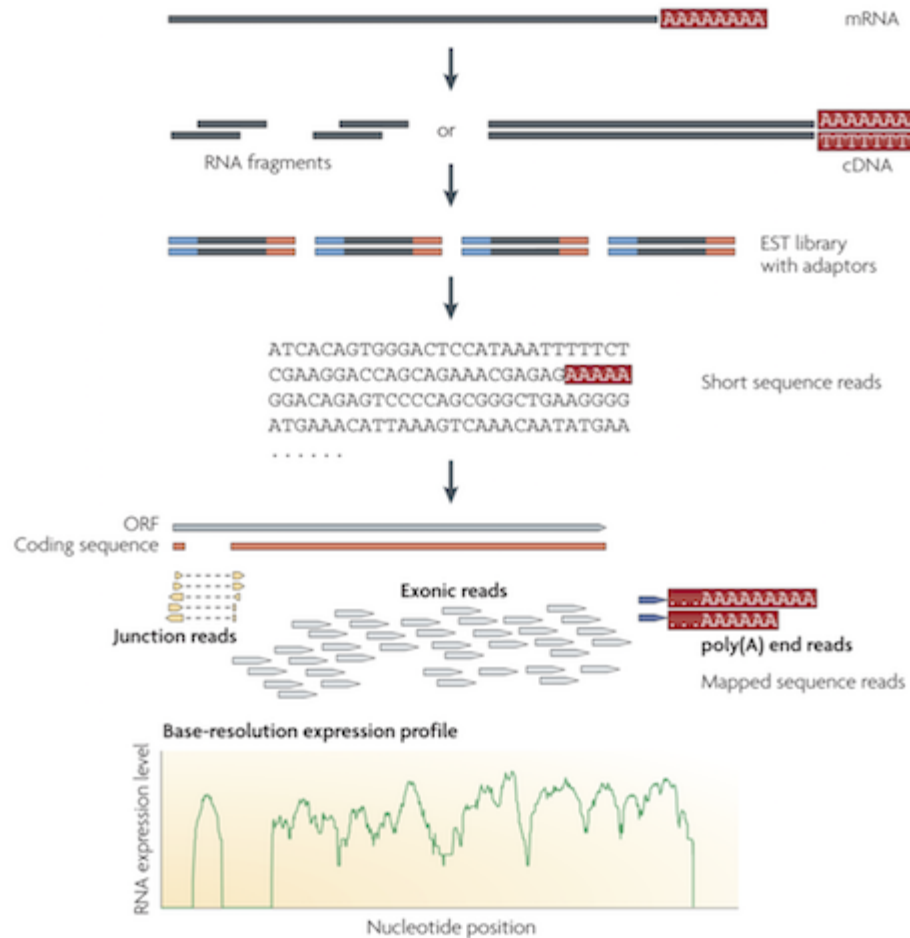


# RNASeq analyses

# RNASeq procedure

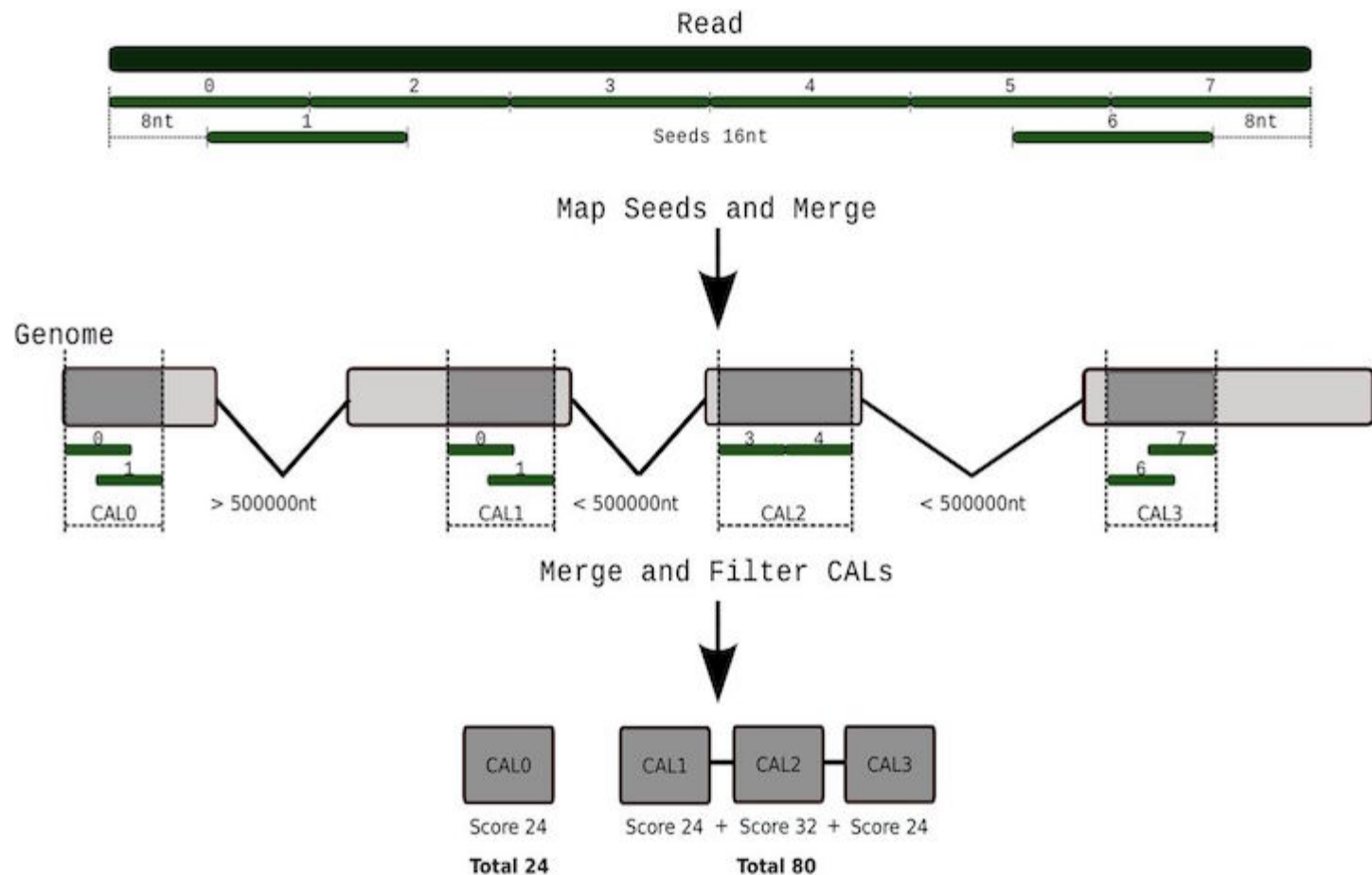
Wang et al. Nat Rev Genetics. 2009. doi:[10.1038/nrg2484](https://doi.org/10.1038/nrg2484)



