

BGGN 213

Foundations of Bioinformatics

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<http://thegrantlab.org/bggn213>

Recap From Last Time:

25 Responses:

<https://tinyurl.com/bggn213-02-F17>

ALIGNMENT FOUNDATIONS

- **Why...**
 - ▶ Why compare biological sequences?
- **What...**
 - ▶ Alignment view of sequence changes during evolution
(matches, mismatches and gaps)
- **How...**
 - ▶ Dot matrices
 - ▶ Dynamic programming
 - Global alignment
 - Local alignment
 - ▶ BLAST heuristic approach

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Basic Idea: Display one sequence above another with spaces (termed **gaps**) inserted in both to reveal **similarity** of nucleotides or amino acids.

Seq1: C A T T C A C

Seq2: C T C G C A G C

Basic Idea: Display one sequence above another with spaces (termed **gaps**) inserted in both to reveal **similarity** of nucleotides or amino acids.

Seq1: C A T T C A C

| |

Seq2: C T C G C A G C

↑↑ mismatch
match

Two types of character correspondence

Basic Idea: Display one sequence above another with spaces (termed **gaps**) inserted in both to reveal **similarity** of nucleotides or amino acids.

Seq1 : C A T - T C A - C
| | | | |
Seq2 : C - T C G C A G C

match
mismatch

Add gaps to increase number of matches

gaps

Basic Idea: Display one sequence above another with spaces (termed **gaps**) inserted in both to reveal **similarity** of nucleotides or amino acids.

Seq1 : C A T - T C A - C
| | | | |
Seq2 : C - T C G C A G C

Gaps represent 'indels'
mismatch represent mutations

match
mismatch } mutation

insertion
deletion } indels

Why compare biological sequences?

- To obtain **functional or mechanistic insight** about a sequence by inference from another potentially better characterized sequence
- To find whether two (or more) genes or proteins are **evolutionarily related**
- To find **structurally or functionally similar regions** within sequences (e.g. catalytic sites, binding sites for other molecules, etc.)
- Many practical bioinformatics applications...

Practical applications include...

- **Similarity searching of databases**
 - Protein structure prediction, annotation, etc...
- **Assembly of sequence reads** into a longer construct such as a genomic sequence
- **Mapping sequencing reads to a known genome**
 - "Resequencing", looking for differences from reference genome - SNPs, indels (insertions or deletions)
 - Mapping transcription factor binding sites via ChIP-Seq (chromatin immuno-precipitation sequencing)
 - Pretty much all next-gen sequencing data analysis

Practical applications include...

- **Similarity searching of databases**
 - Protein structure prediction
- **Assembly of sequences**
construct such
- **Mapping of new sequences to a known genome**
 - Pairwise sequence alignment is arguably the most fundamental operation of bioinformatics!
 - Looking for differences from reference
 - SNPs, indels (insertions or deletions)
 - Mapping transcription factor binding sites via ChIP-Seq (chromatin immuno-precipitation sequencing)
 - Pretty much all next-gen sequencing data analysis

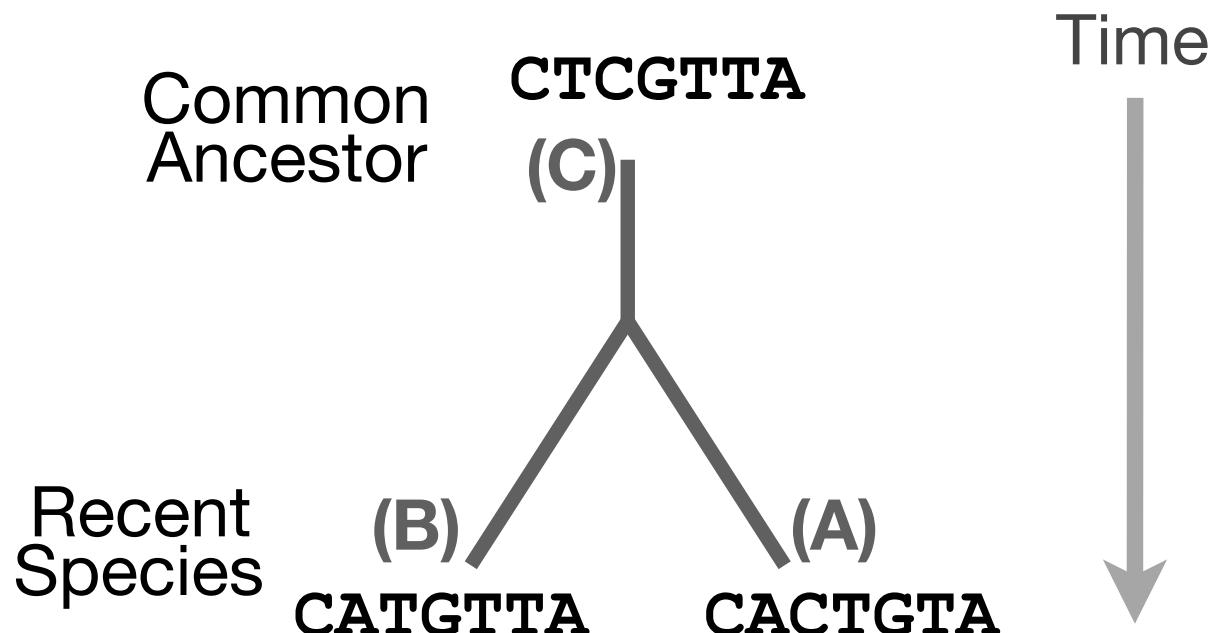
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Sequence changes during evolution

There are three major types of sequence change that can occur during evolution.

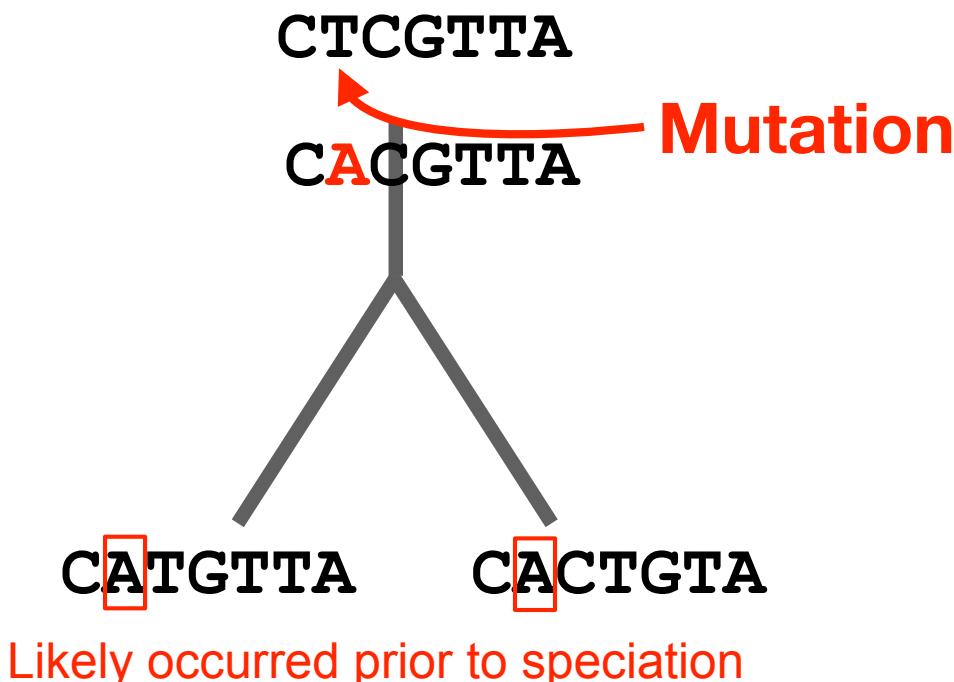
- Mutations/Substitutions
- Deletions
- Insertions



Mutations, deletions and insertions

There are three major types of sequence change that can occur during evolution.

- **Mutations/Substitutions** CTCGTTA → **CACGTTA**
- Deletions
- Insertions

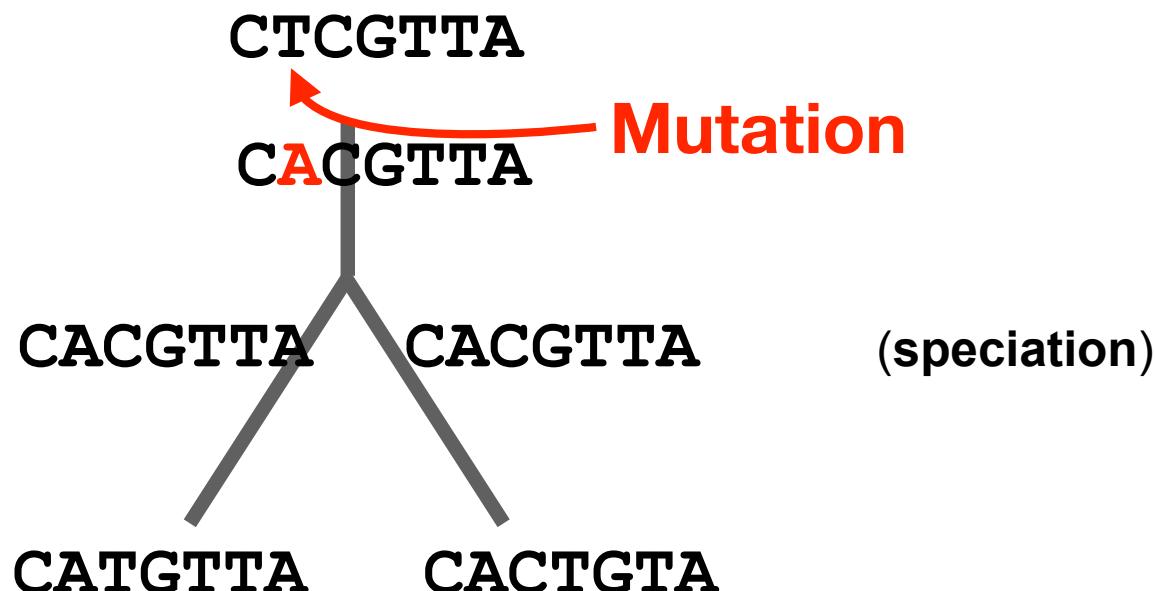


Mutations, deletions and insertions

There are three major types of sequence change that can occur during evolution.

- Mutations/Substitutions
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- Insertions

CTCGTTA → CACGTTA

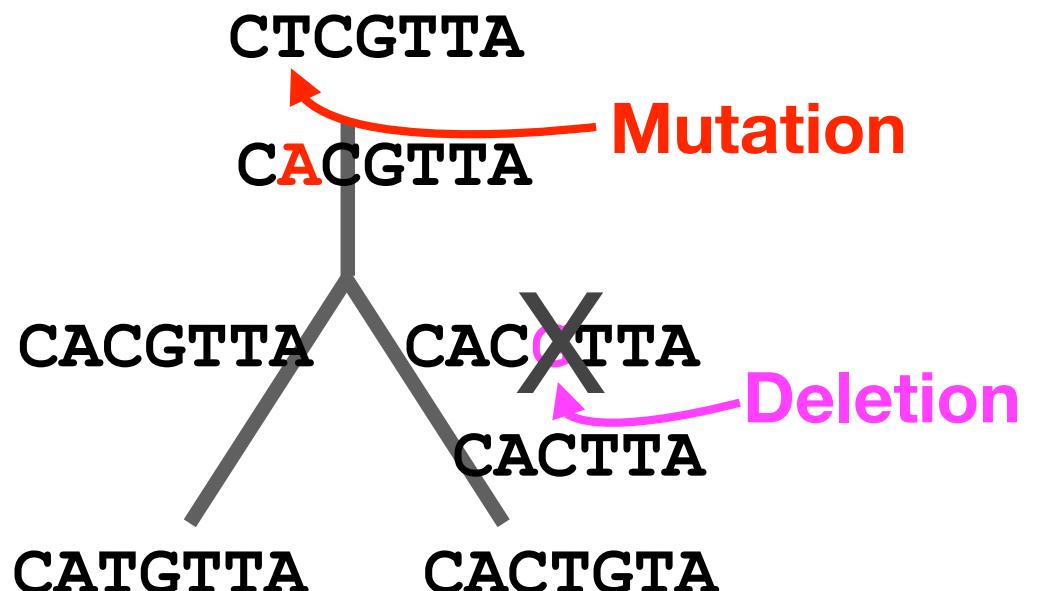


Mutations, deletions and insertions

There are three major types of sequence change that can occur during evolution.

- Mutations/Substitutions
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- Insertions

$\text{CTCGTTA} \rightarrow \text{C}\textcolor{red}{A}\text{CGTTA}$
 $\text{CAC}\textcolor{magenta}{GTTA} \rightarrow \text{CACTTA}$

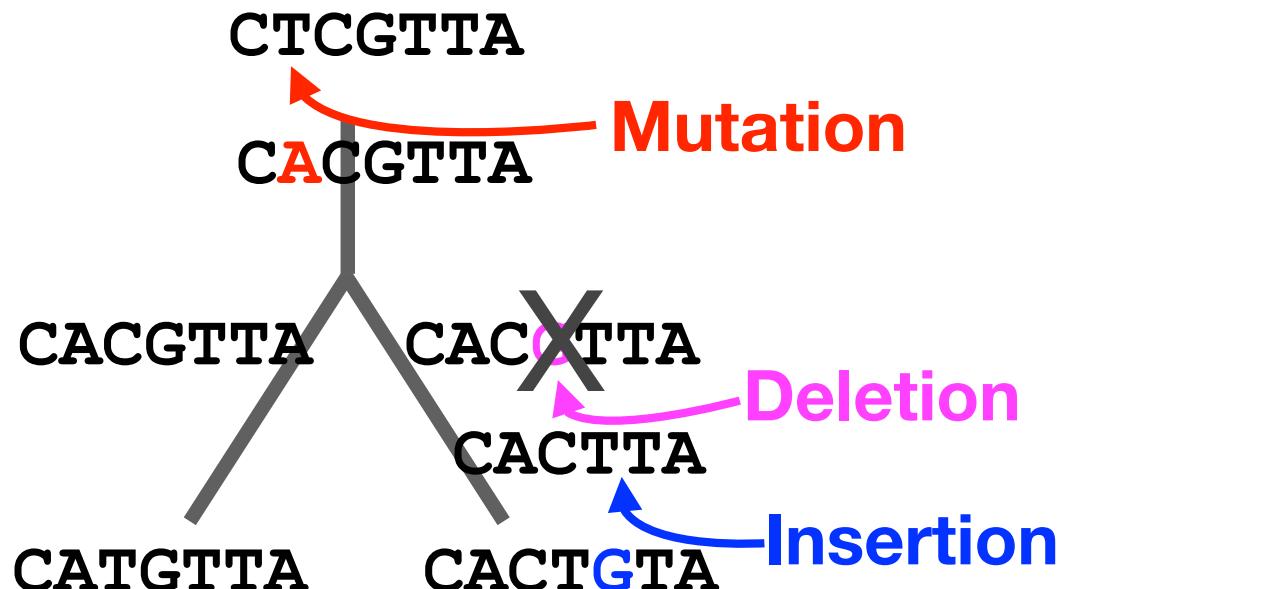


Mutations, deletions and insertions

There are three major types of sequence change that can occur during evolution.

- Mutations/Substitutions
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- Insertions

CTCGTTA → CACGTTA
CACGTTA → CACTTA
CACTTA → CACTGTA

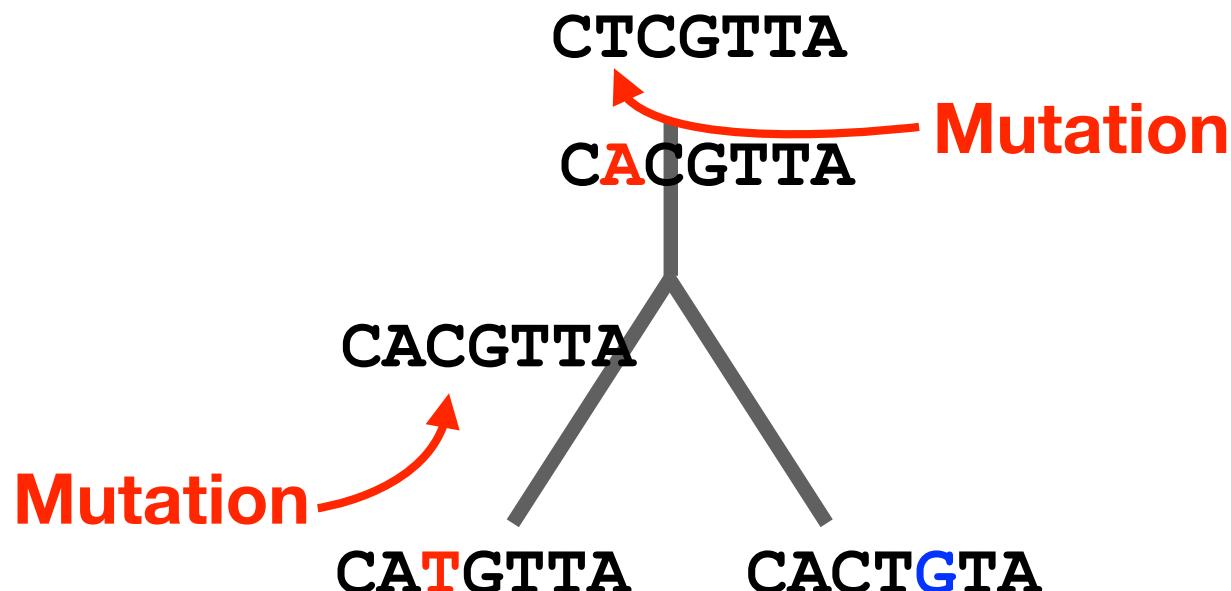


Mutations, deletions and insertions

There are three major types of sequence change that can occur during evolution.

- **Mutations/Substitutions**
- Deletions
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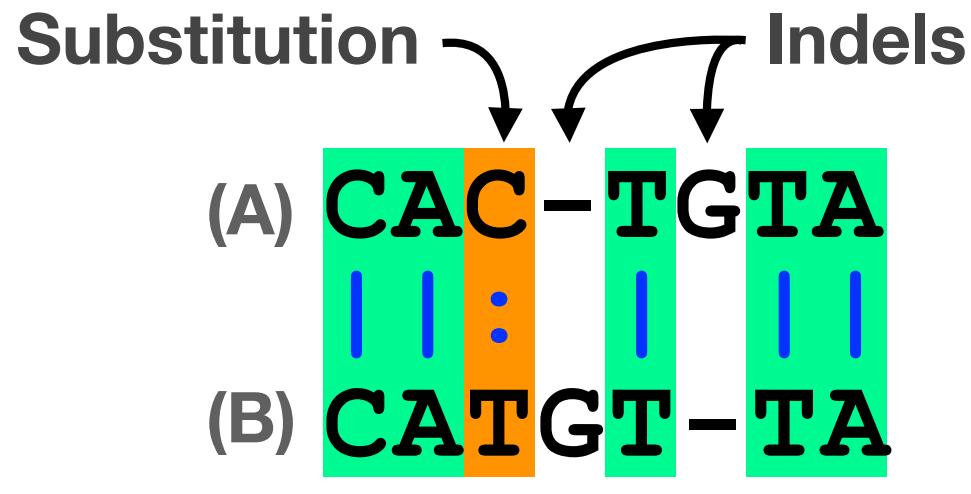
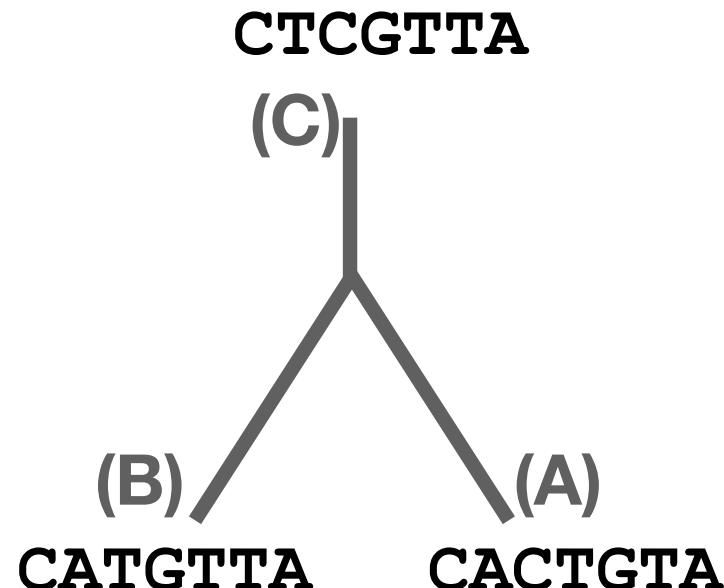
$\text{CTCGTTA} \rightarrow \text{CACGTTA}$
 $\text{CACGTTA} \rightarrow \text{CATGTAA}$



Alignment view

Alignments are great tools to visualize sequence similarity and evolutionary changes in homologous sequences.

- **Mismatches** represent mutations/substitutions
- **Gaps** represent insertions and deletions (indels)



Match	Mismatch	Gap
5	1	2

Alternative alignments

- Unfortunately, finding the correct alignment is difficult if we do not know the evolutionary history of the two sequences

Q. Which of these 3 possible alignments is best?

1.

CA	CTG	TA			
	:	:	:		
CAT	GT	TA			

2.

CA	CTG	T	A		
CA	-	TGTTA			

3.

CAC	-	T	G	TA		
	:					
CAT	GT	-	TA			

Alternative alignments

- One way to judge alignments is to compare their number of matches, insertions, deletions and mutations

● 4 matches
● 3 mismatches
○ 0 gaps

● 6 matches
● 0 mismatches
○ 2 gaps

● 5 matches
● 1 mismatch
○ 2 gaps

CACTGTA
||: : : ||
CATGTAA

This sequence alignment shows a green box for CACTGTA and an orange box for CATGTAA. There are 4 matches (green) between the first four bases of both sequences. A blue colon indicates a mismatch between the fifth base of each sequence. The last two bases are identical, indicated by blue double bars.

CACTGTA
|| | -A
CA-TGTAA

This sequence alignment shows a green box for CACTGTA and an orange box for CA-TGTAA. There are 6 matches (green) between the first five bases of both sequences. The sixth base of the orange sequence is a mismatch (orange). The last three bases are identical, indicated by blue double bars.

CAC-TGTA
|| : |
CATGT-TA

This sequence alignment shows a green box for CAC-TGTA and an orange box for CATGT-TA. There are 5 matches (green) between the first four bases of both sequences. The fifth base of the orange sequence is a mismatch (orange). The last two bases are identical, indicated by blue double bars.

Scoring alignments

- We can assign a score for each match (+3), mismatch (+1) and indel (-1) to identify the **optimal alignment *for this scoring scheme***

● 4 (+3)
● 3 (+1)
○ 0 (-1) = 15

● 6 (+3)
● 0 (+1)
○ 2 (-1) = 16

● 5 (+3)
● 1 (+1)
○ 2 (-1) = 14

CACTGTA
||: : : ||
CATGT TA

A sequence alignment showing two rows of DNA sequence. The top row is CACTGTA and the bottom row is CATGT TA. Vertical blue lines indicate matches between A, C, T, G, T, and A. Vertical orange lines indicate a mismatch between the second T and the bottom T, and an indel (deletion of T) in the bottom sequence. The alignment is highlighted with green boxes around the matching segments.

