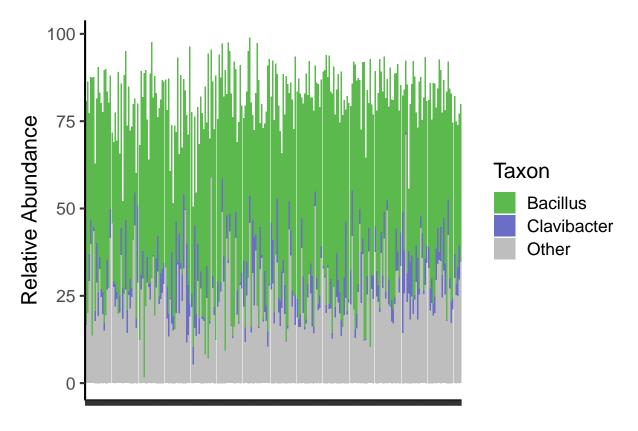
parathaa_kSGB_genus_plot
```



Parathaa SEED

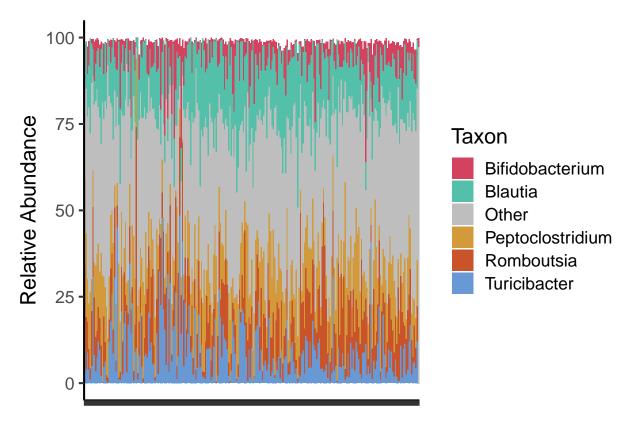
```
parathaa_seed_genus_plot <- parathaa_seed_Genus_melt %>%
   ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
   theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
   xlab("") + ylab("Relative Abundance") +
   scale_fill_manual(values=Genera_colors_5)

parathaa_seed_genus_plot
```



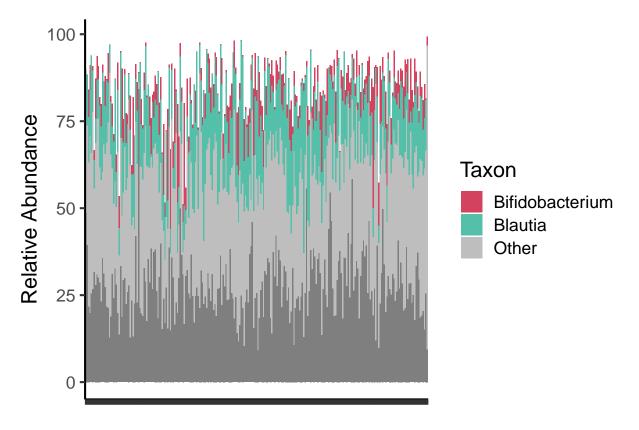
dada2 full

```
dada2_full_Genus_plot <- dada2_full_Genus_melt %>%
   ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
   theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
   xlab("") + ylab("Relative Abundance") +
   scale_fill_manual(values=Genera_colors_5)
dada2_full_Genus_plot
```



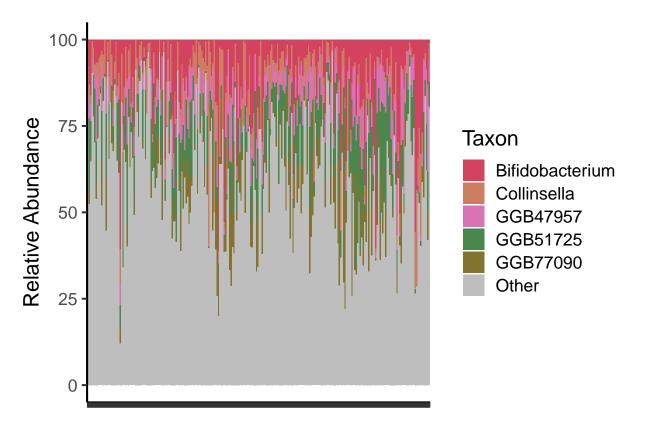
dada2 SEED

