

Using R tools for analysis of primary biodiversity data provided by SBDI

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Introduction

Biodiversity resources are increasingly international. The SBDI has made an effort to canalize biodiversity data and resources to help the research community access and analyze Swedish primary biodiversity data. Each research question draws its own challenges which are unique in themselves. Our aim here is to provide a few examples that prompt questions that may be asked at different stages of the process. The validity and appropriateness of a particular method depends on the individual researcher(s). For a comprehensive workflow on how to treat and analyze PBD please refer to our tutorial on [biodiversity analysis tool](#) where we go through the complete workflow Data -> Cleaning -> Fitness evaluation -> Analysis

R and Mirroreum

The present tutorial is focused on the statistical programming language R. R is a free software environment for statistical computing and graphics that is widely used within the scientific community and where the complete analysis workflow can be documented in a fully reproducible way.

At SBDI we provide access for researchers and students to [Mirroreum](#) – an online web-based environment for Reproducible Open Research in the area of biodiversity analysis. Mirroreum is based on a Free and Open Source stack of software. Logging in, you immediately get access to a web-based version of R Studio with a large number of pre-installed packages such as all the packages offered from ROpenSci and more.

Compared to running R Studio on your own machine, Mirroreum offers more computational resources and a standardized environment where you can rely on all the relevant packages being installed and the configuration parameters being set appropriately. To know more about Mirroreum or to request an account please visit the [SBDI documentation site](#)

SBDI4R - R package to search an access data

The SBDI4R package enables the R community to directly access data and resources hosted by SBDI. The goal is to enable observations of species to be queried and output in a range of standard formats. It includes some filter functions that allow you to filter prior to download. It also includes some simple summary functions, and some function for some simple data exploration. The examples included in this tutorial also show you how you can continue exploring and analyzing using other R package.

Please refer to the [package documentation](#) for details on how to install it. Once installed the SBDI4R package must be loaded for each new R session:

```
library(SBDI4R)
```

Customizing SBDI4R

Various aspects of the SBDI4R package can be customized.

Caching

SBDI4R can cache most results to local files. This means that if the same code is run multiple times, the second and subsequent iterations will be faster. This will also reduce load on the web servers. By default, this caching is session-based, meaning that the local files are stored in a temporary directory that is automatically deleted when the R session is ended. This behaviour can be altered so that caching is permanent, by setting the caching directory to a non-temporary location. For example, under Windows, use something like:

```
sbdi_config(cache_directory = file.path("c:", "mydata", "sbdi_cache")) ## Windows
```

or for Linux:

```
sbdi_config(cache_directory = "~/mydata/sbdi_cache") ## Linux
```

Note that this directory must exist (you need to create it yourself).

All results will be stored in that cache directory and will be used from one session to the next. They won't be re-downloaded from the server unless the user specifically deletes those files or changes the caching setting to “refresh”.

If you change the `cache_directory` to a permanent location, you may wish to add something like this to your `.Rprofile` file, so that it happens automatically each time the SBDI4R package is loaded:

```
setHook(packageEvent("SBDI4R", "onLoad"),
        function(...) sbdi_config(cache_directory=file.path("~", "mydata", "sbdi_cache")))
```

Caching can also be turned off entirely by:

```
sbdi_config(caching="off")
```

or set to “refresh”, meaning that the cached results will re-downloaded from the SBDI servers and the cache updated. (This will happen for as long as caching is set to “refresh” — so you may wish to switch back to normal “on” caching behavior once you have updated your cache with the data you are working on).

E-mail address

Each download request to SBDI servers is also accompanied by an “e-mail address” string that identifies the user making the request. You will need to provide an email address registered with the SBDI. You can create an account [here](#). Once an email is registered with the SBDI, it should be stored in the config:

```
sbdi_config(email="your.valid@emailaddress.com")
```

Else you can provide this e-mail address as a parameter directly to each call of the function `occurrences()`.

