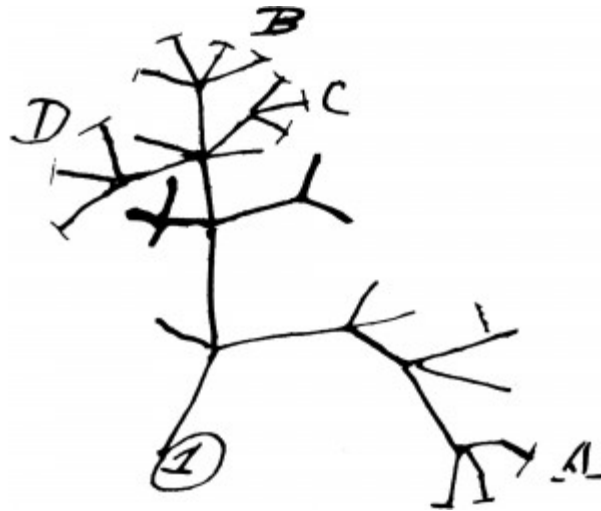


Phylogenetics and data processing

Phylogenetics Trees

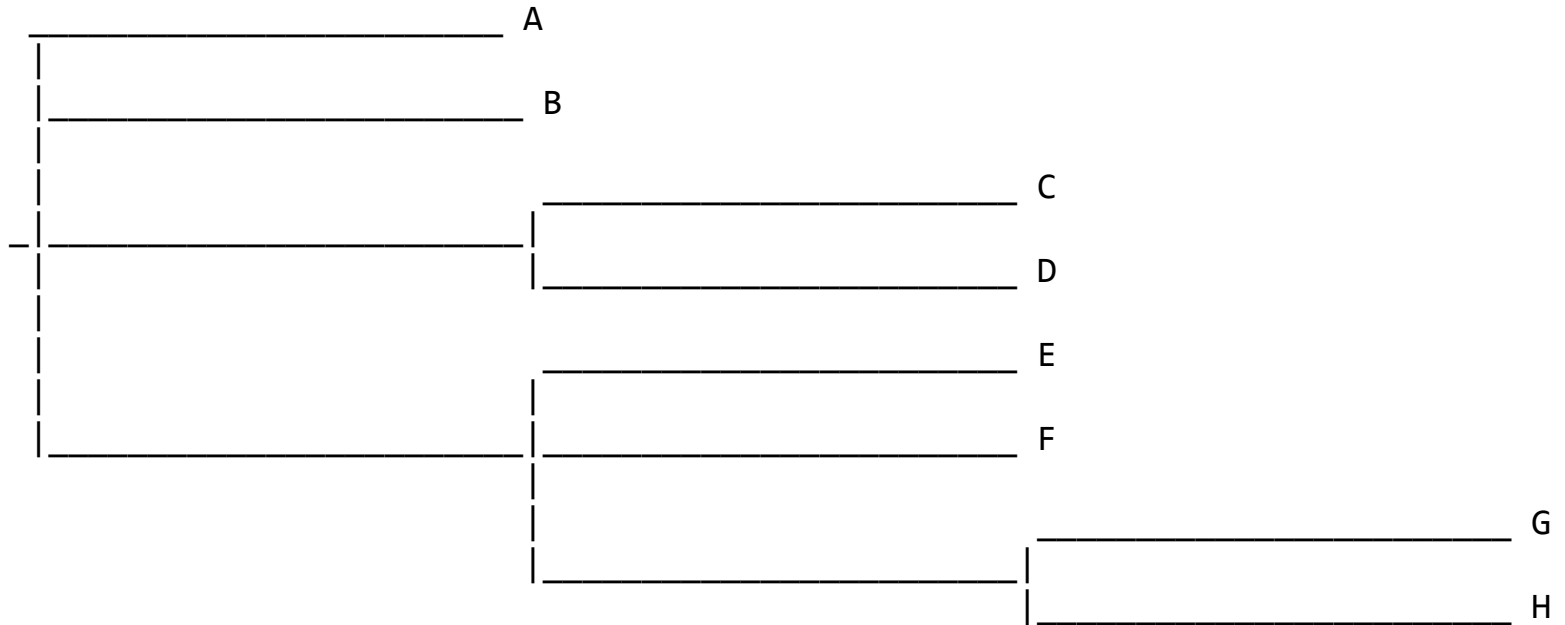
- Representation of relationships of species, genes, groups
- Adjacent branches are more closely related entities than ones further away
- Generate trees based on computed distances or more complex likelihood models



