

OF THE SOUTHERN OCEAN

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THE BIOGEOGRAPHIC ATLAS OF THE SOUTHERN OCEAN

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10.5. Phylogeography and population genetics

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1. Introduction

Simply put, phylogeography is a sub-discipline and extension of biogeography that looks at the distribution of taxa and their various degrees of relatedness simultaneously. One of the hallmarks of phylogeography as a discipline that differentiates it from biogeography is that the latter concentrates on patterns of distribution exclusively whereas the former also takes into account the processes that were involved in the creation of these patterns (Avise 2000). While many studies on biogeography will also discuss the implication of their findings for our understanding of the processes involved (Clarke 2008), phylogeography does this more explicitly so by making the evolutionary history of the taxon under study (e.g. a phylogenetic tree or network) part of the data input already. Not all authors have adopted the term 'phylogeography' for this approach but use others (e.g. historical biogeography) instead (Queiroz 2005, González-Wevar et al. 2010, McGaughran et al. 2011, Sanmartin 2012, Sanmartin et al. 2008, Arbogast & Kenagy 2008).

Decades of internationally coordinated sampling have begun to fill in important geographical as well as taxonomic gaps in our knowledge about the identity and distribution of the fauna in the Southern Ocean. The advent and widespread use of molecular tools on the other hand have allowed us to tap into unprecedented resolution of heritable traits within as well as among species.

In concert, new tools and an increased availability of samples have led to a vastly increased knowledge about the species inventory and their distribution since the synopsis of Hedgpeth (1969).

One of the most popular applications of phylogeography that results from widespread generation of molecular barcodes are phylogeography networks that integrate intraspecific molecular variants, their relative abundances and the locations where they have been found into a single diagram. Since networks and geographical patterns inside of species are covered in detail in the two phylogeographic case studies that follow, the main focus of this chapter on phylogeography lies on how exactly the incorporation of spatial distributions into the context of molecular phylogenies has advanced our understanding of the processes that shaped the evolutionary history of Southern Ocean organisms.

2. Cryptic species and their implications

One of the most visible outcomes of the use of molecular tools in the Southern Ocean is the frequent discovery of genetically distinct species that have previously been classified as a single species on the grounds of their similar morphologies (Held 2003, Held & Wägele 2005, Janosik & Halanych 2010). What makes cryptic species different from the discovery of genuinely new species is the fact that they have been caught and handled before, some very often indeed, only the close resemblance to some seemingly well-known species has prevented recognising a species-level differentiation. Even before molecular barcoding became institutionalised, (pseudo)cryptic species were reported from the Southern Ocean using morphological evidence, but the increasingly widespread use of molecular methods has greatly increased the rate of discovery of new species and cryptic species.

Initially the splitting of well-known benthic species on the basis of the unfamiliar molecular data was greeted with skepticism and the first findings considered as exceptions, but since then numerous cryptic species complexes were discovered, many of them rather unexpectedly and as a result of ongoing barcoding efforts (Grant et al. 2010, Dettai et al. 2011, Linse et al. 2013). It is currently somewhat unclear if there are more such cryptic species discovered in the Southern Ocean than elsewhere. A large-scale global survey correcting for regional differences in sampling effort etc. suggests that there is currently no evidence for higher numbers of discoveries of cryptic species inside the SO than outside (Pfenninger & Schwenk 2007). Considering the logistic impediments to working in the Southern Ocean as well as the fact that this survey included data until the year 2006 thus limiting cryptic species reports to merely 15 (one order of magnitude smaller than for other zoogeographic regions), this result has to be considered preliminary for the SO.

But it is clear that by now the discoveries of cryptic species inside of known nominal species have graduated from an accidental finding that could be attributed to occasional oversights in taxonomy to one of the major sources of discovery of new species in the Southern Ocean and elsewhere (Bickford *et al.* 2007). Even though the most direct impact of the discovery that one species may in reality be two or more species concerns the taxonomy, in practice the refined taxonomic insight has many implications that extend far beyond taxonomy itself.

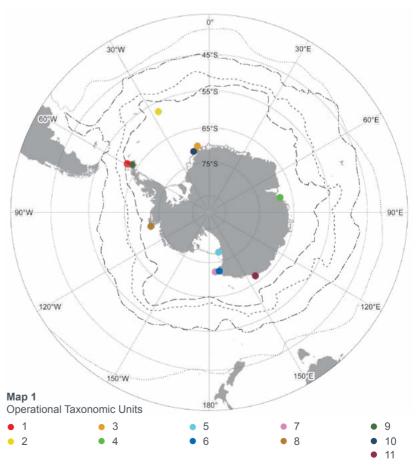
The pattern of new species discovery on the Antarctic shelf where cryptic species have been found in every higher taxon that has been investigated is rather different from the continental slope and the Southern Ocean deep sea where cryptic species discoveries are less common (Raupach & Wägele 2006, Raupach et al. 2007, Brökeland & Raupach 2008) and most new species tend to be first discoveries (Brandt et al. 2007) but this difference is most

likely due to the fact that for depths exceeding 1000 m very little knowledge existed to begin with (Ellingsen et al. 2007; Brandt et al. 2012).

On the Antarctic shelf, cryptic species do not appear to be randomly distributed across the tree of life or in space. A large number of the cryptic species that have been identified so far in the Southern Ocean divide one large distribution area of the more inclusive super taxon into several smaller ones, which are often allopatric (Held 2003, Held & Wägele 2005, Wilson et al. 2007, Linse et al. 2007, Leese et al. 2008a, Leese et al. 2008b, Krabbe et al. 2009, Mahon et al. 2010, Hunt et al. 2010, Smith et al. 2011). There is, however, no general pattern that is common to all newly delimited species and their distributions. Not all widespread species disintegrate into more regionally distributed species when put under molecular scrutiny, some are genuinely widespread on continental or global scales (Schiaparelli et al. 2008, Arango et al. 2010, Hemery et al. 2012, Allcock et al. 2011, Strugnell et al. 2012, Brandt et al. 2012).



Photo 1 Glyptonotus sp. MOTU 3, Eastern Weddell Sea. Image © C. Held.



Phylogeography Map 1 Cryptic diversity in *Glyptonotus* Eights 1852. Sampling localities for molecular operational taxonomic units (MOTUs) in *Glyptonotus* Eights, 1852; candidate species based on mitochondrial COI barcode data (Held 2005; Held & Agrawal unpubl.). MOTU 1 (red dot; South Shetland Islands) is the type locality and most likely represents *Glyptonotus antarcticus* Eights, 1852 that was long considered a prime example of circumpolar distribution.