Setting the download reason

SBDI requires that you provide a reason when downloading occurrence data (via the SBDI4R `occurrences()` function). You can provide this as a parameter directly to each call of `occurrences()`, or you can set it once per session using:

```
sbdi_config(download_reason_id = "your_reason_id")
```

(See `sbdi_reasons()` for valid download reasons, e.g. * 3 for “education”, * 7 for “ecological research”, * 8 for “systematic research/taxonomy”, * 10 for “testing”)

NO other personal identification information is sent. You can see all configuration settings, including the the user-agent string that is being used, with the command:

```
sbdi_config()
```

Other options

If you make a request that returns an empty result set (e.g. an un-matched name), by default you will simply get an empty data structure returned to you without any special notification. If you would like to be warned about empty result sets, you can use:

```
sbdi_config(warn_on_empty=TRUE)
```

Other packages needed

Some additional packages are needed for these examples. Install them if necessary with the following script.

```
to_install <- c("BIRDS", "colorRamps", "cowplot", "dplyr", "ggplot2", "leaflet",  
              "maps", "mapdata", "maptools", "sf", "sp", "rgeos", "tidyr", "xts")  
to_install <- to_install[!sapply(to_install, requireNamespace, quietly=TRUE)]  
if(length(to_install)>0)  
  install.packages(to_install, repos="http://cran.us.r-project.org")
```

Your collaboration is appreciated

Open Source also means that you can contribute. You don't need to know how to program but every input is appreciated. Did you find something that is not working? Have suggestions for examples or text? you can always

1. Reach to us via the [support center](#)
2. Submit and issue to the GitHub code repository [see how](#)
3. Or contribute with your code or documents modifications by “forking” the code and submitting a “pull request”

The repositories you can contribute to are:

- Mirroreum <https://github.com/mskyttner/mirroreum>
- SBDI4R <https://github.com/biodiversitydata-se/SBDI4R> (NOTE: we may not develop this package but instead move to a new one)
- the general analysis workflows <https://github.com/biodiversitydata-se/biodiversity-analysis-tools>
- these tutorial <https://github.com/biodiversitydata-se/r-tools-tutorial>

1 Example with fish data from SERS

In this example we are interested in exploring data from a specific data resource – Swedish Electrofishing Registry - SERS (Institutionen för akvatiska resurser, SLU). This data base has 2.8 M observations starting in the 1950's.

As you may already know, SBDI is a collection of many biodiversity databases. We start by searching for the data resource we are interested in using the function `pick_filter()`. This is an interactive query guiding you through the many resources available to filtering your query (data resources, spatial layers, and curated species lists).

```
library(SBDI4R)  
fq_str <- pick_filter("resource")  
# follow the instructions
```

Follow the instruction. Your choices here would have been “in3” -> “dr10”. Your variable `fq_str` will now contain a string “data_resource_uid:dr10”.

But we are not interested in the complete database, but on the last 10 years of data. for this we concatenate (add to a vector) another filter string. These will be treated as AND factors.

```

y1 <- 2008
y2 <- 2012
fq_str <- c(fq_str, paste0("year:[", y1, " TO ", y2, "]"))
# Note the square brackets are hard limits

```

For references on how to use the filters see SBDI APIS [documentation](#).

Using the function `occurrences()` we can the query for the observations fulfilling our filter. If you haven't specified that in the `sbd_i_config()` before, you need to pass your email and the download reason.

```

library(SBDI4R)
xf <- occurrences(fq = fq_str,
                  email = "sbd_i4r-test@biodiversitydata.se",
                  download_reason_id = 10)

```

```

## Registered S3 methods overwritten by 'ALA4R':
##   method      from
##   subset.occurrences SBDI4R
##   summary.occurrences SBDI4R
##   unique.occurrences SBDI4R

```

```

# Simply summarise all records by data source
table(xf$data$dataResourceName)

```

```

##
## SLU Aqua   Institute of Freshwater Research Swedish Electrofishing Registry - SERS
##                                                    95082

```

```

table(xf$data$dataResourceID)

