

THE BIOGEOGRAPHIC ATLAS OF THE SOUTHERN OCEAN

The "Biogeographic Atlas of the Southern Ocean" is a legacy of the International Polar Year 2007-2009 (www.ipy.org) and of the Census of Marine Life 2000-2010 (www.coml.org), contributed by the Census of Antarctic Marine Life (www.caml.aq) and the SCAR Marine Biodiversity Information Network (www.scarmarbin.be; www.biodiversity.aq).

The "Biogeographic Atlas" is a contribution to the SCAR programmes Ant-ECO (State of the Antarctic Ecosystem) and AnT-ERA (Antarctic Thresholds- Ecosystem Resilience and Adaptation) (www.scar.org/science-themes/ecosystems).

Edited by:

Claude De Broyer (Royal Belgian Institute of Natural Sciences, Brussels)

Philippe Koubbi (Université Pierre et Marie Curie, Paris)

Huw Griffiths (British Antarctic Survey, Cambridge)

Ben Raymond (Australian Antarctic Division, Hobart)

Cédric d'Udekem d'Acoz (Royal Belgian Institute of Natural Sciences, Brussels)

Anton Van de Putte (Royal Belgian Institute of Natural Sciences, Brussels)

Bruno Danis (Université Libre de Bruxelles, Brussels)

Bruno David (Université de Bourgogne, Dijon)
Susie Grant (British Antarctic Survey, Cambridge)
Julian Gutt (Alfred Wegener Institute, Helmoltz Centre for Polar and Marine Research, Bremerhaven)

Christoph Held (Alfred Wegener Institute, Helmoltz Centre for Polar and Marine Research, Bremerhaven)
Graham Hosie (Australian Antarctic Division, Hobart)

Falk Huettmann (University of Alaska, Fairbanks)

Alix Post (Geoscience Australia, Canberra)

Yan Ropert-Coudert (Institut Pluridisciplinaire Hubert Currien, Strasbourg)

Published by:

The Scientific Committee on Antarctic Research, Scott Polar Research Institute, Lensfield Road, Cambridge, CB2 1ER, United Kingdom (www.scar.org).

Publication funded by:

- The Census of Marine Life (Albert P. Sloan Foundation, New York)
- The TOTAL Foundation, Paris

The "Biogeographic Atlas of the Southern Ocean" shared the Cosmos Prize awarded to the Census of Marine Life by the International Osaka Expo'90 Commemorative Foundation, Tokyo, Japan.

Publication supported by:

- The Belgian Science Policy (Belspo), through the Belgian Scientific Research Programme on the Antarctic and the "biodiversity.aq" network (SCAR-MarBIN/ANTABIF)
- The Royal Belgian Institute of Natural Sciences (RBINS), Brussels, Belgium
 The British Antarctic Survey (BAS), Cambridge, United Kingdom
- The Université Pierre et Marie Curie (UPMC), Paris, France
- The Australian Antarctic Division, Hobart, Australia
- The Scientific Steering Committee of CAML, Michael Stoddart (CAML Administrator) and Victoria Wadley (CAML Project Manager)

Mapping coordination and design: Huw Griffiths (BAS, Cambridge) & Anton Van de Putte (RBINS, Brussels)

Editorial assistance: Henri Robert, Xavier Loréa, Charlotte Havermans, Nicole Moortgat (RBINS, Brussels)

Printed by: Altitude Design, Rue Saint Josse, 15, B-1210 Brussels, Belgium (www.altitude-design.be)

Lay out: Sigrid Camus & Amélie Blaton (Altitude Design, Brussels).

Cover design: Amélie Blaton (Altitude Design, Brussels) and the Editorial Team.

Cover pictures: amphipod crustacean (Epimeria rubrieques De Broyer & Klages, 1991), image © T. Riehl, University of Hamburg; krill (Euphausia superba Dana, 1850), image © V. Siegel, Institute of Sea Fisheries, Hamburg; fish (*Chaenocephalus* sp.), image © C. d'Udekem d'Acoz, RBINS; emperor penguin (*Aptenodytes forsteri* G.R. Gray, 1844), image © C. d'Udekem d'Acoz, RBINS; Humpback whale (*Megaptera novaeangliae* (Borowski, 1781)), image © L. Kindermann, AWI

Online dynamic version:

A dynamic online version of the Biogeographic Atlas is available on the SCAR-MarBIN / AntaBIF portal: atlas.biodiversity.aq.

Recommended citation:

De Broyer C., Koubbi P., Griffiths H.J., Raymond B., Udekem d'Acoz C. d', Van de Putte A.P., Danis B., David B., Grant S., Gutt J., Held C., Hosie G., Huettmann F., Post A., Ropert-Coudert Y. (eds.), 2014. Biogeographic Atlas of the Southern Ocean. Scientific Committee on Antarctic Research, Cambridge, XII + 498 pp.

For individual chapter:

(e.g.) Crame A., 2014. Chapter 3.1. Evolutionary Setting. In: De Broyer C., Koubbi P., Griffiths H.J., Raymond B., Udekem d'Acoz C. d', et al. (eds.). Biogeographic Atlas of the Southern Ocean. Scientific Committee on Antarctic Research, Cambridge, pp. xx-yy.

ISBN: 978-0-948277-28-3



This publication is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License



10.6. Phylogeographic patterns of the Lysianassoidea (Crustacea: Peracarida: Amphipoda)

Charlotte Havermans

Royal Belgian Institute of Natural Sciences, Brussels, Belgium

1. Introduction

The Lysianassoidea Dana, 1849 is the largest superfamily belonging to the Gammaridea and currently comprises more than 1000 described species, 172 genera and 18 families worldwide (Horton & De Broyer 2013). Lysianassoids cover a vast range of habitats and are characterised by a wide array of morphological adaptations. Many genera comprise species that are bentho-pelagic or demersal scavengers of which some occupy abyssal and hadal depths. Other genera comprise soft-bottom deposit-feeders or algal-dwellers (Lowry & Stoddart 1983). Commensal life styles are also well documented in several genera, with species living on other invertebrates (e.g. sponges, ascidians), as well as ectoparasitism on fish (Lowry & Stoddart 1983).

This superfamily is a typical cold-water group, occurring primarily at water temperatures below 10°C (Barnard & Karaman 1991). They are common in both deep-sea basins (e.g. Thurston 1979) and high-latitude shallow waters (e.g. Vader & Romppainen 1986), where they significantly contribute to the benthic species number. In terms of composition, the Antarctic and sub-Antarctic amphipod fauna is mostly dominated by representatives of the Lysianassoidea (158 spp. of 57 different genera; De Broyer et al. 2007; Horton & De Broyer 2013). Both in polar oceans and in the world's deep seas, lysianassoids comprise one of the most abundant and widespread macrofaunal scavenger groups (Slattery & Oliver 1986). It has been shown recently that the morphological traits associated with the scavenger mode of feeding evolved several times independently within the Lysianassoidea (Havermans et al. 2010, Corrigan et al. 2013). Scavenging can be considered as an adaptation to the generally or seasonally low food supply in terms of primary production in the deep sea and polar environments, respectively. The diversification of several scavenging lysianassoid lineages was found to coincide with the Eocene/Oligocene cooling, characterized by an increased productivity and the availability of novel food sources, representing an ecological opportunity for specialized scavengers (Corrigan et al. 2013).



Photo 1 Eurythenes gryllus (Polarstern ANTXXIII-8, st. 683-1). Image © C.d'Udekem (RBINS), Brussels.

The Antarctic food web generates significant quantity of food falls, which are less seasonally dependent, composed of fishes, seabirds, seals and cetaceans (Dauby et al. 2001). As a consequence, the Antarctic slope is richer in scavenger species compared to similar depth ranges elsewhere (De Broyer et al. 2004). The occurrence in high numbers of lysianassoids at food falls has been widely documented by means of catches with baited traps (e.g. Stockton 1982, De Broyer et al. 2004, Jamieson et al. 2011, Horton et al. 2013) and baited camera studies (e.g. Klages et al. 2001, Soltwedel et al. 2003, Premke et al. 2003), from below the ice shelf in polar regions up to hadal depths in world oceans.