CACTGT-A
|| | | | |
CA-TGT TA

A sequence alignment showing two rows of DNA sequence. The top row is CACTGT-A and the bottom row is CA-TGT TA. Vertical blue lines indicate matches between C, A, T, G, T, and A. Vertical orange lines indicate a mismatch between the first C and the bottom C, and an indel (deletion of C) in the bottom sequence. The alignment is highlighted with green boxes around the matching segments.

CAC-TGTA
|| : | |
CATGT-TA

A sequence alignment showing two rows of DNA sequence. The top row is CAC-TGTA and the bottom row is CATGT-TA. Vertical blue lines indicate matches between C, A, C, T, G, T, and A. Vertical orange lines indicate a mismatch between the second C and the bottom C, and an indel (deletion of C) in the bottom sequence. The alignment is highlighted with green boxes around the matching segments.

Optimal alignments

- Biologists often prefer **parsimonious alignments**, where the number of postulated sequence changes is minimized.

- 4 matches
- 3 mismatches
- 0 gaps

CACTGTA
||: : : ||
CATGTAA

A sequence alignment showing two DNA strands. The top strand is CACTGTA and the bottom strand is CATGTAA. Vertical blue lines indicate matches between A, C, G, T, and A. Vertical orange lines indicate a mismatch between the second T and G. Horizontal blue lines above and below the strands indicate gaps.

- 6 matches
- 0 mismatches
- 2 gaps

CACTGT-A
|| | | | |
CA-TGTAA

A sequence alignment showing two DNA strands. The top strand is CACTGT-A and the bottom strand is CA-TGTAA. Vertical blue lines indicate matches between all positions. Horizontal blue lines above and below the strands indicate gaps.

- 5 matches
- 1 mismatch
- 2 gaps

CAC-TGTA
|| : | |
CATGT-TA

A sequence alignment showing two DNA strands. The top strand is CAC-TGTA and the bottom strand is CATGT-TA. Vertical blue lines indicate matches between C, A, C, T, and A. Vertical orange lines indicate a mismatch between the third G and T. Horizontal blue lines above and below the strands indicate gaps.

Optimal alignments

- Biologists often prefer **parsimonious alignments**, where the number of postulated sequence changes is minimized.

- 4 matches
- 3 mismatches
- 0 gaps

CA	CTG	TA
	:	:
CAT	GTTA	

- 6 matches
- 0 mismatches
- 2 gaps

CA	CTG	T	-	A
CA	-	TGTTA		

- 5 matches
- 1 mismatch
- 2 gaps

CAC	-	TG	TA
	:		
CAT	GT	-	TA

Optimal alignments

- Biologists often prefer **parsimonious alignments**, where the number of postulated sequence changes is minimized.

- 4 matches
- 3 mismatches
- 0 gaps

CA	CTG	TA
	:	:
CAT	GTTA	

- 6 matches
- 0 mismatches
- 2 gaps

CA	CTG	-	TA
CA	-TGTTA		

- 5 matches
- 1 mismatch
- 2 gaps

CAC	-T	GTA
	:	
CAT	GT-	TA

Optimal alignments

- Biologists often prefer **parsimonious alignments**, where the number of sequence changes is minimized.

● 4 matches
● 3 mismatches
● 2 gaps

Warning: There may be more than one optimal alignment and these may not reflect the true evolutionary history of our sequences!

C	A	C	T	G	T	-	A
C	A	-	T	G	T	T	A

C	A	C	-	T	G	T	A
		:					
C	A	T	G	T	-	T	A

ALIGNMENT FOUNDATIONS

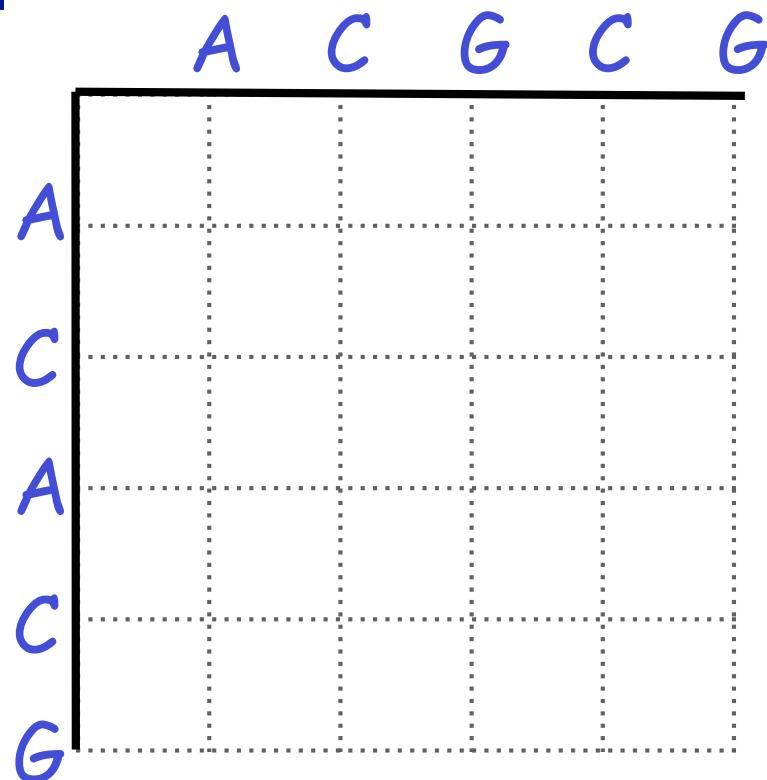
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 - ▶ How do we compute the optimal alignment between two sequences?
 - ▶ BLAST heuristic approach

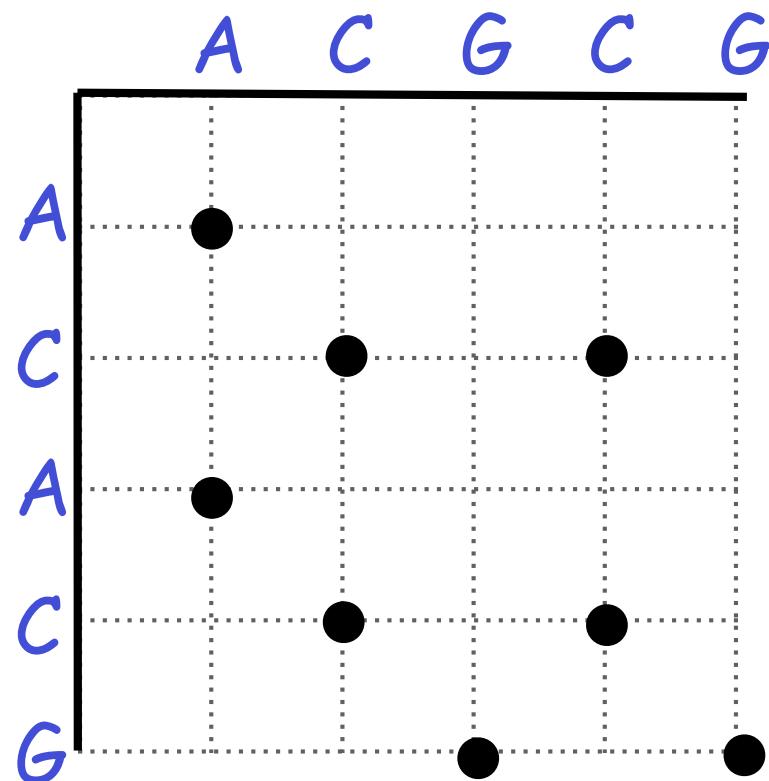
Dot plots: simple graphical approach

- Place one sequence on the vertical axis of a 2D grid (or matrix) and the other on the horizontal



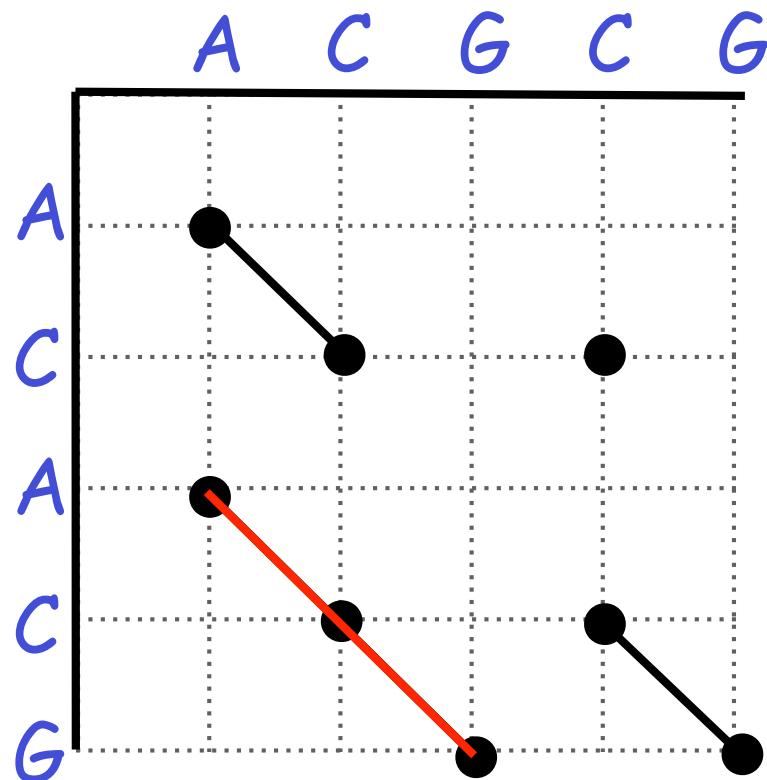
Dot plots: simple graphical approach

- Now simply put dots where the horizontal and vertical sequence values match



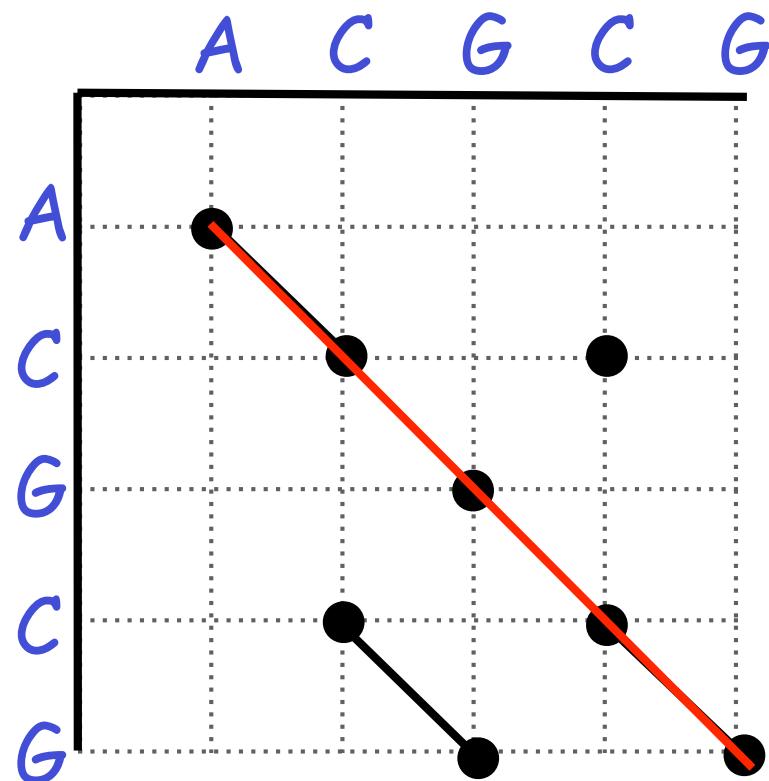
Dot plots: simple graphical approach

- Diagonal runs of dots indicate matched segments of sequence



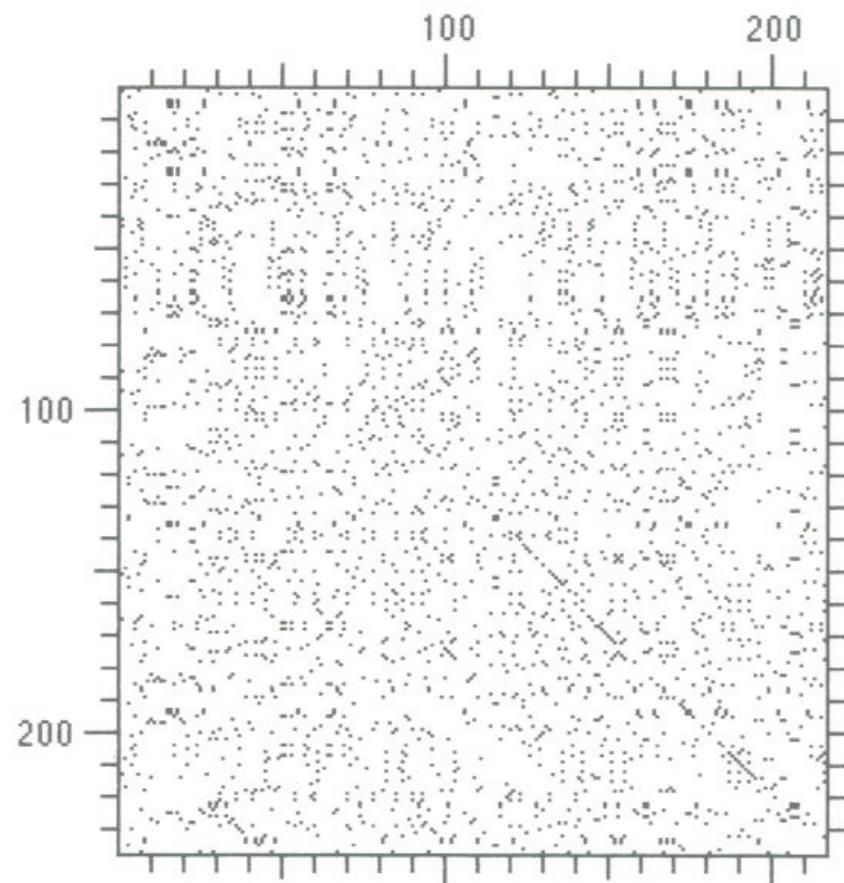
Dot plots: simple graphical approach

Q. What would the dot matrix of a two identical sequences look like?



Dot plots: simple graphical approach

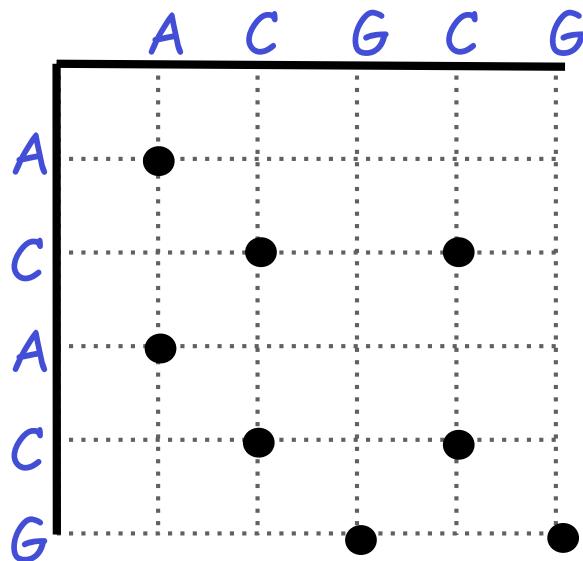
- Dot matrices for long sequences can be noisy



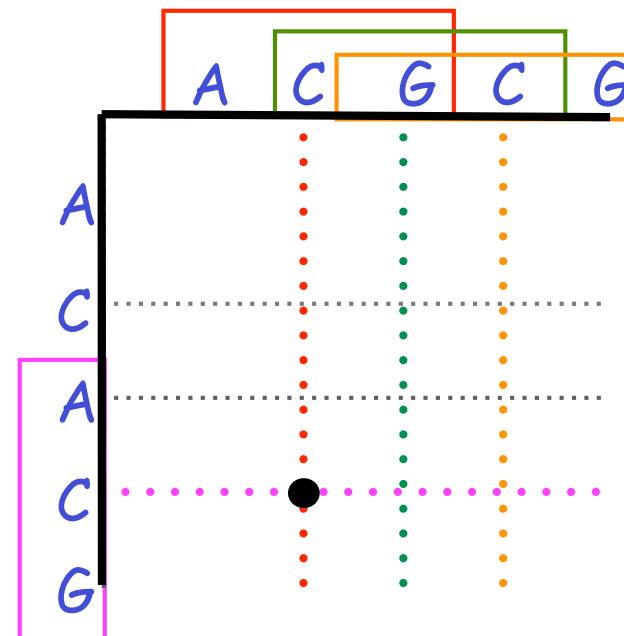
Dot plots: window size and match stringency

Solution: use a window and a threshold

- compare character by character within a window
- require certain fraction of matches within window in order to display it with a dot.
 - You have to choose window size and stringency



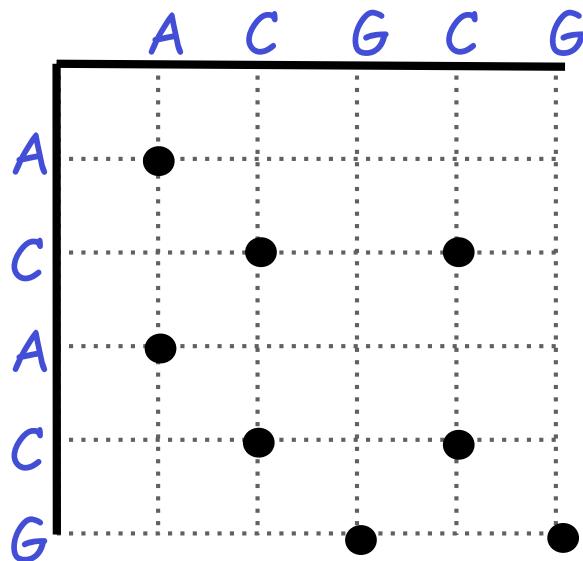
Filter
Window = 3
Stringency = 3



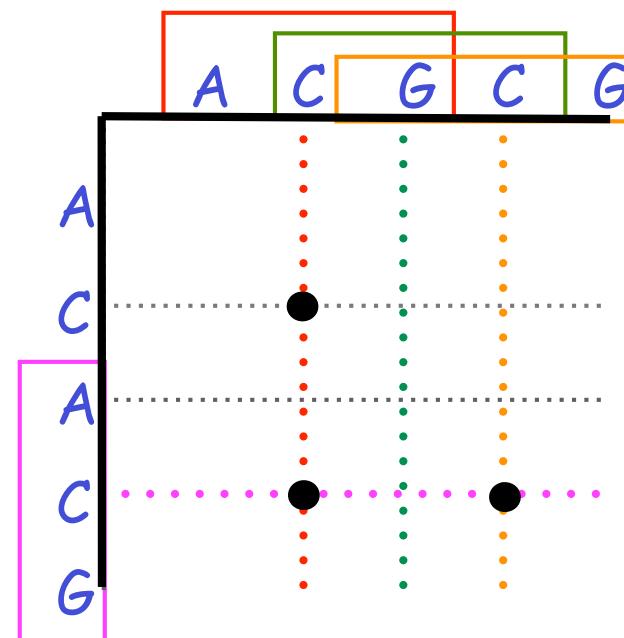
Dot plots: window size and match stringency

Solution: use a window and a threshold

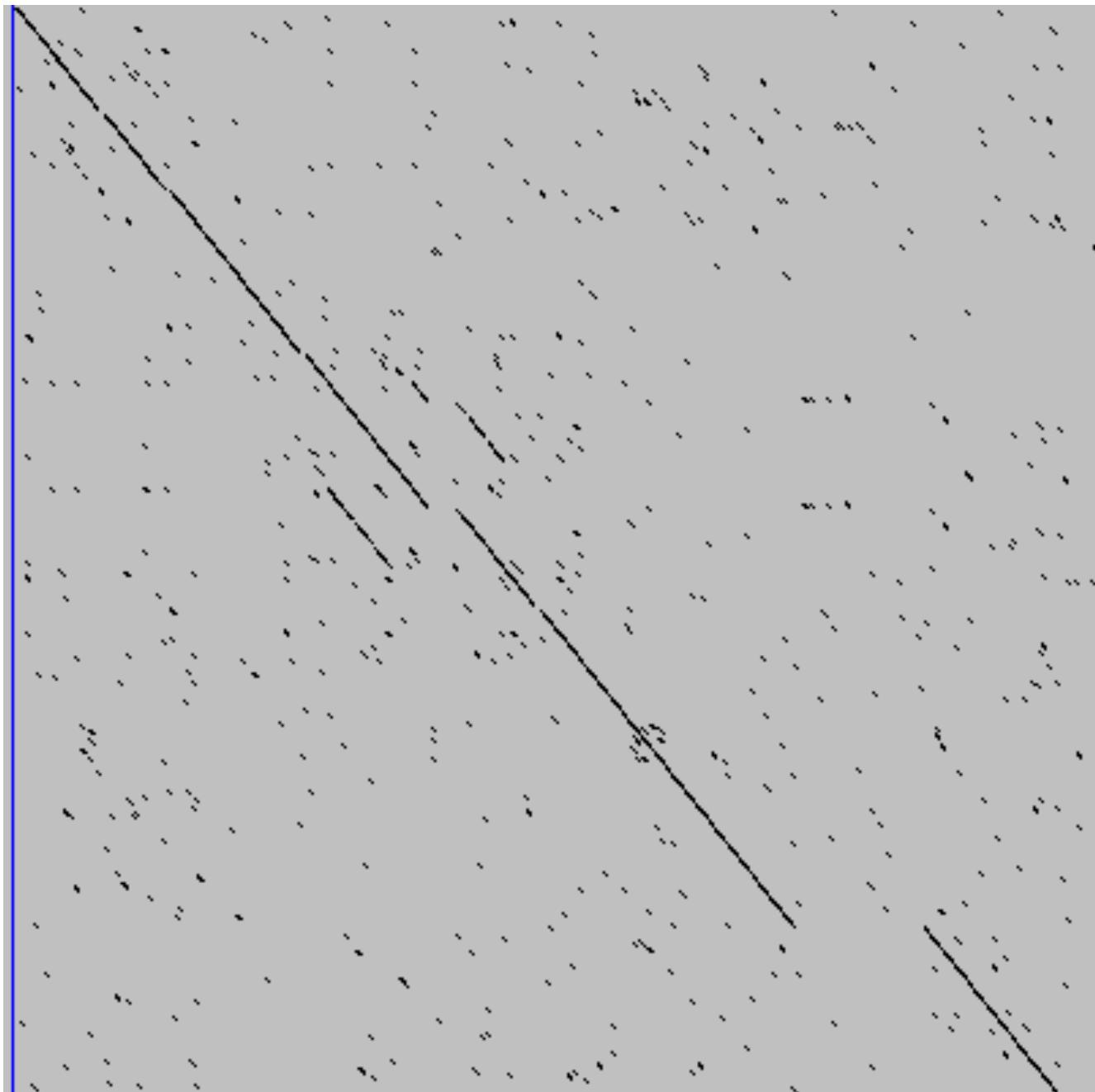
- compare character by character within a window
- require certain fraction of matches within window in order to display it with a dot.
 - You have to choose window size and stringency