One of the biggest impacts of cryptic species is that their discovery makes literature data untrustworthy because species reported in earlier publications will refer to the previous, more inclusive taxon (wrongly) carrying a taxonomic name, which after the taxonomic revision rightfully belongs to only one of the newly delimited species contained within (e.g. Janosik *et al.* 2010). According to a literature and database survey carried out under the auspices of the Census of Marine Life (www.coml.org) and utilising the data collected until and including the International Polar Year 2006/2007 (IPY), a total of 235 species were known to occur in both polar oceans (http://www.coml.org/comlfiles/press/CoML_lce_Oceans_Public_Release_02.15.2009.pdf). A small subset of these candidate bipolar species have been analysed in detail and evidence for reproductively isolated but cryptic species endemic to one polar ocean has been found in some (Hunt *et al.* 2010, Uriz *et al.* 2011) whereas evidence for a single species with bipolar distribution was found in others (Kuklinski & Barnes 2010, Pawlowski *et al.* 2007, Havermans *et al.* 2013). See Chapter 10.4.

3. Gene flow and the brooder vs. broadcaster debate

Even without divergent natural selection at work, genetic drift acting independently on isolated populations will over time result in growing dissimilarity of the two populations. Gene flow connecting the two populations, on the other hand, is the strongest antagonist of genetic drift in this scenario because each migrant acts as a vector transporting alleles from one population to the other, thus potentially reducing their uniqueness. For this reason, species with dispersal propagules in the form of pelagic larvae spending a long time in the water column are often expected to be genetically more homogeneous compared to species without such dedicated dispersal stages (brooders versus broadcasters debate).

During the early phase of Antarctic research it was commonly assumed that many if not most species are circumpolar and that gaps in their distribution were mostly due to incomplete sampling. Until the end of the last century it was commonly assumed that '...the distribution of most of the Antarctic fauna is circumpolar' (Arntz *et al.* 1994, Hedgpeth 1969, Clarke & Johnston 1996, Knox 1994, Dayton 1990).

This assumption was based on the many homogenising abiotic factors, most notably in the form of circumpolar currents (ACC and ACoC). Much of the detailed information about patchiness of geomorphology that makes seafloor habitats appear less homogenous today and are now known to have a structuring effect on benthic communities (Post *et al.* 2011) was still unknown at the time.

Species possessing larval dispersal stages (e.g. Matschiner et al. 2009, González-Wevar et al. 2010) demonstrate genetic connectivity on regional scales, which conforms to the expectations for broadcasting species. Some brooding species without planktonic larvae are also known to have large distributions and presumably strong genetic exchange, especially species that are good swimmers as adults (Baird et al. 2012, Havermans et al. 2013) although it should not be ignored that usually all species in these studies are roughly equally mobile but only some are widely distributed whereas others can contain cryptic species, sometimes on rather small geographic scales. In some species, the predictions based on larval or adult dispersal potential do not match the observed data and require additional processes to explain the observed connectivity patterns. Nominal species spanning the Drake Passage (DP) between South America and the Antarctic Peninsula were found to comprise cryptic species, each restricted to either side of the DP, irrespective of whether they were brooders (Hunter & Halanych 2008) or broadcasters (Thornhill et al. 2008) indicating that dispersal is not the only parameter that determines genetic exchange across a major oceanic barrier. Janko et al. (2007) investigated benthic and pelagic Antarctic notothenioid fish and found a stronger signature of recent population growth in the supposedly more mobile pelagic species than among the benthic feeders that were assumed to be more affected by habitat loss during the glacial maxima. Further examples include significant local genetic differentiation in a species with a planktonic dispersal stage (Hoffman et al. 2010, Hunter & Halanych 2010; Hoffman et al. 2012) or species integrity and gene flow over thousands of kilometers in benthic isopods and algae without any dispersal propagules or the ability to swim or float (Leese et al. 2010, Fraser et al. 2013). In both cases passive rafting on buoyant substrate in major ocean currents has been inferred as the most likely means of dispersal. Testing multiple hypotheses in a statistical framework and using fast-evolving microsatellite markers, Leese et al. (2010) showed that unidirectional gene flow in direction of the ACC was the model with the highest explanatory power, thus demonstrating that dispersal among sub-Antarctic islands in this immobile brooder happens in the upper water layers because the unidirectional ACC is predominantly a surface phenomenon. The means of dispersal in the direct-developing seaslug Doris kerguelensis is less obvious but it, too, shows genetic uniformity across thousands of kilometers despite lacking obvious dispersal capabilities (Wilson et al. 2009).

Developing fast-evolving molecular markers may be more difficult than using universal primers (Held & Leese 2007) but in turn it makes reconstructions of events possible that are too rare to make direct observation feasible (Leese & Held 2011) and for which purely biotic predictors can be very poor (see above).

The consequence of the widespread discovery of cryptic species in the Southern Ocean on the one hand and empirical studies of genetic connectivity on the other hand is that there is no distribution pattern that is a defining characteristic of all Antarctic benthos but rather a multitude of different distributions, ranging from extremely local differentiation to global genetic connectivity. Distribution patterns need to be demonstrated rather than assumed

keeping in mind that predictions based on dispersal potential (larval ecology) can be accurate for some taxa but can be amazingly misleading for others.

4. Eurybathy

A widely held opinion about the distribution of fauna on the Antarctic shelf that may deserve another look in the light of current phylogeographic and molecular methods is that eurybathy is more widespread in the Southern Ocean than elsewhere. In a classic study, Brey et al. (1996) demonstrated that species were indeed reported from a wider depth range in the Southern Ocean than comparative taxa in European waters. The assumption of a eurybathic benthic fauna of the Southern Ocean has never seriously been questioned since and is widely considered a defining characteristic of the Antarctic marine ecosystem as a whole. Most authors interpret the apparent eurybathy as a historical legacy of the glacial history during which the shallow-water fauna had to retreat partly or fully from the shelves to the continental slope or the adjacent deep sea (Thatje et al. 2005; Clarke & Crame 2010; Allcock & Strugnell 2012). The deeper waters ensured the survival of many species during the glacial maxima according to this interpretation of Antarctic eurybathy. The recolonisation of the shallow-water habitats took place from the refugia on the slope, however, only incompletely so, adding some of the deeper water habitats permanently to the species' vertical distributions even after the retreat of the ice on the shelves.

Another reason for the strong ties between Antarctic shallow and deepwater faunas might be the fact that around 70% of the world's deep-sea water is formed in the Antarctic especially under the great ice-shelves. From here the cold, well-oxygenated and saline water sinks into the deep sea forming the Antarctic Bottom Water (AABW) that covers much of the slope and deep-sea seafloor and reaches far into the Northern Hemisphere (Mantyla & Reid 1983). The thermohaline circulation creates both a vehicle for transportation and ensures strong physical similarities (e.g. salinity, temperature) from the Antarctic shelf to the deep sea that might make it easier for shallow-water fauna to descend and adapt to the conditions in the deep sea from the Antarctic shelf than from lower latitude shelves and vice versa.

