# Finding Orthologs and Paralogs

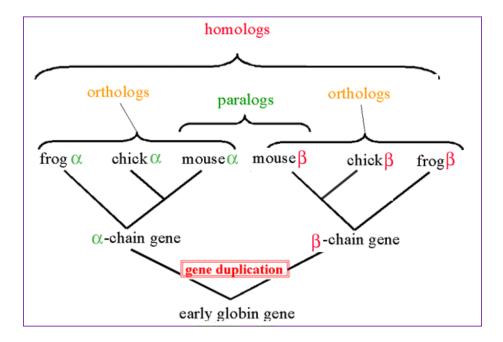


Figure 1: Orthologs

#### Gene families and Orthology

Problem: How to find "same" genes across multiple species.

Genes can duplicate (Paralogs) and can be identical due to descent (Ortholog)

#### Methods

- BLAST: 1 way BLAST (Gene A in Species X, what is best hit in Species Y)
- BLAST: reciprocal BLAST

#### Trees can help resolve relationships

Best hits can sometimes be wrong (B) though it can be resolved with phylogenetics.

### Reciprocal Searches

• Bi-directional or Reciprocal BLAST

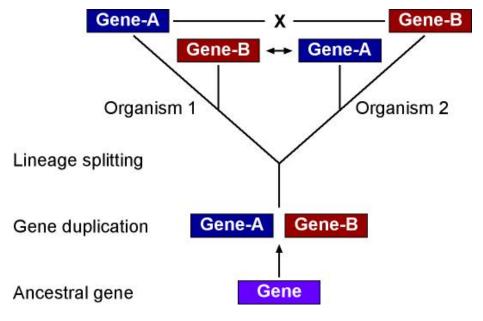


Figure 2: orthologs

#### Implement Bidirectional

Method to find best top hit in one direction and the reverse.

Let's walk through the code

Will write this in Python in Class

### Clustering

- Lumping genes together based on similarity linkage
- Single-linkage means if there is a link between A-B then they are in a cluster

#### Code up single-linkage

Let's look at some code.