Filter
Window = 3
Stringency = 2



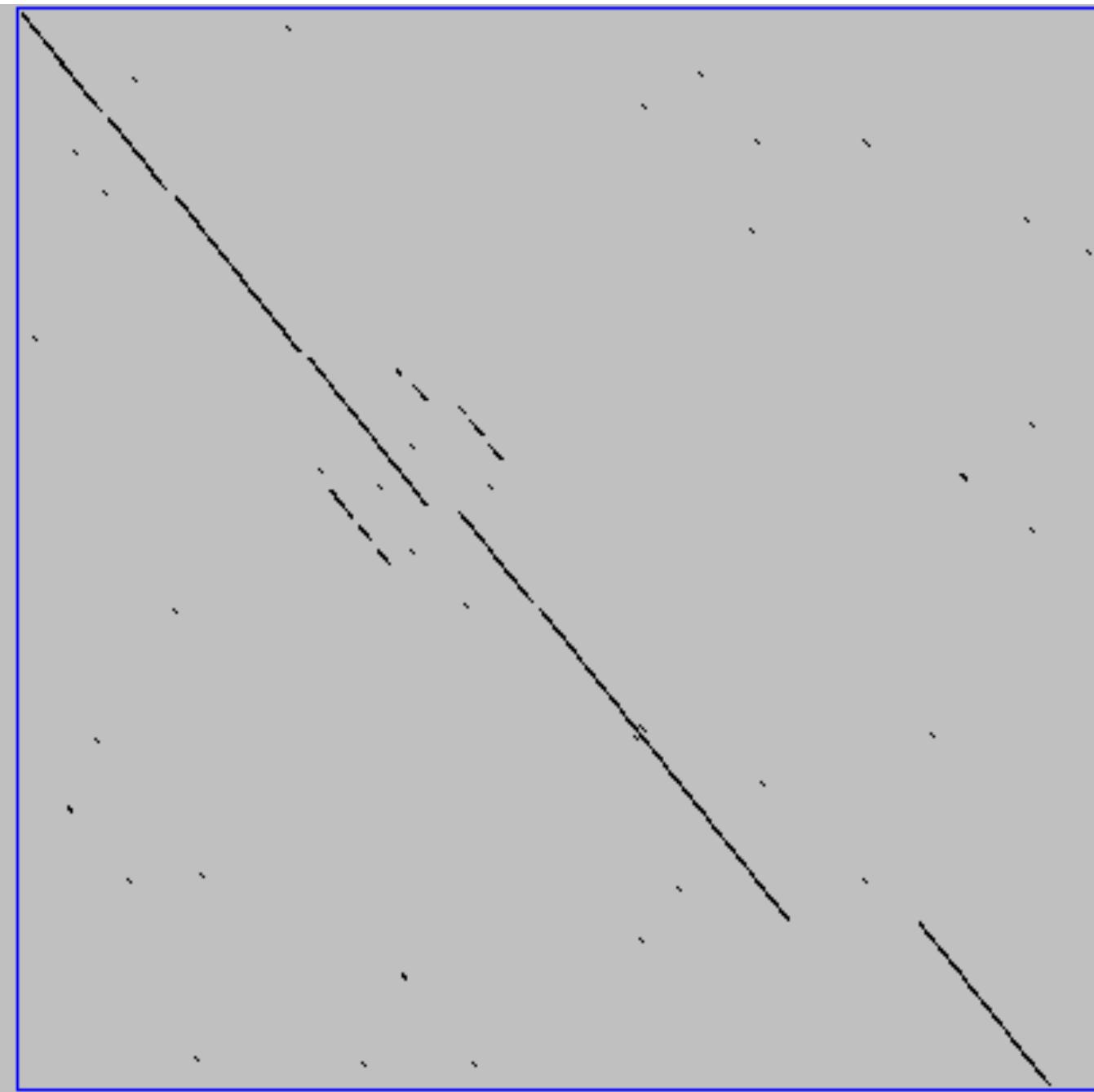
Window size = 5 bases



A dot plot simply puts a dot where two sequences match. In this example, dots are placed in the plot if 5 bases in a row match perfectly. Requiring a 5 base perfect match is a **heuristic** – only look at regions that have a certain degree of identity.

Do you expect evolutionarily related sequences to have more word matches (matches in a row over a certain length) than random or unrelated sequences?

Window size = 7 bases

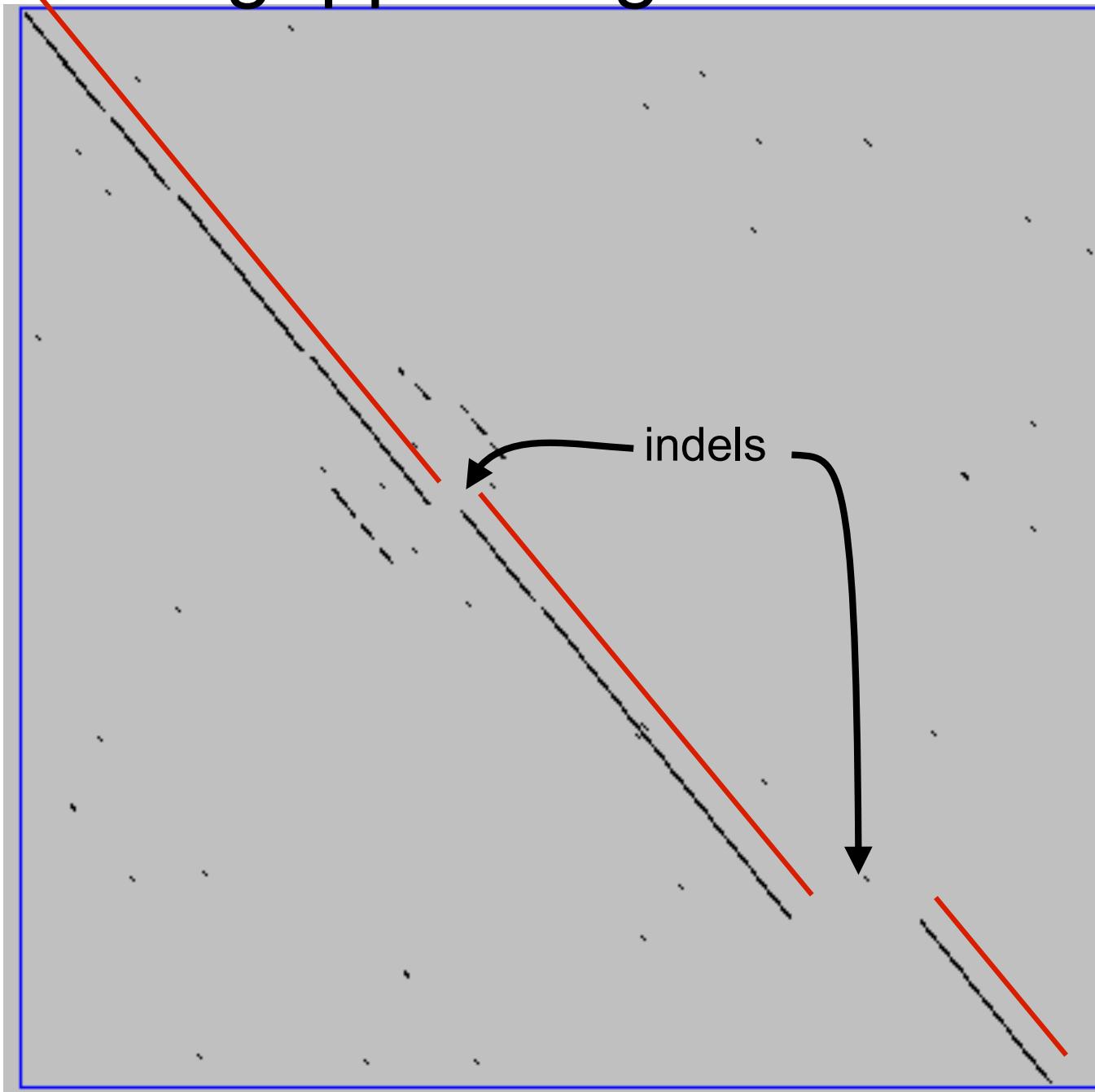


This is a dot plot of the same sequence pair. Now 7 bases in a row must match for a dot to be placed. Noise is reduced.

Using windows of a certain length is very similar to using words (kmers) of N characters in the heuristic alignment search tools

Bigger window (kmer)
fewer matches to consider

Ungapped alignments



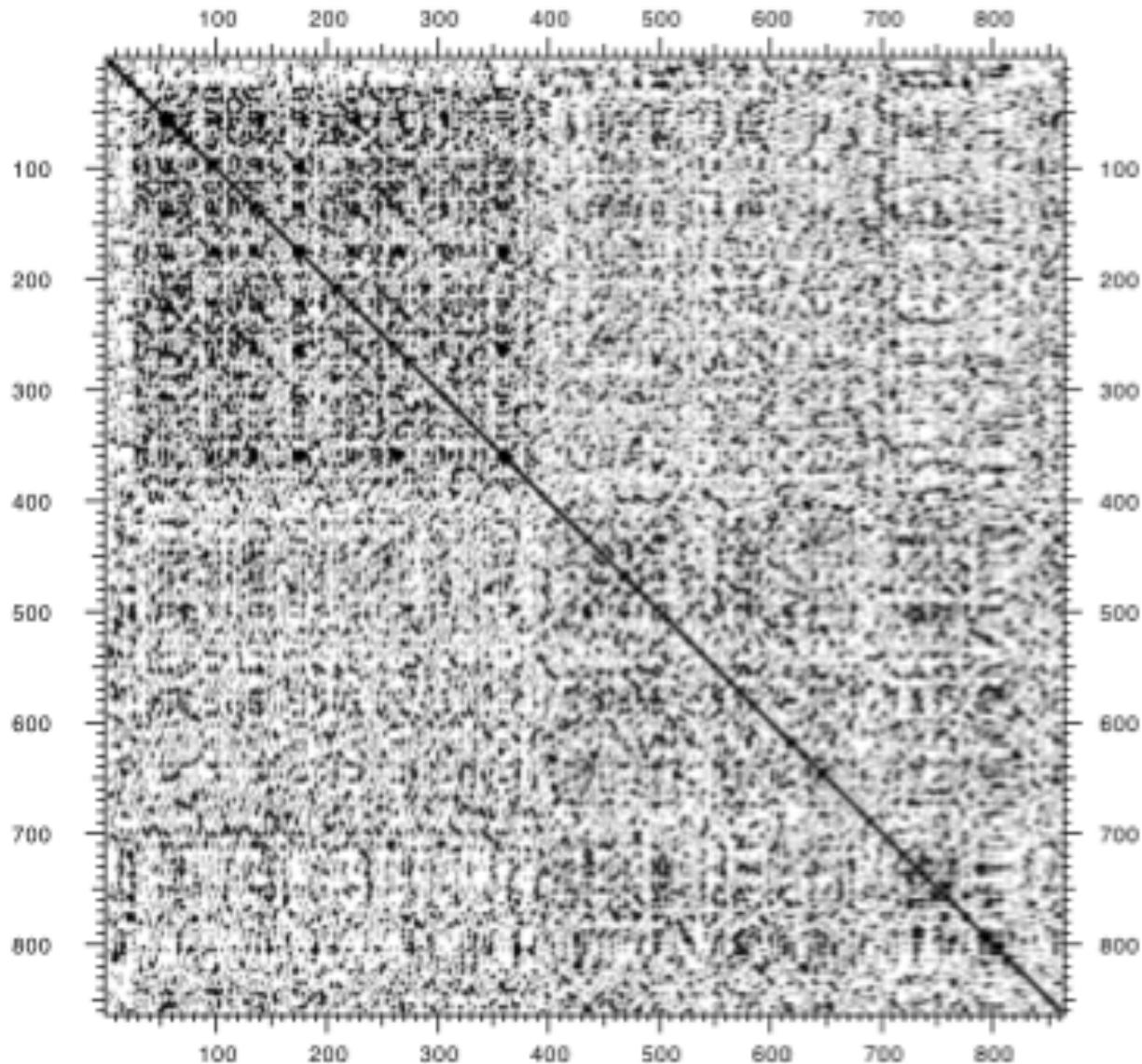
Only **diagonals** can be followed.

Downward or rightward paths represent **insertion** or **deletions** (gaps in one sequence or the other).

Uses for dot matrices

- Visually assessing the similarity of two protein or two nucleic acid sequences
- Finding local repeat sequences within a larger sequence by comparing a sequence to itself
 - Repeats appear as a set of diagonal runs stacked vertically and/or horizontally

Repeats

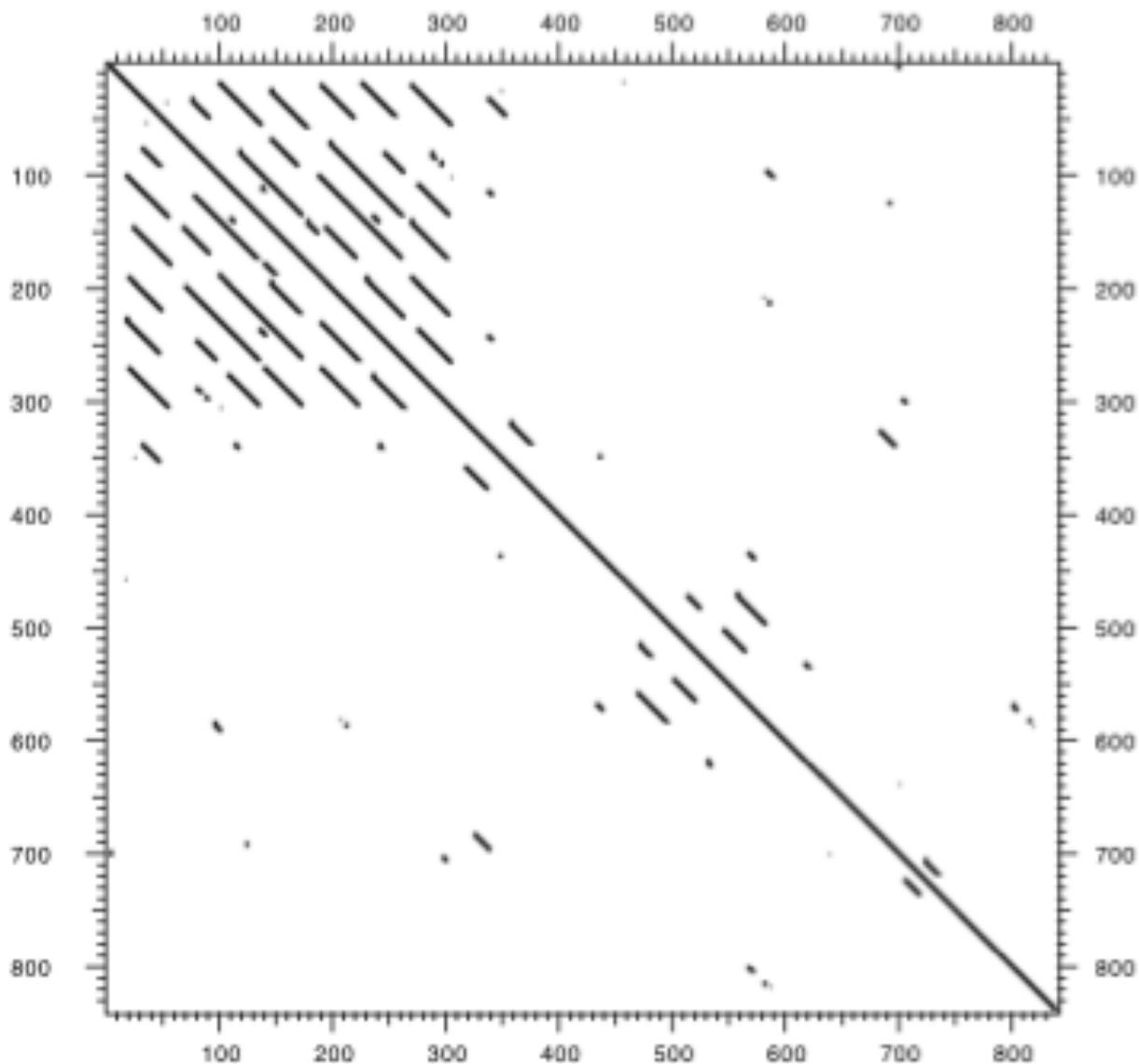


Human LDL receptor
protein sequence
(Genbank P01130)

$$\begin{aligned} W &= 1 \\ S &= 1 \end{aligned}$$

(Figure from Mount, "Bioinformatics sequence and genome analysis")

Repeats



Human LDL receptor
protein sequence
(Genbank P01130)

$$\begin{aligned} W &= 23 \\ S &= 7 \end{aligned}$$

(Figure from Mount, "Bioinformatics sequence and genome analysis")

Your Turn!

Exploration of dot plot parameters (hands-on worksheet **Section 1**)

<http://bio3d.ucsd.edu/dotplot/>

<https://bioboot.shinyapps.io/dotplot/>

BGGN-213: Dot Plot Comparison of Two Sequences

Dot plots are a simple graphical approach for the visual comparison of two sequences. They have a long history (see [Maizel and Lenk 1981](#) and references therein) and entail placing one sequence on the vertical axis of a 2D grid (or matrix) and the other on the horizontal. In its simplest form, a dot is placed where the horizontal and vertical sequence values match. That is a dot is produced at position (i,j) if character number i in the first sequence is the same as character number j in the second sequence. More elaborate forms use 'sliding windows' composed of multiple characters and a threshold value, or 'match stringency' for two windows to be considered as matched.

Dot Plot Parameters

Alter the parameters below to change the displayed protein and DNA dot plots. It is important to have a good feel for these parameters when we get to alignment heuristic approaches later.

Window Size:

Moving window step size:

Match stringency:

Match stringency specifies the number of match characters required per window. It should not be larger than your window size!

Protein Dot Plot
wsize = 3 wstep = 3 , nmatch = 2

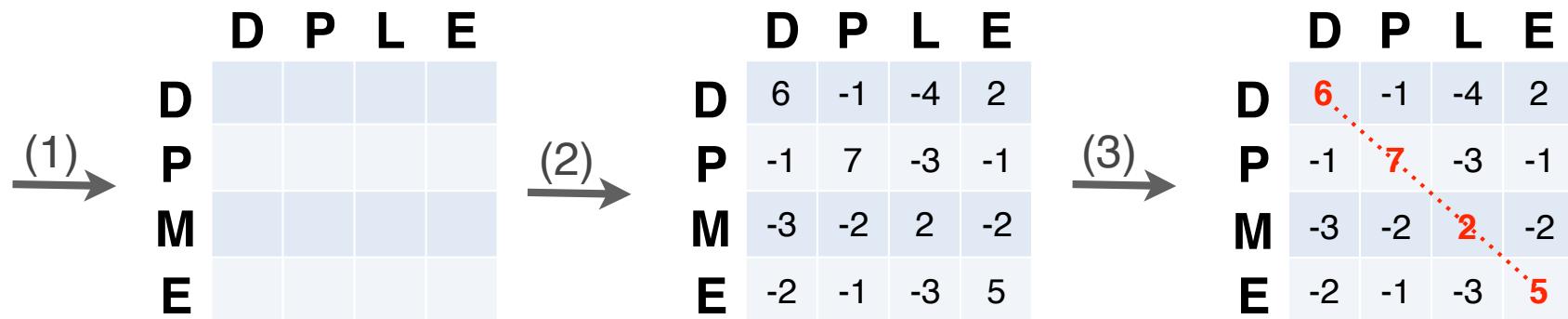
DNA Dot Plot
wsize = 3 wstep = 3 , nmatch = 2

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The Dynamic Programming Algorithm

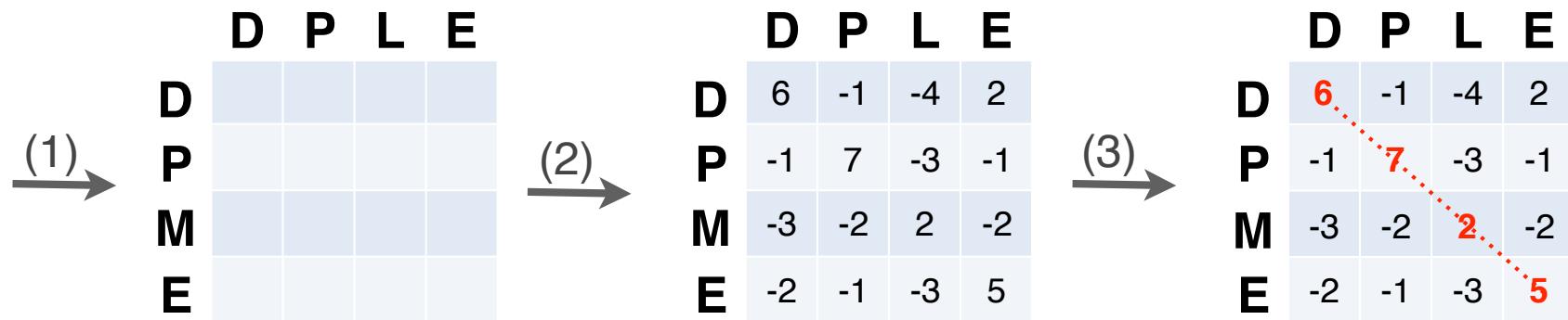
- The dynamic programming algorithm can be thought of an extension to the dot plot approach
 - One sequence is placed down the side of a grid and another across the top
 - Instead of placing a dot in the grid, we **compute a score** for each position
 - Finding the optimal alignment corresponds to finding the path through the grid with the **best possible score**



Needleman, S.B. & Wunsch, C.D. (1970) "A general method applicable to the search for similarities in the amino acid sequences of two proteins." J. Mol. Biol. 48:443-453.

Algorithm of Needleman and Wunsch

- The Needleman–Wunsch approach to global sequence alignment has three basic steps:
 - (1) setting up a 2D-grid (or **alignment matrix**),
 - (2) **scoring the matrix**, and
 - (3) identifying the **optimal path** through the matrix



Needleman, S.B. & Wunsch, C.D. (1970) "A general method applicable to the search for similarities in the amino acid sequences of two proteins." J. Mol. Biol. 48:443-453.