However, the frequent discovery of cryptic species questions the basis upon which much of the support for a eurybathic benthic fauna in the Southern Ocean rests because an unknown number of the taxa included in Brev et al. (1996) study may be affected by this, i.e. be a collection of closely related species rather than a single species. Although in the last decade a lot of progress has been made, still only a small fraction of the taxa has been examined in enough detail. What we do know at this point is that, similar to the patterns of horizontal distribution discussed earlier, several species that were thought to be well-known and distributed vertically across a wide range of water depths, have been found to comprise several reproductively fully isolated species. There is some evidence that depth stratification is present at least in some of cryptic species (Held & Wägele 2005, Brandão et al. 2010, Schüller 2011, Havermans et al. 2013). Currently the number of well-studied cases is still low, hence it is too early to draw far-ranging conclusions, but if it turned out that more precisely delimited cryptic species frequently occupy a) different water depths and b) each species only a fraction of the depth range of the more inclusive taxon, then the status of eurybathy as an unquestioned characteristic of the Antarctic benthic fauna might be in danger just as circumpolarity may have already lost its role as the model of horizontal distribution pattern (see also Downey et al. 2012).

5. Submergence/emergence and connection to the deep sea

Despite the uncertainties regarding a wider depth range for single species in the Antarctic benthos (eurybathy; see above) there is strong phylogeographic support for a close connection between the Antarctic shelf and the adjacent deep sea in several higher taxa. Using molecular phylogenetics, Held (2000) found unequivocal evidence that deep-sea serolids were most closely related to shallow-water species on the Antarctic shelf. The occurrence of vestigial eyes in some of these deep-sea isopods in addition to completely blind species had already suggested that the deep sea was colonised by shallow-water ancestors (polar submergence) and not vice versa. However, the molecular phylogeographic approach established two new findings: the center of origin was the Antarctic shelf rather than shallow-waters of other continents and the deep sea was colonised by serolids several times independently since the deep-sea species are not each others' closest relatives, suggesting processes that promote or facilitate the transition from the Antarctic shelf into the deep sea. Strugnell et al. (2008) found a similar pattern of polar submergence for Antarctic octopus but in this lineage the deep-sea species formed a monophylum, supporting that submergence into the deep happened once in the ancestral lineage followed by subsequent radiation in deep waters rather. Strugnell et al. (2008) correlated the date for the ancestral submergence and the subsequent radiation with the opening of the Drake Passage and the large-scale production of AABW, respectively.

Molecular phylogeographic support for the opposite case, i.e. polar emergence, has also been found in several taxa independently (Strugnell *et al.* 2011, Riehl & Kaiser 2012). More recently, molecular data are appearing that suggest that refugia allowing survival at shallower depths might have been present even during glacial maxima (Rogers 2007; Allcock & Strugnell 2012). If so, continuous survival would not have been limited to taxa that were able to evade the ice by either taking refuge in deeper waters or lower latitudes. Even survival on the shelf at high latitudes might have been possible in geothermal refugia (Fraser *et al.* 2014).

6. Radiations and molecular clocks

The high rate of endemism in the SO (Griffiths et al. 2009) already indicates that the fauna of the Antarctic is fairly isolated from that of lower latitudes but phylogenetic and phylogeographic research has provided more detailed insight into how these differences evolved. The nothothenioid fish are not only the dominant group of fish in the Southern Ocean today but they have also evolved as a group in situ (Eastman & McCune 2000; Near & Cheng 2008; Dettai et al. 2012). A similar pattern of a monophyletic group of Antarctic species that are more closely related to each other than any of them is to the more basal ancestors outside the SO has been found in a number of groups (see compilation in Lecointre et al. 2013 and references therein). These radiations have undoubtedly evolved in response to the drastically different environmental conditions in the SO but they can only be recognised fully if a phylogenetic tree is known. If only the biogeographic pattern without information of relatedness was known, any secondary exportations of derived taxa from inside the Antarctic radiation out of the Southern Ocean to lower latitudes (Held 2000: Strugnell et al. 2008; Göbbeler & Klussmann-Kolb 2010) would prevent the recognition of the radiation itself.

A phylogeographic approach has been used repeatedly to calculate the age of taxa as well as the speed of molecular substitution. Bargelloni et al. (1994) analysed a monophyletic group of Antarctic notothenioid fishes characterised by the possession of antifreeze glycopeptides in their blood that is absent in their relatives outside the Southern Ocean. The unexpectedly low level of divergence in the mitochondrial marker genes suggested either a radiation date that was much younger than the calibration date of 23 Mya for the age of the Drake Passage (Kennett 1977) or much slower accumulation rates of molecular substitutions than what was known from outside Antarctica. The 'molecular slow-down hypothesis' received support from physiology (Clarke & Johnston 1996) in view of the fact that the reduced metabolic rate of polar fish might also slow the production of oxygen radicals.

The assumption that interference of oxygen radicals with the fidelity of DNA replication during cell division directly might translate into reduced rates of evolutionary novelties at subzero temperatures where ROS production is low, has not materialised or at least turned out to be not universally true. In a two-cluster test (a variant of the relative rate test comparing groups of taxa rather than single species), Held (2001) found Antarctic serolid isopods to have mitochondrial substitution rates indistinguishable from their South American relatives. Notably, the use of phylogeographic data makes the results independent from assumptions about the absolute rate of molecular clocks and errors therein. It appears that the marked differences in temperature at which the two groups of invertebrates evolved for many millions of years have not resulted in measurably different rates of their molecular clocks, suggesting that a younger evolutionary origin of taxa including that of the radiation of notothenioids may be a more appropriate interpretation. More recent studies looking at the genetic divergence between pairs of species in South America and the Southern Ocean served to reinforce the impression that many Antarctic species have evolved much more recently than previously thought while estimates concerning the opening of the Drake Passage that separates the these species pairs physically today have shifted from around 23 Mya (Kennett 1977) to much older dates (Lawver & Gahagan 2003, Scher & Martin 2006. Barker et al. 2007).

7. Demography and genetic diversity

The size of a population is one of the most important parameters controlling its genetic diversity (Cutter et al. 2013) and clearly some taxa in the Southern Ocean have enormous population sizes, most notably in Antarctic krill (Euphausia superba). Extensive regional sampling and modeling of population genetics parameters demonstrates that the genetic diversity is exceptionally high in this species (Goodall-Copestake et al. 2010). Even when large sample sizes per replicate are considered thus eliminating a common error in diversity estimates in high abundance species, almost no genotypes are shared among 504 specimens (haplotype diversity 0.9981), indicating that its genetic diversity is not nearly adequately represented even in this large sample.