```
dada2_seed_genus_plot <- dada2_seed_Genus_melt %>%
   ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
   theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
   xlab("") + ylab("Relative Abundance") +
   scale_fill_manual(values=Genera_colors_5)
dada2_seed_genus_plot
```

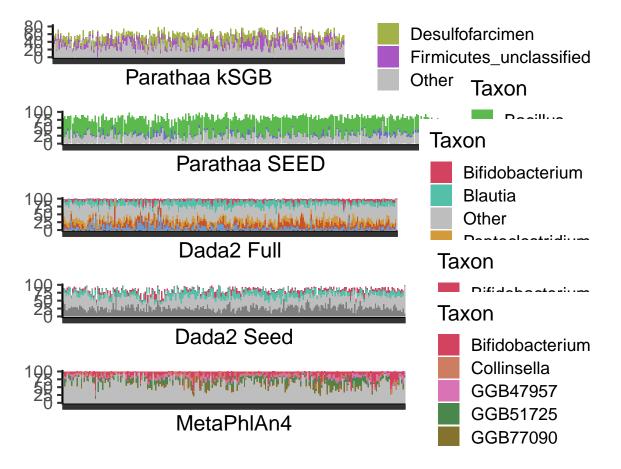


MetaPhlAn4

```
m4_genus_plot <- m4_genus_melt %>%
    ggplot(aes(x=variable, y=value, fill=Taxon)) + geom_col()+
    theme_classic(base_size = 16) + theme(axis.text.x = element_blank()) +
    xlab("") + ylab("Relative Abundance") +
    scale_fill_manual(values=Genera_colors_5)
m4_genus_plot
```



Combined



PCA Analysis

```
#takes in a list of feature tables
#takes in a distance type
#generates PCA
\#its not the perfect analysis since B/C doesn't do well with multi-assignments.
generate_PCA <- function(feat_tables, distance="bray"){</pre>
  merged_data <- Reduce(function(x, y) merge(x, y, by="Taxon", all=T), feat_tables)</pre>
  rownames(merged_data) <- merged_data$Taxon</pre>
  merged_data <- merged_data[,-1]</pre>
  merged_data[is.na(merged_data)] <- 0</pre>
  merged_data <- data.frame(t(merged_data))</pre>
  merged_dist <- vegdist(merged_data, method=distance)</pre>
  merged_PCA <- cmdscale(merged_dist, k=2, eig=T)</pre>
  plot_df <- data.frame(sample=rownames(merged_PCA$points),</pre>
                       PC1=merged_PCA$points[,1],
                       PC2=merged_PCA$points[,2])
  plot_df <- plot_df %>% mutate(Method = case_when(
    grepl("P_kSGB_", sample) ~ 'Parathaa_kSGB',
    grepl("P SEED ", sample) ~ 'Parathaa SEED',
    grepl("D_Full_", sample) ~ 'Dada2_Full',
    grepl("D_SEED_", sample) ~ 'Dada2_SEED',
```

```
grepl("M4_", sample) ~ 'MetaPhlAn4'
))

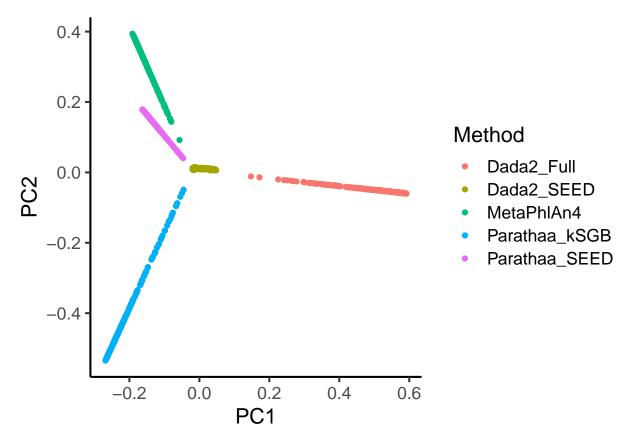
ret_plot <- plot_df %>% ggplot(aes(x=PC1, y=PC2, colour=Method)) + geom_point() +
    theme_classic(base_size=16)
    return(ret_plot)
}
```

Generate data