Scoring the alignment matrix

- Start by filling in the first row and column – these are all indels (gaps).
 - Each step you take you will add the **gap penalty** to the score ($S_{i,j}$) accumulated in the previous cell

		j	Sequence 2				
		-	D	P	L	E	
i		-	0	-2	-4	-6	-8
D	-						
P							
M							
E							

Scores: match = +1, mismatch = -1, gap = -2

Scoring the alignment matrix

- Start by filling in the first row and column – these are all indels (gaps).
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		j	Sequence 2				
		-	D	P	L	E	
i		-	0	-2	-4	-6	-8
D	-	-2					
P	-	-4					
M	-	-6					
E	-	-8					

Scores: match = +1, mismatch = -1, gap = -2

$$S_{i+4} = (-2) + (-2) + (-2) + (-2)$$

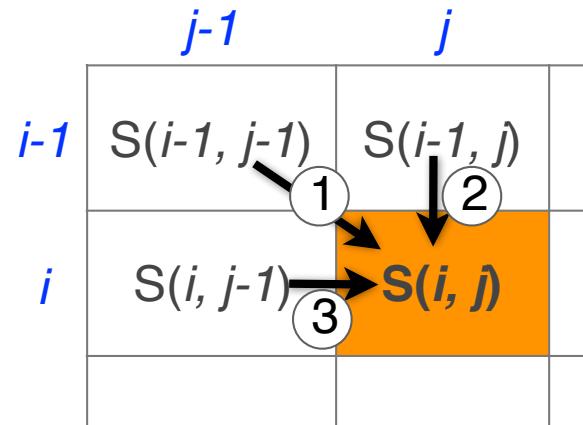
Seq1 : DPME
Seq2 : ----

Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
 - Now can ask which of the three directions gives the highest score?
 - keep track of this score and direction

	-	<i>j</i>	D	P	L	E
-	0	-2		-4	-6	-8
<i>i</i>	D	-2	?			
P	-4					
M	-6					
E	-8					

Scores: match = +1, mismatch = -1, gap = -2



Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
 - Now can ask which of the three directions gives the highest score?
 - keep track of this score and direction

		<i>j</i>	D	P	L	E	
		-	0	-2	-4	-6	-8
-	i	D	-2	?			
P		-4					
M		-6					
E		-8					

Scores: match = +1, mismatch = -1, gap = -2

$$S(i, j) = \text{Max} \left\{ \begin{array}{l} S(i-1, j-1) + (\text{mis})\text{match} \\ S(i-1, j) + \text{gap penalty} \\ S(i, j-1) + \text{gap penalty} \end{array} \right.$$

1
2
3

Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
 - Now can ask which direction gives the highest score
 - keep track of direction and score

	-	D	P	L	E
-	0	-2	-4	-6	-8
i	D	-2	1		
P	-4				
M	-6				
E	-8				

Scores: match = +1, mismatch = -1, gap = -2

① $(0) + (+1) = +1 \text{ <= (D-D) match!}$

Alignment

D
D

② $(-2) + (-2) = -4$

→ ③ $(-2) + (-2) = -4$

Scoring the alignment matrix

- At each step, the score in the current cell is determined by the scores in the neighboring cells
 - The maximal score and the direction that gave that score is stored (we will use these later to determine the optimal alignment)

		j				
		D	P	L	E	
-		-	-2	-4	-6	-8
i	D	-2	1 → -1			
P	-4					
M	-6					
E	-8					

Scores: match = +1, mismatch = -1, gap = -2

① $(-2)+(-1) = -3$ <= (D-P) mismatch!

Alignment

② $(-4)+(-2) = -6$

D-
DP

③ $(1)+(-2) = -1$

Scoring the alignment matrix

- We will continue to store the alignment score ($S_{i,j}$) for all possible alignments in the alignment matrix.

	-	D	P	L	j	E
-	0	-2	-4	-6	-8	
i	D	-2	1 → -1 → -3			
P		-4				
M		-6				
E		-8				

Scores: match = +1, mismatch = -1, gap = -2

① $(-4)+(-1) = -5 \leq (D-L)$ mismatch
Alignment

② $(-6)+(-2) = -8$
 $\begin{array}{l} D-- \\ DPL \end{array}$

③ $(-1)+(-2) = -3$

Scoring the alignment matrix

- For the highlighted cell, the corresponding score ($S_{i,j}$) refers to the score of the optimal alignment of the first i characters from sequence1, and the first j characters from sequence2.

		j				
		-	D	P	L	E
-		0	-2	-4	-6	-8
D	-2	1	-1	-3	-5	
P	-4	-1	2	0		
M	-6					
E	-8					

Scores: match = +1, mismatch = -1, indel = -2

- Alignment
DP-
DPL
- ① $(-1)+(-1) = -2$
 - ② $(-3)+(-2) = -5$
 - ③ $(2)+(-2) = 0$

Scoring the alignment matrix

- At each step, the score in the current cell is determined by the scores in the neighboring cells
 - The maximal score and the direction that gave that score is stored

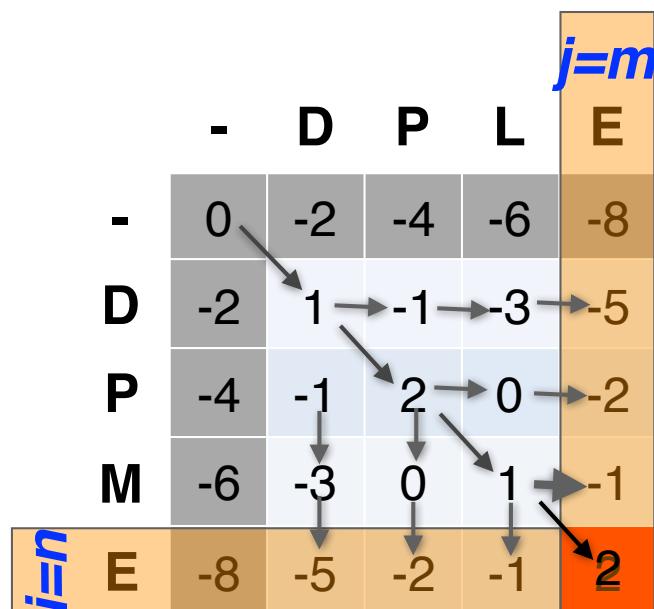
		j				
		-	D	P	L	E
-		0	-2	-4	-6	-8
D	-2	1	-1	-3	-5	
P	-4	-1	2	0	-2	
M	-6	-3	0	1		
E	-8					

Scores: match = +1, mismatch = -1, indel = -2

- ① $(2)+(-1) = 0 \leq \text{mismatch}$ Alignment
DPM
- ② $(0)+(-2) = -2$ DPL
- ③ $(0)+(-2) = -2$

Scoring the alignment matrix

- The score of the best alignment of the entire sequences corresponds to $S_{n,m}$
 - (where n and m are the length of the sequences)



Scores: match = +1, mismatch = -1, indel = -2

① $(+1) + (+1) = +2$

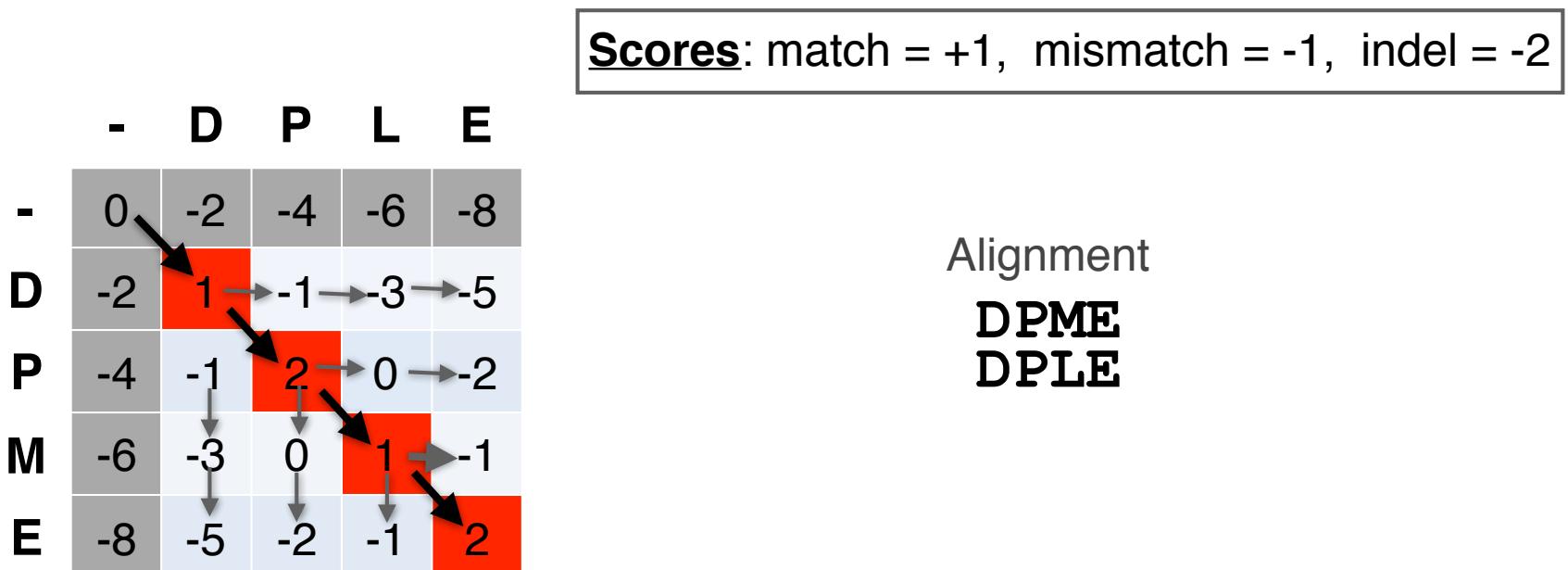
② $(-1) + (-2) = -3$

③ $(-1) + (-2) = -3$

Alignment
DPME
DPLE

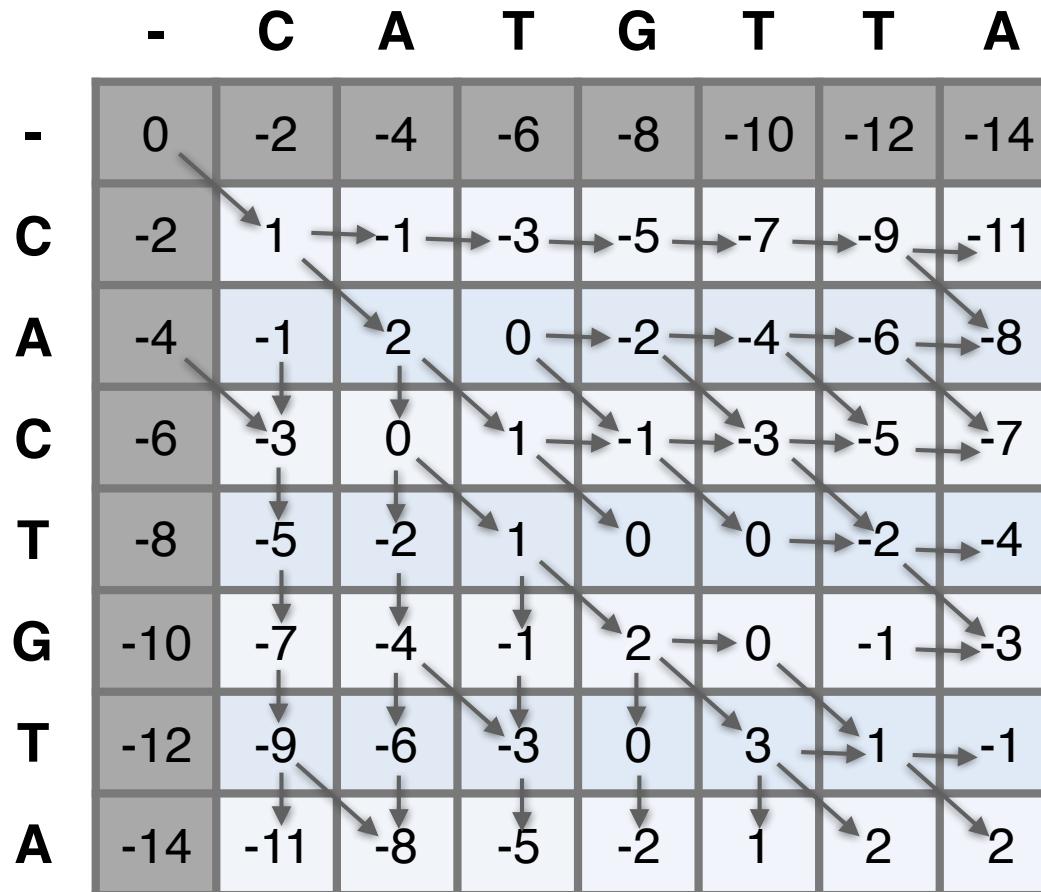
Scoring the alignment matrix

- To find the best alignment, we retrace the arrows starting from the bottom right cell
 - N.B. The optimal alignment score and alignment are dependent on the chosen scoring system



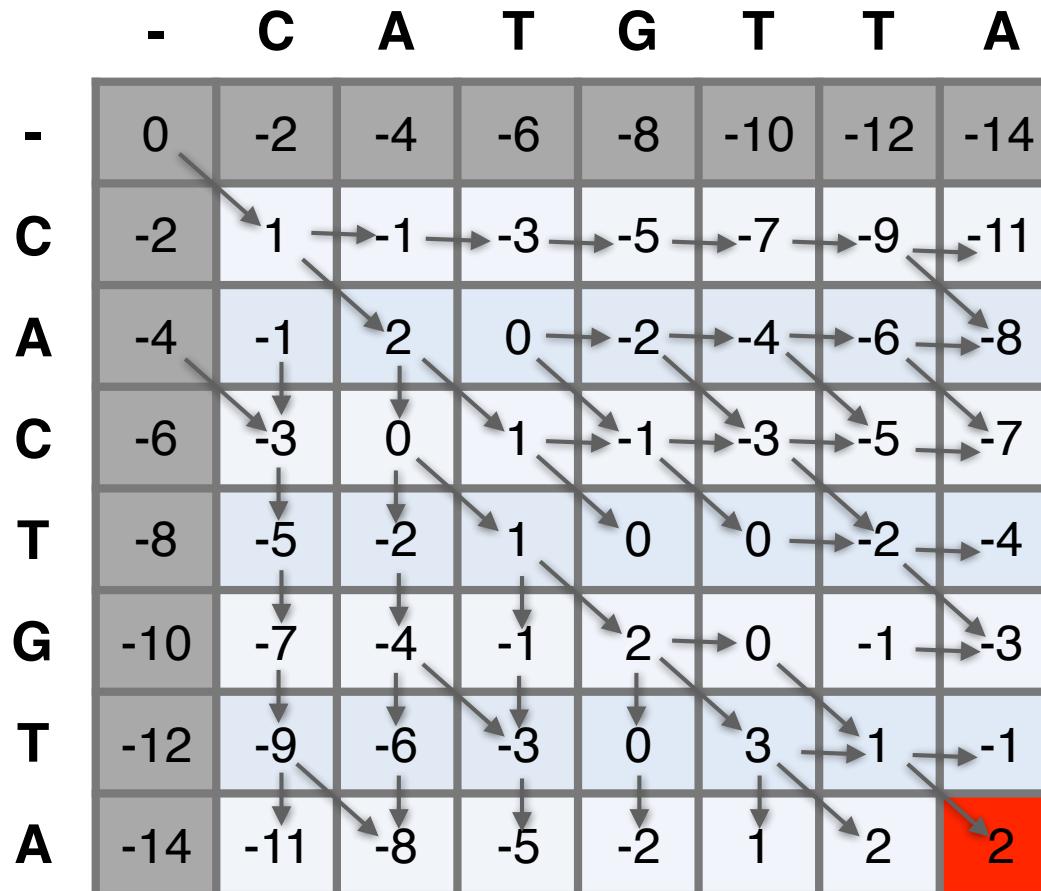
Questions:

- What is the optimal score for the alignment of these sequences and how do we find the optimal alignment?



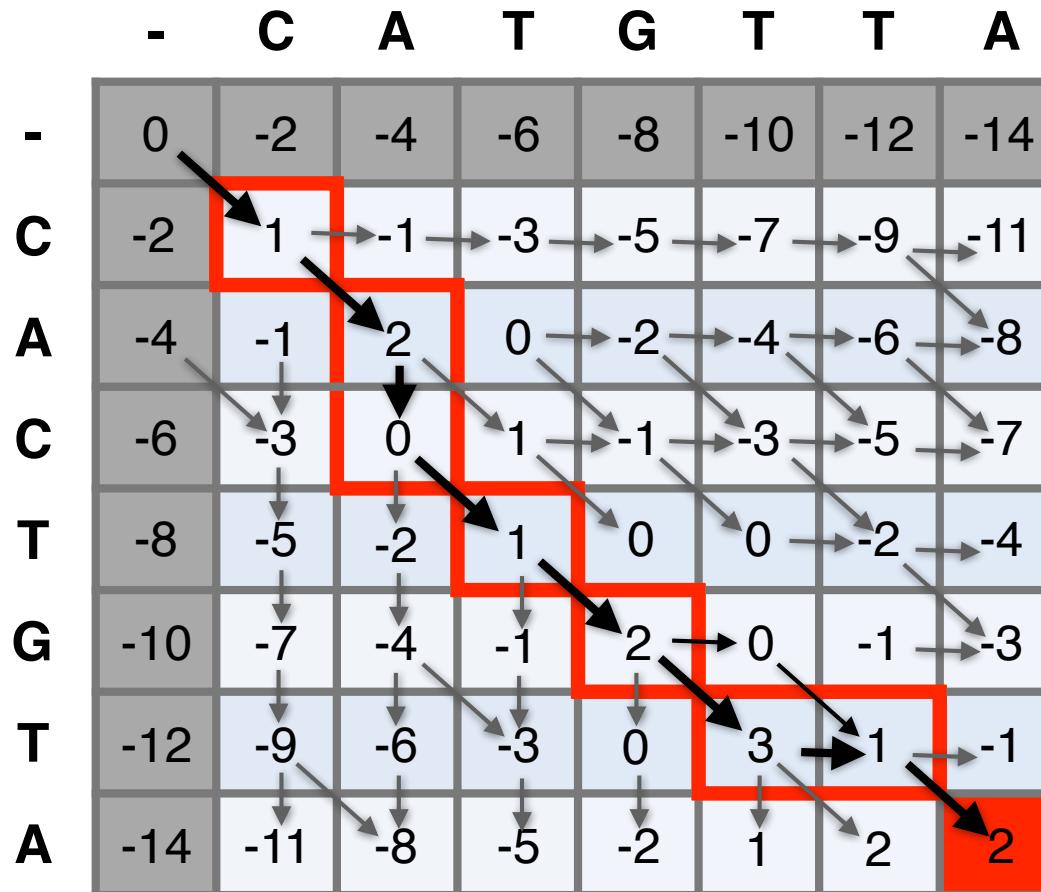
Questions:

- What is the optimal score for the alignment of these sequences and how do we find the optimal alignment?



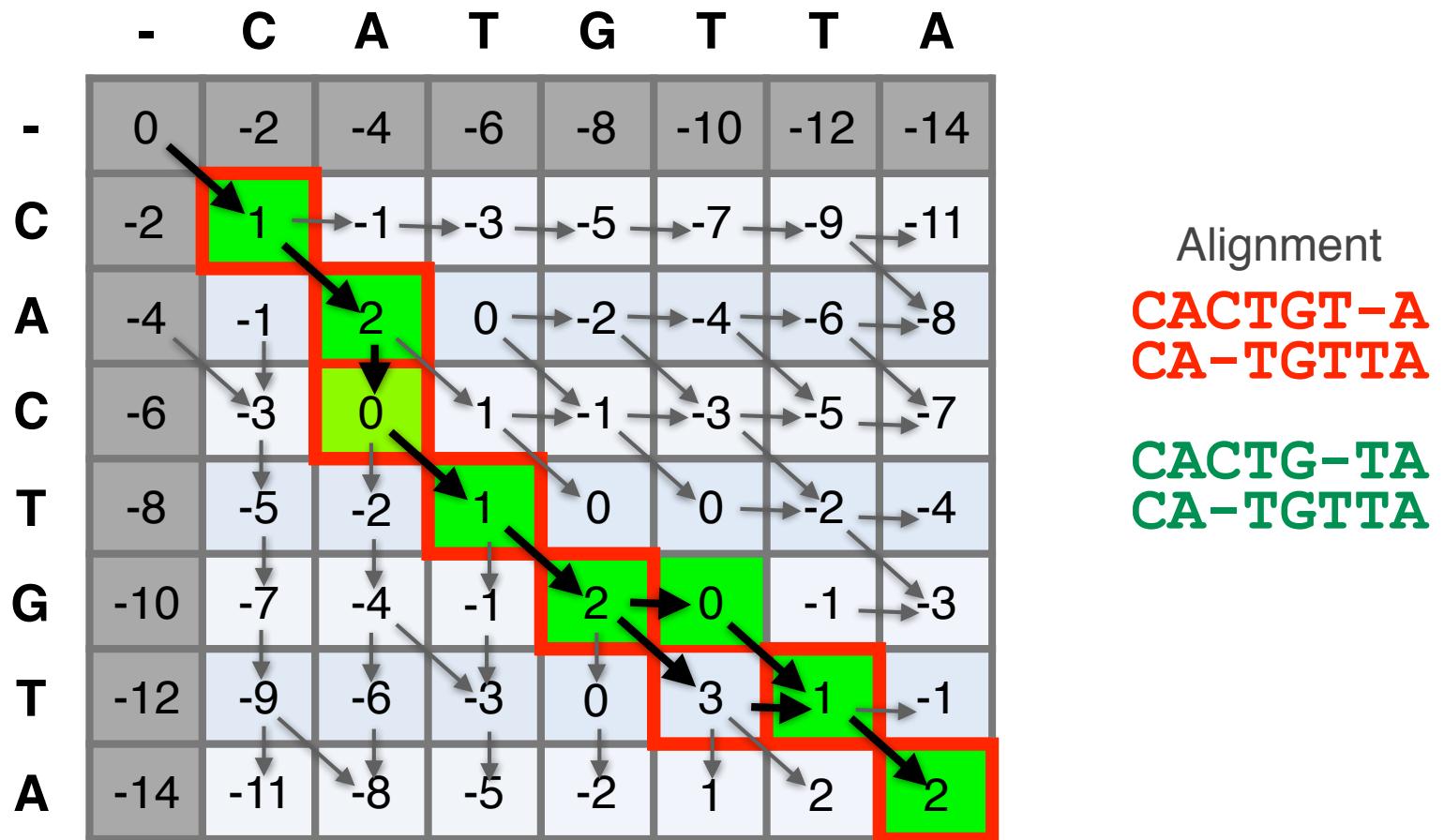
Questions:

- To find the best alignment we retrace the arrows starting from the bottom right cell