Despite the strong evidence for well-mixed krill populations on a regional scale, it is not inconceivable that on larger spatial scales even seemingly unrestricted pelagic species may depart from panmixia and show some geographic structure. Nevertheless, previous reports of significant population structuring on a continental scale in E. superba (Zane et al. 1998) may have been the result of sampling error and were not confirmed in a later study involving multiple markers and more samples (Bortolotto et al. 2011).

While the absolute population size may be hard to determine from genetic data without detailed knowledge about how many individuals actually contribute genetically to the next generation (effective population size vs census size), analysing the patterns how the genetic diversity is precisely distributed within and amongst the individuals yields information about the recent developmental trends of the population. A common pattern that is present in almost all Antarctic datasets is a signal of marked, recent population growth (e.g. Janko et al. 2007; Bortolotto et al. 2011; González-Wevar et al. 2013). Similar trends are known from Arctic datasets, which is consistent with a recovery of populations after the Last Glacial Maximum due to the vast increase of habitable shelf habitat following the retreat of the ice (Hewitt 2000). The relative increase or decrease of the population size can be modeled, but is expressed in terms of generations. Translating it into years before present requires additional assumptions about generation time to be made. Constraining the timing of the widespread population growth to last glacial maximum

(LGM) is therefore difficult and error-prone, especially if a single mitochondrial marker is used for inference, and usually assumed rather than explicitly verified. Besides the plausibility of a causal connection between ice retreat and population growth there is circumstantial evidence supporting a connection between growth and ice retreat (though not necessarily the last) in that shelf taxa tend to show more prominent signals of recent population growth than taxa inhabiting deeper waters on the slope or the adjacent deep sea, which have been less impacted by the glacial cycles than shallower habitats (Raupach et al. 2010).

8. Molecules and morphology

It is worth noting that although molecular data are not the only possible source for phylogeographically motivated studies, in practice their widespread use has given the field of phylogeography a major boost for various reasons. Although the information density may be lower than that of a phylogenetic tree or a genealogical network, information about relatedness is also contained in the delineation of higher taxonomic units (genera, families etc), which is still the main source of information about endemisms. What often muddles the use of taxonomy as a source of information for phylogeography, however, is the fact that this is only strictly true if the underlying taxonomy builds on the foundation of modern phylogenetic theory and more specifically on the identification of monophyletic groups rather than pragmatically identifiable units of similarlooking taxa. Frequently the modern taxonomic treatment of taxa is incomplete, leaving taxonomic 'collecting bins', i.e. para- or polyphyletic groups that are nevertheless united under a valid taxonomic label but one that does not express relatedness. A literature research that takes taxonomic composition of higher taxa for a statement of relatedness without making sure they consist of monophyla is thus conceptually flawed, but carefully constructed phylogenies based on morphology have been used successfully in phylogeographic studies (Wägele 1994, Brandt 1991; but see Held 2000).

At the intraspecific level, molecular data are almost required to prove that morphological variance is based on heredity and not on phenotypic plasticity within species.

9. Synopsis

The widespread use of molecular phylogenetics, phylogeography and population genetics have allowed putting many hypotheses about the common patterns in the Southern Ocean fauna and the driving forces behind them to the test. The growing consensus after more than a decade of molecular scrutiny is that there are far fewer defining characteristics that have proven true for all or even many Southern Ocean species than previously thought. Instead many different, taxon-specific responses have been identified. Species identities and non-identities can be (and need to be) demonstrated rather than assumed, which is a prerequisite upon which almost all subsequent inference critically depends. The distribution and evolutionary history of species in the SO and their response to the same environmental cues is much less uniform than previously believed and often a taxon-specific mix of more than one driving factor.

On the other hand, the application of molecular tools makes it possible to not only correlate observed patterns to general predictions but to formulate hypotheses in a statistical framework and to identify the one with the highest explanatory power among competing alternative hypotheses (Leese et al. 2010). In the near future, high-throughput next-generation sequencing and more advanced analysis techniques will become more widely used and help reduce the effect of sampling error and at the same time further expand the spectrum of questions that can be tackled (Moroz et al. 2014; Hoffman et al.

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References

Allcock, A. L., Barratt, I., Eléaume, M., Linse, K., Norman, M. D., Smith, P. J., Steinke, D., Stevens, D. W., Strugnell, J. M., 2011. Cryptic speciation and the circumpolarity debate: A case study on e Southern Ocean octopuses using the COI barcode of life. *Deep-Sea Research Part II: Topical Studies in Oceanography* **58**, 242-249.

Allcock, A. L., Strugnell, J. M., 2012. Southern Ocean diversity: new paradigms from molecular ecology.

Trends in Ecology and Evolution, 27, 520-528.

Arango, C. P., Soler Membrives, A., Miller, K. J., 2010. Genetic differentiation in the Circum-Antarctic sea spider Nymphon australe (Pycnogonida; Nymphonidae). Deep Sea Research Part II: Topical Studies in Oceanography 58, 212-219.

Arbogast, B. S., Kenagy, G. J., 2008. Comparative phylogeography as an integrative approach to historical biogeography. *Journal of Biogeography* 28, 819-825.

Arntz, W., Brey, T., Gallardo, V. A., 1994. Antarctic zoobenthos. *Oceanography and Marine Biology* 32,

Avise, J. C., 2000. *Phylogeography: the history and formation of species.* Harvard University Press

Baird, H. P., Miller, K. J., Stark, J. S., 2012. Genetic population structure in the Antarctic benthos: insights from the widespread amphipod, Orchomenella franklini. PLoS One 7, e34363.
 Bargelloni, L., Ritchie, P. A., Patarnello, T., Battaglia, B., Lambert, D. M., Meyer, A., 1994. Molecular

evolution at subzero temperatures; mitochondrial and nuclear phylogenies of fishes from Antarctica (suborder Notothenioidei), and the evolution of antifreeze glycopeptides. *Molecular Biology and Evolution* 11, 854-863.

Barker, P. F., Filippelli, G. M., Florindo, F., Martin, E. E., Scher, H. D., 2007. Onset and role of the Antarctic Circumpolar Current. Deep Sea Research Part II: Topical Studies in Oceanography 54, 2388-2398.
Bickford, D., Lohman, D. J., Sodhi, N. S., Ng, P. K. L., Meier, R., Winker, K., Ingram, K. K., Das, I., 2007. Cryptic species as a window on diversity and conservation. Trends in Ecology & Evolution 22, 148-155

Bortolotto, E., Bucklin, A., Mezzavilla, M., Zane, L., Patarnello, T., 2011. Gone with the currents: lack of genetic differentiation at the circum-continental scale in the Antarctic krill Euphausia supering BMC Genetics 12, 32.