```
parathaa_kSGB_Species <- generate_tax_level_table(parathaa_abundance_table_kSGB, level="Species", prop=
colnames(parathaa_kSGB_Species)[-ncol(parathaa_kSGB_Species)] <-</pre>
  paste("P kSGB ", colnames(parathaa kSGB Species)[-ncol(parathaa kSGB Species)], sep="")
parathaa_seed_Species <- generate_tax_level_table(parathaa_abundance_table_seed, level="Species", prop=
colnames(parathaa_seed_Species)[-ncol(parathaa_seed_Species)] <-</pre>
  paste("P_SEED_", colnames(parathaa_seed_Species)[-ncol(parathaa_seed_Species)], sep="")
dada2_full_Species <- generate_tax_level_table(dada2_abundance_table, level="Species", prop=0)
colnames(dada2_full_Species)[-ncol(dada2_full_Species)] <-</pre>
  paste("D_Full_", colnames(dada2_full_Species)[-ncol(dada2_full_Species)], sep="")
dada2_seed_Species <- generate_tax_level_table(dada2_seed_abundance_table, level="Species", prop=0)
colnames(dada2_seed_Species)[-ncol(dada2_seed_Species)] <-</pre>
  paste("D_SEED_", colnames(dada2_seed_Species)[-ncol(dada2_seed_Species)], sep="")
m4_Species <- generate_tax_level_table(m4_filt_spec, level="Species", prop=0)
colnames(m4 Species)[-ncol(m4 Species)] <-</pre>
  paste("M4_", colnames(m4_Species)[-ncol(m4_Species)], sep="")
spec_feat_tabs <- list(parathaa_kSGB_Species, parathaa_seed_Species, dada2_full_Species, dada2_seed_Spe</pre>
                       m4 Species)
```

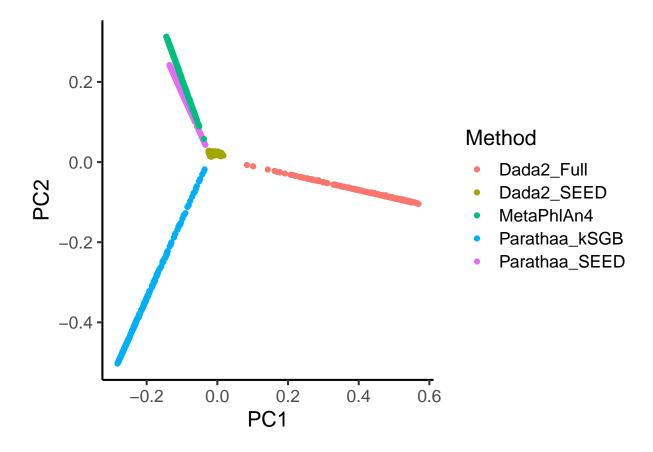
Bray

```
Spec_PCA_bray <- generate_PCA(spec_feat_tabs)
Spec_PCA_bray</pre>
```



Jaccard

Spec_PCA_Jac <- generate_PCA(spec_feat_tabs, distance="jaccard")
Spec_PCA_Jac</pre>



Deeper investigation