Molecular methods unveiled more cryptic species than expected in the Southern Ocean (SO) benthos, in view of the generally assumed circum-Antarctic species distributions (e.g. Held 2003, Held & Wägele 2005, Wilson et al. 2007). In addition, cryptic species have also been observed along depth gradients within species considered to be eurybathic (e.g. Brandao et al. 2010, Schüller 2011). In this respect, amphipods, as well as other peracarids, represent an interesting group to test these hypotheses on faunal distributions. Peracarids are brooders and the lack of drifting stages might considerably limit their dispersal, rendering them particularly subject to cryptic speciation processes. However, even though lysianassoids are brooders, a number of representatives of this superfamily are known to be very mobile. For example, species of the genera Abyssorchomene De Broyer, 1984 and Pseudorchomene Schellenberg, 1926 can form swarms of thousands of individuals, capable of considerable migrations in search of pelagic prey or food falls on the seafloor (Dauby et al. 2001, Horton et al. 2013). Eurythenes gryllus (Lichtenstein, 1822) is known for its impressive swimming speeds, being able to swim against slow currents (Laver et al. 1985, Takeuchi & Watanabe 1998). Many species are thought to be circum-Antarctic (Orchomenella (Orchomenella) pinguides) (Walker, 1903) and/or eurybathic (e.g. Orchomenella (Orchomenopsis) cavimanus) (Stebbing, 1888), some even cosmopolitan (e.g. Abyssorchomene chevreuxi (Chevreux, 1903), Eurythenes gryllus), and there is evidence for gene flow between distinct deep-sea basins in the case of Abyssorchomene species (France 1994). Finally, by representing a coldwater group that has diversified both in polar regions and the deep sea, lysianassoids are of particular interest for molecular studies aiming to test the connectivity between the Southern Ocean and surrounding deep seas.

2. Molecular results

2.1. Genetic structure within and between species of *Orchomene* sensu lato

DNA barcoding was used to test the hypotheses of circumpolarity and eurybathy within the complex of genera *Orchomene sensu lato* (Havermans *et al.* 2011, Havermans 2012). Orchomenid species' distributions were evaluated by comparing intra- versus interspecific distances of COI sequences and linking these to geographic and bathymetric data. For all lysianassoid species investigated, intra- and interspecific COI divergences followed a bimodal distribution with a mean intraspecific variation of 1.7% and a mean interspecific divergence of 14.8 % separated by a barcoding gap of 1.5% (Kimura 2-parameter distance model, K2P; Kimura 1980) (Havermans 2012).

Within this group of closely related species, distinct genetic structures could be observed. Genetic divergences between clusters observed within *Orchomenella* (*Orchomenopsis*) acanthurus (Schellenberg, 1931), O. (O.) cavimanus, O. (*Orchomenella*) franklini (Walker, 1903) and O. (O.) pinguides (Walker, 1903), were congruent with species-level divergences observed between other closely related species of the orchomenid genus complex (K2P: 6.3 – 20.1%; Havermans et al. 2011) (Fig. 1). Furthermore, several cases of sympatry were observed: specimens of the same or nearby sampling sites were found in distinct clusters. Both the bimodal distribution of intra- and interspecific divergences and the prevalence of species-level differentiation of sympatric specimens represent criteria for the presence of cryptic or over-

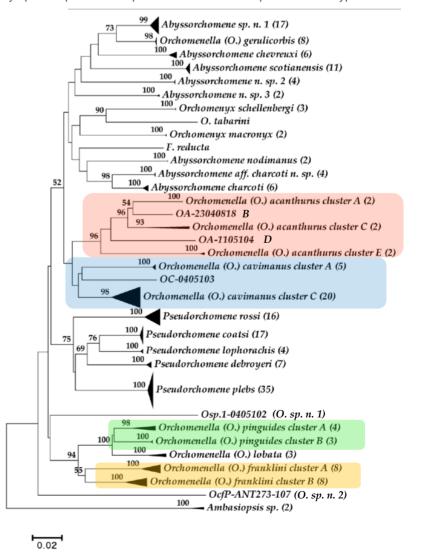
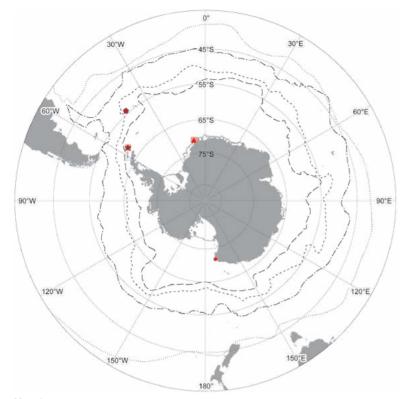
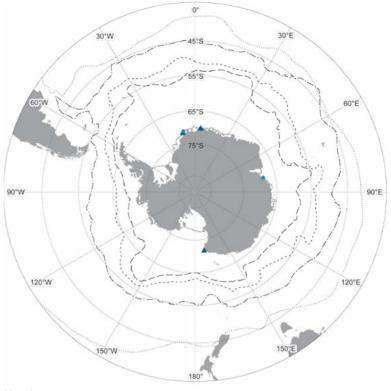


Figure 1 Neighbour-Joining tree of COI sequences of species of *Orchomene sensu lato* based on the Kimura 2-parameter distance model. Clusters were collapsed and the number of specimens in every cluster is indicated in parentheses. Species complexes are indicated with coloured rectangles. Bootstrap support (number of replicates = 2000) is indicated at each node when higher than 50%.



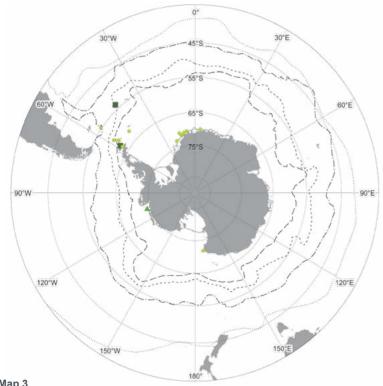


- Orchomenella (Orchomenopsis) acanthurus
- ▲ Orchomenella (Orchomenopsis) acanthurus
- Orchomenella (Orchomenopsis) acanthurus
- Orchomenella (Orchomenopsis) acanthurus ★ Orchomenella (Orchomenopsis) acanthurus



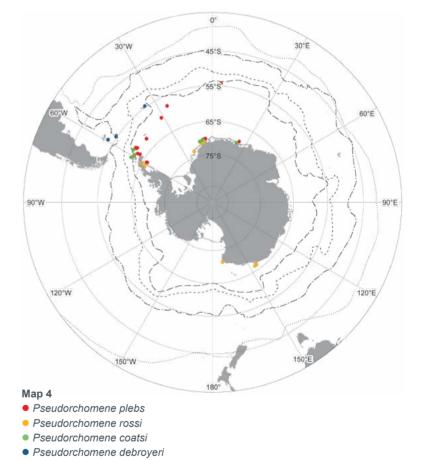
Map 2

- ▲ Orchomenella (Orchomenella) pinguides
- Orchomenella (Orchomenella) pinguides





- Orchomenella (Orchomenopsis) cavimanus
- ▲ Orchomenella (Orchomenopsis) cavimanus
- Orchomenella (Orchomenopsis) cavimanus



Lysianassoidea Maps 1-4 Distribution of the five species-level lineages observed within Orchomenella (Orchomenopsis) acanthurus based on molecular data. Map 2 Distribution of the two species-level lineages observed within *Orchomenella* (*Orchomenella*) *pinguides* based on molecular data. Map 3 Distribution of the three species-level lineages observed within *Orchomenepla* (*Orchomenopsis*) *cavimanus* based on molecular data. Map 4 Distribution of *Pseudorchomene plebs*, *P. rossi*, *P. coatsi* and *P. debroyeri* based on molecular data.