SCAR

- Brandt, A., 1991. Zur Besiedlungsgeschichte des antarktischen Schelfes am Beispiel der Isopoda (Crustacea, Malacostraca) [Colonization of the Antarctic shelf by the Isopoda (Crustacea, Malacostraca)]. Berichte zur Polarforschung (Reports on Polar Research) 98, 1-240.

 Brandt, A., Blażewicz-Paszkowycz, M., Bamber, R. N., Mühlenhardt-Siegel, U., Malyutina, M. V., Kaiser,
- S., De Broyer, C., Havermans, C., 2012. Are there widespread peracarid species in the deep sea (Crustacea: Malacostraca)? *Polish Polar Research* **33**, 139-162.

 Brandt, A., De Broyer, C., De Mesel, I., Ellingsen, K. E., Gooday, A. J., Hilbig, B., Linse, K., Thomson,
- M. R. A., Tyler, P. A., 2007. The biodiversity of the deep Southern Ocean benthos. Philosophical
- Transactions of the Royal Society of London, B: Biological Sciences, 362, 39-66.

 Brandt, A., De Broyer, C., Ebbe, B., Ellingsen, K. E., Gooday, A.J., Janussen, D., Kaiser, S., Linse, K., Schueller, M., Thomson, M. R. A., Tyler, P. A., Vanreusel, A., 2012. Southern Ocean Deep Benthic Biodiversity. In: Rogers A.D., Johnston N.M., Murphy E.G., Clarke A. (eds.). *Antarctic Ecosystems: An extreme environment in a changing world.* Chichester, UK: John Wiley & Sons. pp. 291-334.

 Brey, T., Dahm, C., Gorny, M., Klages, M., Stiller, M., Arntz, W. E., 1996. Do Antarctic benthic invertebrates
- show an extended level of eurybathy? *Antarctic Science* **8**, 3-6.

 Brökeland, W., Raupach, M. J., 2008. A species complex within the isopod genus *Haploniscus* (Crustacea:
- Malacostraca: Peracarida) from the Southern Ocean deep sea: a morphological and molecular
- approach. Zoological Journal of the Linnean Society 152, 655-706.
 Clarke, A., 2008. Antarctic marine benthic diversity: patterns and processes. Journal of Experimental Marine Biology and Ecology 366, 48-55.
- Clarke, A., Crame, J. A., 2010. Evolutionary dynamics at high latitudes: speciation and extinction in polar marine faunas. *Philosophical Transactions of the Royal Society, B Biological Sciences* 365, 3655-
- Clarke, A., Johnston, I. A., 1996. Evolution and adaptive radiation of Antarctic fishes. Trends in Ecology & Evolution 11, 212-218.
- Cutter, A. D., Jovelin, R., Dey, A., 2013. Molecular hyperdiversity and evolution in very large populations
- Molecular Ecology 22, 2074-2095.

 Dayton, P. K., 1990. Polar benthos. In: Smith, W. (ed.). Polar Oceanography: Academic Press. pp. 631-685.
- Dettai, A., Adamowizc, S. J., Allcock, A. L., Arango, C. P., Barnes, D. K., Barratt, I., Chenuil, A., Couloux, A., Cruaud, C., David, B., Denis, F., Denys, G., Díaz, A., Eléaume, M., Féral, J.-P., Froger, A., Gallut, C., Grant, R., Griffiths, H. J., Held, C., Hemery, L. G., Hosie, G., Kuklinski, P., Lecointre, G. Linse, K., Lozouet, P., Mah, C., Monniot, F., Norman, M. D., O'Hara, T., Ozouf-Costaz, C., Piedallu C., Pierrat, B., Poulin, E., Puillandre, N., Riddle, M., Samadi, S., Saucéde, T., Schubart, C. D. Smith, P. J., Stevens, D. W., Steinke, D., Strugnell, J. M., Tarnowska, K., Wadley, V., Améziane Smith, P. J., Stevens, D. W., Steinke, D., Strughell, J. M., Tarnowska, K., Wadiey, V., Afriezlarie, N., 2011. DNA barcoding and molecular systematics of the benthic and demersal organisms of the CEAMARC survey. *Polar Science* 5, 298-312.
 Downey, R. V., Griffiths, H. J., Linse, K., Janussen, D., 2012. Diversity and distribution patterns in high southern latitude sponges. *PLoS One* 7, e41672.
 Eastman, J. T., McCune, A. R., 2000. Fishes on the Antarctic continental shelf: evolution of a marine species flock? *Journal of Fish Biology*, 57, 84-102.
 Ellingsen, K. E., Brandt, A., Ebbe, B., Linse, K., 2007. Diversity and species distribution of polychaetes, isonods and bivalves in the Atlantic sector of the deep Southern Ocean. *Polar Biology*, 30, 1265-

- isopods and bivalves in the Atlantic sector of the deep Southern Ocean. Polar Biology 30, 1265-
- Fraser, C. I., Terauds, A., Smellie, J., Convey, P., Chown, S. L., 2014. Geothermal activity helps life survive
- glacial cycles. *Proceedings of the National Academy of Sciences of the United States of America*, . Fraser, C. I., Zuccarello, G. C., Spencer, H. G., Salvatore, L. C., Garcia, G. R., Waters, J. M., 2013. Genetic affinities between trans-oceanic populations of non-buoyant macroalgae in the high latitudes of the Southern Hemisphere. PLoS One 8, e69138.
- González-Wevar, C. A., Nakano, T., Cañete, J. I., Poulin, E., 2010. Molecular phylogeny and historical biogeography of *Nacella* (Patellogastropoda: Nacellidae) in the Southern Ocean. *Molecular* Phylogenetics and Evolution 56, 115-124
- González-Wevar, C. A. G., Saucède, T., Morley, S.A., Chown, S. L., Poulin, 2013. E. Extinction and recolonization of maritime Antarctica in the limpet *Nacella concinna* (Strebel, 1908) during the last glacial cycle: toward a model of Quaternary biogeography in shallow Antarctic invertebrates Molecular Ecology 22, 5221-5236.

 Goodall-Copestake, W. P., Pérez-Espona, S., Clark, M. S., Murphy, E. J., Seear, P. J., Tarling, G. A., 2010.
- Swarms of diversity at the gene cox1 in Antarctic krill. *Heredity* **104**, 513-518.

