# Trendy: Breakpoint analysis of time course expression data

Rhonda Bacher, Ning Leng, Ron Stewart

# Contents

Overview	1
Fhe model  Installation Install via GitHub Install locally Load the package	1
del         1           tion         2           l via GitHub         2           l locally         3           the package         3           s         3           segmented regressions         3           slize trends of the top dynamic genes         4           lize individual genes         7           specific estimates         10           upoint distribution over the time course         10           dvanced analysis         11           course with non-uniform sampling         11           tional options         12	
Install via GitHub	2
Install locally	3
Load the package	3
Analysis	_
Input	3
Run segmented regressions	3
Visualize trends of the top dynamic genes	4
Visualize individual genes	7
Gene specific estimates	10
Breakpoint distribution over the time course	10
More advanced analysis	11
Time course with non-uniform sampling	11
Additional options	12
Trendy shiny app	13
SessionInfo	16

# Overview

Trendy is a R package that can be used to perform breakpoint analysis on microarray or RNA-seq expression data with ordered conditions (e.g. time course, spatial course). For each gene or other features, Trendy estimates the optimal number of breakpoints as well as the breakpoints by fitting a set of segmented regression models. The top dynamic genes are then identified by taking genes that can be well profiled by its gene-specific segmented regression model. Trendy also implements functions to visualize the dynamic genes and their trends, to order dynamic genes by their trends, and to compute breakpoint distribution at different time points (e.g. detect time points with a large number of expression changes).

# The model

To illustrate Trendy, here we use time course gene expression data as an example. Note Trendy may also be applied to other types of features (e.g. isoform or exon expression) and/or other types of experiments with ordered conditions (e.g. spatial course).

Denote the normalized gene expression of gene g and sample s is  $X_{g,s}$ . Denote the total number of genes as G and the total number of samples as S. For each gene, Trendy fits segmented regression models with

varying numbers of breakpoints from 1 to  $n_k$ . In which  $n_k$  defaults to 3 but can also be specified by the user. The model with k breakpoints can then be written as:

$$\begin{split} M_g^k: X_g \sim \beta_0^k + \beta_1^k * I\{s: s \geq 1, s \leq b_{g,1}^k\} * s + \beta_2^k * I\{s: s \geq b_{g,1}^k + 1, s \leq b_{g,2}^k\} * (s - b_{g,1}^k) +, ..., \\ + \beta_{k+1}^k * I\{s: s \geq b_{g,k}^k + 1, s \leq S\} * (s - b_{g,k}^k) \end{split}$$

For each k, the segmented regression estimates k breakpoints  $(b_{g,1}^k, b_{g,2}^k, \dots b_{g,k}^k)$  between 1 and S. The segmented regression also estimate k+2  $\beta$ s. In which  $\beta_0^k$  indicates the intercept, and the other  $\beta$ s indicate slopes for the k+1 segments separated by the k breakpoints. We denote the adjusted  $R^2$  for this model as  $r_g^k$ .

For a given gene, among the models with varying k, SegeReg picks the optimal number of breakpoints for this gene by comparing the adjusted  $R^2$ s:

$$\tilde{k_g} = \operatorname{argmax}_{k=1,\dots,n_k}(r_g^k)$$

To avoid overfitting, the optimal number of breakpoints will be set as  $\tilde{k_g} = \tilde{k_g} - 1$  if any of the following happens: at least of one segments having less than  $c_{num}$  samples, or  $r_g^{\tilde{k}} - r_g^{\tilde{k}-1} < c_{diff}$ . The thresholds  $c_{num}$  and  $c_{diff}$  can be specified by the user; defaults are 5 and 0.1, respectively.