looked species (Held 2003). Hence, these four species, consisting of at least two lineages, represent complexes of cryptic or pseudocryptic species, of which the distribution was either more restricted than previously assumed (e.g. for O. (O.) acanthurus) or rather widespread (cluster A of O. (O.) franklini, two clusters of O. (O.) pinguides and cluster C of O. (O.) cavimanus) (Havermans 2012). Within O. (O.) acanthurus (Fig. 1), three well-supported clusters can be observed, as well as two divergent single sequences (pairwise sequence divergences ranging from 5.8 up to 22.6%). These five specieslevel lineages all seemed to be restricted in distribution to a single Antarctic shelf or shallow-water region (max. depth of 400 m) (Map 1). Conversely, both species-level clades (A and B) of O. (O.) pinguides cover a large geographic distribution (Map 2). Specimens of cluster A were found on opposite sides of the Antarctic continent whilst specimens of cluster B were found in the Eastern Weddell Sea and on the East Antarctic coast. (O.) (O.) cavimanus consisted

of three well-supported species-level lineages, of which two lineages (A and B) seemed to be confined to a particular region whilst cluster C encompassed a wide geographic and bathymetric range (Map 3). Specimens of lineage A were found in shallow waters in the Scotia Sea and near King George Island; lineage B was composed of a single specimen (OC-0405103) from a shallowwater locality in the Amundsen Sea (Fig. 1). On the contrary, cluster C comprised specimens from the Scotia Sea, localities near the South Orkney and South Shetland Islands, the eastern Weddell Sea, the Ross Sea and the Magellan region (Burdwood Bank) (Map 3), covering a large bathymetric range from 130 to more than 3600 m depth. However, no subclusters based on a geographic or bathymetric segregation could be observed within this cluster, characterised by a gradient of intraspecific distances (K2P: 0.0 - 5.6%).

Conversely, in several species of the genera Abyssorchomene and Pseudorchomene (A. sp. 1, A. chevreuxi, P. plebs, P. rossi, P. coatsi), a genetic homogeneity was found between specimens from remote sampling sites in the SO (and Atlantic for *Abyssorchomene* species), indicating a widespread or even a truly circum-Antarctic and/or eurybathic distribution (Map 4). In *Pseudorchomene plebs* (Hurley, 1965), a low genetic variation was observed between specimens from the Scotia Sea, the Antarctic Peninsula, the eastern Weddell Sea and the shelf of Bouvet Island, across a depth gradient from 40 up to 2890 m (Havermans *et al.* 2011). The same was true for *P. rossi* (Walker, 1903), between sites in the Weddell and Ross seas, off the George V coast and near the Antarctic Peninsula (Havermans 2012). Specimens of *P. debroyeri* d'Udekem & Havermans, 2012 from shallow-water South Georgia and sub-Antarctic localities (Falkland Islands, Burdwood Bank) were separated by low genetic divergences, across the Polar Front (Map 4). *P. coatsi* (Chilton, 1912) was characterised by a genetic homogeneity between localities in the Scotia, Weddell and King Haakon VII seas (Havermans 2012).

For a number of species, molecular results confirmed the existence of species distributions extending from the SO to surrounding deep-sea basins. Low genetic divergences (K2P: 0.1 to 1.5%) were observed for an undescribed *Abyssorchomene* species (*A.* sp. 1, Map 5) between specimens from the Antarctic Peninsula, Weddell, Ross and Amundsen seas and the Argentine abyssal Basin, at depths ranging from 300 to more than 4500 m (Havermans 2012). This species thus represents a truly circum-Antarctic and eurybathic species, with its distribution extended to the Atlantic abyssal basins. *Abyssorchomene chevreuxi* was confirmed to be a candidate for a cosmopolitan distribution, since only low genetic divergences (K2P: 0.0 to 1.7%) separated specimens from the Weddell Sea, the Antarctic Peninsula region, the Southwest Atlantic and the North Atlantic (Map 5), covering depths from 3070 m to more than 4800 m (Havermans 2012 and unpublished data).

2.2. Phylogeography of Eurythenes gryllus sensu lato

Eurythenes gryllus (Lichtenstein, 1822) is one of world's most widely distributed amphipods, occurring in every major ocean over a depth range covering the bathyal, abyssal and hadal zones (De Broyer et al. 2007; Brandt et al. 2012; and references therein). Previously, the assumption of a cosmopolitan and eurybathic distribution was questioned due to the presence of several genetically and morphologically divergent lineages (Barnard 1961; Bucklin et al. 1987; France & Kocher 1996a, b; Escobar-Briones et al. 2010). This was further tested on a global scale by means of phylogeographic and species delimitation analyses on mitochondrial (COI, 16S rDNA) and nuclear (28S rDNA) sequence data (Havermans et al. 2013).

Nine genetic lineages within Eurythenes gryllus were revealed based on the most extensive dataset of 16S sequences, of which five were corroborated by the three-gene dataset (COI, 16S, 28S) (Havermans et al. 2013). Bimodal distributions of intra- and interspecific divergences were observed for all three genes and for COI a clear barcoding gap could be distinguished. Tree-construction methods on the three-gene dataset revealed five clades within E. gryllus (Map 6): Eg1, comprising specimens from bathyal depths (<3000 m) in various sites in the Arctic and Southern oceans, Eg2, comprising specimens from Antarctic abyssal (>3000 m) sites, Eg3, grouping specimens from the Antarctic and Atlantic at abyssal depths, Eg4 and Eg5, both comprising specimens from the abyssal Brazil Basin but separated by high genetic divergences. For the 16S dataset, based on sequences from Havermans et al. (2013) and sequences from previous studies (France & Kocher 1996b; Escobar-Briones et al. 2010), the same clades were recovered, complemented by four additional well-supported clades (Eg6-9). Eg3 now also comprised specimens from abyssal sites in the North Atlantic and the Pacific Ocean. In addition to the clades Eg4 and Eg5, another abyssal cluster (Eg6) was revealed comprising a specimen from the Brazil Basin and one from the Gulf of Mexico. Clade Eg8 grouped specimens from bathyal depths in the Iceland Basin and the Bahamas, whilst Eg9 comprised specimens at abyssal depths (3193 m) on the slope of a seamount in the Pacific. Finally, Eg7 represents a single sequence from a specimen from the same seamount but sampled at a different depth (3982 m).

A morphological analysis on the available specimens revealed small but consistent phenotypical differences between Eg1, Eg2, Eg3, Eg4+5 and Eg6 and for the specimen(s) of each clade, a unique combination of character states could be identified. Overall, five putative morphospecies were recognized within *E. gryllus* based on the material examined but no differences were detected between the clades Eg4 and Eg5, although characterised by relatively high genetic divergences. Specimens from the bipolar clade (Eg1), collected in proximity of the type locality (Greenland Sea), were the most similar to the holotype of *E. gryllus* (illustrated by Stoddart & Lowry 2004).

Results of the statistical parsimony network analysis (Fig. 2) recovered eight of these lineages as unconnected networks but Eg4 and Eg5 were grouped in one single network. Since these two sympatric clades were well-supported with a clear barcoding gap between intra- and interclade divergences for COI, but not for 16S and 28S, this might represent a case of recent or ongoing speciation. The bathyal network Eg1 (Map 6) comprised specimens from both Arctic (Svalbard, Canada Basin, Eastern Fram Strait) and SO regions (King George Island, Antarctic Peninsula, South Sandwich Islands, Weddell Sea) sampled from 839 to 2743 m depth. Additional specimens from the Kerguelen Islands (1947 m depth) were also found to cluster in Eg1 (Havermans, unpublished data). This bipolar clade grouped specimens of *E. gryllus* separated by distances up to 16,750 km but characterised by low genetic divergences (COI K2P: 0.8–2.0%) and sharing a single 16S haplotype.

More genetic lineages were encountered in abyssal than in bathyal regions: two species-level clades could be distinguished above 3000 m and at

least six species-level clades below 3000 m (Fig. 2). The two bathyal species-level clades identified here are both characterised by a very wide geographic range: one is bipolar and the other is found to occur both in the Bahamas and in the Iceland Basin. Conversely, all abyssal lineages but Eg3 and Eg6, seemed to be restricted to a single ocean basin. Finally, networks showed no overlap between bathyal and abyssal depths: two networks grouped specimens exclusively from above 3000 m depth each and six networks comprised only specimens from below 3000 m (Fig.2). This break around 3000 m also appeared in the phylogenetic trees.