```

```

##
## dr10
## 95082

```

1.1 Plotting data on a map

You can quickly plot all the observations as a PDF file with the function `occurrence_plot()`, one page per species:

```

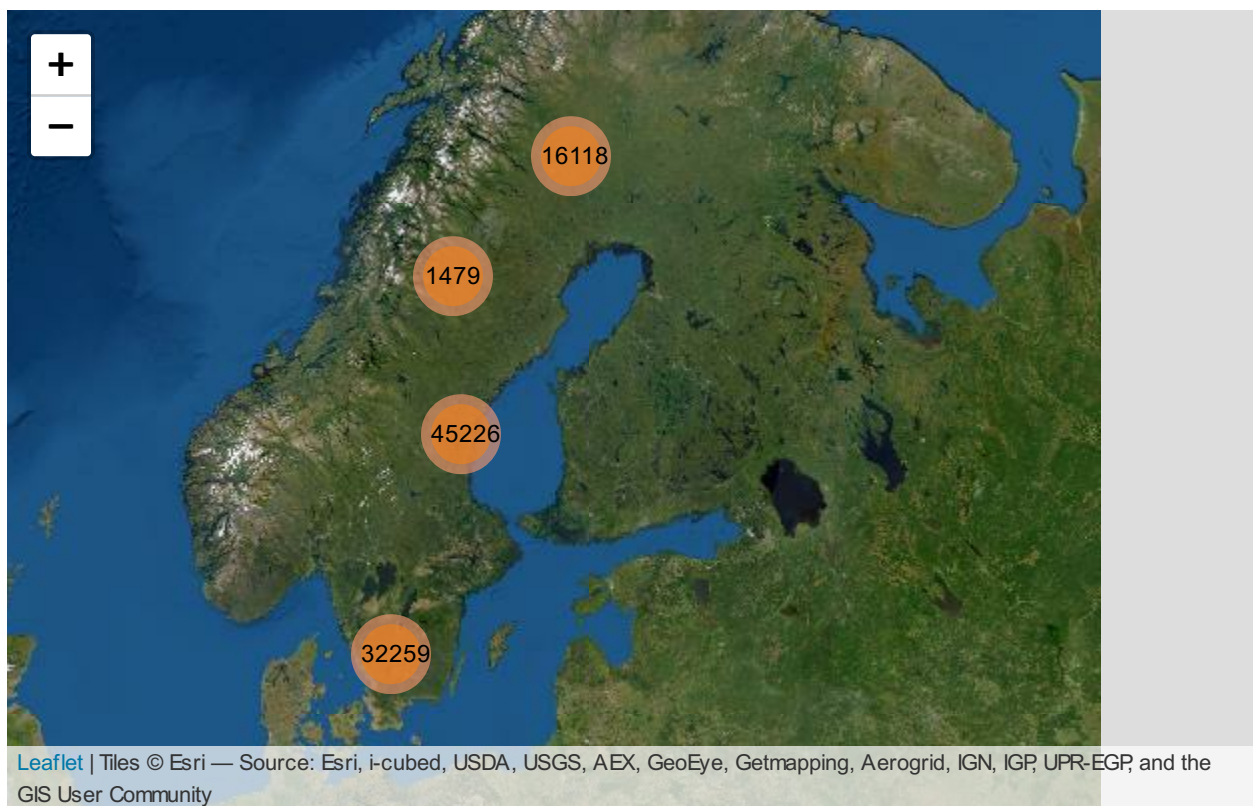
occurrences_plot(xf, "obsPlot.pdf",
                  grouped=FALSE,
                  taxon_level="species",
                  pch='+')

```

Note that the plot is saved to a pdf file in the current working directory. You can find that with `getwd()`.

1.1.0.1 Leaflet There are many other ways of producing spatial plots in R. The leaflet package provides a simple method of producing browser-based maps with panning, zooming, and background layers:

```
library(leaflet)
# drop any records with missing lat/lon values
xfl <- xfl$data[!is.na(xfl$data$longitude) | !is.na(xfl$data$latitude),]
marker_colour <- rep("#d95f02", nrow(xfl))
# blank map, with imagery background
leaflet(width = "100%") %>%
  addProviderTiles("Esri.WorldImagery") %>%
  # add markers
  addCircleMarkers(xfl$longitude, xfl$latitude,
    radius = 1,
    fillOpacity = .5,
    opacity = 1,
    col=marker_colour,
    clusterOptions = markerClusterOptions())
```