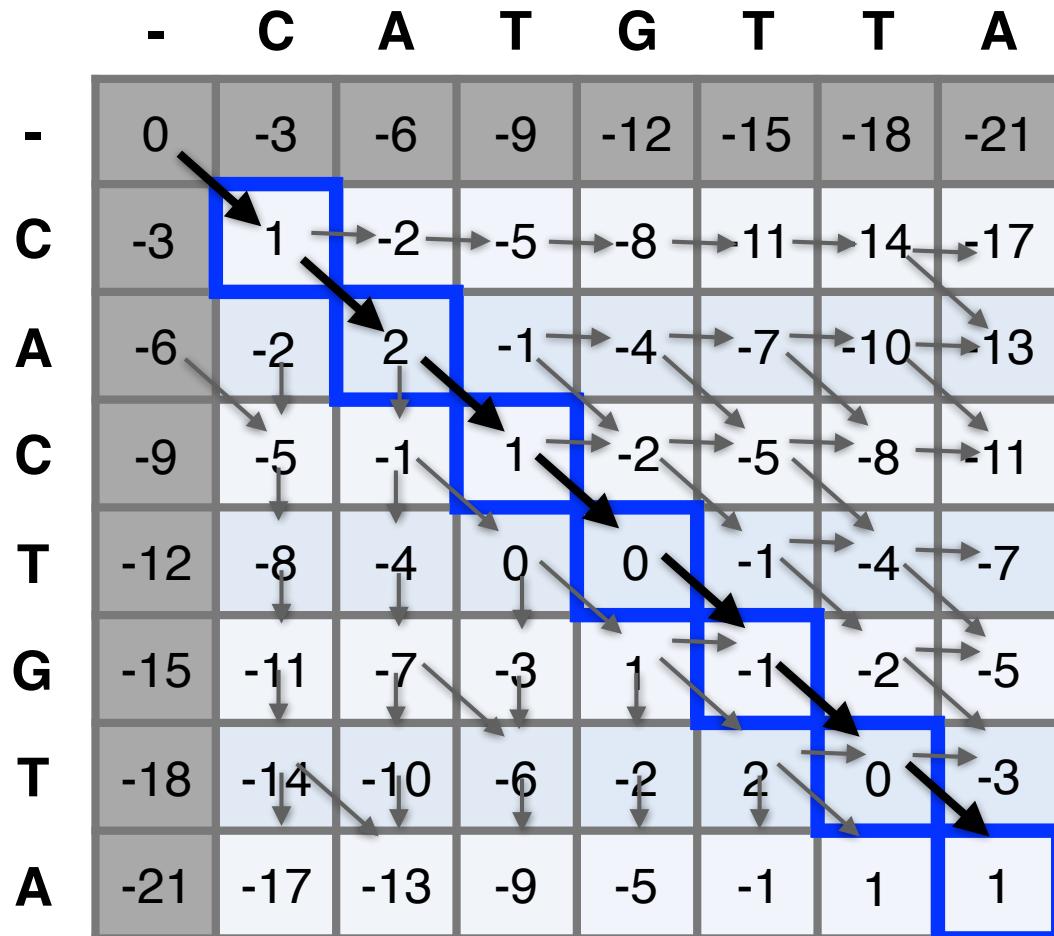
More than one alignment possible

- Sometimes more than one alignment can result in the same optimal score



The alignment and score are dependent on the scoring system

- Here we increase the gap penalty from -2 to -3



Alignment

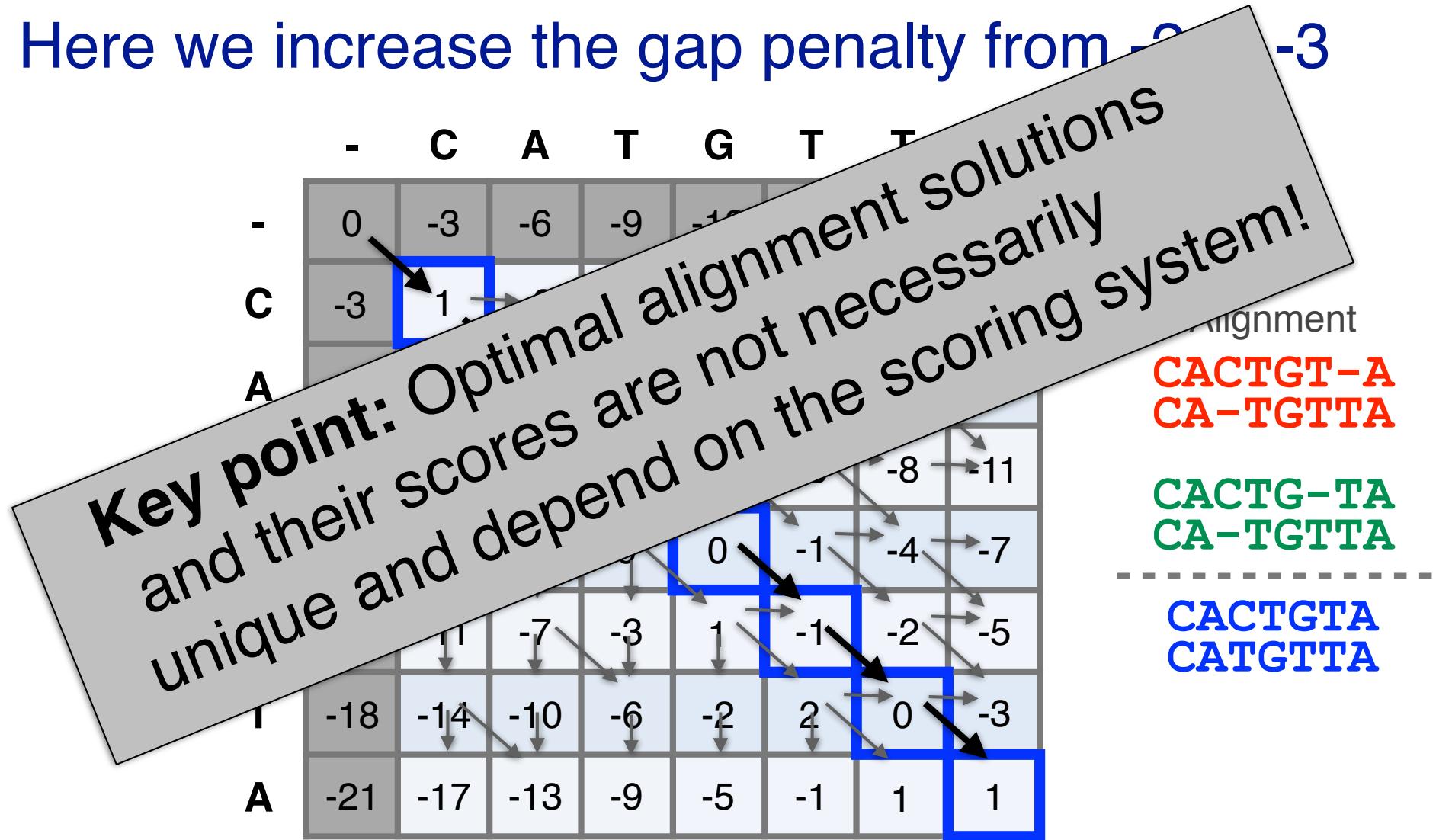
CACTGT-A
CA-TGTTA

CACTG-TA
CA-TGTTA

CACTGTA
CATGTTA

The alignment and score are dependent on the scoring system

- Here we increase the gap penalty from -2 to -3



Your Turn!

Hands-on worksheet **Sections 2 & 3**

Match: +2

Mismatch: -1

Gap: -2

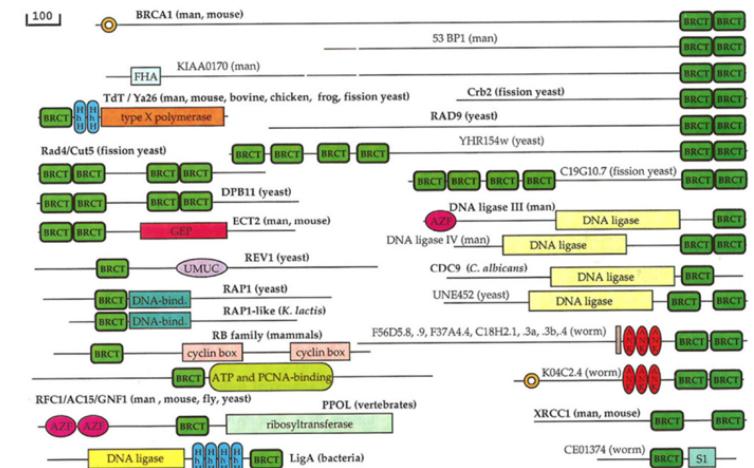
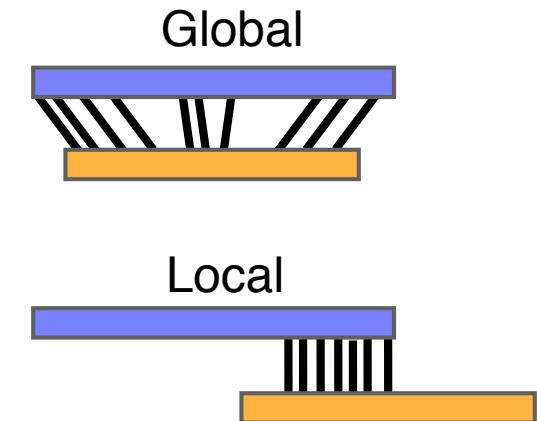
	A	G	T	T	C
0					
A					
T					
T					
G					
C					

ALIGNMENT FOUNDATIONS

- **Why...**
 - Why compare biological sequences?
- **What...**
 - Alignment view of sequence changes during evolution
(matches, mismatches and gaps)
- **How...**
 - ▶ Dot matrices
 - ▶ Dynamic programming
 - Global alignment
 - Local alignment
 - ▶ BLAST heuristic approach

Global vs local alignments

- Needleman-Wunsch is a **global alignment** algorithm
 - Resulting alignment spans the complete sequences end to end
 - This is appropriate for closely related sequences that are similar in length
- For many practical applications we require **local alignments**
 - Local alignments highlight sub-regions (e.g. protein domains) in the two sequences that align well



Local alignment: Definition

- Smith & Waterman proposed simply that a local alignment of two sequences allow arbitrary-length segments of each sequence to be aligned, with no penalty for the unaligned portions of the sequences. Otherwise, the score for a local alignment is calculated the same way as that for a global alignment

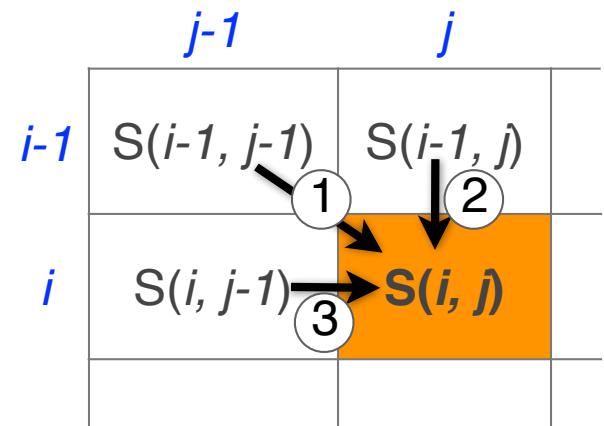
Smith, T.F. & Waterman, M.S. (1981) “Identification of common molecular subsequences.” J. Mol. Biol. 147:195-197.

The Smith-Waterman algorithm

- Three main modifications to Needleman-Wunsch:
 - Allow a node to start at 0
 - The score for a particular cell cannot be negative
 - if all other score options produce a negative value, then a zero must be inserted in the cell
 - Record the highest- scoring node, and trace back from there

$$S(i, j) = \text{Max} \left\{ \begin{array}{l} S(i-1, j-1) + (\text{mis})\text{match} \\ S(i-1, j) - \text{gap penalty} \\ S(i, j-1) - \text{gap penalty} \\ 0 \end{array} \right.$$

(1)
(2)
(3)
(4)



Sequence 1

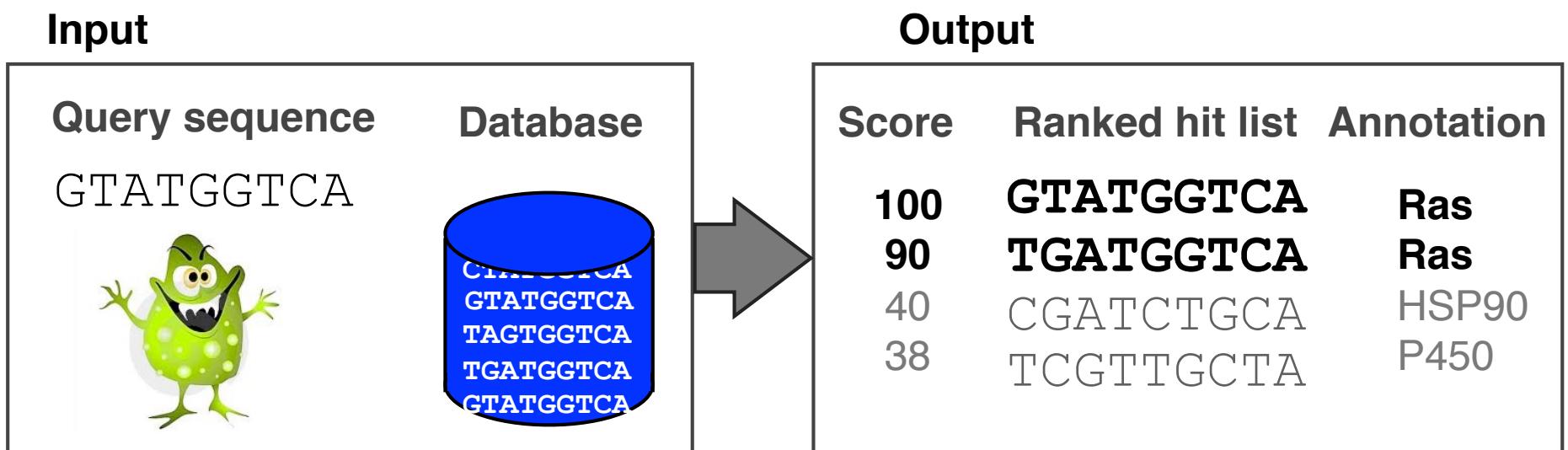
	-	C	A	G	C	C	U	C	G	C	U	U	A	G
-	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
A	0.0	0.0	1.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	1.0	0.0
A	0.0	0.0	1.0	0.7	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	1.0	0.7
U	0.0	0.0	0.0	0.7	0.3	0.0	1.0	0.0	0.0	0.0	1.0	1.0	0.0	0.7
G	0.0	0.0	0.0	1.0	0.3	0.0	0.0	0.7	1.0	0.0	0.0	0.7	0.7	1.0
C	0.0	1.0	0.0	0.0	2.0	1.3	0.3	1.0	0.3	2.0	0.7	0.3	0.3	0.3
C	0.0	1.0	0.7	0.0	1.0	3.0	1.7	1.3	1.0	1.3	1.7	0.3	0.0	0.0
A	0.0	0.0	2.0	0.7	0.3	1.7	2.7	1.3	1.0	0.7	1.0	1.3	1.3	0.0
U	0.0	0.0	0.7	1.7	0.3	1.3	2.7	2.3	1.0	0.7	1.7	2.0	1.0	1.0
U	0.0	0.0	0.3	0.3	1.3	1.0	2.3	2.3	2.0	0.7	1.7	2.7	1.7	1.0
G	0.0	0.0	0.0	1.3	0.0	1.0	1.0	2.0	3.3	2.0	1.7	1.3	2.3	2.7
A	0.0	0.0	1.0	0.0	1.0	0.3	0.7	0.7	2.0	3.0	1.7	1.3	2.3	2.0
C	0.0	1.0	0.0	0.7	1.0	2.0	0.7	1.7	1.7	3.0	2.7	1.3	1.0	2.0
G	0.0	0.0	0.7	1.0	0.3	0.7	1.7	0.3	2.7	1.7	2.7	2.3	1.0	2.0
G	0.0	0.0	0.0	1.7	0.7	0.3	0.3	1.3	1.3	2.3	1.3	2.3	2.0	2.0

Local alignment

GCC-AUG
GCCUCGC

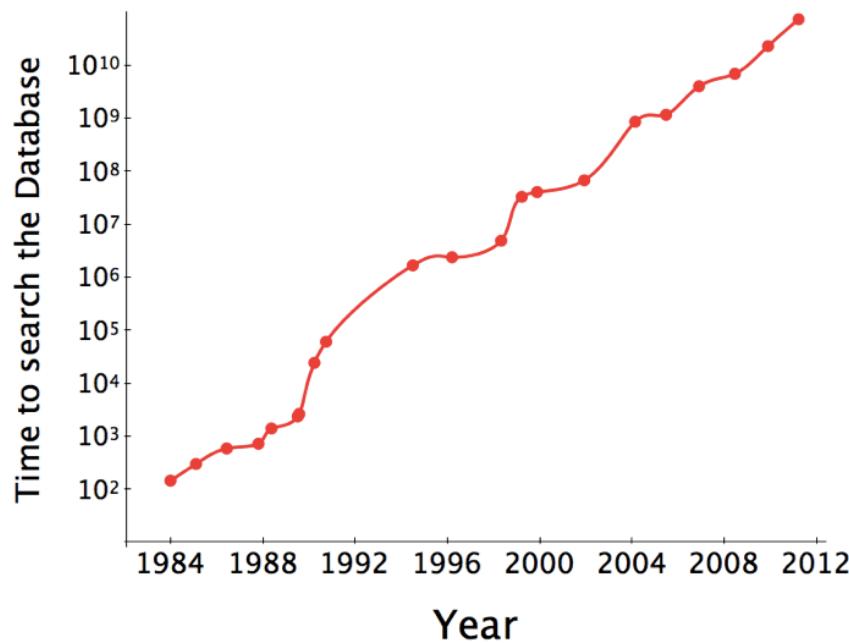
Local alignments can be used for database searching

- **Goal:** Given a query sequence (Q) and a sequence database (D), find a list of sequences from D that are most similar to Q
 - **Input:** Q, D and scoring scheme
 - **Output:** Ranked list of hits



The database search problem

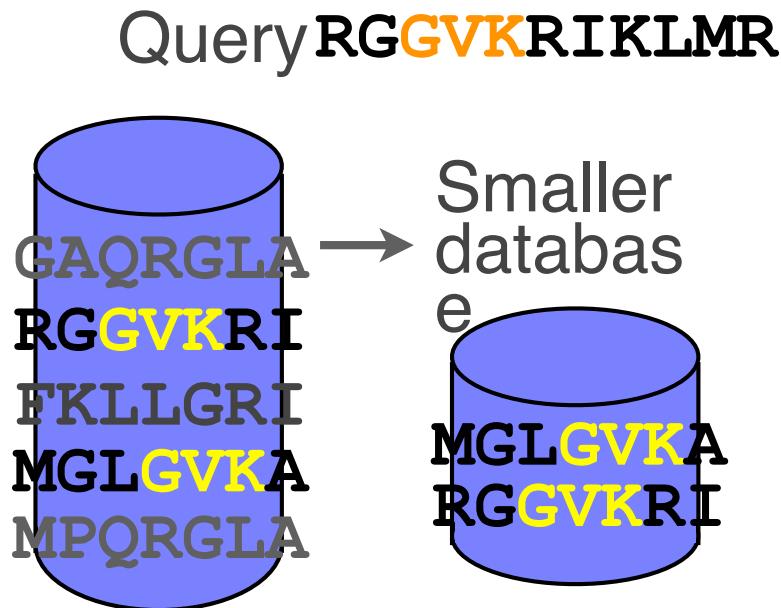
- Due to the rapid growth of sequence databases, search algorithms have to be both efficient and sensitive
 - Time to search with SW is proportional to $m \times n$ (m is length of query, n is length of database), **too slow for large databases!**



To reduce search time **heuristic algorithms**, such as BLAST, first remove database sequences without a strong local similarity to the query sequence in a quick initial scan.

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To reduce search time **heuristic algorithms**, such as BLAST, first remove database sequences without a strong local similarity to the query sequence in a quick initial scan.

ALIGNMENT FOUNDATIONS

- **Why...**
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 - Local alignment
 - ▶ BLAST heuristic approach

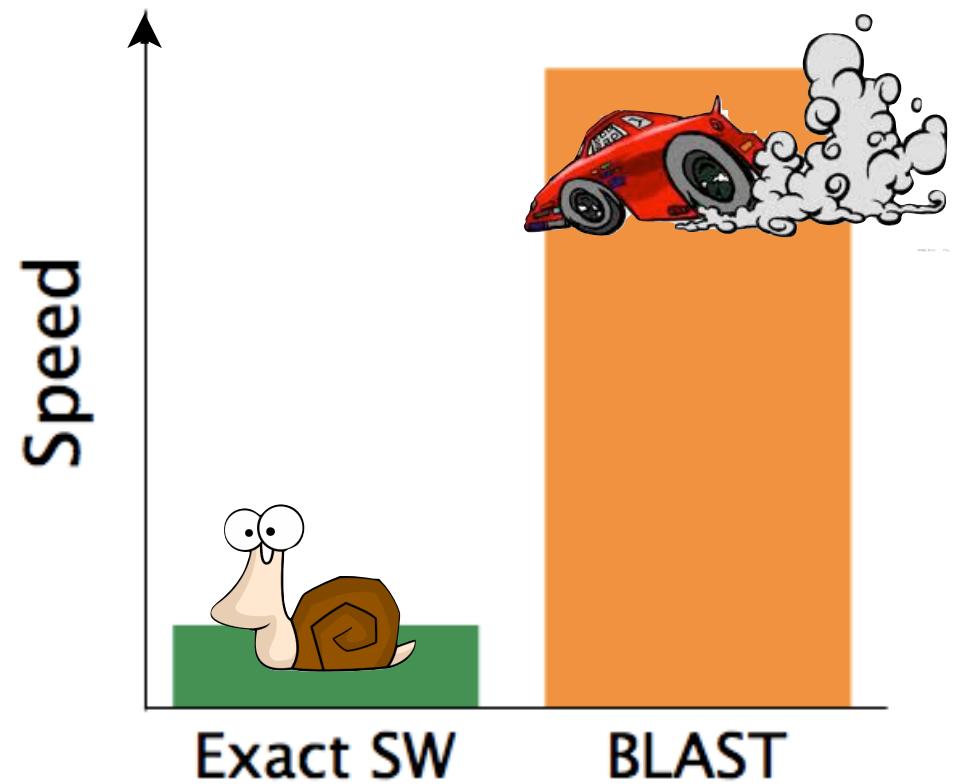
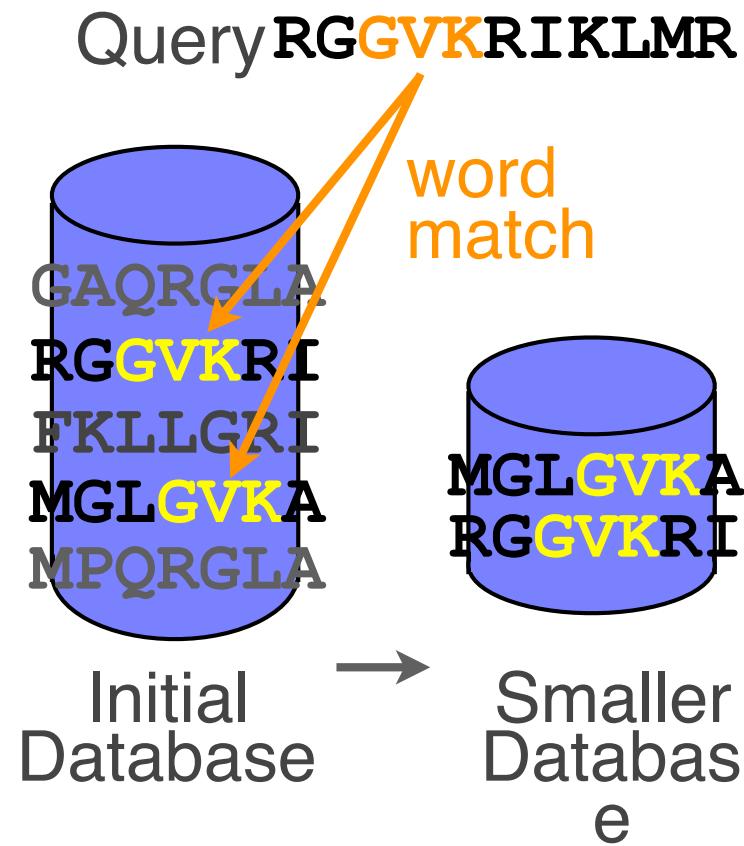
Rapid, heuristic versions of Smith–Waterman: **BLAST**

- BLAST (Basic Local Alignment Search Tool) is a simplified form of Smith-Waterman (SW) alignment that is popular because it is **fast** and **easily accessible**
 - BLAST is a heuristic approximation to SW - It examines only part of the search space
 - BLAST saves time by restricting the search by scanning database sequences for likely matches before performing more rigorous alignments
 - Sacrifices some sensitivity in exchange for speed
 - In contrast to SW, BLAST is not guaranteed to find optimal alignments