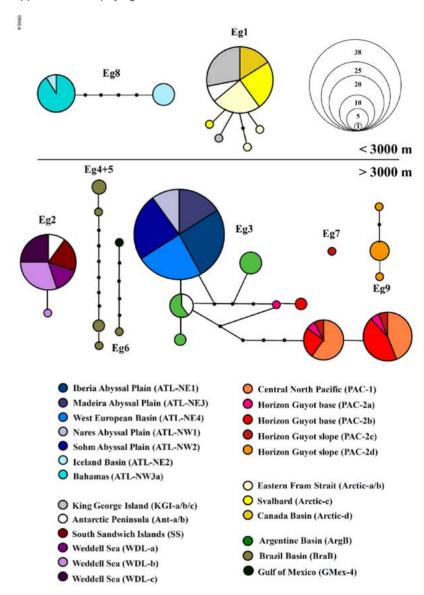


Figure 2 Statistical parsimony haplotype networks based on the 16S rDNA sequences of *Eurythenes gryllus* (from Havermans *et al.* 2013). The dataset includes sequences from this study, that of France and Kocher (1996b) and Escobar-Briones *et al.* (2010). The area of each circle is proportional to the frequency of the haplotype in our sampling (a scale is presented). Each line represents a single substitution, nodes represent hypothetical haplotypes and colors refer to the sampling localities. Haplotype networks (95% probability threshold) are numbered (Eg1-9) according to the different clusters identified in Bayesian trees (see Havermans *et al.* 2013).

2.3. (Pseudo)cryptic speciation in other lysianassoid species

Similar patterns of genetic structure as in Orchomene sensu lato have been observed in other (non-orchomenid) lysianassoid taxa. For the circum-Antarctic species Waldeckia obesa (Chevreux, 1905), two well-supported clusters were revealed with overlapping distributional ranges, separated by genetic distances (K2P: min. 15%) in the range of interspecific divergences known for lysianassoid taxa (Fig. 3) (Havermans 2012). One species-level lineage (B) was truly circum-Antarctic, occurring in the Lazarev, King Haakon VII, Amundsen and Dumont d'Urville seas, whilst the other (A) seemed to be restricted to the Atlantic sector of the SO (Map 7). Previously, small morphological differences have been reported between specimens from different regions (Chapelle pers. comm.). Hence, an in-depth morphological study is needed to confirm whether these lineages represent pseudocryptic species. Furthermore, the closely related epifaunal lysianassid species Parawaldeckia kidderi (Smith, 1876) represents a single species across the sub-Antarctic and a single mitochondrial haplotype was shared by Kerguelen, Macquarie, Marion and Possession Islands (Nikula et al. 2010).

Based on a limited sampling, two geographical lineages could be distinguished within *Uristes murrayi* (Walker, 1903) (Fig. 3): cluster A grouped specimens from the eastern Weddell and the King Haakon VII seas, while cluster B grouped specimens from the Ross Sea (Havermans 2012). By means of a multigene molecular study based on a more extensive sampling, Feldkamp (2010) observed the presence of three distinct species-level lineages within *U. murrayi*. Using morphometrics and scanning electron microscopy, consistent phenotypical differences were detected between specimens of these clades (Seefeldt 2012).

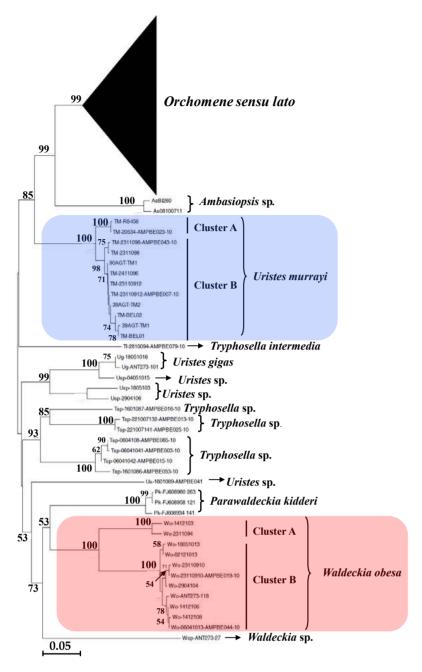


Figure 3 Neighbour-Joining tree of COI sequences of lysianassoid species based on K2P distances. (number of replicates = 2000) is indicated at each node when higher than 50%. Species where a high intraspecific variation was detected are indicated with coloured rectangles and the clusters are indicated next to the tree.

3. Geographic and bathymetric distributions

3.1. Circum-Antarctic, bipolar and cosmopolitan species

Molecular studies have shown evidence for circumpolarity in the species Abyssorchomene sp. 1, Pseudorchomene rossi and one species-level clade of Waldeckia obesa (Havermans 2012). Parawaldeckia kidderi exhibited a wide-scale circum-Antarctic distribution throughout a number of sampled sub-Antarctic islands (Nikula et al. 2010). Even though several species complexes were revealed as anticipated for Antarctic benthos, there was a limited number of cases where the distinct lineages showed a more restricted distribution than originally thought (e.g. all lineages of Orchomenella (Orchomenopsis) acanthurus, lineage A of Waldeckia obesa). Several complexes comprised lineages which were characterised by a widespread or even circumpolar distribution (cluster C of O. (O.) cavimanus, cluster B of W. obesa). Abyssorchomene chevreuxi represents a single species throughout sampling sites in the SO, the South and North Atlantic, indicating this species as a candidate for a truly cosmopolitan species. The bathyal species-level clades within Eurythenes gryllus (Eg1, Eg8) are characterised by distributions encompassing several ocean basins, as well as the abyssal lineage Eg3, comprising specimens of the Atlantic, Pacific and Southern oceans. The bathyal clade Eg1 represents the first case of a bipolar distribution in a macro-benthic deep-sea organism confirmed by genetic data (Havermans et al. 2013). Finally, the distribution of both the species-level clade C of O. (O.) cavimanus and P. debroyeri encompasses sub-Antarctic and Antarctic regions across the Antarctic Polar Front, suggesting that this physical boundary has not been an absolute barrier through time (Havermans 2012).

3.2. Eurybathy versus vertical segregation

Several SO orchomenid species were confirmed to be truly eurybathic, of which a number cover both bathyal (<3000 m, according to Smith et al. 2008) and abyssal (>3000 m) zones (Abyssorchomene sp. 1, A. scotianensis, species-level lineage C of Orchomenella (Orchomenopsis) cavimanus). Although encompassing a wide bathymetric range, Abyssorchomene chevreuxi was confirmed to be restricted to exclusively abyssal depths (3070 – 4800 m).

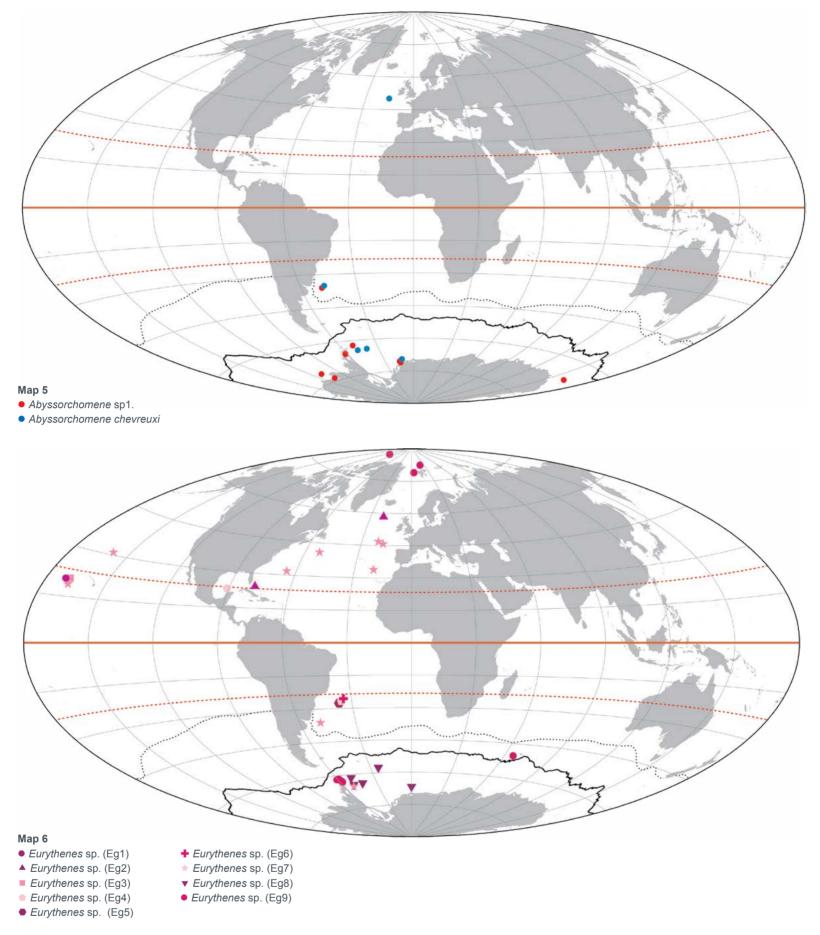
Some species were shown to cover a much wider depth range than initially thought (e.g. *Orchomenella* (*O.*) *franklini*, up to more than 1100 m while literature records were limited to sites of 250 m depth).