 Göbbeler, K., Klussmann-Kolb, A., 2010. Out of Antarctica? New insights into the phylogeny and biogeography of the Pleurobranchomorpha (Mollusca, Gastropoda). *Molecular Phylogenetics and*
- Evolution, 55, 996-1007.
 Grant, R. A., Griffiths, H. J., Steinke, D., Wadley, V., Linse, K., 2010. Antarctic DNA barcoding; a drop in the ocean? Polar Biology 34, 775-780.
- Griffiths, H. J., Barnes, D. K., Linse, K., 2009. Towards a generalized biogeography of the Southern Ocean benthos. *Journal of Biogeography* 36, 162-177.
- Havermans, C., Sonet, G., d'Udekem d'Acoz, C., Nagy, Z. T., Martin, P., Brix, S., Riehl, T., Agrawal, S. Held, C., 2013. Genetic and morphological divergences in the cosmopolitan deep-sea amphipod Eurythenes gryllus reveal a diverse abyss and a bipolar species. PLoS One 8, e74218.
- Hedgpeth, J. W., 1969. Introduction to Antarctic zoogeography. In: Bushnell, V. C., Hedgpeth, J. W. (eds.) Distribution of selected groups of marine invertebrates in waters south of 35°S:. Antarctic Map Folio Series, American Geographical Society. Folio 11, 1-29.

 Held, C., 2000. Phylogeny and biogeography of serolid isopods (Crustacea, Isopoda, Serolidae) and
- the use of ribosomal expansion segments in molecular systematics. Molecular Phylogenetics and
- Evolution **15**, 165-178.

 Held, C., 2001. No evidence for slow-down of molecular substitution rates at subzero temperatures in
- Antarctic serolid isopods (Crustacea, Isopoda, Serolidae). *Polar Biology* **24**, 497-501. C., 2003. Molecular evidence for cryptic speciation within the widespread Antarctic crustacean *Ceratoserolis trilobitoides* (Crustacea, Isopoda). In: Huiskes, A. H. L., Gieskes, W. W. C., Rozema, J., Schorno, R. M. L., van der Vies, S. M., Wolff, W. J. (eds.). Antarctic biology in a global context. Leiden: Backhuys. pp. 135-139.

 Held, C., Leese, F., 2007. The utility of fast evolving molecular markers for studying speciation in the
- Antarctic benthos. Polar Biology 30, 513-521.
- Held, C., Wägele, J. W., 2005. Cryptic speciation in the giant Antarctic isopod *Glyptonotus antarcticus* (Isopoda, Valvifera, Chaetiliidae). *Scientia Marina* 69, 175-181.
 Hemery, L. G., Eléaume, M., Roussel, V., Améziane, N., Gallut, C., Steinke, D., Cruaud, C., Couloux,
- A., Wilson, N. G., 2012. Comprehensive sampling reveals circumpolarity and sympatry in seven mitochondrial lineages of the Southern Ocean crinoid species *Promachocrinus kerguelensis* (Echinodermata). *Molecular Ecology* **21**, 2502-2518.
- Hewitt, G. M., 2000. The genetic legacy of the Quaternary ice ages. *Nature*, **405**, 907-913.

 Hoffman, J. I., Clarke, A., Fretwell, M. S., Peck, P. A., Lloyd, L., 2012. Unexpected fine-scale population structure in a broadcast-spawning Antarctic marine mollusc. *PLoS One* **7**, e32415.
- Hoffman, J. I., Peck, L. S., Linse, K., Clarke, A., 2010. Strong population genetic structure in a broadcast-spawning Antarctic marine invertebrate. *Journal of Heredity* 102, 55-66.Hoffman, J. I., Simpson, F., David, P., Rijks, J. M., Kuiken, T., Thorne, M. A. S., Lacy, R. C., Dasmahapatra,
- K. K., 2014. High-throughput sequencing reveals inbreeding depression in a natural population. *Proceedings of the National Academy of Sciences* 111, 3775-3780.

 Hunt, B., Strugnell, J. M., Bednarsek, N., Linse, K., Nelson, R. J., Pakhomov, E., Seibel, B., Steinke, D.,
- Würzberg, L., 2010. Poles Apart: The "bipolar" pteropod species *Limacina helicina* is genetically distinct between the Arctic and Antarctic oceans. *PLoS One* **5**, e9835.

 Hunter, R. L., Halanych, K. M., 2008. Evaluating connectivity in the brooding brittle star *Astrotoma agassizii*
- across the Drake Passage in the Southern Ocean. Journal of Heredity 99, 137-148.
- Hunter, R. L., Halanych, K. M., 2010. Phylogeography of the Antarctic plantotrophic brittle star *Ophionotus* victoriae reveals genetic structure inconsistent with early life history. *Marine Biology* **157**, 1693-1704
- Janko, K., Lecointre, G., DeVries, A., Couloux, A., Cruaud, C., Marshall, C., 2007, Did glacial advances during the Pleistocene influence differently the demographic histories of benthic and pelagic Antarctic shelf fishes? - Inferences from intraspecific mitochondrial and nuclear DNA sequence diversity. BMC Evolutionary Biology 7, 220. ik, A. M., Halanych, K. M., 2010. Unrecognized Antarctic biodiversity: A case study of the genus Odontaster (Odontasteridae; Asteroidea). Integrative and Comparative Biology 50, 981-992.
- Kennett, J. P., 1977. Cenozoic evolution of Antarctic glaciation, the circum-Antarctic Ocean, and their

- impact on global paleoceanography. *Journal of Geophysical Research* **82**, 3843-3860.

 Knox, G. A., 1994. *The biology of the Southern Ocean*. New York: Cambridge University Press.

 Krabbe, K., Leese, F., Mayer, C., Tollrian, R., Held, C., 2009. Cryptic mitochondrial lineages in the widespread pycnogonid *Colossendeis megalonyx* Hoek, 1881 from Antarctic and Subantarctic waters. *Polar Biology* **33**, 281-292.
- Kuklinski, P., Barnes, K. A., 2010. First bipolar benthic brooder. Marine Ecology Progress Series 401,
- 15-20.

 Lawver, L. A., Gahagan, L. M., 2003. Evolution of Cenozoic seaways in the circum-Antarctic region.
- Palaeogeography Palaeoclimatology Palaeoecology 198, 11-37.