Rapid, heuristic versions of Smith–Waterman: BLAST

- BLAST (Basic Local Alignment Search Tool) is a simplified form of Smith-Waterman (SW). It is popular because it is **fast**.
 - BLAST finds regions of similarity between two sequences
 - BLAST does not find all matches, but it finds many matches by scanning for local alignments
- “The central idea of the BLAST algorithm is to confine attention to sequence pairs that contain an initial **word pair match**”
Altschul et al. (1990)
- The sensitivity in exchange for speed
- In contrast to SW, BLAST is not guaranteed to find optimal alignments

- BLAST uses this pre-screening heuristic approximation resulting in an approach that is about 50 times faster than the Smith-Waterman



How BLAST works

- Four basic phases
 - Phase 1: compile a list of query word pairs ($w=3$)

generate list
of $w=3$
words for
query

RGGVKRI Query sequence

RGG

GGV

GVK

VKR

KRI

Blast

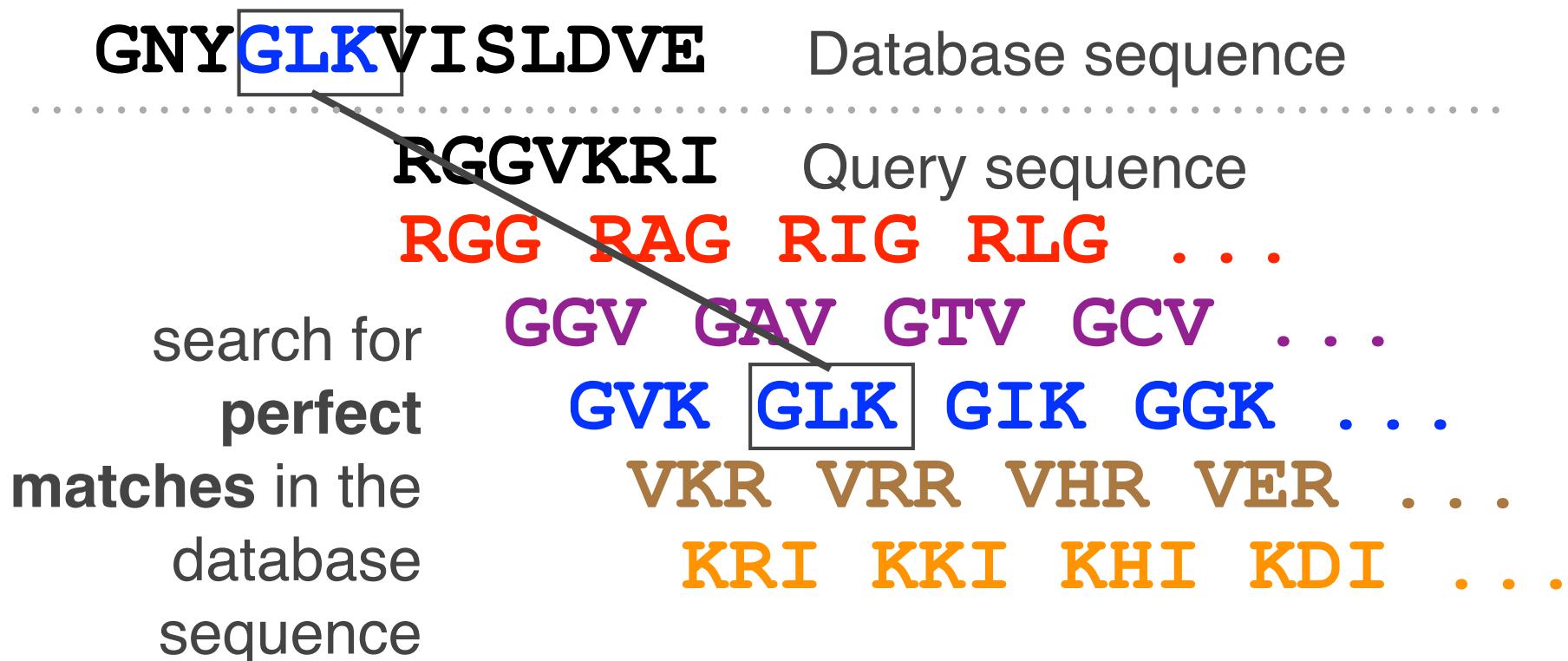
- Phase 2: expand word pairs to include those similar to query (defined as those above a similarity threshold to original word, i.e. match scores in substitution matrix)

extend list of
words similar
to query

RGGVKRI Query sequence
RGG RAG RIG RLG . . .
GGV GAV GTV GCV . . .
GVK GAK GIK GGK . . .
VKR VRR VHR VER . . .
KRI KKI KHI KDI . . .

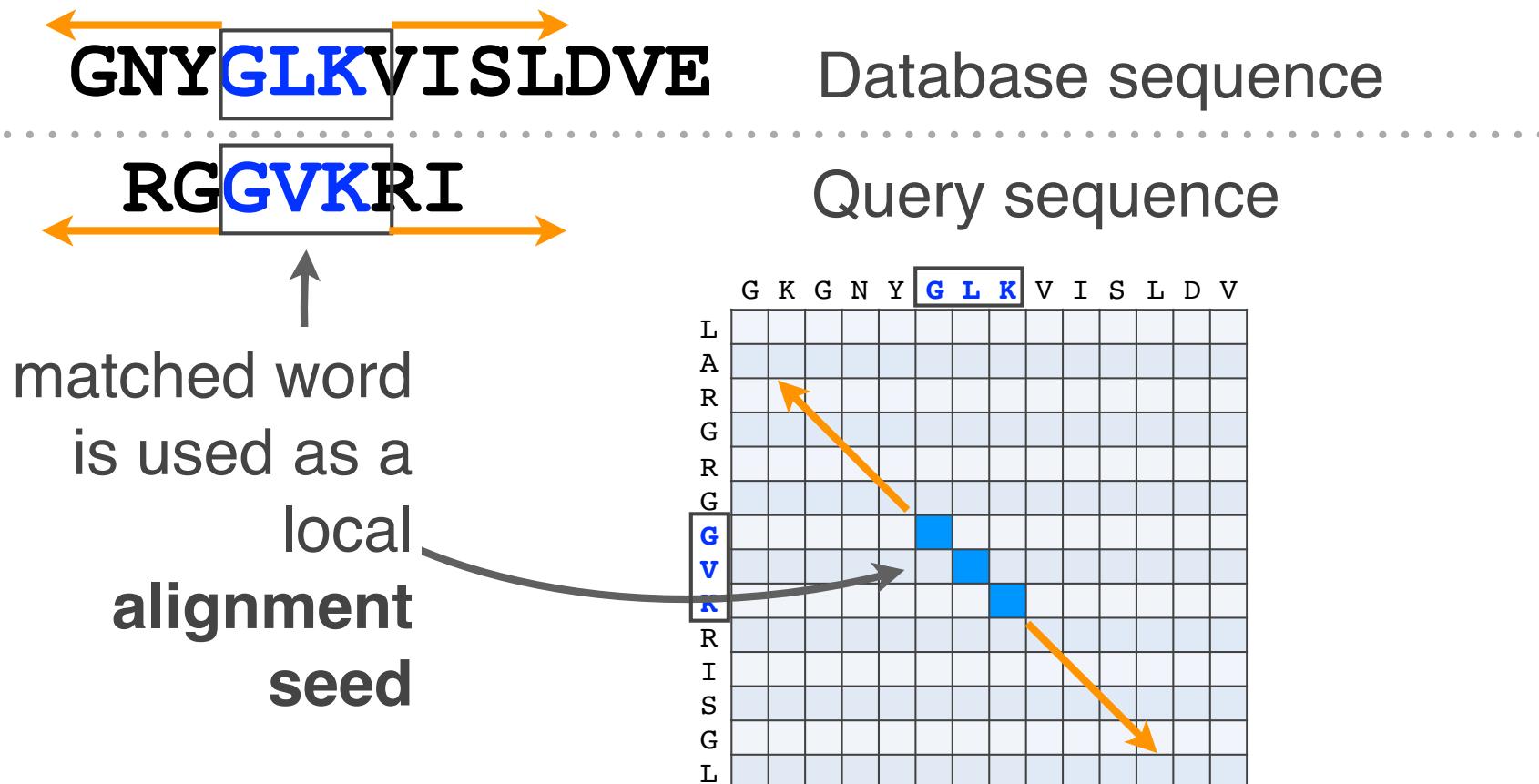
Blast

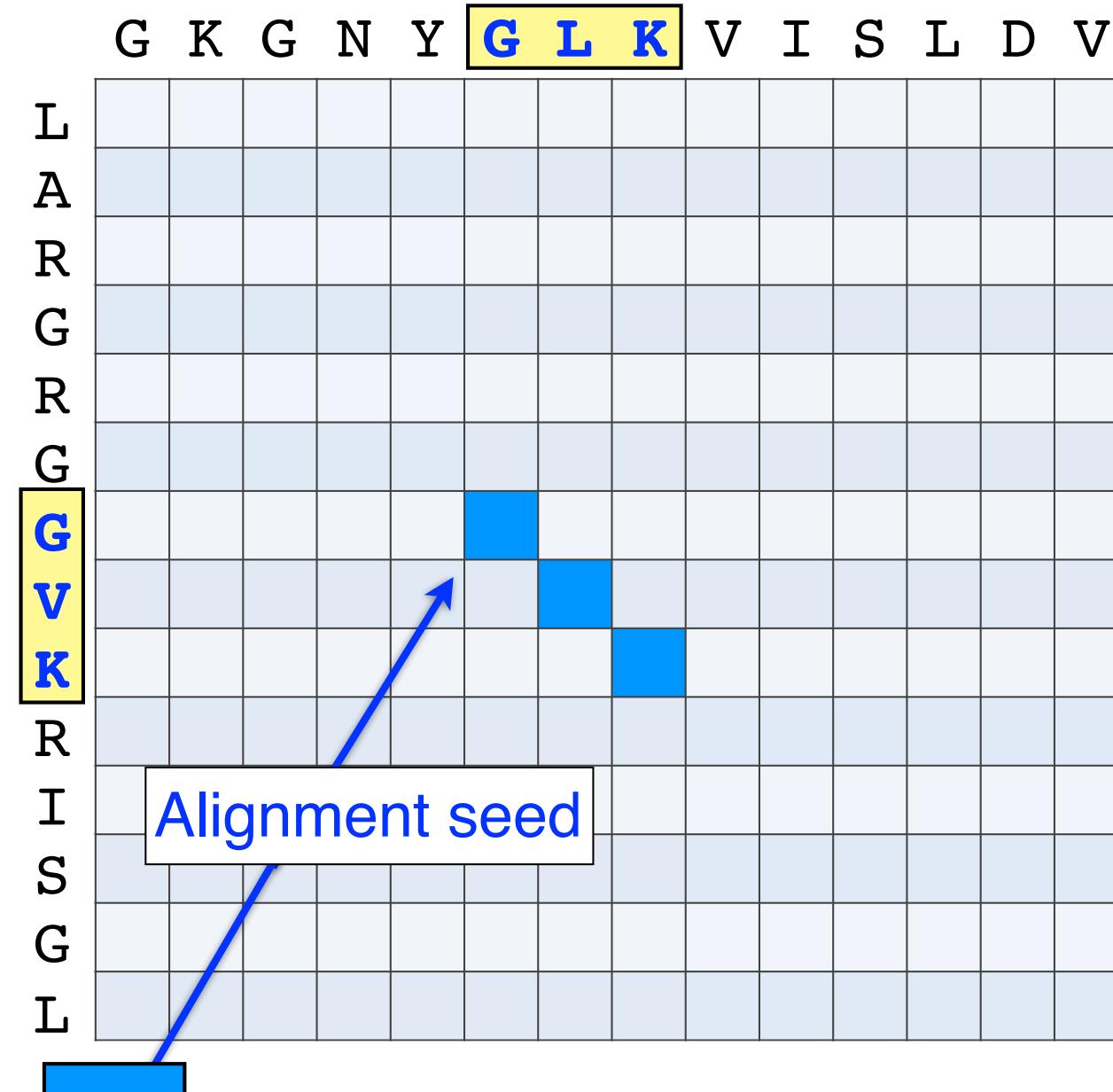
- Phase 3: a database is scanned to find sequence entries that match the compiled word list



Blast

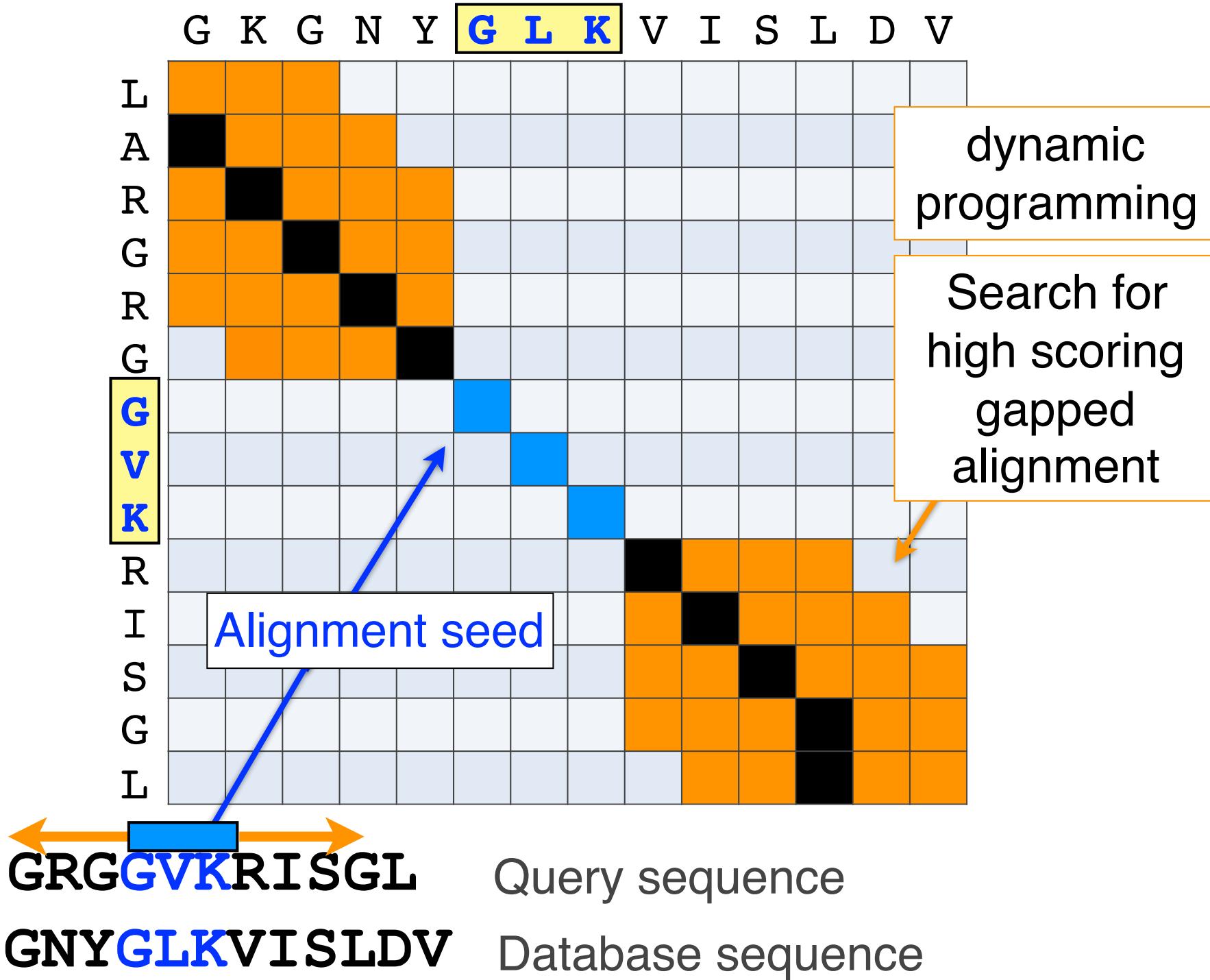
- Phase 4: the initial database hits are extended in both directions using dynamic programming

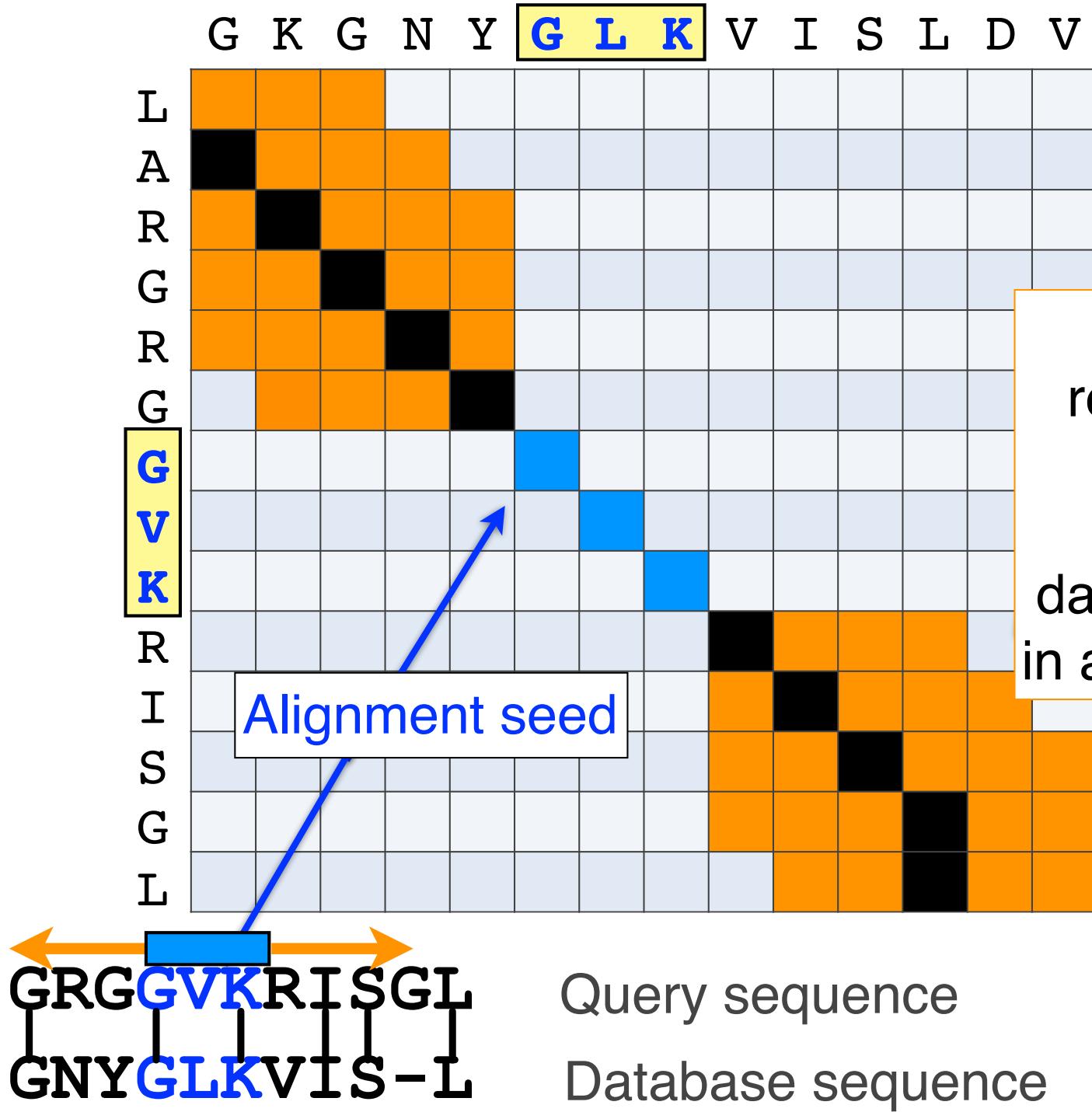




GRGGVK**RISGL** Query sequence

GNYGLK**VISLDV** Database sequence





BLAST returns the highest scoring database hits in a ranked list

BLAST output

- BLAST returns the highest scoring database hits in a ranked list along with details about the target sequence and alignment statistics

Description	Max score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	100%	0	98%	AAA20133.1
Kinesin-14 heavy chain [Danio rerio]	595	88%	0	78%	XP_00320703
hypothetical protein EGK_18589	48.2	40%	0.03	32%	ELK35081.1
mKIAA4102 protein [Mus musculus]	42.7	38%	3.02	24%	EHH28205.1

Statistical significance of results

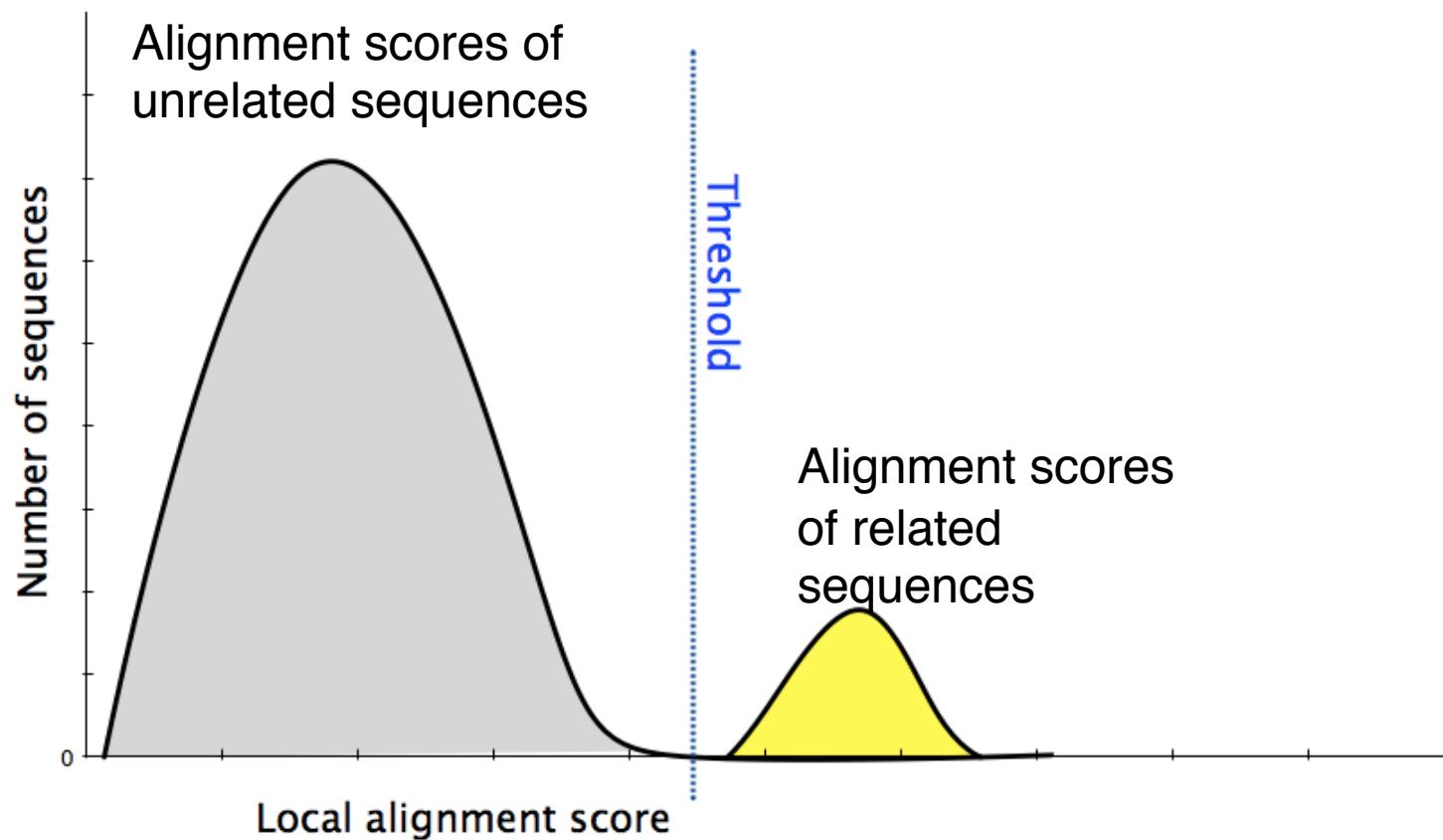
- An important feature of BLAST is the computation of statistical significance for each hit. This is described by the **E value** (expect value)