Compact Tree Representation

Newick format

`(A,B,(C,D),(E,F,(G,H)))`;



Molecular Phylogenetics

- Identify homologous sequences (e.g. genes)
- Align sequence with Multiple alignment software
- Potentially trim sequence alignment or poor alignment region
- Construct phylogenetic tree

Identify homologous sequences

- Reciprocal BLAST
- Orthology Clustering
- Syntenic information (shared flanking regions)
- Molecular approaches (PCR, Clone, Sequencing)
- DNA and Protein considerations
- Store in FASTA file

Extract those sequences

```
$ module load cdbfasta  
$ cdbfasta DATABASE  
# make a file which has the sequence names, one each line  
$ cdbyank DATABASE.cidx < listseqids > sequencefile.fa
```

Multiple Sequence Alignment

- Construct MSAs: MUSCLE

```
$ module load muscle  
$ muscle -in VMS1.aa.fasta -out VMS1.aa.fasaln  
$ muscle -clw -in VMS1.aa.fasta -out VMS1.aa.aln
```

- Construct MSA of coding sequence: [T-Coffee](#)
- Align the sequence on Codon Boundaries (e.g. w/ knowledge of the amino-acid it codes for)

```
$ module load tcoffee  
$ t_coffee VMS1.cds.fasta -method cdna_fast_pair
```

Trimming alignments

- [Trimal](#) or Gblocks to remove poorly aligning regions

```
$ module load trimal  
$ trimal -in VMS1.aa.fasaln -out VMS1.aa.trim -automated1
```

- additional [options for trimming](#)

```
# remove spurious sequences ('good site's are 75% seqs share a site;  
# 80% of the sites in sequence must be good to keep the sequence)  
$ trimal -in inputfile -out outputfile -resoverlap 0.75 -seqoverlap 80
```

- Trimal can also convert formats of files (fasta to PHYLIP, NEXUS, etc)

Phylogenetic analyses

- Neighbor-Joining: Rapid, fast tree building. Distance based

```
#!/usr/bin/bash
#SBATCH --nodes 1 --ntasks 1
module load fasttree
FastTree VMS1.aa.trim > VMS1.aa.nj.tre
```

- FastTree automatically bootstraps 1000x
- FastTreeMP will run on multiple processors
- Many parameters to specify if nucleotide sequences, distributions of site variance, etc

Maximum Likelihood

- [RAxML](#) or [IQTree](#) on the cluster

```
#!/usr/bin/bash
#SBATCH --ntasks 2 --nodes 1
module load IQ-TREE
iqtree-omp -s VMS1.aa.trim -nt 2
```

```
#!/usr/bin/bash
#SBATCH --ntasks 4 --nodes 1
module load RAXML
raxmlHPC-PTHREADS-SSE3 -m PROTGAMMAAUTO -T 4 -s VMS1.aa.trim -n VMS1_Run1 -
```

Viewing trees

- iTOL - <https://itol.embl.de/>
- Figtree - <http://tree.bio.ed.ac.uk/software/figtree/>
<https://github.com/rambaut/figtree/>

```
# with X11 enabled on your laptop  
$ module load figtree  
$ figtree file.tre
```

Advanced Models

- Model fit - to identify most likely model of seq evolution : ModelTest, jModelTest, ProtTest
- Concatenated gene sets, allow partitioning of data : Partition Finder

more you can do in BioPython

BioPython tutorial <http://biopython.org/wiki/Phylo> and Cookbook [http://biopython.org/wiki/Phylo cookbook](http://biopython.org/wiki/Phylo_cookbook)

- Read / Write Trees
- Construct trees from MSA (distance and parsimony)
- Bootstraps

Traverse a tree

```
#!/usr/bin/env python3

from Bio import Phylo
trees = Phylo.parse("treefile1.tre", "newick")

def all_parents(tree):
    parents = {}
    for clade in tree.find_clades(order='level'):
        for child in clade:
            parents[child] = clade
    return parents

for tree in trees:
    print(tree)
    Phylo.draw_ascii(tree)

    for tip in tree.get_terminals():
        print("terminal tip", tip)
        term_names = [term.name for term in tree.get_terminals()]

    parents = all_parents(tree)
    clades = tree.find_clades("E")
    for myclade in clades: # get first instance of 'E'
        parent_of_myclade = parents[myclade]
```