

# Short Sequencing Read Mapping

## BWA for short read alignment

### Index genome

It is necessary to index the genome in preparation of alignment.

```
#SBATCH -p short -N 1 -n 2 --mem 2gb
module load bwa
GENOME=S_enterica_CT18.fasta
bwa index $GENOME
```

### Align reads

```
#!/usr/bin/bash
#SBATCH -p short -N 1 -n 16 --mem 4gb

module load bwa
module load samtools
CPU=16
mkdir -p ~/bigdata/Short_read_aligning
cd ~/bigdata/Short_read_aligning
mkdir -p fastq
ln -s /bigdata/gen220/shared/data/S_enterica/*.fastq.gz fastq
ln -s /bigdata/gen220/shared/data/S_enterica/S_enterica_CT18.fasta
ln -s /bigdata/gen220/shared/data/S_enterica/acc.txt
GENOME=S_enterica_CT18.fasta
if [ ! -f $GENOME.sa ]; then
    bwa index $GENOME
fi

for acc in $(cat acc.txt)
do
    FWDREAD=fastq/${acc}_1.fastq.gz
    REVREAD=fastq/${acc}_2.fastq.gz

    bwa mem -t $CPU $GENOME $FWDREAD $REVREAD > ${acc}.sam
    samtools fixmate -O bam ${acc}.sam ${acc}_fixmate.bam
    samtools sort --threads $CPU -O BAM -o ${acc}.bam ${acc}_fixmate.bam
    samtools index ${acc}.bam
done
```

## Visualizing depth of coverage

Interactively - you can use `samtools`

```
module load samtools
samtools tview SRR10574912.bam
```

## SNP calling

There are many standardized SNP calling pipelines. [GATK](#) provides a robust pipeline that can be used.

[Samtools/BCFTools](#) are also useful and straight forward.

[freebayes](#) is another very useful pipeline for non-model systems.

## Samtools/BCFTools SNP and INDEL calling

[Workflows from the htlib](#)

```
#SBATCH -p batch -N 1 -n 4 --mem 16gb
module unload perl
module load samtools
module load bcftools
GENOME=S_enterica_CT18.fasta

# need to make a string which is all the bam files you want to process
# but if we do *.bam it will catch the intermediate bam files that are in the folder
for a in $(cat acc.txt)
do
    m="$a.bam $m"
done

VCF=Salmonella.vcf.gz
VCFFILTER=Salmonella.filtered.vcf.gz
bcftools mpileup -Ou -f $GENOME $m | bcftools call -vm0 z -o $VCF
tabix -p vcf $VCF
bcftools stats -F $GENOME -s - $VCF > $VCF.stats
mkdir -p plots
plot-vcfstats -p plots/ $VCF.stats
bcftools filter -O z -o $VCFFILTER -s LOWQUAL -i '%QUAL>10' $VCF
```

## Genome Browsers

We will do more on genome browsers later in the course. But if you want to see how to visualize genome you can see some of these tools.

### IGV

[IGV](#) - High-performance visualization tool for interactive exploration of large, integrated genomic datasets. It supports a wide variety of data types, including array-based and next-generation sequence data, and genomic annotations.

### JBrowse

[JBrowse2](#) provides google-maps style interface to genomes

## Public genome browsers

Many browsers allow upload of aligned data (bam files) to integrate local data with public genome resources.

- [Ensembl](#), [Ensembl Genomes](#)
- [UCSC Genome Browser](#)
- [WormBase](#), [FlyBase](#)
- [TAIR](#) - [Arabidopsis](#), [Phytozome](#)
- [EuPathDB](#), [JGI Genomes](#)
- [IMG/M](#) - [JGI](#)

## Displaying data in Ensembl

Go to [Ensembl Site for Salmonella enterica subsp. enterica serovar Typhi str. CT18](#)

See [the Ensembl tutorial](#) on how to add a BAM file track (note this only works if you have aligned reads to the SAME ASSEMBLY that is in Ensembl).

Click on “Display your data in Ensembl Bacteria”

Make a link on the web for your data. Follow directions on [HPCC site](#)

```
mkdir -p ~/.html/share
cd ~/.html/share
ln -s ~/bigdata/Short_read_aligning . # or wherever you were doning
```

Now you can use the URL [http://cluster.hpcc.ucr.edu/~YOURLOGIN/share/Short\\_read\\_aligning](http://cluster.hpcc.ucr.edu/~YOURLOGIN/share/Short_read_aligning) and the .bam files that are in there.