Description	Max score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	100%	0	100%	NP_004512.1
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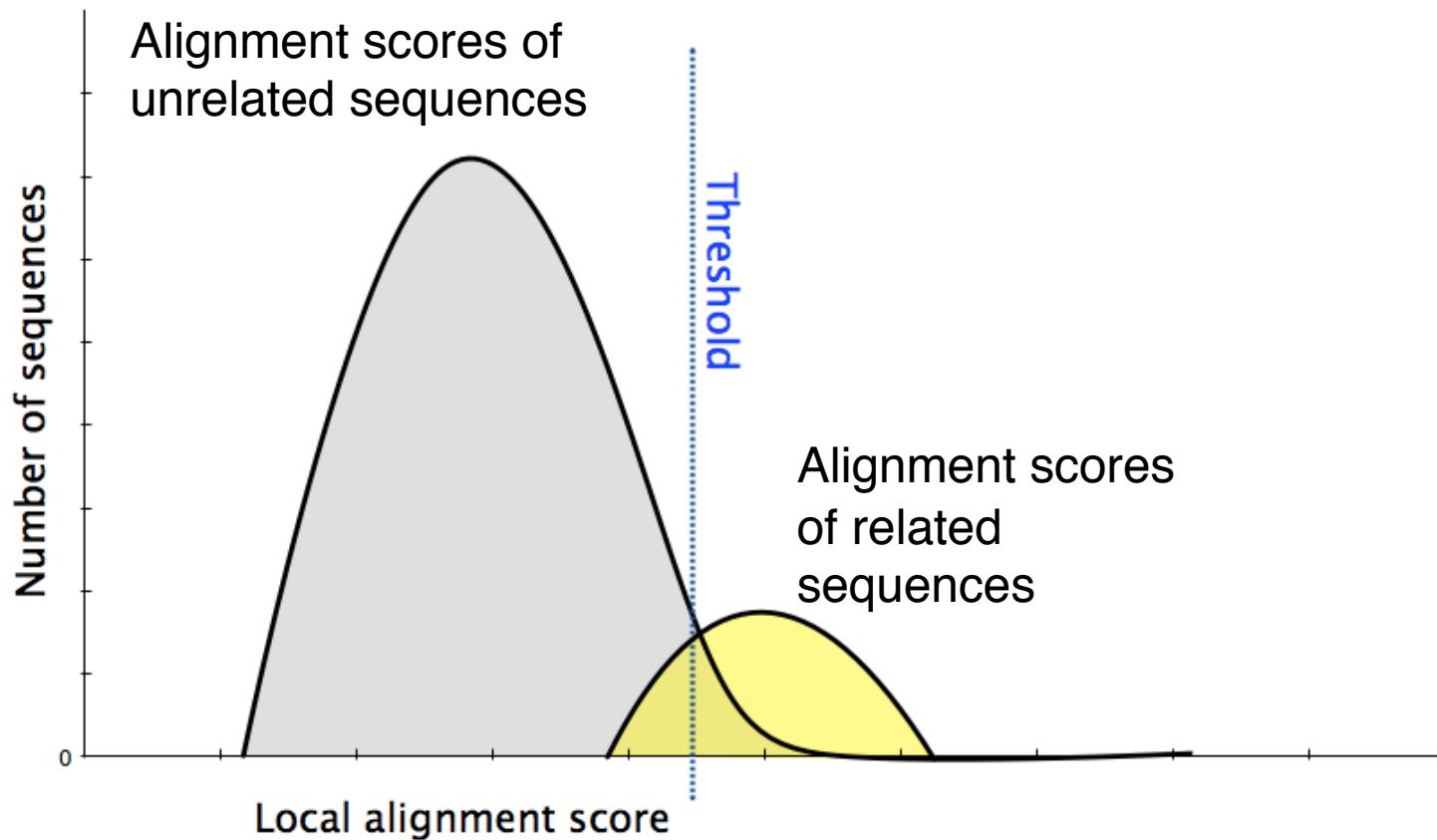
BLAST scores and E-values

- The **E value** is the **expected** number of hits that are as good or better than the observed local alignment score (with this score or better) if the query and database are **random** with respect to each other
 - *i.e.* the number of alignments expected to occur by chance with equivalent or better scores
- Typically, only hits with E value **below** a significance threshold are reported
 - This is equivalent to selecting alignments with score above a certain score threshold

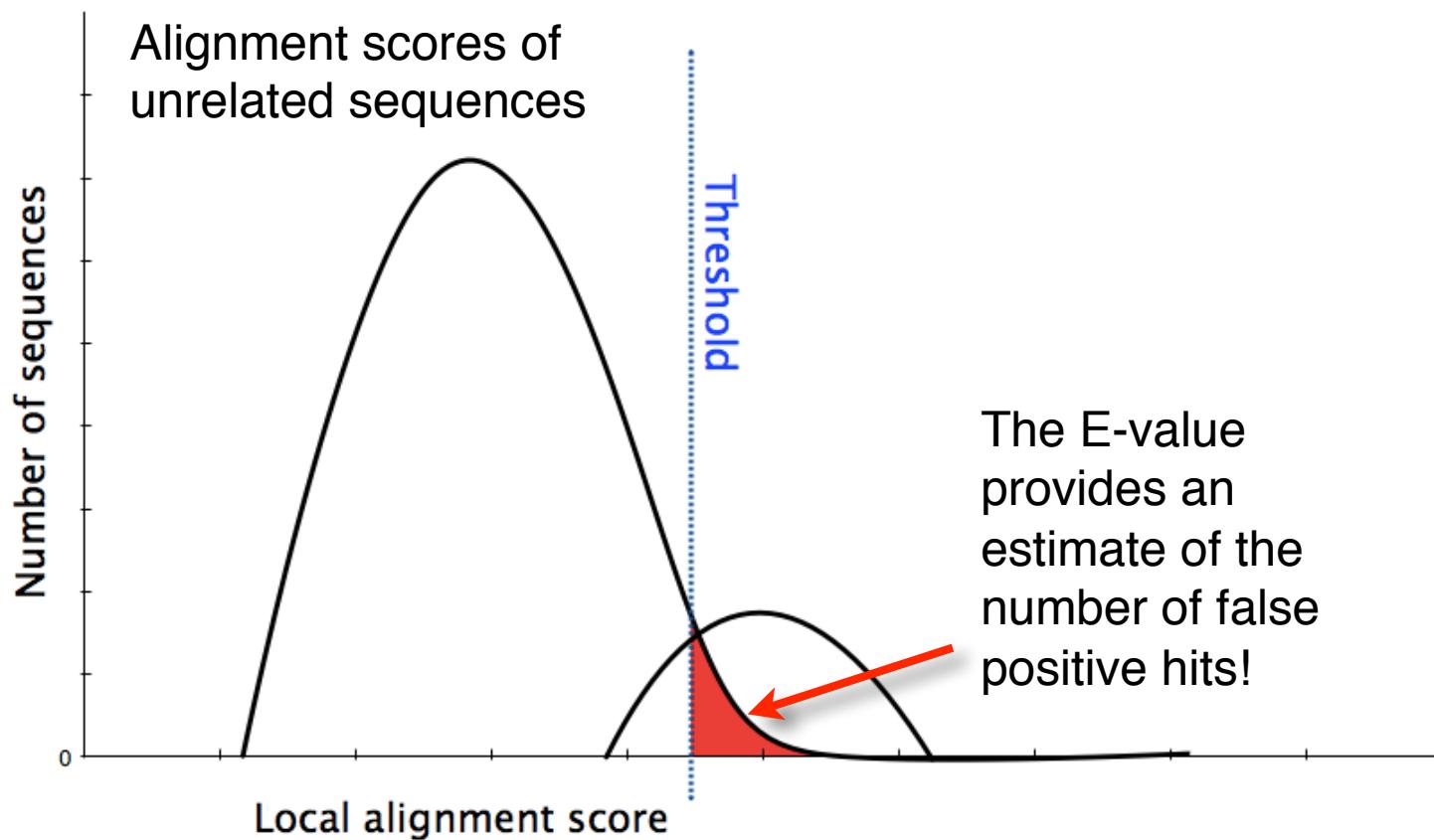
- Ideally, a threshold separates all query related sequences (yellow) from all unrelated sequences (gray)



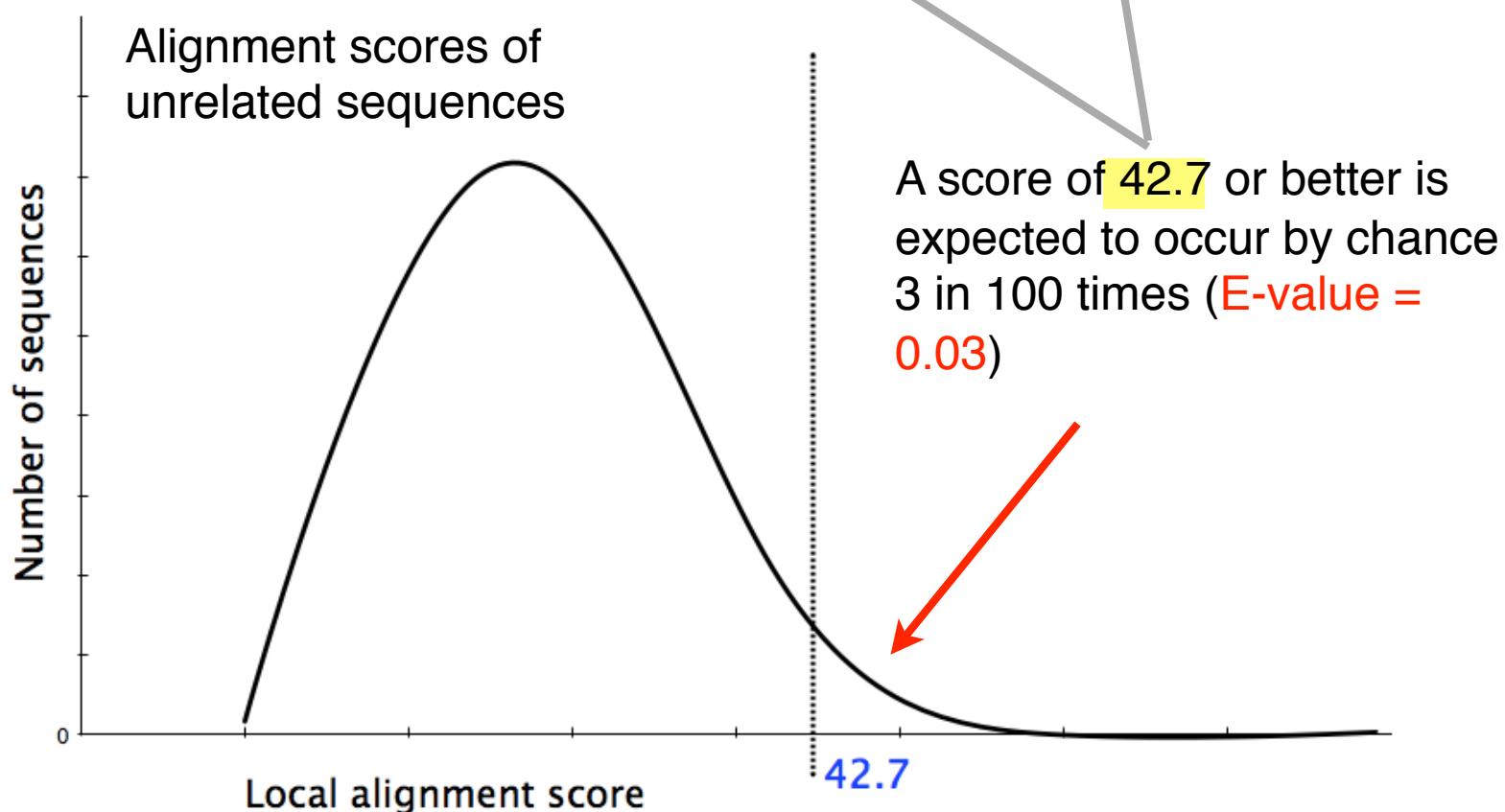
- Unfortunately, often both score distributions overlap
 - The E value describes the expected number of hits with a score above the threshold if the query and database are unrelated



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 - The E value describes the expected number of hits with a score above the threshold if the query and database are unrelated



Description	Max score	Query cover	E value	Max ident	Accession
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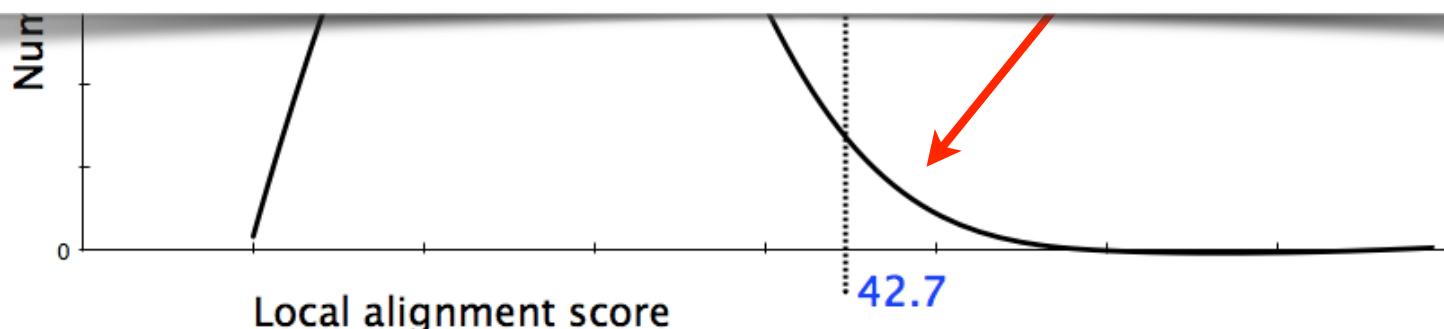


Description	Max score	Total score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo	677	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	676	100%	0	98%	AAA2013.3 1

In general E values < 0.005 are usually significant.

To find out more about E values see: “*The Statistics of Sequence Similarity Scores*” available in the help section of the NCBI BLAST site:

<http://www.ncbi.nlm.nih.gov/blast/tutorial/Altschul-1.html>

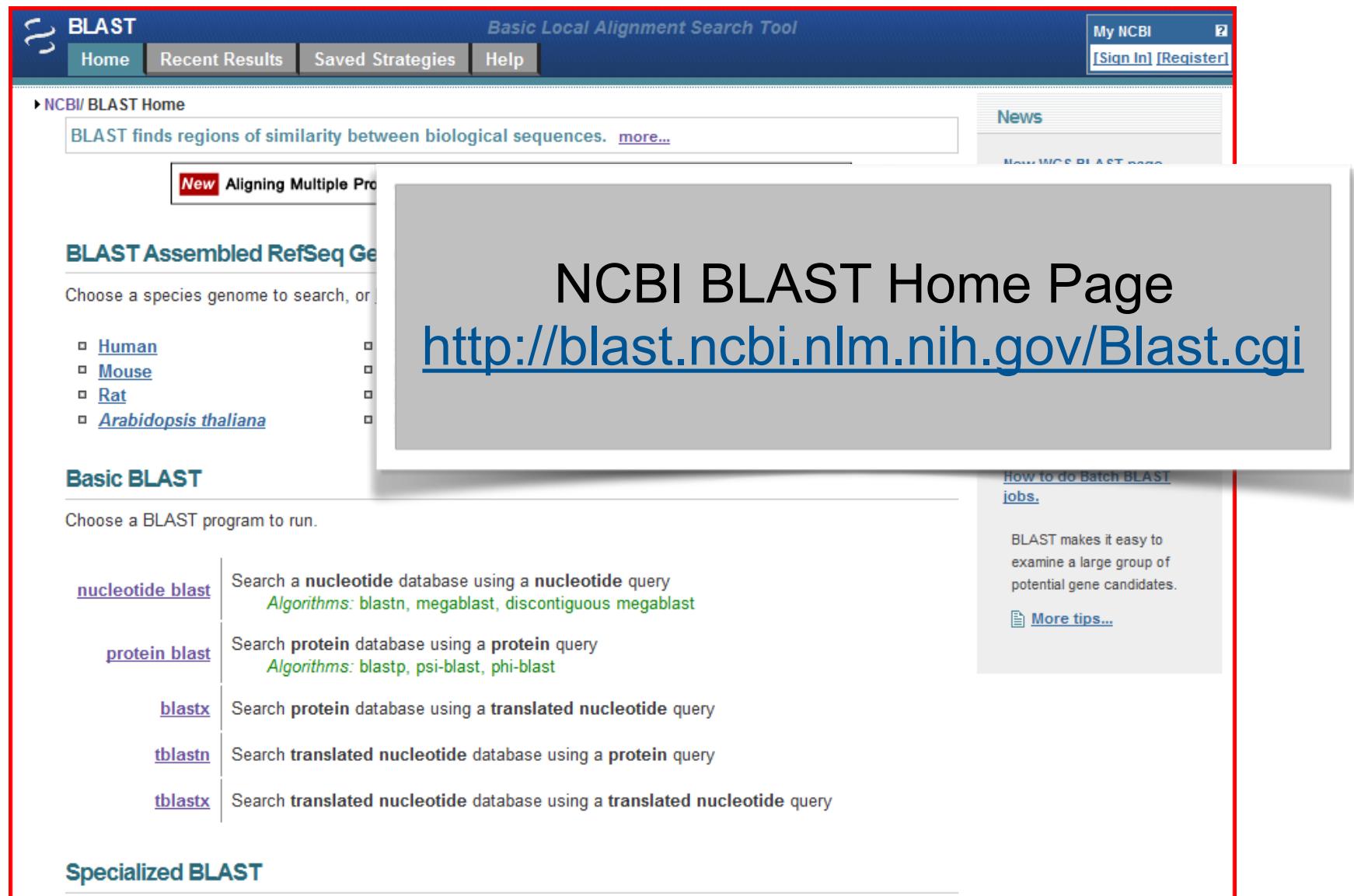


Your Turn!

Hands-on worksheet **Sections 4 & 5**

- Please do answer the last lab review question (Q19).
- We encourage discussion and exploration!

Practical database searching with BLAST



The screenshot shows the NCBI BLAST Home Page. The top navigation bar includes links for Home, Recent Results, Saved Strategies, Help, My NCBI, Sign In, and Register. A banner at the top states "Basic Local Alignment Search Tool". The main content area features a large central box with the text "NCBI BLAST Home Page" and the URL "http://blast.ncbi.nlm.nih.gov/Blast.cgi". To the left, there are sections for "BLAST Assembled RefSeq Genomes" (with a list of species: Human, Mouse, Rat, Arabidopsis thaliana) and "Basic BLAST" (with options for nucleotide blast, protein blast, blastx, tblastn, and tblastx). To the right, there is a sidebar with a "How to do Batch BLAST jobs" section and a "More tips..." link.

NCBI BLAST Home Page
<http://blast.ncbi.nlm.nih.gov/Blast.cgi>

BLAST Assembled RefSeq Genomes

Choose a species genome to search, or...
Human
Mouse
Rat
Arabidopsis thaliana

Basic BLAST

Choose a BLAST program to run.

nucleotide blast Search a nucleotide database using a nucleotide query
Algorithms: blastn, megablast, discontiguous megablast

protein blast Search protein database using a protein query
Algorithms: blastp, psi-blast, phi-blast

blastx Search protein database using a translated nucleotide query

tblastn Search translated nucleotide database using a protein query

tblastx Search translated nucleotide database using a translated nucleotide query

Specialized BLAST

How to do Batch BLAST jobs.