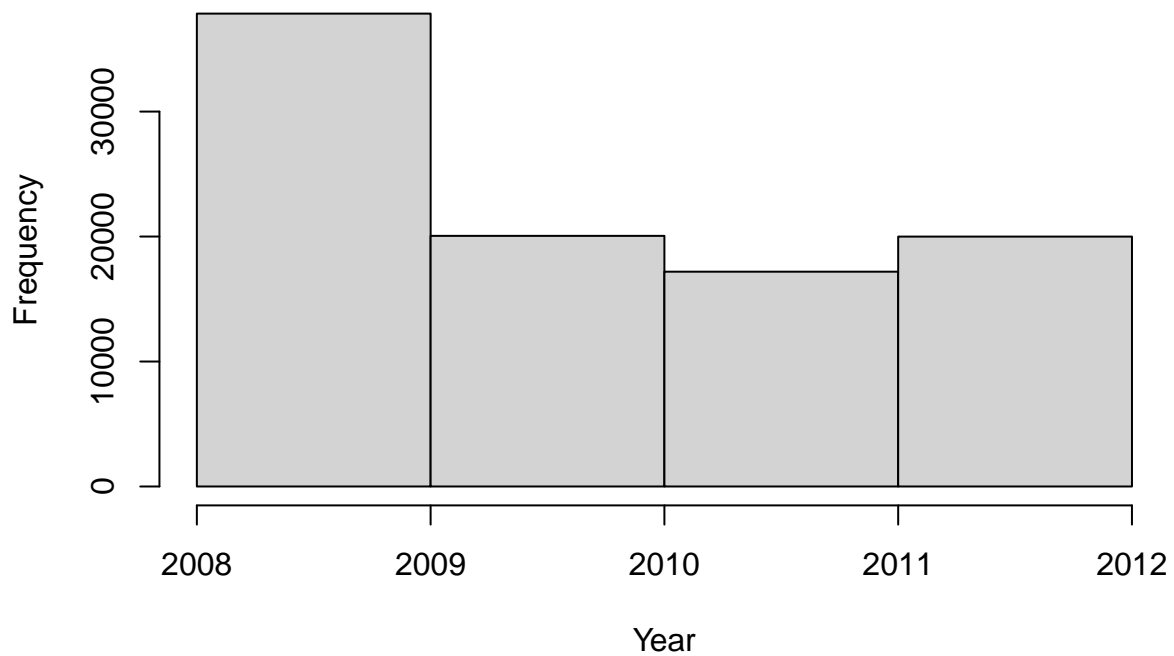
1.2 Temporal summary

A quick summary over the years reveal a drop in number of records over time.

```
table(xf$data$year)
```

```
##  
## 2008 2009 2010 2011 2012  
## 18168 19674 20055 17188 19997
```

```
hist(xf$data$year,  
     breaks = seq(y1, y2),  
     xlab = "Year",  
     main = "")
```



1.3 Species summary

In the same way we can summaries the number of observations for each species, by common or scientific name.

```
sppTab <- table(xf$data$commonName)  
sppDF <- as.data.frame(sppTab)  
colnames(sppDF)[1] <- "species"  
head(sppDF)
```

```
##           species Freq  
## 1                66
```

```
## 2 Alpine bullhead 4615
## 3 American burbot 7081
## 4      Aral asp    6
## 5      Arctic char 46
## 6      aurora trout 856
```

```
sppTab <- table(xf$data$scientificName)
sppDF <- as.data.frame(sppTab)
colnames(sppDF)[1] <- "species"
head(sppDF)
```

```
##               species Freq
## 1      Abramis brama (Linnaeus, 1758) 61
## 2      Alburnus alburnus (Linnaeus, 1758) 660
## 3      Anguilla anguilla (Linnaeus, 1758) 2140
## 4               Astacidae 100
## 5      Astacus astacus (Linnaeus, 1758) 618
## 6 Barbatula barbatula (Linnaeus, 1758) 620
```

Perhaps, you need to send this table as a .CSV file to a colleague.

```
write.csv(sppDF, "SERS_species_summary.csv")
# NOTE: again this will be saved on your working directory
```