```
region_tree_kSGB <- "~/Repos/Parathaa2_0P3/M4_0ct22/M4_kSGB_V3V4/region_specific.tree"
taxafile_kSGB <- "~/Repos/Parathaa2_0P3/M4_0ct22/taxmapper.tsv"
load("~/Repos/Parathaa2_0P3/M4_0ct22/M4_kSGB_V3V4/resultTree_bestThresholds.RData")
Parathaa_kSGB_tree <- resultData$tax_bestcuts
Parathaa_kSGB_placements <- read.jplace("~/Repos/Parathaa2_0P3/M4_0ct22/M4_kSGB_V3V4/Fiber_assignments/specific_tree"
taxafile_SEED <- "~/Repos/Parathaa2_0P3/SILVA_run_V3V4/region_specific.tree"
taxafile_SEED <- "~/Repos/Parathaa/parathaa/input/silva_v138/taxmap_slv_ssu_ref_138.1.txt"
load("~/Repos/Parathaa2_0P3/SILVA_run_V3V4/resultTree_bestThresholds.RData")
Parathaa_SEED_tree <- resultData$tax_bestcuts
Parathaa_SEED_placements <- read.jplace("~/Repos/Parathaa2_0P3/SILVA_run_V3V4/Fiber_assignments/merged_
```

Phylum investigation...

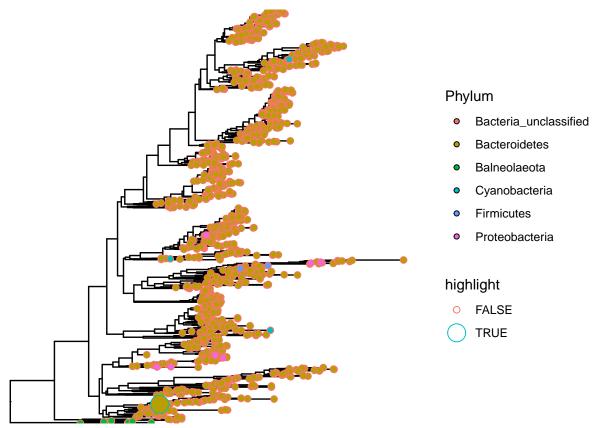
```
merged_parathaa_assignments <- full_join(parathaa_ksgb_assignments, parathaa_seed_assignments, by="quer_phylum_non_agreement <- merged_parathaa_assignments[which(merged_parathaa_assignments$Phylum.x != merged_parathaa_assignments$Phylum.y),]
setdiff(phylum_non_agreement$Phylum.x, phylum_non_agreement$Phylum.y)
```

[1] "Bacteroidetes"

```
[2] "Actinobacteria"
##
## [3] "Bacteroidetes; Cyanobacteria"
## [4] "Acidobacteria"
## [5] "Cyanobacteria; Tenericutes"
   [6] "Tenericutes"
## [7] "Bacteria unclassified"
## [8] "Chlorobi"
## [9] "Ignavibacteriae"
## [10] "Candidatus_Kryptonia; Ignavibacteriae; Synergistetes"
## [11] "Candidatus_Margulisbacteria"
## [12] "Synergistetes"
## [13] "Nitrospirae"
## [14] "Fusobacteria"
## [15] "Candidatus_Eremiobacteraeota"
setdiff(phylum_non_agreement$Phylum.y, phylum_non_agreement$Phylum.x)
   [1] "Actinobacteriota"
                                           "Bacteroidota"
   [3] "Myxococcota"
                                           "Desulfobacterota"
## [5] "Acidobacteriota"
                                           "Bdellovibrionota"
## [7] "Campylobacterota"
                                           "Nitrospirota"
## [9] "Synergistota"
                                           "Nitrospinota"
## [11] "Acidobacteriota; Actinobacteriota" "Chloroflexi"
## [13] "Fusobacteriota"
                                           "Fibrobacterota"
## [15] "Patescibacteria"
                                           "Gemmatimonadota"
## [17] "Dependentiae"
                                           "Marinimicrobia (SAR406 clade)"
## [19] "Caldatribacteriota"
phylum_non_agreement$Phylum.y[which(phylum_non_agreement$Phylum.y=="Actinobacteriota")] <- "Actinobacte
phylum non agreement$Phylum.y[which(phylum non agreement$Phylum.y=="Bacteroidota")] <- "Bacteroidetes"
phylum_non_agreement$Phylum.y[which(phylum_non_agreement$Phylum.y=="Acidobacteriota")] <- "Acidobacteri
phylum non agreement$Phylum.y[which(phylum non agreement$Phylum.y=="Fusobacteriota")] <- "Fusobacteria"
phylum_non_agreement <- phylum_non_agreement[which(phylum_non_agreement$Phylum.x !=</pre>
                                                     phylum non agreement$Phylum.y),]
Bacterodietes dive...
non_agree_bacts <- phylum_non_agreement[which(phylum_non_agreement$Phylum.x=="Bacteroidetes"),]</pre>
non_agree_bacts$query.name[1]
## [1] "008424a175a5cc75c61aa1a547a93f2b"
non_agree_bacts$query.name[2]
## [1] "00fd4ac0447c6b6539db1bcc40240e81"
non_agree_bacts[which(non_agree_bacts$query.name=="008424a175a5cc75c61aa1a547a93f2b"),]
008424a175a5cc75c61aa1a547a93f2b
                                                     Phylum.x
##
                           query.name Kingdom.x
## 5 008424a175a5cc75c61aa1a547a93f2b Bacteria Bacteroidetes Flavobacteriia
              Order.x
                               Family.x
                                                Genus.x Species.x maxDist.x
## 5 Flavobacteriales Flavobacteriaceae Salinimicrobium
                                                             <NA> 0.04222775
```