BLAST makes it easy to examine a large group of potential gene candidates.
[More tips...](#)

Practical database searching with BLAST

- There are four basic components to a traditional BLAST search
 - (1) Choose the sequence (query)
 - (2) Select the BLAST program
 - (3) Choose the database to search
 - (4) Choose optional parameters
- Then click “BLAST”

Step 1: Choose your sequence

- Sequence can be input in FASTA format or as accession number

NCBI Resources How To My N

Protein Translations of Life

Search: Protein Limits Advanced search Help

Display Settings FASTA Send to: Change region shown

hemoglobin subunit beta [Homo sapiens]

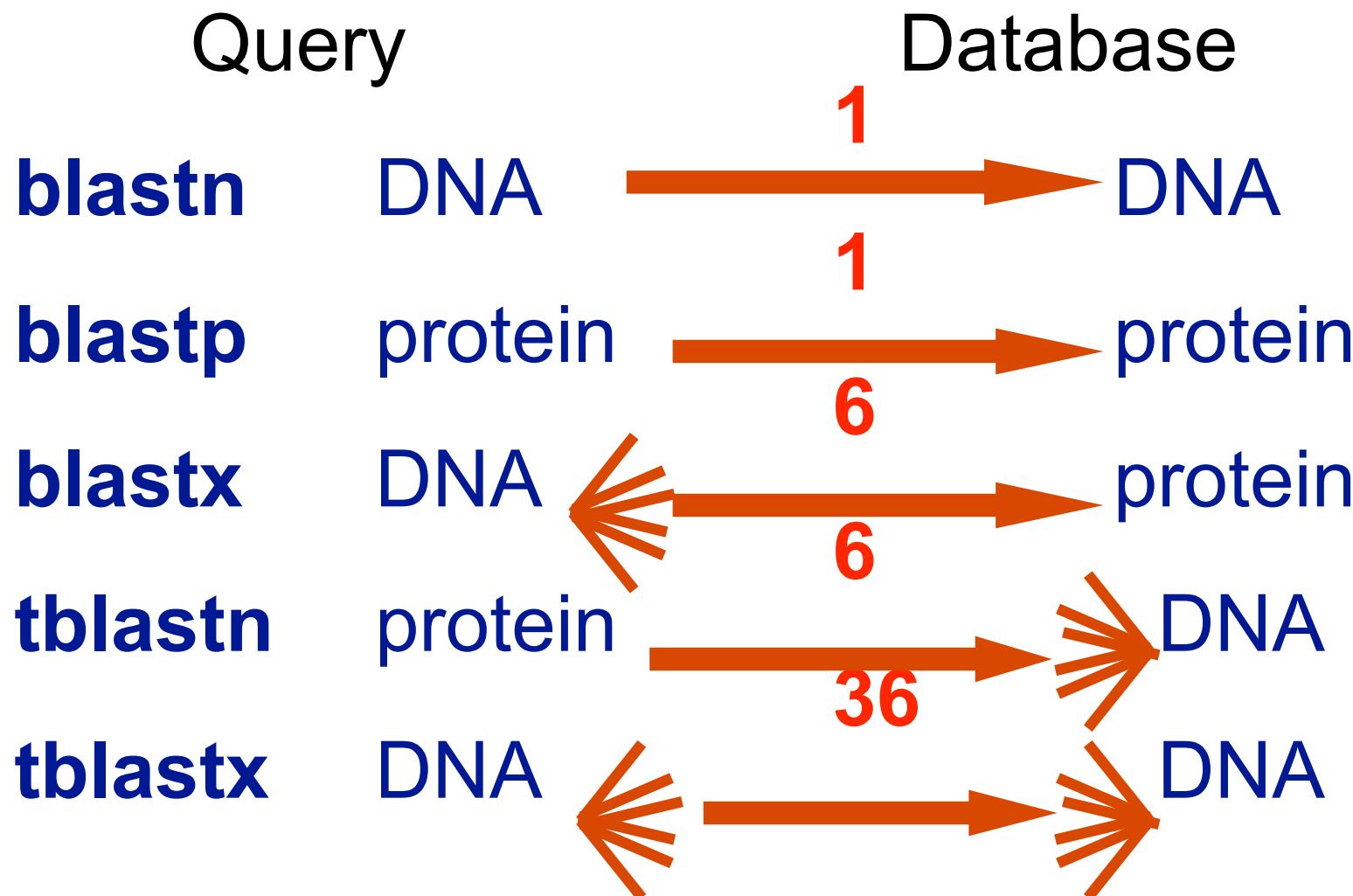
NCBI Reference Sequence **NP_000509.1**

[GenPept](#) [Graphics](#)

>gi|4504349|ref|NP_000509.1| hemoglobin subunit beta [Homo sapiens]
MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFESFGDLSTPDAVMGNPKVKAHGKKVLG
AFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLGNVLVCVLAHHFGKEFTPVQAAYQKVVAGVAN
ALAHKYH

Analyze this sequence
Run BLAST
Identify Conserved Domains
Find in this Sequence

Step 2: Choose the BLAST program



DNA potentially encodes six proteins

5' CAT CAA

5' ATC AAC

5' TCA ACT



5' CATCAACTACAACCTCCAAAGACACCCCTTACACATCAACAAACCTACCCAC 3'

3' GTAGTTGATGTTGAGGTTCTGTGGGAATGTGTAGTTGGATGGGTG 5'

5' GTG GGT

5' TGG GTA

5' GGG TAG



Protein BLAST: search protein databases using a protein query

blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastp&BLAST_PROGRAMS=blastp&PAGE_TYPE=Blast+Search

Reader

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [?](#)

>gi|4504349|ref|NP_000509.1| hemoglobin subunit beta [Homo sapiens]
MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVYPWTQRFESFGDLSTPDAMGNPKVKAHGK
KVLGAFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLGNVLVCVLAHHFGKEFTPVQAAYQK
VVACVANALAHKYH

Clear

Query subrange [?](#)

From
To

Or, upload file no file selected [?](#)

Job Title
Enter a descriptive title for your BLAST search [?](#)

Align two or more sequences [?](#)

Choose Search Set

Database Non-redundant protein sequences (nr) [?](#)

Organism [Optional](#) Exclude [+](#)
Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown. [?](#)

Exclude [Optional](#) Models (XM/XP) Uncultured/environmental sample sequences

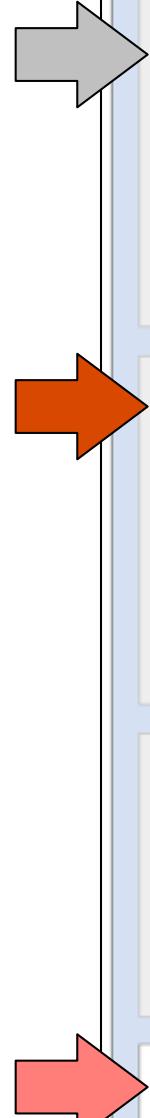
Entrez Query [Optional](#)
Enter an Entrez query to limit search [?](#)

Program Selection

Algorithm blastp (protein-protein BLAST)
 PSI-BLAST (Position-Specific Iterated BLAST)
 PHI-BLAST (Pattern Hit Initiated BLAST)
 DELTA-BLAST (Domain Enhanced Lookup Time Accelerated BLAST)
Choose a BLAST algorithm [?](#)

BLAST Search database Non-redundant protein sequences (nr) using Blastp (protein-protein BLAST)
 Show results in a new window

[Algorithm parameters](#)



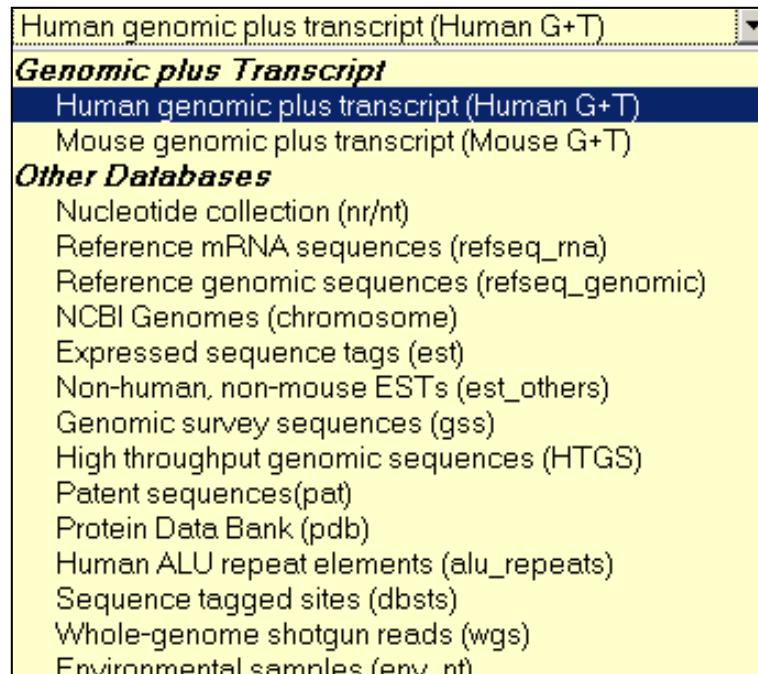
Step 3: Choose the database

nr = non-redundant (most general database)

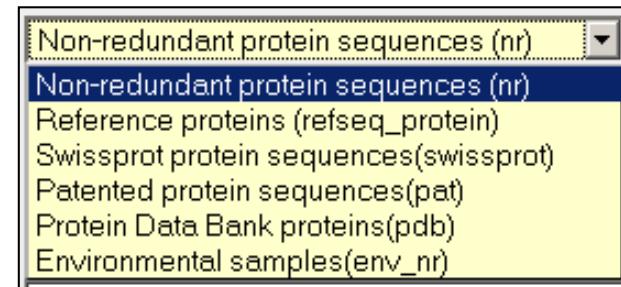
dbest = database of expressed sequence tags

dbsts = database of sequence tag sites

gss = genomic survey sequences



nucleotide databases



protein databases

Protein BLAST: search protein databases using a protein query

blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastp&BLAST_PROGRAMS=blastp&PAGE_TYPE=BlastSearch

Reader

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s)

Query subrange

Or, upload file no file selected

Job Title

Align two or more sequences

Choose Search Set

Database Non-redundant protein sequences (nr)

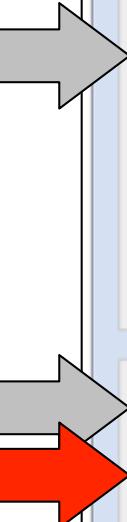
Organism Exclude +
Optional Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown.

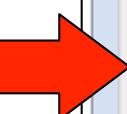
Exclude Models (XM/XP) Uncultured/environmental sample sequences
Optional

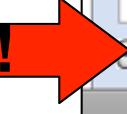
Entrez Query
Optional Enter an Entrez query to limit search

Program Selection

Algorithm blastp (protein-protein BLAST)
 PSI-BLAST (Position-Specific Iterated BLAST)
 PHI-BLAST (Pattern Hit Initiated BLAST)
 DELTA-BLAST (Domain Enhanced Lookup Time Accelerated BLAST)
Choose a BLAST algorithm

Organism 

Entrez 

Settings! 

BLAST Show results in a new window

+ Algorithm parameters

Step 4a: Select optional search parameters

Algorithm parameters

General Parameters

Max target sequences: 100
Select the maximum number of aligned sequences to display ⓘ

Short queries: Automatically adjust parameters for short input sequences ⓘ

Expect threshold: 10 ← **Expect**

Word size: 3 ← **Word size**

Max matches in a query range: 0

Scoring Parameters

Matrix: BLOSUM62 ← **Scoring matrix**

Gap Costs: Existence: 11 Extension: 1

Compositional adjustments: Conditional compositional score matrix adjustment

Filters and Masking

Filter: Low complexity regions ⓘ

Mask: Mask for lookup table only ⓘ
 Mask lower case letters ⓘ

BLAST

Search database Non-redundant protein sequences (nr) using Blastp
 Show results in a new window

Step 4: Optional parameters

- You can...
 - choose the organism to search
 - change the substitution matrix
 - change the expect (E) value
 - change the word size
 - change the output format

Results page

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gi|4504349|ref|NP_000509.1| hemoglobin

Query ID: Icl|84677 Database Name: nr
Description: gi|4504349|ref|NP_000509.1| hemoglobin subunit Description: All non-redundant GenBank CDS
beta [Homo sapiens] translations+PDB+SwissProt+PIR+PRF excluding environmental samples from WGS projects
Molecule type: amino acid Program: BLASTP 2.2.27+ > Citation
Query Length: 147

Other reports: > Search Summary [Taxonomy reports] [Distance tree of results] [Related Structures] [Multiple alignment]

New DELTA-BLAST, a more sensitive protein-protein search Go

Graphic Summary

Show Conserved Domains

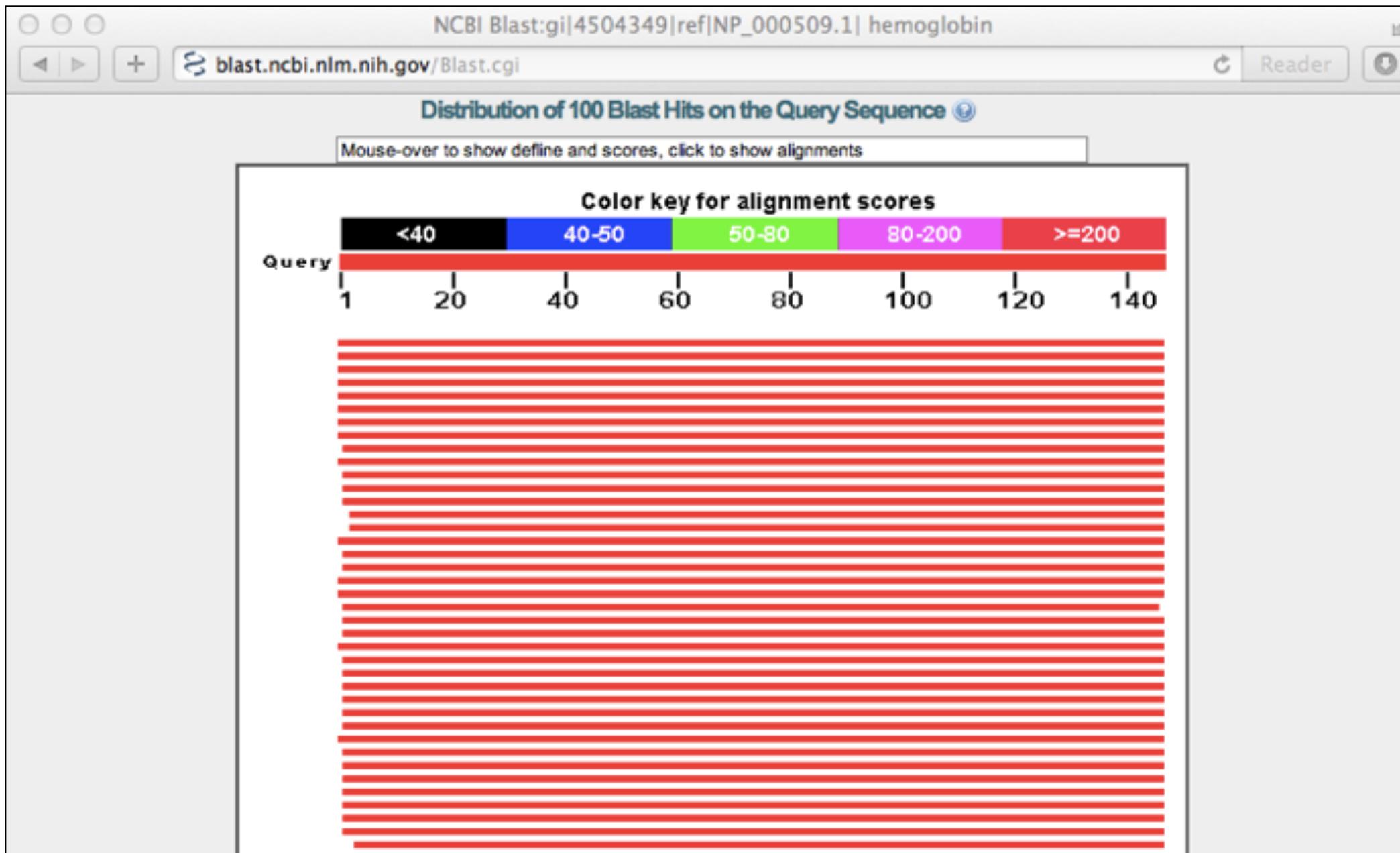
Putative conserved domains have been detected, click on the image below for detailed results.

Query seq. 1 25 50 75 100 125 147
Specific hits: hem-binding site globin
Superfamilies: globin_like superfamily

Distribution of 100 Blast Hits on the Query Sequence ⓘ

Mouse over to show details and scores ⌂ to show elements

Further down the results page...



Further down the results page...

NCBI Blast:gi|4504349|ref|NP_000509.1| hemoglobin

blast.ncbi.nlm.nih.gov/Blast.cgi

Select: All None Selected:0

Alignments Download GenPept Graphics Distance tree of results Multiple alignment

Description Max score Total score Query cover E value Max ident Accession

<input type="checkbox"/> hemoglobin beta [synthetic construct]	301	301	100%	9e-103	100%	AAX37051.1
<input type="checkbox"/> hemoglobin beta [synthetic construct]	301	301	100%	1e-102	100%	AAX29557.1
<input type="checkbox"/> hemoglobin subunit beta [Homo sapiens] >ref XP_508242.1 PREDICTED: hemoglobin s	301	301	100%	1e-102	100%	NP_000509.1
<input type="checkbox"/> RecName: Full=Hemoglobin subunit beta; AltName: Full=Beta-globin; AltName: Full=Hemoglobin subunit beta	300	300	100%	4e-102	99%	P02024.2
<input type="checkbox"/> beta globin chain variant [Homo sapiens]	299	299	100%	5e-102	99%	AAN84548.1
<input type="checkbox"/> beta globin [Homo sapiens] >gb AAZ39781.1 beta globin [Homo sapiens] >gb AAZ39781.1 beta globin [Homo sapiens]	299	299	100%	5e-102	99%	AAZ39780.1
<input type="checkbox"/> beta-globin [Homo sapiens]	299	299	100%	5e-102	99%	ACU56984.1
<input type="checkbox"/> hemoglobin beta chain [Homo sapiens]	299	299	100%	6e-102	99%	AAD19696.1
<input type="checkbox"/> Chain B, Structure Of Haemoglobin In The Deoxy Quaternary State With Ligand Bound At The Beta Subunit	298	298	99%	9e-102	100%	1COH_B
<input type="checkbox"/> hemoglobin beta subunit variant [Homo sapiens] >gb AAA88054.1 beta-globin [Homo sapiens] >gb AAA88054.1 beta-globin [Homo sapiens]	298	298	100%	1e-101	99%	AAF00489.1
<input type="checkbox"/> Chain B, Human Hemoglobin D Los Angeles: Crystal Structure >pdb 2YRS D Chain D, H	298	298	99%	2e-101	99%	2YRS_B
<input type="checkbox"/> Chain B, High-Resolution X-Ray Study Of Deoxy Recombinant Human Hemoglobins Synthesized In Escherichia Coli	297	297	99%	3e-101	99%	1DXU_B
<input type="checkbox"/> Chain B, Analysis Of The Crystal Structure, Molecular Modeling And Infrared Spectroscopic Characterization Of Human Hemoglobin D Los Angeles	297	297	99%	3e-101	99%	1HDB_B



Further down the results page...

NCBI Blast:gi|4504349|ref|NP_000509.1| hemoglobin

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hemoglobin subunit beta [Homo sapiens]
Sequence ID: ref|NP_000509.1| Length: 147 Number of Matches: 1
► See 84 more title(s)

Range 1: 1 to 147 GenPept Graphics			▼ Next Match	▲ Previous Match	
Score	Expect	Method	Identities	Positives	Gaps
301 bits(770)	1e-102	Compositional matrix adjust.	147/147(100%)	147/147(100%)	0/147(0%)

Query 1 MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAMGNPK 60
Sbjct 1 MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAMGNPK 60

Query 61 VKAHGKKVLGAFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLLGTVLCVLAHHFG 120
Sbjct 61 VKAHGKKVLGAFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLLGTVLCVLAHHFG 120

Query 121 KEFTPQVAAQKVVAAGVANALAHKYH 147
Sbjct 121 KEFTPQVAAQKVVAAGVANALAHKYH 147

Related Information

[Gene](#) - associated gene details
[UniGene](#) - clustered expressed sequence tags
[Map Viewer](#) - aligned genomic context
[Structure](#) - 3D structure displays
[PubChem Bio](#)
[Assay](#) - bioactivity screening

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RecName: Full=Hemoglobin subunit beta; AltName: Full=Beta-globin; AltName: Full=Hemoglobin beta chain
Sequence ID: sp|P02024.2|HBB_GORGO Length: 147 Number of Matches: 1

Range 1: 1 to 147 GenPept Graphics			▼ Next Match	▲ Previous Match	
Score	Expect	Method	Identities	Positives	Gaps
300 bits(767)	4e-102	Compositional matrix adjust.	146/147(99%)	147/147(100%)	0/147(0%)

Related Information

Different output formats are available

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Formatting options

Show	Alignment as	HTML	<input type="checkbox"/> Old View	Reset form to defaults		
Alignment View	Query-anchored with letters for identities					
Display	<input checked="" type="checkbox"/> Graphical Overview	<input checked="" type="checkbox"/> Sequence Retrieval	<input type="checkbox"/> NCBI-gi	Reformat		
Masking	Character:	Lower Case	Color:	Grey		
Limit results	Descriptions:	50	Graphical overview:	50	Alignments:	50
Organism Type common name, binomial, taxid, or group name. Only 20 top taxa will be shown.						
Enter organism name or id--completions will be suggested				<input type="checkbox"/> Exclude	+	Reformat
Entrez query:						
Expect Min:		Expect Max:		Reformat		
Percent Identity Min:			Percent Identity Max:			
Format for	<input type="checkbox"/> PSI-BLAST	with inclusion threshold:				

gi|4504349|ref|NP_000509.1| hemoglobin

E.g. Query anchored alignments

NCBI Blast:gi|4504349|ref|NP_000509.1| hemoglobin

blast.ncbi.nlm.nih.gov/Blast.cgi Reader

Query	Score	Sequence	Length
AAX37051	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
AAX29557	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
NP_000509	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
P02024	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
AAN84548	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
AAZ39780	1	MVHLTPKEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
ACU56984	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
AAD19696	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
1COH_B	1	VHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
AAF00489	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
2YRS_B	1	VHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1DXU_B	1	MHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1HDB_B	1	VHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1DXV_B	2	HLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
3KMF_C	2	HLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
AAL68978	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
1NQP_B	1	VHLTPEEKSAVTALWGKVNDEVGGKALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1K1K_B	1	VHLTPKEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
AAN11320	1	MVHLTPVEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
XP_002822173	1	MVHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
1Y85_B	1	VHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1YE0_B	1	MHLTPEEKSAVTALWGKVNDEVGGEALGRLLAVYPWTQRFFESFGDLSTPDAVMGNPK	59
1O1O_B	1	MHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
CAA23759	1	MVHLTPVEEKSAVTAXWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
1YE2_B	1	MHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVFPWTQRFFESFGDLSTPDAVMGNPK	59
1Y5F_B	1	MHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1A00_B	1	MHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1HBS_B	1	VHLTPVEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1ABY_B	1	MHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59
1CMY_B	1	VHLTPEEKSAVTALWGKVNDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	59

... and alignments with dots for identities

NCBI Blast:gi|4504349|ref|NP_000509.1| hemoglobin

blast.ncbi.nlm.nih.gov/Blast.cgi Reader

Query	Length	Sequence	Score
AAX37051	1	MVHLTPPEEKSAVTALWGKVNVDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK	60
AAX29557	1	60
NP_000509	1	60
P02024	1	60
AAN84548	1	60
AAZ39780	1K.....	60
ACU56984	1K.....	60
AAD19696	1L.....	60
1COH_B	1	59
AAF00489	1	60
2YRS_B	1	59
1DXU_B	1	M.....	59
1HDB_B	1	59
1DXV_B	2	59
3KMF_C	2	59
AAL68978	1	60
1NQP_B	1K.....	59
1K1K_B	1K.....	59
AAN11320	1V.....	60
XP_002822173	1	60
1Y85_B	1	59
1YE0_B	1	M.....A.....	59
1O1O_B	1	M.....	59
CAA23759	1V.....X.....	60
1YE2_B	1	M.....F.....	59
1Y5F_B	1	M.....	59
1A00_B	1	M.....Y.....	59

Common problems

- Selecting the wrong version of BLAST
- Selecting the wrong database
- Too many hits returned
- Too few hits returned
- Unclear about the significance of a particular result - are these sequences homologous?

How to handle too many results

- Focus on the question you are trying to answer
 - select “refseq” database to eliminate redundant matches from “nr”
 - Limit hits by organism
 - Use just a portion of the query sequence, when appropriate
 - Adjust the expect value; lowering E will reduce the number of matches returned

How to handle too few results

- Many genes and proteins have no significant database matches
 - remove Entrez limits
 - raise E-value threshold
 - search different databases
 - try scoring matrices with lower BLOSUM values (or higher PAM values)
 - use a search algorithm that is more sensitive than BLAST (*e.g.* PSI-BLAST or HMMer)

Summary of key points

- Sequence alignment is a fundamental operation underlying much of bioinformatics.
- Even when optimal solutions can be obtained they are not necessarily unique or reflective of the biologically correct alignment.
- Dynamic programming is a classic approach for solving the pairwise alignment problem.
- Global and local alignment, and their major application areas.
- Heuristic approaches are necessary for large database searches and many genomic applications.

FOR NEXT CLASS....

Check out the online:

- Reading**: Sean Eddy's "What is dynamic programming?"
- Homework**: (1) [Quiz](#), (2) [Alignment Exercise](#).

To Update!

Homework Grading

Both (1) quiz questions and (2) alignment exercise carry equal weights (*i.e.* 50% each).

(Homework 2) Assessment Criteria	Points	
Setup labeled alignment matrix	1	
Include initial column and row for GAPs	1	
All alignment matrix elements scored (<i>i.e.</i> filled in)	1	
Evidence for correct use of scoring scheme	1	
Direction arrows drawn between all cells	1	
Evidence of multiple arrows to a given cell if appropriate	1	D
Correct optimal score position in matrix used	1	C
Correct optimal score obtained for given scoring scheme	1	B
Traceback path(s) clearly highlighted	1	A
Correct alignment(s) yielding optimal score listed	1	A+