 Lecointre, G., Améziane, N., Boisselier, M. -C., Bonillo, C., Busson, F., Causse, R., Chenuil, A., Couloux, A., Coutanceau, J. -P., Cruaud, C., d'Udekem d'Acoz, C., De Ridder, C., Denys, G., Dettai, A., Duhamel, G., Eléaume, M., Féral, J.-P., Gallut, C., Havermans, C., Held, C., Hemery, L., Lautrédou, .-C., Martin, P., Ozouf-Costaz, C., Pierrat, B., Pruvost, P., Puillandre, N., Samadi, S., Saucède T., Schubart, C. D., David, B., 2013. Is the species flock concept operational? The Antarctic shelf case. PloS One. 8, e68787.
- Leese, F., Agrawal, S., Held, C., 2010. Long-distance island hopping without dispersal stages: transportation across major zoogeographic barriers in a Southern Ocean isopod. *Naturwissenschaften* **97**, 583-594
- Leese, F., Held, C., 2011. Analyzing intraspecific genetic variation: A practical guide using mitochondrial DNA and microsatellites. In: Koenemann, S., Schubart, C. D. (eds.) *Analyzing intraspecific genetic variation: A practical guide using mitochondrial DNA and microsatellites*. CRC Press. pp. 3-30.
- Leese, F., Kop, A., Agrawal, S., Held, C., 2008a. Isolation and characterization of microsatellite markers from the marine isopods Serolis paradoxa and Septemserolis septemcarinata (Crustacea: Peracarida). Molecular Ecology Resources 8, 818-821.
- Leese, F., Kop, A., Wägele, J. W., Held, C., 2008b. Cryptic speciation in a benthic isopod from Patagonian and Falkland Island waters and the impact of glaciations on its population structure. *Frontiers in* Zoology 5, 19
- Linse, K., Cope, T., Lörz, A.-N., Sands, C. J., 2007, Is the Scotia Sea a centre of Antarctic marine diversification? Some evidence of cryptic speciation in the circum-Antarctic bivalve *Lissarca notorcadensis* (Arcoidea: Philobryidae). *Polar Biology* **1047**, 1059-1068.
- Linse, K., Griffiths, H. J., Barnes, D. K., Brandt, A., Davey, N., David, B., De Grave, S., d'Udekem d'Acoz, C., Eléaume, M., Glover, A. G., Hemery, L. G., Mah, C., Martín-Ledo, R., Munilla, T., O'Loughlin, M., Pierrat, B., Saucède, T., Sands, C. J., Strugnell, J. M., Enderlein, P., 2013. The macro- and megabenthic fauna on the continental shelf of the eastern Amundsen Sea, Antarctica. Continental Shelf Research 68, 80-90.
- Mahon, A. R., Thornhill, D. J., Norenburg, J. L., Halanych, K. M., 2010. DNA uncovers Antarctic ne biodiversity and exposes a decades-old cold case of asymmetric inventory. Polar Biology 33, 193-202
- Mantyla, A. W., Reid, J. L., 1983. Abyssal characteristics of the World Ocean waters. Deep Sea Research Part A. Oceanographic Research Papers 30, 805-833.
- Matschiner, M., Hanel, R., Salzburger, W., 2009. Gene flow by larval dispersal in the Antarctic notothenioid fish *Gobionotothen gibberifrons. Molecular Ecology* 18, 2574-2587.

 McGaughran, A., Stevens, M. I., Hogg, I. D., Carapelli, A., 2011. Extreme glacial legacies: a synthesis of
- the Antarctic springtail phylogeographic record. *Insects* **2**, 62-82. L. L., Kocot, K. M., Citarella, M. R., Dosung, S., Norekian, T. P., Povolotskaya, I. S., Grigorenko, A. P., Dailey, C., Berezikov, E., Buckley, K. M., Ptitsyn, A., Reshetov, D., Mukherjee, K., Moroz, T. P., Bobkova, Y., Yu, F., Kapitonov, V. V., Jurka, J., Bobkov, Y. V., Swore, J. J., Girardo, D. O., Fodor, A., Gusev, F., Sanford, R., Bruders, R., Kittler, E., Mills, C. E., Rast, J. P., Derelle, R., Solovyev, V. V., Kondrashov, F. A., Swalla, B. J., Sweedler, J. V., Rogaev, E. I., Halanych, K. M., Kohn, A. B., 2014.
- The ctenophore genome and the evolutionary origins of neural systems. *Nature*, **510**, 109-114.

 T. J., Cheng, C. -H. C., 2008. Phylogenetics of notothenioid fishes (Teleostei: Acanthomorpha): Inferences from mitochondrial and nuclear gene sequences. *Molecular Phylogenetics and* Evolution, 47, 832-840.
- Pawlowski, J., Fahrni, J., Lecroq, B., Longet, D., Cornelius, N., Excoffier, L., Cedhagen, T., Gooday, A.J., 2007. Bipolar gene flow in deep-sea benthic foraminifera. *Molecular Ecology* **16**, 4089-4096. Pfenninger, M., Schwenk, K., 2007. Cryptic animal species are homogeneously distributed among taxa
- and biogeographical regions. *BMC Evolutionary Biology* **7**, 121.

 Post, A. L., Beaman, R. J., O'Brien, P. E., Eléaume, M., Riddle, M. J., 2011. Community structure and benthic habitats across the George V Shelf, East Antarctica: Trends through space and time. *Deep*
- Sea Research Part II: Topical Studies in Oceanography 55, 105-118.

 Queiroz, A., 2005. The resurrection of oceanic dispersal in historical biogeography. Trends in Ecology & Evolution 20, 68-73.
- Raupach, M., Thatje, S., Dambach, J., Rehm, P., Misof, B., Leese, F., 2010. Genetic homogeneity and circum-Antarctic distribution of two benthic shrimp species of the Southern Ocean, *Chorismus antarcticus* and *Nematocarcinus lanceopes. Marine Biology* 157, 1783-1797.
- Raupach, M. J., Malyutina, M., Brandt, A., 2007. Molecular data reveal a highly diverse species flock within the munopsoid deep-sea isopod *Betamorpha fusiformis* (Barnard, 1920) (Crustacea : Isopoda Asellota) in the Southern Ocean. *Deep-Sea Research II* **54**, 1820-1830.
- Raupach, M. J., Wägele, J. W., 2006. Distinguishing cryptic species in Antarctic Asellota (Crustacea: Isopoda) a preliminary study of mitochondrial DNA in *Acanthaspidia drygalskii*. *Antarctic Science* **18,** 191-198.
- Riehl, T., Kaiser, S., 2012. Conquered from the deep sea? A new deep-sea isopod species from the
- Antarctic shelf shows pattern of recent colonization. *PLoS One* **7**, e49354.

