# **Python Packages**

### Python 3

To use Python 3 on the cluster you can use a module that has it as the default.

```
$ module load miniconda3
```

This will load an environment where python3 is the python installed. You can check that this works by doing python -V and see the version is 3.XX

#### You can setup your own conda environment

This will allow you to install python packages as well as any other pkgs you might want to.

Do this one-time

```
mkdir ~/bigdata/.conda
ln -s ~/bigdata/.conda ~/.conda
conda create -y -n gen220 python=3
source activate gen220
conda config --add channels defaults
conda config --add channels bioconda
conda config --add channels conda-forge
conda install biopython bcbio-gff
```

#### Python Libraries

Python has many built in packages already installed and a whole host of contributions that span everything from plotting data, interfacing with other datatypes.

- https://docs.python.org/3/library/gzip.html
- https://docs.python.org/3/library/csv.html
- PyBed tools (processing BED files) https://daler.github.io/pybedtools/
- BioPython GFF parsing
- There is a nice list of available libraries https://wiki.python.org/moin/Use fulModules

#### Gzip

Can open a file that is compressed or write to a file already compressed. This can save space for large files or when you get data from a resource without having to decompress it.

```
import gzip
with gzip.open(file,"rt") as fh:
    for line in fh:
        print("The first line from uncompressed")
    break
```

#### URL / Web requests directly

```
# this is a URL at the UniProt database to get a protein sequence based on
# accession number
import urllib.request
url="https://www.uniprot.org/uniprot/P10127.fasta"
seqdata = urllib.request.urlopen(url)
for line in seqdata:
    linestrip = line.decode('UTF-8').strip()
    print(linestrip)
```

#### BioPython - a library of modules for bioinformatics

BioPython Tutorial

Modules for Sequence data, BLAST parsing, Multiple alignments

Already installed on biocluster

To installed on own computer you control use Python tool 'pip'

```
$ conda create -y -n gen220 # only need to do this once
$ source activate gen220
$ conda install biopython # if you forget to do the line above it will fail
```

### Simple BioPython

```
import Bio
from Bio.Seq import Seq
my_seq = Seq("ATGAGTACACTAGGGTAA")
print(my_seq)

rc = my_seq.reverse_complement()
pep = my_seq.translate()
print("revcom is", rc)
print(pep)
```

#### Parsing sequence files

more /bigdata/gen220/share/data/E3Q6S8.fasta >tr|E3Q6S8|E3Q6S8\_COLGM RNAse P Rpr2/Rpp21/SNM1 subunit domain-containing protein OS=Collete MAKPKSESLPNRHAYTRVSYLHQAAAYLATVQSPTSDSTTNSSQPGHAPHAVDHERCLET NETVARRFVSDIRAVSLKAQIRPSPSLKQMMCKYCDSLLVEGKTCSTTVENASKGGKKPW ADVMVTKCKTCGNVKRFPVSAPRQKRRPFREQKAVEGQDTTPAVSEMSTGAD

To process this file:

```
import sys
import Bio
```

```
from Bio import SeqIO
from Bio.Seq import Seq
# segfile
filename = "/bigdata/gen220/shared/data/E3Q6S8.fasta"
for seq_record in SeqIO.parse( filename , "fasta"):
    print(seq_record.id)
    print(repr(seq_record.seq))
    print(seq_record.seq)
    print(len(seq_record))
This will output:
tr|E3Q6S8|E3Q6S8_COLGM
Seq('MAKPKSESLPNRHAYTRVSYLHQAAAYLATVQSPTSDSTTNSSQPGHAPHAVDH...GAD',
SingleLetterAlphabet())
MAKPKSESLPNRHAYTRVSYLHQAAAYLATVQSPTSDSTTNSSQPGHAPHAVDHERCLETNETVARRFVSDIRAVSLKAQIRPSPSLKQMM
172
GenBank files: another sequence format
LOCUS
            AJ240084
                                    1905 bp
                                               DNA
                                                       linear PRI 03-FEB-2000
DEFINITION Homo sapiens TRIM gene, promoter.
ACCESSION
            AJ240084
VERSION
            AJ240084.1 GI:6911579
KEYWORDS
            T-cell receptor interacting molecule; TRIM gene.
SOURCE
            Homo sapiens (human)
  ORGANISM Homo sapiens
      Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi;
     Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini;
      Catarrhini; Hominidae; Homo.
REFERENCE
          Hubener, C., Mincheva, A., Lichter, P., Schraven, B. and Bruyns, E.
AUTHORS
TITLE
          Genomic organization and chromosomal localization of the human gene
          encoding the T-cell receptor-interacting molecule (TRIM)
          Immunogenetics 51 (2), 154-158 (2000)
JOURNAL
 PUBMED
          10663578
REFERENCE
           2 (bases 1 to 1905)
```

Submitted (06-MAY-1999) Huebener C., Immunomodulation Laboratory,

Institute for Immunology, University of Heidelberg, Im Neuenheimer

/organism="Homo sapiens"
/mol\_type="genomic DNA"

Feld 305, Heidelberg, 69120, GERMANY

Location/Qualifiers

AUTHORS

JOURNAL

source

TITLE

**FEATURES** 

Huebener, C.