Conversely, in the deep-sea species Eurythenes gryllus, which was originally thought to cover bathyal, abyssal and even hadal zones, a genetic break has been observed around 3000 m (Havermans et al. 2013). Although no species-level lineages covered both bathyal and abyssal zones, most of them covered vast bathymetric gradients of 1000 to 2000 m (e.g. Eg3: 3070 - 5770 m). Hence, this genetic break within *E. gryllus* around 3000 m, ubiquitously present in each sampled ocean region, might represent evidence for an ecotone between the bathval and abvssal zone (i.e. a narrow transition of a distinct species composition between two habitats; Jamieson et al. 2011). A similar barrier for gene flow around this depth was observed for a deep-sea bivalve species (Zardus et al. 2006). Vertical partitioning of closely related species has also been observed in other deep-sea benthic (Blankenship et al. 2006, Duffy et al. 2012) and pelagic crustaceans (Laakmann et al. 2012). In deep-sea lysianassoids, several examples of differing depth ranges for congeneric species exist, e.g. for Paralicella (Duffy et al. 2012). This is thought to result from competition for resources (Blankenship & Levin 2007, Laakmann et al. 2012) or to be linked with the occurrence of favorable habitats at particular depths. Suitable habitats could be determined by the physiological limits of a species in relation to the synergistic effects of high pressure and low temperature (Brown & Thatje 2012). For example, the maximum pressure tolerated at a certain temperature by the bathyal lysianassoid Stephonyx biscayensis (Chevreux, 1908), is consistent with its bathymetric and geographic distribution (Brown & Thatje 2011).

4. Dispersal capacities and historical events

The feeding ecology of a species could have a major influence on its dispersal capacities, which in turn, can explain the genetic patterns observed across a species' distributional range. Feeding ecology strongly differs between SO lysianassoid species, and even within closely related species, varying from commensals of sponges or ascidians (Orchomenyx De Broyer, 1984) to more common feeding types such as opportunistic scavenging and/ or deposit feeding (Orchomenella (O.) franklini, O. (O.) pinguides, Abyssorchomene chevreuxi) to exclusive scavenging (e.g. Pseudorchomene coatsi, P. plebs) or even predation (P. rossi) (Chevreux 1903, Dauby et al. 2001, Nyssen et al. 2002, Baird & Stark 2013). For commensal species, active swimming might be limited when spending partial or entire life cycles inside the host, severely reducing their dispersal potential. Even though olfactive stimulations trigger a guick swimming reaction in both opportunistic and obligate scavengers, opportunistic ones display a passive sit-and-wait strategy with swimming reactions limited to the vicinity of the bottom while exclusive scavengers and predators are known for an active swimming behaviour, often in swarms, in the search of chemical cues in the water column. It has even been suggested that marine scavengers make use of currents for finding food falls, in order to reduce the energetic costs between subsequent meals (Ruxton & Houston 2004). Drift with currents is known to play a role in amphipod dispersal (e.g. Havermans et al. 2007) and the combination of active swimming and passive transport by currents can significantly increase the extent of dispersal between remote populations. This might explain the genetic homogeneity observed within obligate scavenger species (W. obesa, P. plebs, P. rossi, P. coatsi) between sites all around the Antarctic continent. Passive transportation with the thermohaline circulation, i.e. the Antarctic Bottom Water (AABW). originating in the Weddell Sea and covering much of the world ocean floor (Mantyla & Reid 1983) could facilitate the connectivity within and between deep-water populations of species such as Eg1 and Eg3 of Eurythenes gryllus, A. chevreuxi and A. sp. 1. For less active swimmers, passive dispersal by rafting can also be responsible for gene flow among remote populations (Highsmith 1985, Barnes 2002, Leese et al. 2010, Fraser et al. 2011). Rafting on drifting kelp holdfasts with the Antarctic Circumpolar Current (ACC) has likely been responsible for the circum-Antarctic establishment of the sub-Antarctic kelp-dweller Parawaldeckia kidderi (Nikula et al. 2010) (see Fig. 4). To our knowledge, only two studies aiming to test gene flow within lysianassoid species have been conducted, which confirmed that, even within a group of closely related species, actual dispersal capacities can differ. For unidentified Abyssorchomene species, France (1994) has shown evidence for gene flow between distinct deep-sea basins, supporting their high dispersal capacities. In a population genetic study on O. (O.) franklini focused on the East Antarctic shelf, gene flow was shown to be limited to nearby populations situated at distances in the order of hundreds of metres (Baird et al. 2012).

However, dispersal capacities may not be the only influence on the genetic structure observed within a species, historical events should also be taken into account. (Pseudo)cryptic speciation events can be explained by the repeated glacial and interglacial cycles causing an alternation of periods of population contraction in isolated glacial refugia and periods of population expansion when available habitats increased (e.g. Rogers 2007). The use of different refugia (shelf vs. deep sea) during glacial maxima could provide another explanation for the observed genetic structures in lysianassoids. For species restricted to shallow waters, a migration to deeper waters could have been hampered, for example due to physiological constraints. Hence, these species could only have survived in shelf refugia, and the isolation of populations in these shelters might have favoured allopatric speciation events (Thatje *et al.* 2005). Conversely, eurybathic species could have migrated to deeper waters and established on the less affected slope, or in the deep sea. In these envi-



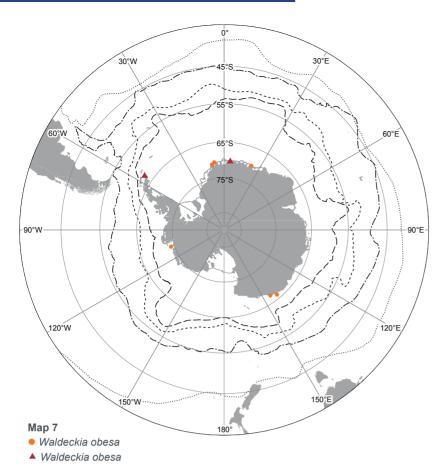
Lysianassoidea Maps 5, 6 Map 5 Distribution of two *Abyssorchomene* species, *A.* sp. 1 and *A. chevreuxi*, based on molecular data. Map 6 Distribution of the nine species-level lineages observed within *Eurythenes gryllus* based on molecular data.

ronments, population size would not have been severely reduced as in shelf refugia, owing to the extent of the available habitat (Allcock & Strugnell 2012). The colonisation of deeper waters may have reduced the probability for allopatric speciation events to occur and might also have favoured these species to occupy the released habitat more quickly after glacial cycles (Thatje *et al.* 2005). This could have been the case for eurybathic species such as O. (O.) cavimanus, A. sp. 1, P. plebs. Recolonisation of populations from shelf refugia will have been comparatively slow, however, widespread dispersal may have been achieved over time, through postglacial colonisation events: this has been hypothesised for O. (O.) franklini (Baird *et al.* 2012) and P. kidderi (Nikula *et al.* 2010). The interaction between dispersal capacities and historical events on the genetic structure of Antarctic lysianassoid species is summarised in Figure 4. Future population genetic studies based on fast-evolving nuclear markers will permit to test these hypotheses.