```
Kingdom.y Phylum.y
                             Class.v
                                              Order.v
                                                             Family.y
## 5 Bacteria Firmicutes Clostridia Oscillospirales Ruminococcaceae Ruminococcus
    Species.y maxDist.y
          <NA> 0.03843471
## 5
DADA2_assignments[which(DADA2_assignments$query.name=="008424a175a5cc75c61aa1a547a93f2b"),]
##
## ATTGGACAATGGACCAAAAGTCTGATCCAGCAATTCTGTGTGCACGATGAAGTTTTTCGGAATGTAAAGTGCTTTCAGTTGGGAAGAAGAAAGTGACGGT
## ATTGGACAATGGACCAAAAGTCTGATCCAGCAATTCTGTGTGCACGATGAAGTTTTTCGGAATGTAAAGTGCTTTCAGTTGGGAAGAAGAAAGTGACGGT
##
## ATTGGACAATGGACCAAAAGTCTGATCCAGCAATTCTGTGTGCACGATGAAGTTTTTCGGAATGTAAAGTGCTTTCAGTTGGGAAGAAGAAAGTGACGGT
##
## ATTGGACAATGGACCAAAAGTCTGATCCAGCAATTCTGTGTGCACGATGAAGTTTTTCGGAATGTAAAGTGCTTTCAGTTGGGAAGAAGAAAGTGACGGT
##
## ATTGGACAATGGACCAAAAGTCTGATCCAGCAATTCTGTGTGCACGATGAAGTTTTTCGGAATGTAAAGTGCTTTCAGTTGGGAAGAAGAAAGTGACGGT
## ATTGGACAATGGACCAAAAGTCTGATCCAGCAATTCTGTGTGCACGATGAAGTTTTTCGGAATGTAAAGTGCTTTCAGTTGGGAAGAAGAAAGTGACGGT
## ATTGGACAATGGACCAAAAGTCTGATCCAGCAATTCTGTGTGCACGATGAAGTTTTTCGGAATGTAAAGTGCTTTCAGTTGGGAAGAAGAAAGTGACGGT
##
## ATTGGACAATGGACCAAAAGTCTGATCCAGCAATTCTGTGTGCACGATGAAGTTTTTCGGAATGTAAAGTGCTTTCAGTTGGGAAGAAGAAAGTGACGGT
HMMM so dada2 assigns it Fusobacterium but the other two do not... that is very concerning...
All blast searches give: Fusobacterium..
Lets look at the tree placements...
parthaa kSGB placements <- get.placements(Parathaa kSGB placements)