 Sanmartin, I., 2012. Historical biogeography: Evolution in time and space. *Evolution: Education and* Outreach 5, 555-568.
- Sanmartin, I., van der Mark, P., Ronquist, F., 2008. Inferring dispersal: a Bayesian approach to phylogeny-based island biogeography, with special reference to the Canary Islands. *Journal of Biogeography*
- 35, 428-449.
- Scher, H. D., Martin, E. E., 2006. Timing and climatic consequences of the opening of Drake Passa Science 312, 428-430. Schiaparelli, S., Olivero, M., Taviani, M., Griffiths, H., Lorz, A.-N., Albertelli, G., 2008. Circumpolar
- distribution of the pycnogonid-ectoparasitic gastropod *Dickdellia labioflecta* (Dell, 1990) (Mollusca: Zerotulidae). *Antarctic Science* **20**, 497-498. Schüller, M., 2011. Evidence for a role of bathymetry and emergence in speciation in the genus Glycera
- (Glyceridae, Polychaeta) from the deep Eastern Weddell Sea. *Polar Biology* **34**, 549-564. P. J., Steinke, D., McMillan, P. J., Stewart, A. L., McVeagh, S. M., Diaz de Astarloa, J. M., Welsford, D., Ward, R. D., 2011. DNA barcoding highlights a cryptic species of grenadier *Macrourus* in the
- Southern Ocean. *Journal of Fish Biology*, **78**, 355-365.

 Strugnell, J. M., Rogers, A. D., Prodohl, P. A., Collins, M. A., Allcock, A. L., 2008. The thermohaline expressway: the Southern Ocean as a centre of origin for deep-sea octopuses. *Cladistics* **24**, 853-860
- Strugnell, J. M., Watts, P. C., Smith, P. J., Allcock, A. L., 2012. Persistent genetic signatures of historic climatic events in an Antarctic Octopus. Molecular Ecology 21, 2775-2787.
- Strugnell, M., Cherel, Y., Cooke, R., Gleadall, G., Hochberg, G., Ibáñez, M., Jorgensen, E., Laptikhovsky, V., Linse, K., Norman, M., Vecchione, M., Voight, R., Allcock, L., 2011. The Southern Ocean: Source and sink? *Deep Sea Research Part II: Topical Studies in Oceanography* 58, 196-204.
 Thatje, S., Hillenbrand, C.-D., Larter, R., 2005. On the origin of Antarctic marine benthic community
- structure. *Trends in Ecology and Evolution* 20, 534-540.

 Thornhill, D. J., Mahon, A. R., Norenburg, J. L., Halanych, K. M., 2008. Open-ocean barriers to dispersal: a test case with the Antarctic Polar Front and the ribbon worm *Parborlasia corrugatus* (Nemertea:
- Lineidae). Molecular Ecology 17, 5104-5117.

 Uriz, M.-J., Gili, J.-M., Orejas, C., Perez-Porro, A.-R., 2011. Do bipolar distributions exist in marine sponges? Stylocordyla chupachups sp. nov. (Porifera: Hadromerida) from the Weddell Sea
- (Antarctic), previously reported as S. borealis (Lovén, 1868). Polar Biology 34, 243-255. ele, J.-W., 1994. Notes on Antarctic and South American Serolidae (Crustacea, Isopoda) with remarks on the phylogenetic biogeography and a description of new genera. Zoologische Jahrbücher.
- Abteilung für Systematik, Ökologie und Geographie der Tiere 121, 3-69.

 Wilson, N. G., Hunter, R. L., Lockhart, P. J., Halanych, K. M., 2007. Multiple lineages and absence of panmixia in the "circumpolar" crinoid Promachocrinus kerguelensis from the Atlantic sector of Antarctica. Marine Biology 152, 895-904.
- Wilson, N. G., Schrödl, M., Halanych, K. M., 2009. Ocean barriers and glaciation: Evidence for explosive radiation of mitochondrial lineages in the Antarctic sea slug *Doris kerguelenensis* (Mollusca,
- Nudibranchia). Molecular Ecology 18, 965-984.

 Zane, L., Ostellari, L., Maccatrozzo, L., Bargelloni, L., Battaglia, B., Patarnello, T., 1998. Molecular evidence for genetic subdivision of Antarctic krill (Euphausia superba Royal Society, B: Biological Sciences 265, 2387-2391.

THE BIOGEOGRAPHIC ATLAS OF THE SOUTHERN OCEAN

Biogeographic information is of fundamental importance for discovering marine biodiversity hotspots, detecting and understanding impacts of environmental changes, predicting future distributions, monitoring biodiversity, or supporting conservation and sustainable management strategies

The recent extensive exploration and assessment of biodiversity by the Census of Antarctic Marine Life (CAML), and the intense compilation and validation efforts of Southern Ocean biogeographic data by the SCAR Marine Biodiversity Information Network (SCAR-MarBIN / OBIS) provided a unique opportunity to assess and synthesise the current knowledge on Southern

The scope of the Biogeographic Atlas of the Southern Ocean is to present a concise synopsis of the present state of knowledge of the distributional patterns of the major benthic and pelagic taxa and of the key communities, in the light of biotic and abiotic factors operating within an evolutionary framework. Each chapter has been written by the most pertinent experts in their field, relying on vastly improved occurrence datasets from recent decades, as well as on new insights provided by molecular and phylogeographic approaches, and new methods of analysis, visualisation, modelling and prediction of biogeographic distributions.

A dynamic online version of the Biogeographic Atlas will be hosted on www.biodiversity.aq.

The Census of Antarctic Marine Life (CAML)

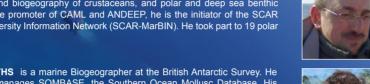
CAML (www.caml.aq) was a 5-year project that aimed at assessing the nature, distribution and abundance of all living organisms of the Southern Ocean. In this time of environmental change, CAML provided a comprehensive baseline information on the Antarctic marine biodiversity as a sound benchmark against which future change can reliably be assessed. CAML was initiated in 2005 as the regional Antarctic project of the worldwide programme Census of Marine Life (2000-2010) and was the most important biology project of the International Polar Year 2007-2009.

The SCAR Marine Biodiversity Information Network (SCAR-MarBIN)
In close connection with CAML, SCAR-MarBIN (www.scarmarbin.be, integrated into www.biodiversity.aq) compiled and managed the historic, current and new information (i.a. generated by CAML) on Antarctic marine biodiversity by establishing and supporting a distributed system of interoperable databases, forming the Antarctic regional node of the Ocean Biogeographic Information System (OBIS, www.iobis.org), under the aegis of SCAR (Scientific Committee on Antarctic Research, www.scar.org). SCAR-MarBIN established a comprehensive register of Antarctic marine species and, with biodiversity.aq provided free access to more than 2.9 million Antarctic georeferenced biodiversity data, which allowed more than 60 million downloads.

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