Acknowledgements

This study was performed in the framework of the Scientific Research Programme on the Antarctic supported by the Belgian Science Policy. The molecular work was funded by an Antarctic Science Bursary. A SYNTHESYS grant (FR-TAF-2102) allowed me to visit the Muséum National d'Histoire Naturelle to study amphipod samples from the Dumont d'Urville Sea. Special thanks go to my host Dr. Nadia Améziane. Samples from the Kerguelen Islands collected by the "programme d'observation des pêcheries australes" under direction of Dr. Guy Duhamel were kindly provided by Charlotte Chazeau and Marc Eléaume from the MNHN and the "service des pêches des TAAF". Samples from the cruise JC062 were kindly provided by Dr. Tammy Horton from the National Oceanography Centre in Southampton. Thanks to Dr. Katrin Linse and Dr. David Barnes from the British Antarctic Survey for making the samples of the





Lysianassoidea Map 7 Distribution of the two species-level lineages observed within Waldeckia obesa based on molecular data

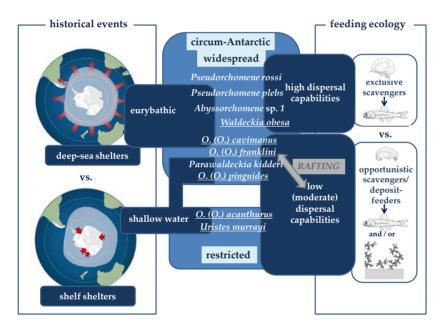


Fig.ure 4 Schematic representation of the hypothetical links between feeding ecology - dispersal capacities and bathymetric distribution - historical events, and their respective influences on lysianassoid species' distributions (widespread or circum-Antarctic versus restricted). Species of which the names are underlined were found to represent of species complexes. Rafting is suggested as a dispersal mechanism for species with relatively lower dispersal capacities. Figures modified with permission from Fraser et al 2012

RRS JC Ross cruise JR 179 available to me. Thanks to Dr. Kareen Schnabel for providing specimens collected during biodiversity surveys of the western Ross Sea undertaken by the National Institute of Water and Atmospheric Research, New Zealand. I thank the Alfred Wegener Institute for Polar and Marine Research, the chief scientists, crew and collectors of the RV Polarstern expeditions ANT XV-3, ANT XIX-5, ARK XIX-3, ANDEEP I-II-III, ANT XXIII-8, ANT XXVII-3, ANT XXVIII-3, as well as the staff of the German Centre of Marine Biodiversity Research (DZMB) for providing the necessary logistics and part of the samples from the DIVA-3 expedition with RV Meteor. Dr Huw Griffiths (BAS, Cambridge) and Dr Anton Van de Putte (RBINS, Brussels) prepared the maps. This is CAML contribution # 154

References

- Allcock, A.L., Strugnell, J.M., 2012. Southern Ocean diversity: new paradigms from molecular ecology Trends in Ecology and Evolution, 27, 520–528.
- Baird, H.P., Miller, K.J., Stark, J.S., 2012. Genetic population structure in the Antarctic benthos: Insights from the widespread amphipod, *Orchomenella franklini*. *PLoS One* **7(3)**, e34363.
 Baird, H.P., Stark, J.S., 2013. Population dynamics of the ubiquitous Antarctic benthic amphipod
- Orchomenella franklini and its vulnerability to environmental change. Polar Biology, 36, 155–167.
 Barnard, J.L., 1961. Gammaridean Amphipoda from depths of 400 to 6000 meters. Galathea Report,
- Barnard, J.L., Karaman, G., 1991. The families and genera of marine Gammaridean Amphipoda (except

- marine Gammaroids). Records of the Australian Museum. 13, 1–866
- Barnes, D.K.A., 2002. Biodiversity: Invasions by marine life on plastic debris. *Nature*, **416**, 808–809. Blankenship, L.E., Levin, L.A. 2007. Extreme food webs: Foraging strategies and diets of scavenging amphipods from the ocean's deepest 5 kilometers. Limnology and Oceanography, 52, 1685-
- Blankenship, L.E., Yayanos, A.A., Cadien, D.B., Levin, L.A. 2006. Vertical zonation patterns of scavenging amphipods from the hadal zone of the Tonga and Kernadec Trenches. Deep-Sea Research Part I: Oceanographic Research Papers, 53, 48–61.
- Brandão, S.N., Sauer, J., Schön, I., 2010. Circumantarctic distribution in Southern Ocean benthos? A genetic test using the genus *Macroscapha* (Crustacea, Ostracoda) as a model. *Molecular* Phylogenetics and Evolution, 55, 1055-1069.
- Brandt, A., Blażewicz-Paszkowycz, M., Bamber, R.N., Mühlenhardt-Siegel, U., Malyutina, M.V., Kaiser, S., De Broyer, C., Havermans, C., 2012. Are there widespread peracarid species in the deep sea (Crustacea: Malacostraca)? Polish Polar Research. 33, 139–162.
- A, Thatje, S. 2011. Respiratory response of the deep-sea amphipod Stephonyx biscayensis indicates bathymetric range limitation by temperature and hydrostatic pressure. PLoS ONE, 6, e28562
- Brown, A., Thatje, S. 2012. Interactive comment on "Deep-sea scavenging amphipod assemblages from the submarine canyons of the Iberian Peninsula" by G.A. Duffy et al. *Biogeosciences*
- Discussions, 9, C1–C3.

 Bucklin, A., Wilson, R.R.Jr., Smith, K.L.Jr., 1987. Genetic differentiation of seamount and basin populations of the deep-sea amphipod Eurythenes gryllus. Deep Sea Research Part A: Oceanographic Research Papers, 34, 1795–1810.

 Chevreux, E., 1903. Campagnes scientifiques de S.A. le P. Albert Ier de Monaco. Note preliminaire sur
- les amphipodes de la famille des Lysianassidae recueillis par la « Princesse Alice » dans les eaux profondes de l'Atlantique et de la Méditerranée. Bulletin de la Société Zoologique de France, 28, 81-97
- Corrigan, L.J., Horton, T., Fotherby, H., White, T.A., Hoelzel, A.R. (2013). Adaptive evolution of deep-sea amphipods from the superfamily Lysianassoidea in the North Atlantic. Evolutionary Biology, doi:10.1007/s11692-013-9255-2
- Dauby, P., Scallteur, Y., De Broyer, C., 2001. Trophic diversity within the eastern Weddell Sea amphipod community. *Hydrobiologia*, 443, 69–86.
- De Broyer, C., Nyssen, F., Dauby, P., 2004. The crustacean scavenger guild in Antarctic shelf, bathyal and abyssal communities. *Deep-Sea Research Part II: Topical studies in Oceanography*, **51**, 1733-1752.
- De Broyer, C., Lowry, J.K., Jażdżewski, K., Robert, H. 2007. Catalogue of the Gammaridean and Corophiidean Amphipoda (Crustacea) of the Southern Ocean with distribution and ecological data. In: De Broyer, C. (ed.), Census of Antarctic Marine Life: Synopsis of the Amphipoda of the Southern Ocean. *Bulletin de l'Institut Royal des Sciences Naturelles de Belgique, Biologie*, 77(suppl. 1), 1-325.
- Duffy, G.A., Horton, T., Billett, D.S.M., 2012. Deep-sea scavenging amphipod assemblages from the submarine canyons of the Iberian Peninsula. *Biogeosciences Discussions*, 9, 7831–7851.
 Escobar-Briones, E., Nájera-Hillman, E., Alvarez, F., 2010. Unique 16S rDNA sequences of *Eurythenes*
- gryllus (Crustacea: Amphipoda: Lysianassidae) from the Gulf of Mexico abyssal plain. Revista Mexicana de Biodiversidad, 81, 177–185.
- Feldkamp, T., 2010. Genetische Struktur und Diversität ausgewählter antarktischer Amphipoden.
- Diplomarbeit, Ruhr-Universität Bochum, 115 pp.
 France, S.C., 1994. Genetic population structure and gene flow among deep-sea amphipods, Abyssorchomene spp., from six California continental Borderland basins. Marine Biology, 118,
- France, S.C., Kocher, T.D., 1996a. DNA sequencing of formalin-fixed crustaceans from archival research
- collections. Molecular Marine Biology and Biotechnology, 5, 304–313.
 France, S.C., Kocher, T.D., 1996b. Geographic and bathymetric patterns of mitochondrial 16S rDNA sequence divergence among deep-sea amphipods, Eurythenes gryllus. Marine Biology, 126, 633–643
- Fraser, C.I., Nikula, R., Waters, J.M., 2011. Oceanic rafting by a coastal community. Proceedings of the
- Royal Society of London, Series B: Biological Sciences, 278, 649–655.