1.4 Spatial biodiversity analysis

Let's now ask: how does the species richness vary across Sweden? In this case we want to summarise occurrences species-wise over a defined grid instead of plotting every observation point. First we need to overlay the observations with a grid. In this case, the standard Swedish grids at 50, 25, 10 and 5 km are provided as data in the SBDI4R package (with Coordinate Reference System = WGS84, EPSG:4326).

```
library(sp) # the function coordinates() and proj4string() are in sp
```

```
## Warning: package 'sp' was built under R version 4.0.3
```

```
library(rgeos) # the function over() is in package rgeos
```

```
## rgeos version: 0.5-5, (SVN revision 640)
## GEOS runtime version: 3.8.0-CAPI-1.13.1
## Linking to sp version: 1.4-2
## Polygon checking: TRUE
```

```
# load some shapes over Sweden's political borders
data("swe_wgs84", package="SBDI4R", envir=environment())
# A standard 50km grid
data("Sweden_Grid_50km_Wgs84", package="SBDI4R", envir=environment())

grid <- Sweden_Grid_50km_Wgs84

# make the observations spatial
```



```

# NOTE: make sure there are no NAs on either column defining the coordinates
# xf$data[!is.na(xf$data$longitude) / !is.na(xf$data$latitude),]

obs <- as.data.frame(xf$data)
coordinates(obs) <- obs[,c("longitude", "latitude")]
wkt <- sf::st_crs(4326)[[2]]
proj4string(obs) <- sp::CRS(wkt) #CRS("+init=epsg:4326")

nObs <- nrow(obs)

# overlay the data with the grid
ObsInGridList <- over(grid, obs, returnList=TRUE)
wNonEmpty <- unname( which( unlist(lapply(ObsInGridList, nrow)) != 0 ) )
if(length(wNonEmpty)==0) message("Observations don't overlap any grid cell.")

```

The result `ObsInGridList` is a list object with a subset of the data on each grid. Now summarise occurrences within grid cells:

```

# check n the total number of observations
sum(unlist(lapply(ObsInGridList, nrow)))

```

```
## [1] 95082
```

```

# apply a summary over the grid cells
nCells <- length(ObsInGridList)

res <- data.frame("nObs"=as.numeric(rep(NA,nCells)),
                 "nYears"=as.numeric(rep(NA,nCells)),
                 "nSpp"=as.numeric(rep(NA,nCells)),
                 row.names = row.names(grid),
                 stringsAsFactors = FALSE)

cols2use <- c("scientificName", "year")

dataRes <- lapply(ObsInGridList[wNonEmpty],
                 function(x){
                   x <- x[,cols2use]
                   colnames(x) <- c("scientificName", "year")
                   return(c("nObs" = length(x[, "scientificName"]),
                           "nYears" = length(unique(x[, "year"])),
                           "nSpp" = length(unique(x[, "scientificName"])))
                   )
                 })

dataRes <- as.data.frame(dplyr::bind_rows(dataRes, .id = "gridID"))

res[wNonEmpty,] <- dataRes[,-1]

resSp <- sp::SpatialPolygonsDataFrame(grid, res)