# Multiple approaches

1. Genome sequenced, align RNAseq reads to genome
2. de novo Assembly of mRNA into transcripts
3. Quantify gene expression from reads aligned to genome or transcripts

# Reads to Genome mapping



# Reads to Genome mapping

Challenges: mRNA is spliced, genome contains introns

Splice-aware short read aligners. Speed and accuracy tradeoffs

- Tophat + Bowtie
- HISAT/HISAT2
- GMAP/GSNAP
- STAR

# Quantify expression

- Count reads overlapping exons
- Table of total read counts per gene
- Normalize counts for gene length and sequencing library depth
- Gene expression then is FPKM - Fragments per Kilobase per Millions of reads
- Tools: htseq-count, stringtie
- BEDtools
- R tools with iRanges

# Evaluating expression differences

Statistical tools for evaluating gene expression differences

- Ballgown [bioconductor package](#)
- DESeq [bioconductor package](#)
- edgeR [bioconductor package](#)

# Alternative approach for Quantifying

Compare reads to **Transcripts** instead of Genome

- Kalisto and Sailfish are common tools
- Bray et al 2016 "Near-optimal probabilistic RNA-seq quantification" doi:[10.1038/nbt.3519](https://doi.org/10.1038/nbt.3519)
- Patro et al 2014 "Sailfish enables alignment-free isoform quantification from RNA-seq reads using lightweight algorithms" doi:[10.1038/nbt.2862](https://doi.org/10.1038/nbt.2862)



# Denovo assembly

[Trinity Assembler](#) for RNASeq

```
$ module load trinity-rnaseq  
$ module switch perl/5.22.0  
$ Trinity --seqType fq --left reads_1.fq --right reads_2.fq --CPU 8 --max_m
```

# ORF identification

## [TransDecoder](#)

- Finds Open Reading Frames in mRNA transcripts

```
$ module load transdecoder  
$ TransDecoder.LongOrfs -t target_transcripts.fasta
```

# RNAseq read mapping

## Using HISAT2

```
# srun --ntasks 8 --pty bash -l
$ mkdir rnaseq; cd rnaseq
$ cp -r /bigdata/gen220/shared/projects/RNAseq ./
$ module load hisat2
$ cd genome
$ ls -l
$ hisat2-build yeast_genome.fasta yeast
$ cd ..
```

# RNAseq read mapping

```
$ hisat2 -x genome/yeast -1 fastq/yeast_RNASeq_1.fq -2 fastq/yeast_RNASeq_2.fq  
-S RNASeq_aln.sam -p 16  
$ module load samtools  
$ samtools view -b RNASeq_aln.sam > RNASeq_aln.bam  
$ samtools sort RNASeq_aln.bam > RNASeq_aln.sort.bam  
$ samtools index RNASeq_aln.sort.bam  
$ samtools flagstat RNASeq_aln.sort.bam
```

# Process BAM files for other tools

- give to htseq-count to get the read depth
- process with stringtie

```
$ module load stringtie  
GTF=genome/genes.gff  
$ stringtie -G $GTF -b stringtie_out -e -o stringtie.gtf -A stringtie.gene_
```