 Fraser, C.I., Nikula, R., Ruzzante, D.E., Waters, J.M., 2012. Poleward bound: biological impacts of Southern Hemisphere glaciation. Trends in Ecology and Evolution, 27, 462–471.
- Havermans, C., De Broyer, C., Mallefet, J., Zintzen, V., 2007. Dispersal mechanisms in amphipods: a case study of *Jassa herdmani* (Crustacea, Amphipoda) in the North Sea. *Marine Biology*, **153**,
- 83-89. rmans C., Nagy Z.T., Sonet G., De Broyer C., Martin P., 2010. Incongruence between molecular phylogeny and morphological classification in amphipod crustaceans: A case study of Antarctic
- lysianassoids. *Molecular Phylogenetics and Evolution*, **55**, 202–209.

 Havermans, C., Nagy, Z.T., Sonet, G., De Broyer, C., Martin, P., 2011. DNA barcoding reveals new insights into the diversity of Antarctic species of Orchomene sensu lato (Crustacea: Amphipoda:
- Lysianassoidea). Deep-Sea Research Part II: Topical studies in Oceanography, **58**, 230–241.

 Havermans, C., 2012. DNA barcoding, phylogeography and phylogeny of the Lysianassoidea (Crustacea: Amphipoda) from the Southern Ocean and the World's deep seas. PhD thesis,
- Université Catholique de Louvain, 423 pp.

 Havermans, C., Sonet, G., d'Udekem d'Acoz, C., Nagy, Z.T., Martin, P., Brix, S., Riehl, T., Agrawal, S. Held, C., 2013. Genetic and morphological divergences in the cosmopolitan deep-sea amphipod Eurythenes gryllus reveal a diverse abyss and a bipolar species. PLoS ONE, 8(9), e74218
- Held, C., 2003. Molecular evidence for cryptic speciation within the widespread Antarctic crustacean Ceratoserolis trilobitoides (Crustacea, Isopoda). In: Huiskes, A.H.L., Gieskes, W.W.C., Rozema, J., Schorno, R.M.L., van der Vies, S.M., Wolff, W.J. (eds.) Proceedings of the SCAR Biology Symposium Amsterdam, Antarctic Biology in a global context. Backhuys Publishers, Leiden, The
- Netherlands, 87–95.
 Held, C., Wägele, J.W., 2005. Cryptic speciation in the giant Antarctic isopod *Glyptonotus antarcticus*
- (Isopoda: Valvifera: Chaetiliidae). *Scientia Marina* 69: 175-181. Highsmith, R.C., 1985. Floating and algal rafting as potential dis invertebrates. *Marine Ecology Progress Series*, **25**, 169–179. dispersal mechanisms in brooding
- Horton, T., De Broyer, C., 2013. Lysianassoidea. In: Horton, T., Lowry, J. & De Broyer, C. (eds.) World Amphipoda database at http://www.marinespecies.org/amphipoda/aphia. php?p=taxdetails&id=176788 on 2013-12-18
- n T., Thurston M., Duffy, G. (2013) Community Composition of Scavenging Amphipods at Bathyal Depths on the Mid-Atlantic Ridge. *Deep-Sea Research Part II: Topical studies in Oceanography*,
- 98, 352–359. http://dx.doi.org/10.1016/j.dsr2.2013.01.032 son, A.J., Kilgallen, N.M., Rowden, A.A., Fujii, T., Horton, T., Lörz, A.-N., Kitazawa, K., Priede, I.G., 2011. Bait-attending fauna of the Kermadec Trench, SW Pacific Ocean: Evidence for an ecotone across the abyssal-hadal transition zone. Deep-Sea Research Part I: Oceanographic Research Papers, **58**, 49–62.

- Kimura, M., 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16, 111–120.
 Klages, M., Vopel, K., Bluhm, H., Brey, T., Soltwedel, T., Arntz, W.E., 2001. Deep-sea food falls: first observation of a natural event in the Arctic Ocean. *Polar Biology*, 24, 292–295.
 Laakmann, S., Auel, H., Kochzius, M., 2012. Evolution in the deep sea: Biological traits, ecology and phylogenetics of pelagic copepods. *Molecular Phylogenetics and Evolution*, 65, 535–546.
- Laver, M.B., Olsson, M.S., Edelman, J.L., Smith, K.L., 1985. Swimming rates of scavenging deep-amphipods recorded with a free vehicle video camera. *Deep-Sea Research*, 32, 1135–1142.
- Leese, F., Agrawal, S., Held, C., 2010. Long-distance island hopping without dispersal stages: transportation across major zoogeographic barriers in a Southern Ocean isopod. Naturwissenschaften, 97, 583–594.
- Lowry, J.K., Stoddart, H.E. (1983). The shallow-water gammaridean Amphipoda of the subantarctic islands of New Zealand and Australia: Lysianassoidea. Journal of the Royal Society of New Zealand 13: 279-394.
- Mantyla, A.W., Reid, J.L., 1983. Abyssal characteristics of the world ocean waters. Deep-Sea Research Part A, 30, 805–833.
- Nikula. R., Fraser, C.I., Spencer, H.G., Waters, J.M., 2010. Circumpolar dispersal by rafting in two subantarctic kelp-dwelling crustaceans. Marine Ecology Progress Series, 405, 221–230

- Nyssen, F., Brey, T., Lepoint, G., Bouquegneau, J.M., De Broyer, C., Dauby, P., 2002. A stable isotope approach to the eastern Weddell Sea trophic web: focus on benthic amphipods. *Polar Biology*, **25**, 280–287.
- Premke, K., Muyakshin, S., Klages, M., Wegner, J., 2003. Evidence for long-range chemoreceptive Regers, A., Muyakshiri, S., Klages, M., Wegher, J., 2005. Evidence for long-range chemioteceptive tracking of food odour in deep-sea scavengers by scanning sonar data. *Journal of Experimental Marine Biology and Ecology*, 285-286, 283–294.
 Rogers, A.D., 2007. Evolution and biodiversity of Antarctic organisms: a molecular perspective. *Philosophical Transactions of the Royal Society of London, Series B*, 362, 2191–2214.
 Ruxton, G.D., Houston, D.C., 2004. Energetic feasability of an obligate marine scavenger. *Marine Ecology Progress Series*, 266, 59–63.

- Schüller, M., 2011. Evidence for a role of bathymetry and emergence in speciation in the genus *Glycera* (Glyceridae, Polychaeta) from the deep Eastern Weddell Sea. *Polar Biology*, **34**, 549–564.
- Seefeldt, M.A., 2012. Morphological analyses to clarify the systematics of *Uristes murrayi* (Walker, 1903) species complex in the Southern Ocean (Crustacea: Amphipoda: Lysianassoidea). Masterarbeit, Ruhr-Universität Bochum, 123 pp.
- Slattery, P.N., Oliver, J.S., 1986. Scavenging and other feeding habits of Lysianassid amphipods (*Orchomene* spp.) from McMurdo Sound, Antarctica. *Polar Biology*, **6**, 171–177.

 Smith, C.R., De Leo, F.C., Bernardino, A.F., Sweetman, A.K., Arbizu, P.M. (2008). Abyssal food limitation,
- ecosystem structure and climate change. *Trends in Ecology and Evolution*, **23**, 518–528. Soltwedel, T., Juterzenka, K.V., Premke, K., Klages, M., 2003. What a lucky shot! Photographic evidence

- for a medium-sized natural food-fall at the deep seafloor. *Oceanological Acta*, **26**, 623–628. Stockton, W.L., 1982. Scavenging amphipods from under the Ross Ice Shelf, Antarctica. *Deep-Sea*
- Stockton, W.L., 1982. Scavenging amphipods from under the Ross fee Shell, Amarchea. Deep-sea Research, 29, 819–835.

 Stoddart, H., Lowry, J., 2004. The deep-sea lysianassoid genus Eurythenes (Crustacea, Amphipoda, Eurytheneidae n. fam.) Zoosystema, 26, 425–468.