### hmmm I think something is going wrong here...
parthaa_kSGB_placement_nodes <- plot_placement(Parathaa_kSGB_tree, "Phylum", Parathaa_kSGB_placements,
                                         label="008424a175a5cc75c61aa1a547a93f2b")
get n parents(Parathaa kSGB tree, 2320, 10)
## [1] 13294
subtree parent <- get subtree plot data(Parathaa kSGB tree, 13294, isTip=FALSE,
                                        highlight lab = "PEMPGKGO 00004 16S ribosomal RNA|M1878371282")
ggtree(as.phylo(subtree_parent)) %<+% subtree_parent + geom_tippoint(aes(fill=Phylum, size=highlight, c
```

Warning: Using size for a discrete variable is not advised.



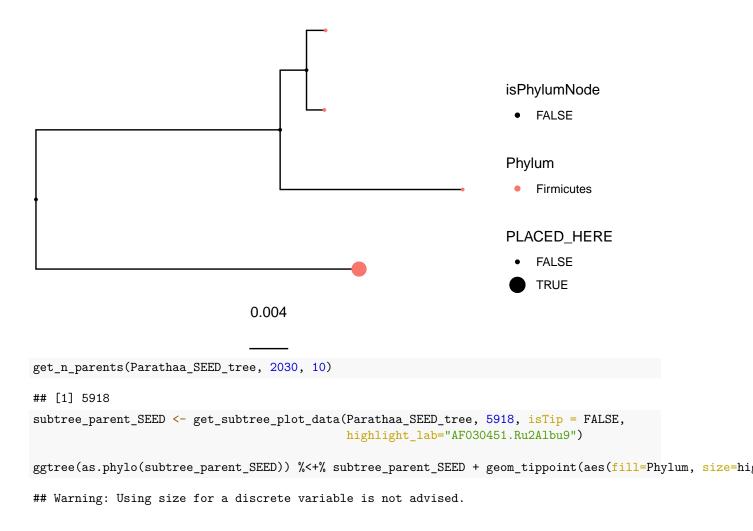
seems the placements around it are all "bacteroidetes" lets blast the sequence at its tip..

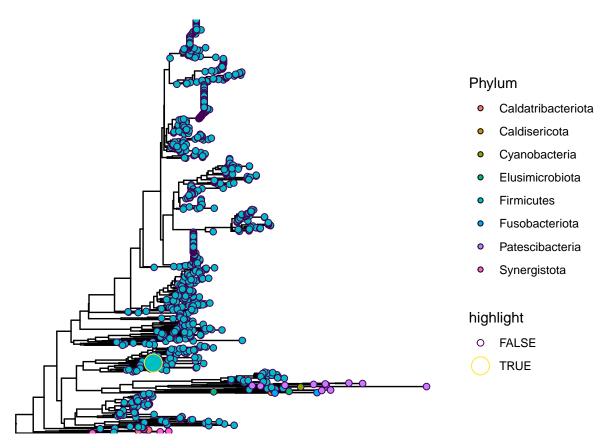
It does blast to Bacteroidetes... so doesn't seem like a huge issue...

Why is it being placed there though????? Something is clearly going wrong here during placement not sure what it is though... Is it the bifurcation enforcement?

 It

Distal length: 0.029911 Pendant length: 0.008





HMMM it places it in the Firmicutes... There are Fuso's in the subtree but there are a bit far away... Again looks like it is a placement issue?

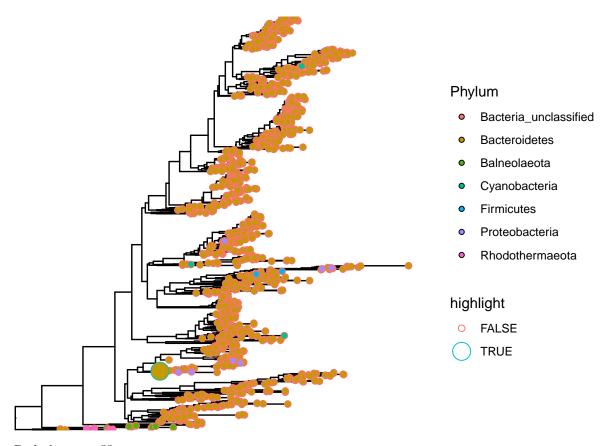
```
non_agree_bacts[which(non_agree_bacts$query.name=="00fd4ac0447c6b6539db1bcc40240e81"),]
```

00 fd 4ac 0447c 6b 6539db 1bcc 40240e 81