Then the gene specific adjusted  $R^2$  and breakpoint estimates are then obtained from this optimal model:  $r_g = r_g^{\tilde{k_g}}; \ (\beta_{g,0},...,\beta_{g,\tilde{k_g}+1}) = (\beta_{g,0}^{\tilde{k_g}},...,\beta_{g,\tilde{k_g}+1}^{\tilde{k_g}})$  and  $(b_{g,1},...,b_{g,\tilde{k_g}}) = (b_{g,1}^{\tilde{k_g}},...,b_{g,\tilde{k_g}}^{\tilde{k_g}})$ . Among all genes, the top dynamic genes are defined as those whose optimal model has high adjusted  $R^2$ s.

To compute the breakpoint distribution over the time course, Trendy calculates:

$$N_s = \sum_{g=1,...,G} \sum_{j=1,...,\tilde{k_g}} I\{b_{g,j} = s\}$$

The time points with high  $N_s$  might be considered as time points with a large amount of expression changes.

Trendy also outputs fitted trend of each gene. For samples between the  $j^{th}$  and  $j+1^{th}$  breakpoint for a given gene, if the t statistic of  $\beta_{g,j+1}$  has p value greater than  $c_{pval}$ , the trend of this segment will be defined as no change. Otherwise the trend of this segment will be defined as up/down based on the coefficient of  $\beta_{g,j+1}$ . The  $c_pval$  defaults to 0.1, but can also be specified by the user.

# Installation

### Install via GitHub

The Trendy package can be installed using functions in the devtools package.

To install, type the following codes in R:

install.packages("devtools")

library(devtools)

install\_github("rhondabacher/Trendy/package/Trendy")

# Install locally

Install packages segmented and gplots:

```
install.packages(c("segmented","gplots"))
library("segmented")
library("gplots")
```

Download the Trendy package from:

https://github.com/rhonda/Trendy/tree/master/package

And install the package locally.

### Load the package

To load the Trendy package:

```
library(Trendy)
```

# **Analysis**

### Input

##

g23

The input data should be a G-by-S matrix containing the expression values for each gene and each sample, where G is the number of genes and S is the number of samples. The samples should be sorted following the time course order. These values should exhibit expression data after normalization across samples. For example, for RNA-seq data, the raw counts may be normalized using MedianNorm and GetNormalizedMat() function in EBSeq. More details can be found in the EBSeq vignette:

http://www.bioconductor.org/packages/devel/bioc/vignettes/EBSeq/inst/doc/EBSeq\_Vignette.pdf

The object TrendyExData is a simulated data matrix containing 50 rows of genes and 40 columns of samples.

```
data(TrendyExData)
str(TrendyExData)
```

```
## num [1:50, 1:40] 240 199 198 239 202 ...
## - attr(*, "dimnames")=List of 2
## ..$ : chr [1:50] "g1" "g2" "g3" "g4" ...
## ..$ : chr [1:40] "s1" "s2" "s3" "s4" ...
```

g8

### Run segmented regressions

The trendy() function can be used to run gene specific segmented regressions. Here we want to only consider up to 2 breakpoints for each gene. To do so we may specify maxk=2:

```
res <- trendy(TrendyExData, maxk=2)
res.top <- toptrendy(res)
# default adjusted R square cutoff is 0.5
res.top$radj

## g3 g1 g28 g20 g15 g2 g10
## 0.9787382 0.9775005 0.9751380 0.9739715 0.9729747 0.9710139 0.9705118
```

g17

g12

g29

g24

g5

```
## 0.9701402 0.9691341 0.9689555 0.9656732 0.9652141 0.9644343 0.9632348
                                                  g11
                                                             g30
##
         g16
                   g22
                              g18
                                        g25
                                                                       g26
## 0.9630272 0.9627092 0.9626837 0.9611528 0.9600736 0.9597989 0.9572072
                                                             g19
                                                                       g27
##
                    g4
                                        g21
          g7
                               g9
                                                   g6
## 0.9529077 0.9420853 0.9377311 0.9304116 0.9291045 0.9259893 0.9183375
##
         g14
                   g13
## 0.8656596 0.8576471
```

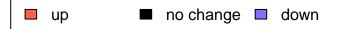
The toptrendy() function may be used to extract top dynamic genes. By default, toptrendy() will extract genes whose adjusted  $R^2$   $r_g$  is greater or equal to 0.5. To change this threshold, a user may specify the r.cut parameter in toptrendy() function. res.top\$radj gives  $r_g$  of the top dynamic genes, sorted decreasingly by  $r_g$ .

