## Run a minimal deconvolution simulation

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This vignette shows how to run minimal deconvolution simulations on some example synthetic data objects.

#### Run a minimal simulation

This section walks through how to set up, run, and analyze a deconvolution simulation series. The entire code to run the simulation is as follows:

```
num.sim <- 1e3
lgv <- random_lgv(gindexv = c(1,2), num.iter = num.sim)
size1 <- 1
size2 <- 100
lsv <- lapply(seq(num.sim), function(ii){c(size1, size2)})
lpv <- make_lpv()
lres <- decon_analysis(lgv = lgv, lpv = lpv, lsv = lsv)</pre>
```

#### Simulation setup

```
num.sim <- 1e3
```

We have set the number of simulations to 1000. We need to define the core deconvolution objects to run the simulation. These are the following 3 list variables:

• lgv: Marker signals. These are used to calculate Z. We can assign these as follows:

```
marker1 <- c(1, 0)
marker2 <- c(0, 1)
lgv <- lapply(seq(num.sim),function(ii){list(marker1, marker2)})</pre>
```

• lsv: Size factors. These are used to transform Z and also to make the Y pseudo-bulked sample. To assign these values, use:

```
size1 <- 1
size2 <- 100
lsv <- lapply(seq(num.sim), function(ii){c(size1, size2)})</pre>
```

• lpv: True prediction values. These are used to make the Y pseudo-bulked sample and to compare predictions returned from deconvolution. We can vary complementary proportions for 2 types using:

```
prop1 <- seq(1e-3, 1, 1e-3)
prop2 <- rev(prop1)
lpv <- lapply(seq(num.sim), function(ii){c(prop1[ii], prop2[ii])})</pre>
```

#### Run simulations

Now run simulations and store the results as lres.

```
lres <- decon_analysis(lgv, lpv, lsv)</pre>
## Loading required package: ggplot2
## Loading required package: dplyr
## Attaching package: 'dplyr'
## The following object is masked from 'package:Biobase':
##
##
       combine
## The following objects are masked from 'package:GenomicRanges':
##
##
       intersect, setdiff, union
## The following object is masked from 'package:GenomeInfoDb':
##
##
       intersect
## The following objects are masked from 'package: IRanges':
##
       collapse, desc, intersect, setdiff, slice, union
##
## The following objects are masked from 'package:S4Vectors':
##
       first, intersect, rename, setdiff, setequal, union
##
##
  The following objects are masked from 'package:BiocGenerics':
##
##
       combine, intersect, setdiff, union
## The following object is masked from 'package:matrixStats':
##
##
       count
## The following objects are masked from 'package:stats':
##
##
       filter, lag
## The following objects are masked from 'package:base':
##
```

intersect, setdiff, setequal, union

##

## Analyze results

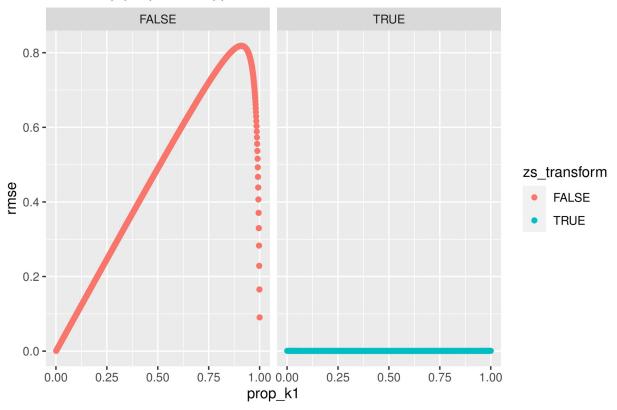
We access the plot objects from lres.