```
##
                           query.name Kingdom.x
                                                      Phylum.x Class.x Order.x
## 9 00fd4ac0447c6b6539db1bcc40240e81 Bacteria Bacteroidetes
                                                                  <NA>
                                                                          <NA>
     Family.x Genus.x Species.x maxDist.x Kingdom.y
                                                           Phylum.y
                                                                           Class.y
         <NA>
                           <NA> 0.1187457 Bacteria Actinobacteria Actinobacteria
## 9
##
                 Order.y Family.y Genus.y Species.y maxDist.y
## 9 Streptosporangiales
                             <NA>
                                     <NA>
                                                <NA> 0.1434061
DADA2_assignments[which(DADA2_assignments$query.name=="00fd4ac0447c6b6539db1bcc40240e81"),]
```

```
##
##
So parathaa kSGB says it should be Bacteroidetes... (no classification after Phylum...) Parathaa SEED
says it should be Actinobacteria...(Order is Streptosporangiales...) DADA2 says it should be Actinobacte-
ria... (Family is Eggerthellaceae, Genus is Asaccharobacter...)
So DADA2 and Parathaa do not agree on Order assignment... lets dig in...
BLAST says: it should be in Eggerthellaceae....
### hmmm I think something is going wrong here...
parthaa_kSGB_placement_nodes <- plot_placement(Parathaa_kSGB_tree, "Phylum", Parathaa_kSGB_placements,
                                 label="00fd4ac0447c6b6539db1bcc40240e81")
parthaa_kSGB_placement_nodes
                 Distal length: 0.021795 Pendant length: 0.122
                                                      isPhylumNode
                                                          FALSE
                                                      Phylum
                                                          Bacteroidetes
                                                      PLACED HERE
                                                          FALSE
                                                          TRUE
                        0.003
get_n_parents(Parathaa_kSGB_tree, 2456, 10)
## [1] 13287
subtree_parent <- get_subtree_plot_data(Parathaa_kSGB_tree, 13287, isTip=FALSE,
                                highlight lab = "MDNOLOHA 04242 16S ribosomal RNA|M1873143409")
ggtree(as.phylo(subtree_parent)) %<+% subtree_parent + geom_tippoint(aes(fill=Phylum, size=highlight, c
```

Warning: Using size for a discrete variable is not advised.

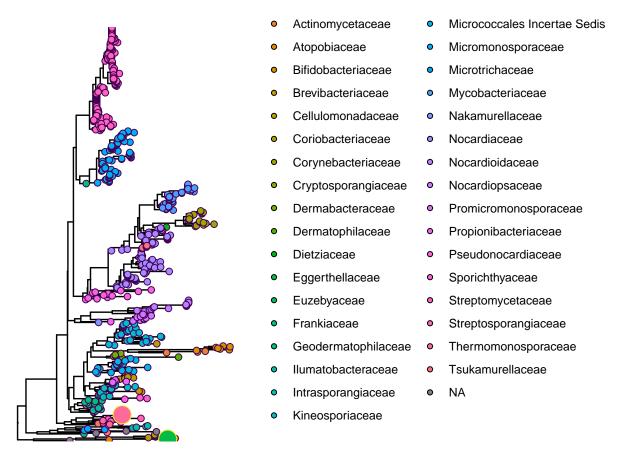


Bad placement??

[1] 6920

ggtree(as.phylo(subtree_parent)) %<+% subtree_parent + geom_tippoint(aes(fill=Phylum, size=highlight, c</pre>

 $\mbox{\tt \#\#}$ Warning: Using size for a discrete variable is not advised.



This placement looks fine at genus level but is pretty far away from Eggerthelleace at the family level (which is its most likely origin given the dada2 and blast search)