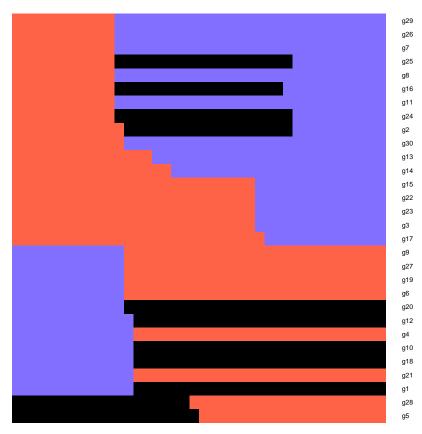
By default the trendy() function only consider genes whose mean expression is greater than 10. To use another threshold, a user may specify the parameter meancut in trendy() function.

# Visualize trends of the top dynamic genes

res.top\$id.sign gives trend specification of the top genes. Function trendheatmap() can be used to display these trends:

```
res.trend <- trendheatmap(res.top)</pre>
```



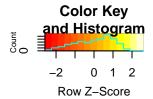


### str(res.trend)

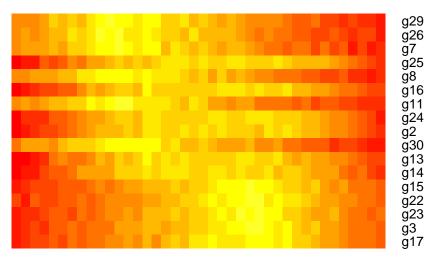
```
## List of 3
## $ firstup : Named num [1:17] 11.4 11.5 11.6 11.6 11.6 ...
## ..- attr(*, "names")= chr [1:17] "g29" "g26" "g7" "g25" ...
## $ firstdown : Named num [1:11] 12.1 12.6 12.6 12.7 12.8 ...
## ..- attr(*, "names")= chr [1:11] "g9" "g27" "g19" "g6" ...
## $ firstnochange: Named num [1:2] 19 20.4
## ..- attr(*, "names")= chr [1:2] "g28" "g5"
```

The trendheatmap() function classify the top dynamic genes into three groups: start with up, start with down and start with no change. Within each group, genes are sorted by the position of the first breakpoint.

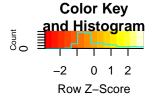
To generate expression heatmap of the first group of genes (first go up):



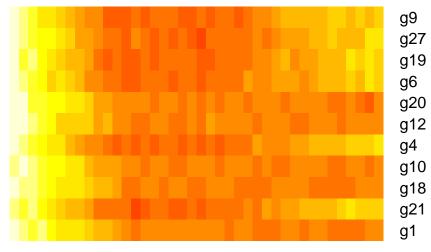
# top genes (first go up)



Similarly, to generate expression heatmap of the second group of genes (first go down):

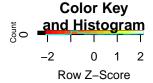


# top genes (first go down)

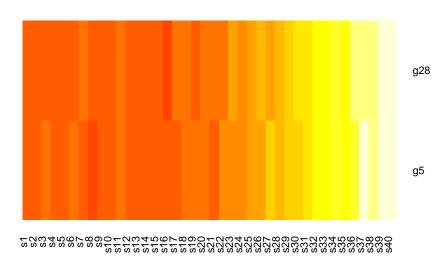


ate expression heatmap of the second group of genes (first no change):