#### lgg <- lres\$lgg</pre>

We can view the scatter plots of NNLS-predicted proportions for type 1 (x-axis) by the root mean squared error (RMSE), grouped on whether the S-transform was first applied to the Z reference, using:

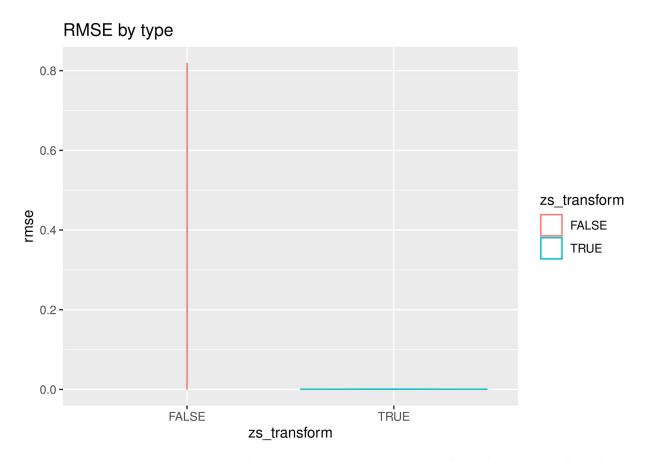
lgg\$ggpt1





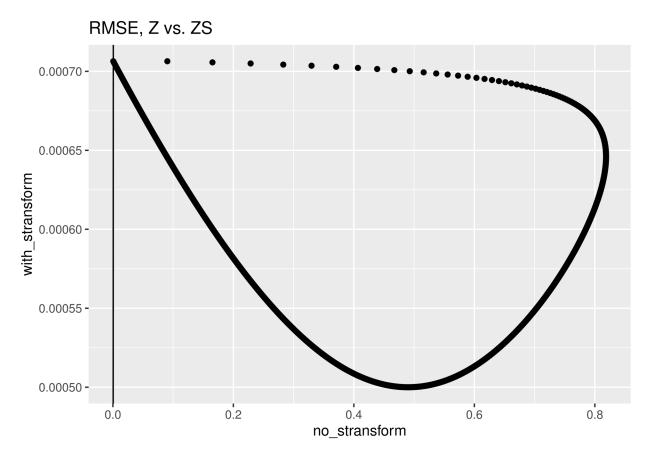
We can view the violin plots of RMSE grouped on S transformation status using:

### lgg\$ggvp



We can view the scatterplot of the RMSEs for the non-transformed data (x-axis) versus the S-tranformed data (y-axis), with a red reference line (y-intercept = 0, slope = 1), using:

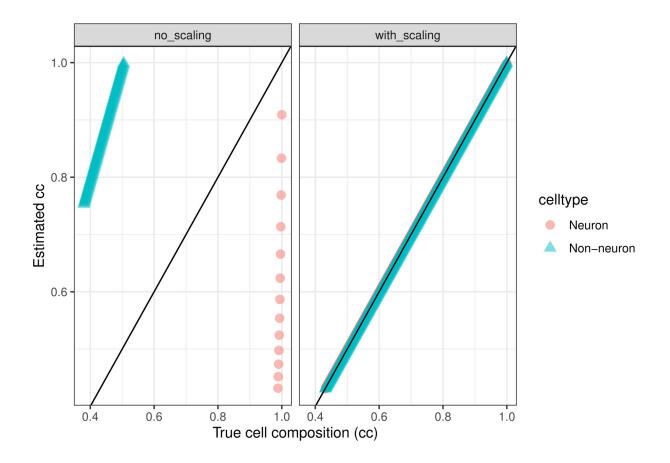
lgg\$ggpt2



The final plot shows scatterplots of the bias, or proportions. The true cell type proportions are on the x-axis, the predicted proportions are on the y-axis, and the reference line with a slope of 1 and intercept of 0 is also shown.

## lgg\$ggpt.bias

## Warning: Removed 2721 rows containing missing values (geom\_point).



# Conclusions

This vignette showed how to perform a minimal simulation and analyze the results.

For general information about the lute R package, see the User's Guide.

For additional simulation examples, see  $\verb|size_factor_experiments.Rmd| vignette.$