```

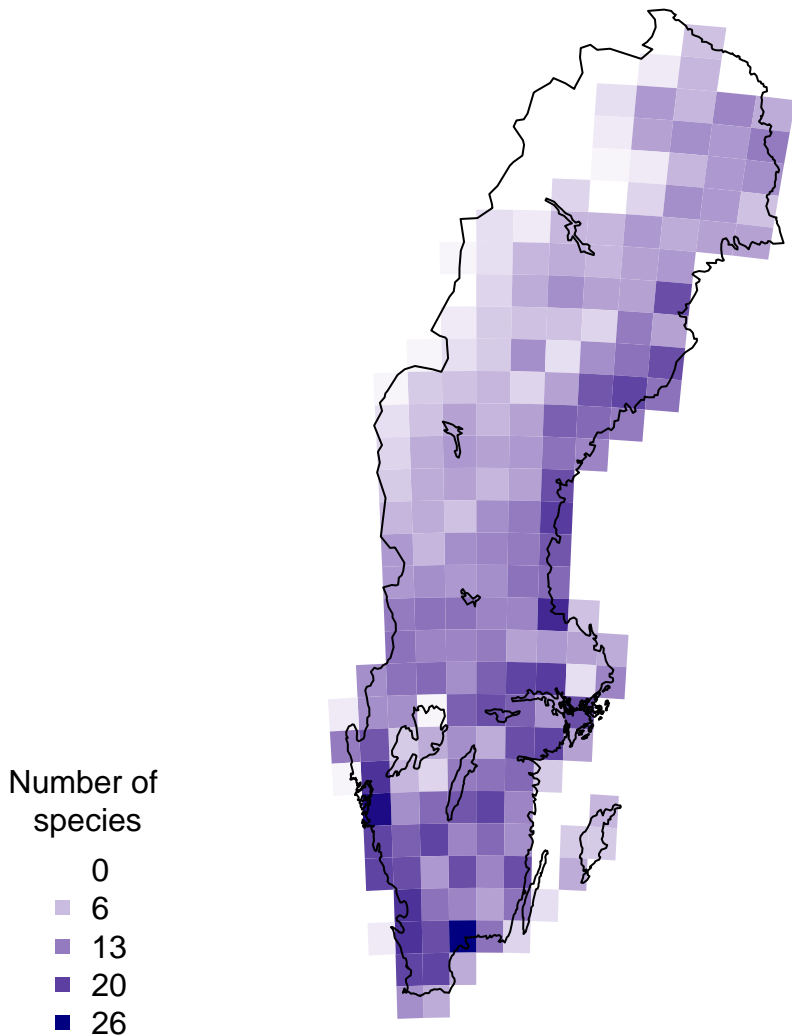
And finally plot the grid summary as a map:

```

palBW <- leaflet::colorNumeric(c("white", "navyblue"),
                              c(0, max(resSp@data$nSpp, na.rm = TRUE)),
                              na.color = "transparent")

oldpar <- par()
par(mar = c(1,1,0,0))
plot(resSp, col=palBW(resSp@data$nSpp), border = NA)
plot(swe_wgs84$Border, border=1, lwd=1, add=T)
legend("bottomleft",
      legend = round(seq(0, max(resSp@data$nSpp, na.rm = TRUE), length.out = 5)),
      col = palBW(seq(0, max(resSp@data$nSpp, na.rm = TRUE), length.out = 5)),
      title = "Number of \nspecies", pch = 15, bty="n")

```



```
suppressWarnings(par(oldpar))
```

We can go further by gathering the observations by latitude.

```

library(dplyr)
library(tidyr)
xgridded <- xf$data %>%
  ## discard genus- and higher-level records
  filter(rank %in% c("species", "subspecies", "variety", "form", "cultivar")) %>%
  mutate(longitude = round(longitude * 4)/4,
         latitude = round(latitude * 4)/4) %>%
  group_by(longitude, latitude) %>%
  ## subset to vars of interest
  select(longitude, latitude, species) %>%
  ## take one row per cell per species (presence)
  distinct() %>%
  ## calculate species richness
  mutate(richness=n()) %>%
  ## convert to wide format (sites by species)
  mutate(present=1) %>%
  do(tidyr::pivot_wider(data=., names_from=species, values_from=present, values_fill=0)) %>%
  ungroup()
## where a species was not present, it will have NA: convert these to 0
sppcols <- setdiff(names(xgridded),
                  c("longitude", "latitude", "richness"))
xgridded <- xgridded %>%
  mutate_at(sppcols, function(z) ifelse(is.na(z), 0, z))