To gener-



# top genes (first no change)

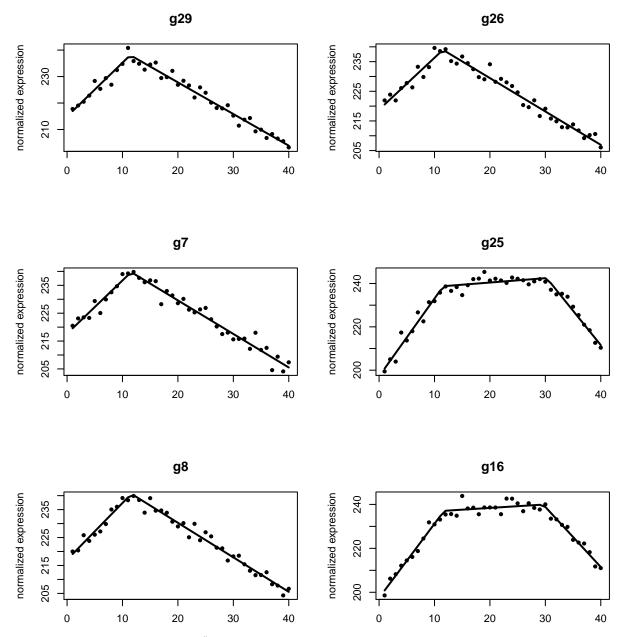


# Visualize individual genes

The plotmarker() function may be used to plot expression of individual genes and the fitted lines.

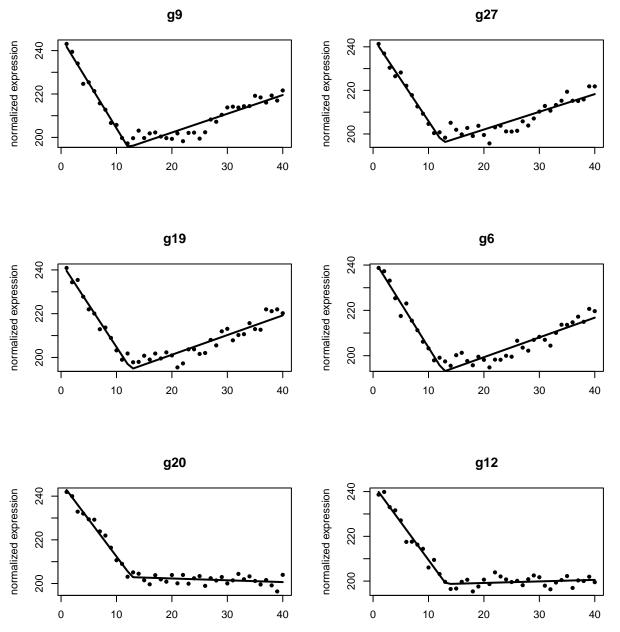
For example, to plot the top 6 genes in the first group of genes (first go up):

plot1 <- plotmarker(TrendyExData,listname=names(res.trend\$firstup)[1:6],fittedres=res)</pre>

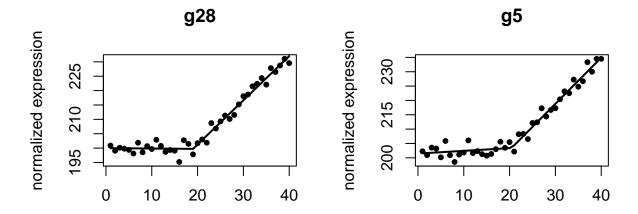


The input of function plotmarker() requires the expression data and a list of genes of interest. The parameter fittedres in function plotmarker() takes trendy() fitted results. If it is not specified, the function plotmarker() will run Trendy model on the genes of interest before plotting. Specifying fitted results obtained from previous steps will save time by avoiding fitting the models again.