Will write this in Python in Class

#### **Issues**

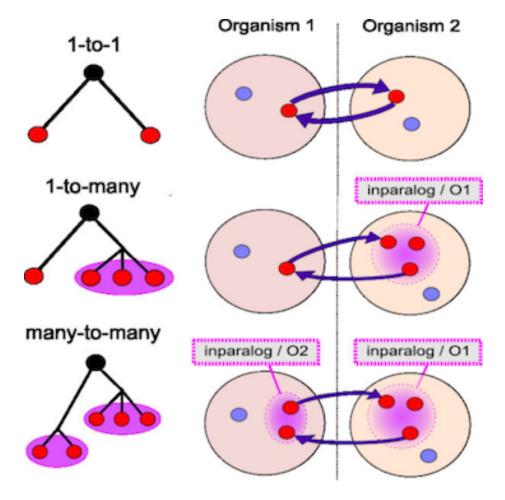


Figure 3: diagramorth

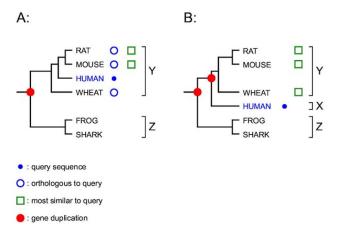


Figure 4: RIO

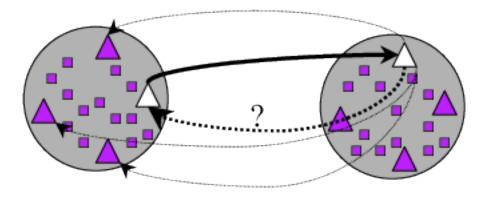


Figure 5: BRH

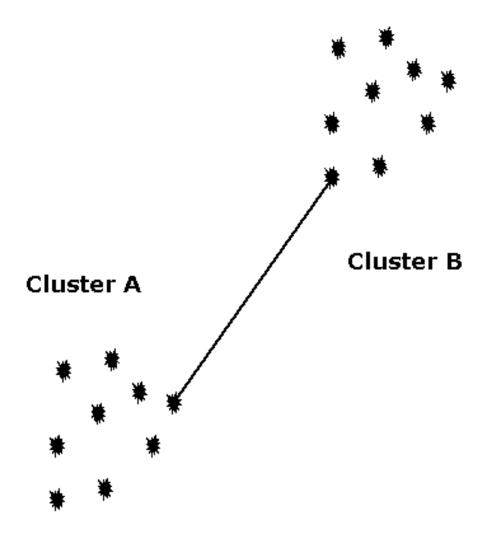


Figure 6: SingleLinkage

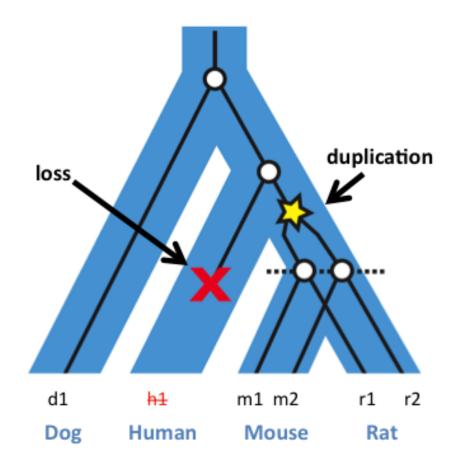


Figure 7: orthologsloss

# Tools to go after Orthologous and Paralogous equences

OrthoFinder

#### Steps to build orthologs on cluster

We will take 3 datasets of annotated Cyanobacteria, download and run analysis to generate Ortholog table.

```
#!/usr/bin/bash
#SBATCH --ntasks 16 --mem 8G -p short
module load ncbi-blast
module load orthofinder
module load miniconda2
CPU=8
mkdir -p cyanobacteria
cd cyanobacteria
curl -L -O ftp://ftp.ensemblgenomes.org/pub/bacteria/release-45/fasta/bacteria_10_collection
curl -L -O ftp://ftp.ensemblgenomes.org/pub/bacteria/release-45/fasta/bacteria_O_collection,
curl -L -O ftp://ftp.ensemblgenomes.org/pub/bacteria/release-45/fasta/bacteria_4_collection,
# uncompress files and name them all *.fasta
for file in *.fa.gz
do
m=$(basename $file .pep.all.fa.gz)
pigz -dc $file > $m.fasta
done
cd ..
```

## Ortholog results

orthofinder.py -a \$CPU -f cyanobacteria

The output file by default will be the date of the anlaysis. Opening the file cyanobacteria/Results\_XXX/Orthogroups.txt but I made a folder in the examples you look over. Here's one table

#### **Format**

```
GroupName\tSp1_Gene1, Sp1_Gene2\tSp2_Gene1, Sp2_Gene2\tSp3_Gene1, Sp3_Gene2

Cyanobacterium_aponinum_pcc_10605.ASM31767v1 Nostoc_punctiforme_pcc_73102.ASM2002v1

DG00000000 EKQ66605, EKQ66611, EKQ66662, EKQ66782, EKQ66954, EKQ66984, EKQ67084, EK
```

EKQ67433, EKQ67590, EKQ67680, EKQ67799, EKQ67807, EKQ67983, EKQ68026, EKQ68032, EKQ68054, I EKQ69279, EKQ69300, EKQ69345, EKQ69368, EKQ69506, EKQ69549, EKQ69629, EKQ69630, EKQ69655, E KQ70786, EKQ70840, EKQ70870, EKQ70894, EKQ71088, EKQ71090, EKQ71265, EKQ71335 AFZ52442, AFZ54265, AFZ54640 ACC78968, ACC78978, ACC79054, ACC79090, ACC79138 81797, ACC82091, ACC82628, ACC82978, ACC83035, ACC83215, ACC83711, ACC84528, ACC84844, ACC84 69971, EKQ69995, EKQ70003, EKQ70556, EKQ70833, EKQ71286 AFZ55137 ACC79344, ACC80485, ACC80595, ACC82143, ACC82836, ACC82962, ACC83849 ACC84972, ACC84974, ACC84981, ACC84982, ACC84983, ACC85032 EKQ66950, EKQ67597, EKQ67615, El DG0000003 AFZ53198 ACC78875, ACC78976, ACC79256, ACC79524, ACC79759, ACC80145, ACC80528 ACC82769, ACC83025, ACC83081, ACC83457, ACC83602, ACC83721, ACC83749, ACC84422, ACC85331 ACC80422, ACC80525, ACC80662, ACC80851, ACC80857, ACC80914, ACC81440, ACC819 6, ACC83981, ACC84622, ACC84732, ACC85457 EKQ66830, EKQ66911, EKQ67039, EKQ67311, EKQ6999 AFZ52318, AFZ52611, AFZ52613, AFZ52925, AFZ52973, AFZ53626, AFZ53840, AFZ53841, OG0000005 CC82559, ACC83603, ACC83674, ACC85005, ACC85009 EKQ67574, EKQ67809, EKQ69976 AFZ52319, AFZ53394, AFZ54017, AFZ54472 ACC79360, ACC79745, ACC79853, ACC80832, DG0000006 478, EKQ67551, EKQ67724, EKQ67810, EKQ68266 ACC79786, ACC80242, ACC80282, ACC80538, ACC80768 DG0000007 AFZ53704, AFZ54461, AFZ54462 2, EKQ68369, EKQ70142, EKQ70145, EKQ71300

The tool also generates summary statistics we can look through.

Could write a script to turn this into a table or use the summary count table provided.

ORTHOLOG_GRP	SP1	SP2		SP3
ORTHO_0001	10	)	5	
ORTHO 0002	1	L	1	