Direct Submission

1..1905

```
/gene="TRIM"
     regulatory
                     1..1746
                     /regulatory_class="promoter"
                     /gene="TRIM"
     5'UTR
                     1747..1902
                     /gene="TRIM"
ORIGIN
   1 ccaaaaattt ccagtcctga aaccctttct ctttccaatg tcctctgtaa gctcgagttg
  61 tgggcatcta ctttgcccat attccaaggt cttgcttagg taacctctgt agtcctttct
 121 tgagcctagg acttctactt ttcttaccag ttaccctctt tcaggaccaa agctcaactc
 181 ctcaaggcca taactaggcc ctctcctctc aaactgattt atcaggtgcc cgaatcttcc
 241 tgaatgtctg ggattcaact tttcagcagt cttcctccct acgttccatc taattctaag
 301 atgaaacctt ctgattcttt gttgtcctct gatccctaca tgaacctgag gctgctgttc
 361 cctgaagtct tgttctgtca gcatccaggc ctgcttcata aaacctgtca ctctgctaat
 421 ggttagcggc tgaacaaaga gtcctctggc caaataagtt tagaaaaact ctgataaaaa
 481 tattatttgg gtttcctttt cgcaggactt acctaagcct ttaatatgca tctacggagg
 541 taaaaataaa gctatatatt ttttccaaag atatttgttg aagaaacatt tgtcttctgc
 601 gtttcttaaa ggccgagtgt tctatggaac atactttaaa aaaccctttt aaagaagctt
 661 agaccagaga atctccaagg tctctttcag ttttacagcc tctgagtcaa cgattcacca
 721 aaaaatattt tggggggaag tgattgaagt ggaaaaattt gttagtgttt agccagcttt
 781 gtccaaagga taagatgcac tgtattttgc ttactaggga gttattttct ataatggaag
 841 acaaagaaag cacaagacac ccatggtttt gtttgttcaa tcactgagag taagtctcaa
 901 ttattgagac ttacgatgtg ccggtgtgct taattctagt tatgaaattt taataatgaa
 961 taatatagat tctattcctt atatgagttt ccaaaagcat tgtccagaac atctatatta
1021 aaatatetta teatataeaa tatatgtaat ttaaaatgea eteagaaaat etgettgtta
1081 aaatgcagat tctagtgctt caccctaaat agtctaattt agacgggccc aggattttaa
1141 actagcatct tatagcatac ttatgtacac caacatgtaa gaactgctgc tattaagatt
1201 ctgggatggt ggttgagaac aggagcttgt tgtcaggtgg ctctagattg gacagagaaa
1261 ctcatactga taaggtgagg attgtcagga aataaggcag gcatctagcc tcgcattaag
1321 atgaggtata gaaggcaact gatacatact aagtgctcaa aaaatattaa ctccctgtcc
1381 tccatcatgg ctcaagaaaa tacaacagct gagcacaccc acgggttgct tactatttac
1441 ttatcagttt agtgtatctt attttgtttc catgtgaatt tacttgtgaa gagatgactg
1501 gattctctcc agagatagga agatccctcc tggtttaatt cctaccttta tttatttatt
1561 tttcaattag actcaggtat tgataaaaat tcaaatgtca gattacaaag gtgtgtggga
1621 tttttcttcc cacgttacac aatttaagtc gactgttttc agatcaaaac tcaagacaac
1681 tccttcacca catttcctgt ttgtaactga aacaaagtac acacaaaaga ttttaagaaa
1741 cagaagagaa aagaatccga ggcacagata aagataagtt ttactgtcat gctgctttta
1801 acataacaga gcaacatcac ctaggaaaaa agtttgtagg aggattttta atccatatat
1861 ttgtcttatg gctagataaa gatttctccg aaaaaaagaa gcatg
```

/db\_xref="taxon:9606"

1..1902

gene

//

/clone\_lib="RPCI1,3-5 Human PAC library"

## Now parse GenBank

To Parse the genbank file - it is the same code! Just change the format.

```
import sys
import Bio
from Bio import SeqIO
from Bio.Seq import Seq
# seqfile
filename = "/bigdata/gen220/shared/data/AJ240084.gbk"
for seq_record in SeqIO.parse( filename , "genbank"):
   print(seq_record.id)
   print(repr(seq_record.seq))
   print(seq_record.seq)
   print(len(seq_record))
Produces
python bp_parse_gbk.py ../data/AJ240084_TRIM.gbk
AJ240084.1
Seq('CCAAAAATTTCCAGTCCTGAAACCCTTTCTCTTTCCAATGTCCTCTGTAAGCTC...ATG',
IUPACAmbiguousDNA())
1905
```

 $\frac{}{\#}$  Parse fea- ${\rm tures}$  ${\rm from}$  $\operatorname{Gen-}$  $\operatorname{Bank}$ file See documenta- ${\rm tion}$ on Se- $\operatorname{qIO}$ here and the tutorial $\overline{\text{urlcolor}}$ :

blue