Similarly, to plot the top 6 genes in the second group of genes (first go down):



To plot the 2 genes in the third group of genes (first no change):



# Gene specific estimates

For a given gene of interest, its estimated parameters can be obtained by (using g2 as an example):

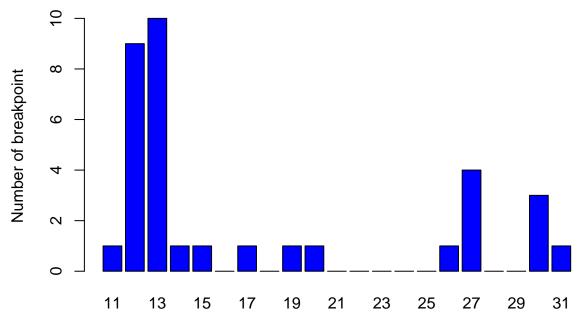
```
print(res.top$bp["g2"]) # break points
## $g2
## psi1.t.use psi2.t.use
     12.47356
                30.14908
print(res.top$radj["g2"]) # adjusted r square
##
## 0.9710139
print(res.top$slp["g2"]) # fitted slopes of the segments
## $g2
## slope1 slope2 slope3
   3.3110 0.0607 -2.9730
print(res.top$slp.pval["g2"]) # p value of each the segment
## $g2
##
       slope1
                  slope2
                             slope3
## 0.01669386 0.31815050 0.02445599
```

The above printouts show that for gene g2, the optimal number of breakpoints is 2. Two estimated breakpoints are close to s12 and s30. The fitted slopes for the 3 segments are 3.31, 0.06 and -2.97.

### Breakpoint distribution over the time course

To calculate number of breakpoints over the time course:

```
res.bp <- bpdist(res.top)
barplot(res.bp, ylab="Number of breakpoint", col="blue")</pre>
```



The bar plot indicates that many genes have breakpoint around s12 and s13.

# More advanced analysis

# Time course with non-uniform sampling

If the samples were collected with different time intervals and the user wants to use the original time (instead of a vector of consecutive numbers), the user may specify it via the t.vect parameter in trendy() function. For example, suppose for the example data, the first 30 samples were collected every hour and the other 10 samples were collected every 5 hours. We may define the time vector as:

```
t.v \leftarrow c(1:30, seq(31,80,5))
names(t.v) <- colnames(TrendyExData)</pre>
print(t.v)
##
                      s5
                          s6
                                   s8
                                       s9 s10 s11 s12 s13 s14 s15 s16 s17
         2
              3
                  4
                       5
                           6
                                7
                                    8
                                        9
                                            10
                                                11
                                                     12
                                                         13
                                                             14
                                                                  15
                                                                      16
                                                                           17
   s19 s20 s21 s22 s23 s24 s25 s26 s27 s28
                                               s29
                                                   s30 s31 s32 s33 s34 s35
    19
        20
             21
                 22
                      23
                          24
                              25
                                   26
                                       27
                                            28
                                                29
                                                    30
                                                         31
                                                             36
                                                                  41
   s37 s38
           s39 s40
    61
        66
            71
                76
To run Trendy model using the empirical collecting time instead of sample ID (1-40):
res2 <- trendy(TrendyExData, t.vect=t.v, maxk=2)</pre>
res.top2 <- toptrendy(res2)</pre>
res.trend2 <- trendheatmap(res.top2, showplot=FALSE)</pre>
str(res.trend2)
## List of 3
##
    $ firstup
                     : Named num [1:18] 9.95 11.36 11.43 11.56 11.66 ...
     ..- attr(*, "names")= chr [1:18] "g30" "g29" "g8" "g26" ...
                    : Named num [1:11] 11 11.2 11.3 11.4 11.4 ...
##
    $ firstdown
##
     ..- attr(*, "names")= chr [1:11] "g9" "g19" "g21" "g4" ...
    $ firstnochange: Named num(0)
```

```
## ..- attr(*, "names")= chr(0)
```

To plot the first 6 genes that have up-regulated pattern at the beginning of the time course, by showing empirical time at x axis:

# Additional options

In trendy() function, the thresholds  $c_{num}$ ,  $c_{diff}$  and  $c_{pval}$  can be specified via parameters min.num.in.seg, cutdiff and pvalcut.