 Takeuchi, I., Watanabe, K., 1998. Respiration rate and swimming speed of the necrophagous amphipod Eurythenes gryllus from Antarctic deep waters. Marine Ecology Progress Series, 163, 285–288.

- Thatje, S., Hillenbrand, C-D, Larter, R., 2005. On the origin of Antarctic marine benthic community structure. *Trends in Ecology and Evolution*, **20**, 534–540.
- Thurston, M.H., 1979. Scavenging abyssal amphipods from the North-East Atlantic Ocean. Marine Biology, **51**, 55–68.

 Vader, W., Romppainen, K., 1986. Notes on Norwegian marine Amphipoda. 10. Scavengers and fish
- associates. Fauna Norvegica, Series A, Volume 6, pp. 3–8.
 Wilson, N.G., Hunter, R.L., Lockhart, S.J., Halanych, K.M., 2007. Multiple lineages and absence of panmixia in the "circumpolar" crinoid *Promachocrinus kerguelensis* from the Atlantic sector of
- Antarctica. *Marine Biology*, **152**, 895–904.

 Zardus, J.D., Etter, R.J., Chase, M. R., Rex, M.A., Boyle, E.E., 2006. Bathymetric and geographic population structure in 1000 and 1100 period of the population structure in 1000 and 1100 period of the population structure. Molecular Ecology, **15**, 639–651.



THE BIOGEOGRAPHIC ATLAS OF THE SOUTHERN OCEAN

Biogeographic information is of fundamental importance for discovering marine biodiversity hotspots, detecting and understanding impacts of environmental changes, predicting future distributions, monitoring biodiversity, or supporting conservation and sustainable management strategies

The recent extensive exploration and assessment of biodiversity by the Census of Antarctic Marine Life (CAML), and the intense compilation and validation efforts of Southern Ocean biogeographic data by the SCAR Marine Biodiversity Information Network (SCAR-MarBIN / OBIS) provided a unique opportunity to assess and synthesise the current knowledge on Southern

The scope of the Biogeographic Atlas of the Southern Ocean is to present a concise synopsis of the present state of knowledge of the distributional patterns of the major benthic and pelagic taxa and of the key communities, in the light of biotic and abiotic factors operating within an evolutionary framework. Each chapter has been written by the most pertinent experts in their field, relying on vastly improved occurrence datasets from recent decades, as well as on new insights provided by molecular and phylogeographic approaches, and new methods of analysis, visualisation, modelling and prediction of biogeographic distributions.

A dynamic online version of the Biogeographic Atlas will be hosted on www.biodiversity.aq.

The Census of Antarctic Marine Life (CAML)

CAML (www.caml.aq) was a 5-year project that aimed at assessing the nature, distribution and abundance of all living organisms of the Southern Ocean. In this time of environmental change, CAML provided a comprehensive baseline information on the Antarctic marine biodiversity as a sound benchmark against which future change can reliably be assessed. CAML was initiated in 2005 as the regional Antarctic project of the worldwide programme Census of Marine Life (2000-2010) and was the most important biology project of the International Polar Year 2007-2009.

The SCAR Marine Biodiversity Information Network (SCAR-MarBIN)
In close connection with CAML, SCAR-MarBIN (www.scarmarbin.be, integrated into www.biodiversity.aq) compiled and managed the historic, current and new information (i.a. generated by CAML) on Antarctic marine biodiversity by establishing and supporting a distributed system of interoperable databases, forming the Antarctic regional node of the Ocean Biogeographic Information System (OBIS, www.iobis.org), under the aegis of SCAR (Scientific Committee on Antarctic Research, www.scar.org). SCAR-MarBIN established a comprehensive register of Antarctic marine species and, with biodiversity.aq provided free access to more than 2.9 million Antarctic georeferenced biodiversity data, which allowed more than 60 million downloads.

The Editorial Team



Claude DE BROYER is a marine biologist at the Royal Belgian Institute of Natural Sciences in Brussels. His research interests cover structural and ecofunctional biodiversity and biogeography of crustaceans, and polar and deep sea benthic ecology. Active promoter of CAML and ANDEEP, he is the initiator of the SCAR Marine Biodiversity Information Network (SCAR-MarBIN). He took part to 19 polar



Huw GRIFFITHS is a marine Biogeographer at the British Antarctic Survey. He created and manages SOMBASE, the Southern Ocean Mollusc Database. His interests include large-scale biogeographic and ecological patterns in space and time. His focus has been on molluscs, bryozoans, sponges and pycnogonids as model groups to investigate trends at high southern latitudes.



Cédric d'UDEKEM d'ACOZ is a research scientist at the Royal Belgian Institute of Natural Sciences, Brussels. His main research interests are systematics of amphipod crustaceans, especially of polar species and taxonomy of decapod crustaceans. He took part to 2 scientific expeditions to Antarctica on board of the *Polarstern* and to several sampling campaigns in Norway and Svalbard.



Bruno DANIS is an Associate Professor at the Université Libre de Bruxelles, where his research focuses on polar biodiversity. Former coordinator of the scarmarbin. be and antabif.be projects, he is a leading member of several international committees, such as OBIS or the SCAR Expert Group on Antarctic Biodiversity Informatics. He has published papers in various fields, including ecotoxicology, physiology, biodiversity informatics, polar biodiversity or information science.



Susie GRANT is a marine biogeographer at the British Antarctic Survey. Her work is focused on the design and implementation of marine protected areas, particularly through the use of biogeographic information in systematic conservation planning.



Christoph HELD is a Senior Research Scientist at the Alfred Wegener Institute Helmholtz Centre for Polar and Marine Research, Bremerhaven. He is a specialis in molecular systematics and phylogeography of Antarctic crustaceans, especially



Falk HUETTMANN is a 'digital naturalist' he works on three poles (Arctic, Anta and Hindu-Kush Himalaya) and elsewhere (marine, terrestrial and atmosphe He is based with the university of Alaska-Fairbank (UAF) and focuses prim on effective conservation questions engaging predictions and open access date.





Philippe KOUBBI is professor at the University Pierre et Marie Curie (Paris, France) and a specialist in Antarctic fish ecology and biogeography. He is the Principal Investigator of projects supported by IPEV, the French Polar Institute. As a French representative to the CCAMLR Scientific Committee, his main input is on the proposal of Marine Protected Areas. His other field of research is on the ecoregionalisation of the high seas.



Ben RAYMOND is a computational ecologist and exploratory data analyst, working across a variety of Southern Ocean, Antarctic, and wider research projects. His areas of interest include ecosystem modelling, regionalisation and marine protected area selection, risk assessment, animal tracking, seabird ecology, complex systems, and remote sensed data analyses.



Anton VAN DE PUTTE works at the Royal Belgian Institute for Natural Sciences (Brussels, Belgium). He is an expert in the ecology and evolution of Antarctic fish and is currently the Science Officer for the Antarctic Biodiveristy Portal www. biodiversity.aq. This portal provides free and open access to Antarctic Marine and terrestrial biodiversity of the Antarctic and the Southern Ocean.



Bruno DAVID is CNRS director of research at the laboratory BIOGÉOSCIENCES, University of Burgundy. His works focus on evolution of living forms, with and more specifically on sea urchins. He authored a book and edited an extensive database on Antarctic echinoids. He is currently President of the scientific council of the Muséum National d'Histoire Naturelle (Paris), and Deputy Director at the CNRS Institute for Ecology and Environment.



Julian GUTT is a marine ecologist at the Alfred Wegener Institute Helmholtz Centre for Polar and Marine Research, Bremerhaven, and professor at the Oldenburg University, Germany. He participated in 13 scientific expeditions to the Antarctic and was twice chief scientist on board Polarstern. He is member of the SCAR committees ACCE and AnT-ERA (as chief officer). Main focii of his work are: biodiversity, ecosystem functioning and services, response of marine systems to climate change, non-invasive technologies, and outreach.



Alexandra POST is a marine geoscientist, with expertise in benthic habitat mapping, sedimentology and geomorphic characterisation of the seafloor. She has worked at Geoscience Australia since 2002, with a primary focus on understanding seafloor processes and habitats on the East Antarctic margin. Most recently she has led work to understand the biophysical environment beneath the Amery Ice Shelf, and to characterise the habitats on the George V Shelf and slope following the successful CAML voyages in that region.

