# Trendy shiny app

The Trendy shiny app requires an RData object output from the trendy() call, which can be obtained by setting saveObject=TRUE.

```
res <- trendy(TrendyExData, maxk=2, saveObject = TRUE, fileName="exampleObject")
Then in R run:</pre>
```

library(shiny)
runGitHub('rhondabacher/Trendy')

# **Trendy**

File is uploaded!

Upload .RData object first, then obtain list of genes according to some pattern or visualize genes one by one.

# Input .Rdata from trendy() run: Browse... exampleObject\_trendyForShin Upload complete Upload File

Upload shiny object

# **Trendy**

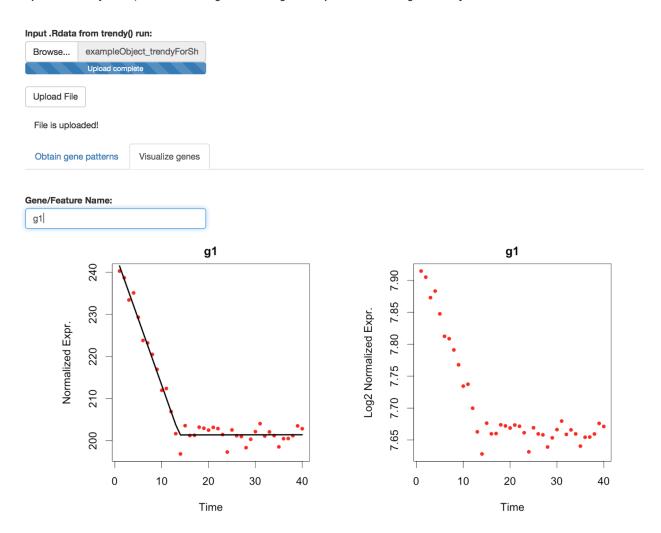
Upload .RData object first, then obtain list of genes according to some pattern or visualize genes one by one.

Input .Rdata from trendy() run:		
Browse	exampleO	bject_trendyForSh
	Upload com	plete
Upload File		
File is uploa	aded!	
Obtain gen	e patterns	Visualize genes
Please selec	t a folder fo	r output :
Select Outp	out Folder	
Enter pattern spaces):	n (separate l	by comma, no
up,down		
ар,чочн		
Only conside squared great		h adjusted R
.5	ater than:	
.5		
Only conside time-point:	er genes wit	h pattern after
0		
Output a plot	t of pattern	nd gange?
<ul><li>Yes</li></ul>	t of patterne	u genes:
○ No		
Output file na	ame (will de	fault to pattern)
		Tauri to pattorn,
Submit for	processing	
Submit for p	processing	J

Find all genes with a given pattern

# **Trendy**

Upload .RData object first, then obtain list of genes according to some pattern or visualize genes one by one.



Search genes individually

# SessionInfo

# print(sessionInfo())

```
## R version 3.3.1 (2016-06-21)
## Platform: x86_64-apple-darwin13.4.0 (64-bit)
## Running under: OS X 10.11.6 (El Capitan)
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
## attached base packages:
## [1] stats
                graphics grDevices utils
                                              datasets methods
                                                                  base
##
## loaded via a namespace (and not attached):
## [1] backports_1.0.4 magrittr_1.5
                                       rprojroot_1.1
                                                      tools_3.3.1
## [5] htmltools_0.3.5 yaml_2.1.14
                                       Rcpp_0.12.8
                                                       stringi_1.1.1
## [9] rmarkdown_1.2
                      knitr_1.15.1
                                       stringr_1.1.0
                                                      digest_0.6.12
## [13] evaluate_0.10
```