```

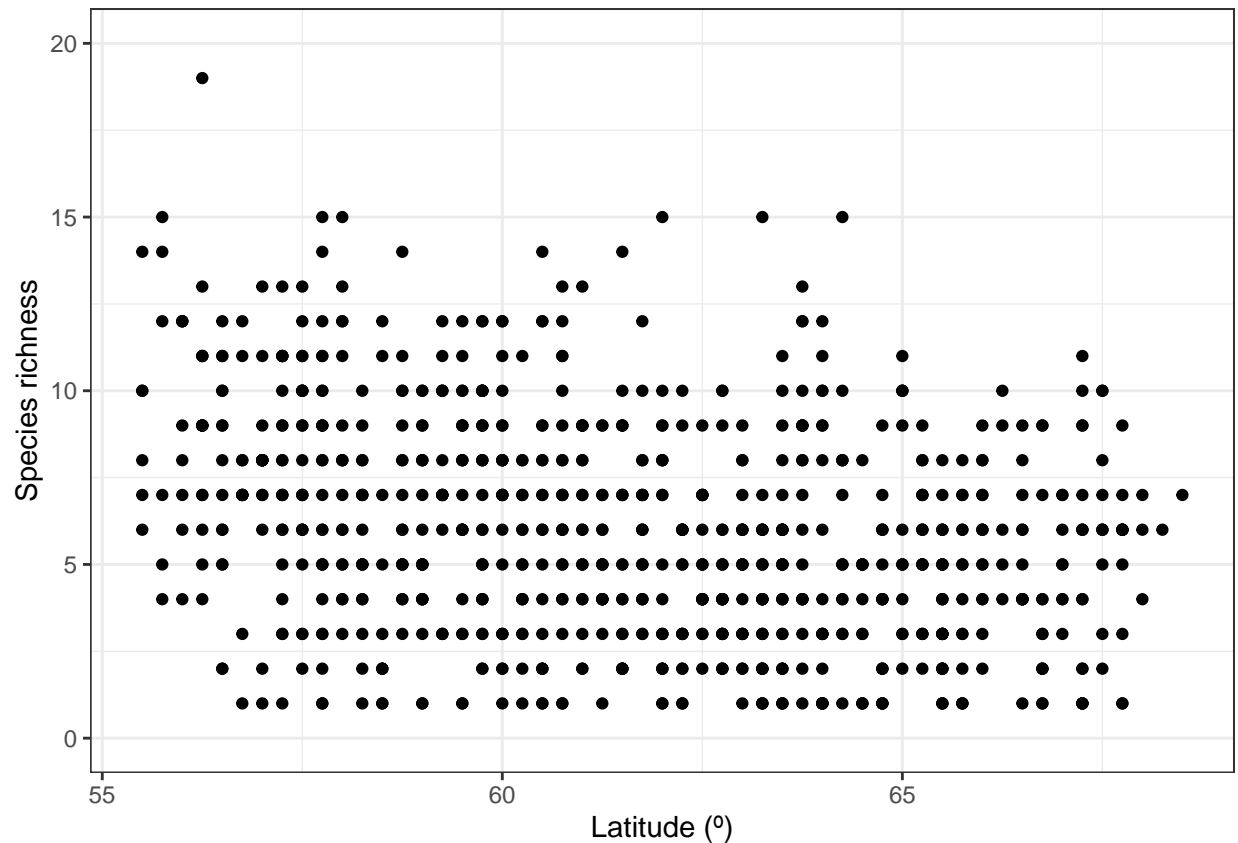
And plot it accordingly

```

library(ggplot2)

ggplot(xgridded, aes(latitude, richness)) +
  labs(x = "Latitude (°)",
       y = "Species richness") +
  lims(y = c(0,20)) +
  geom_point() +
  theme_bw()

```



2 Example with opportunistic data on Dragonflies

In this example we are interested in exploring opportunistically collected data from the Swedish citizen science observation data portal - Artportalen.

2.1 Name searching

To begin, we want be sure there is an unequivocal way to find the species within the order Odonata and nothing else, so let's search for it:

```
library(SBDI4R)
sx <- search_fulltext("odonata")
```

```
## [1] "https://species.biodiversitydata.se/ws/search.json?q=odonata&fq=idxtype%3ATAXON"
```

```
sx
```

```
## Search metadata:
##   totalRecords queryTitle
## 1           5    odonata
##
## Facet results:
```

```
## [1] "NULL"
##
## Search results:
##   name                                     commonNameSingle rank
## 1 "Odonata associated gemycircularvirus 1" ""                "species"
## 2 "Odonata associated gemycircularvirus 2" ""                "species"
## 3 "Bdellodes odonata Wallace & Mahon, 1976" ""              "species"
## 4 "Odonata"                               ""                "order"
## 5 "Ramalina fastigiata var. odonata Hue"   ""                "variety"
##   guid
## 1 "9829523"
## 2 "10072832"
## 3 "8062407"
## 4 "789"
## 5 "7367071"
```

we see there that other taxonomic definitions appear too, but only one order. Let's refine the search. To know the names of the search fields (that may not be the same as returned column names) we can use the function `sbdi_fields(fields_type = "general")`. The search field we are looking for is "order_s".

```
sx <- search_fulltext(fq="order_s:Odonata", page_size = 10)
```

```
## [1] "https://species.biodiversitydata.se/ws/search.json?fq=order_s%3AOdonata&fq=idxtype%3ATAXON&page=1"
```

```
sx$data[,c("name", "scientificName", "guid", "rank")]
```

```
##                                     name
## 1      Gomphomacromia Brauer, 1864
## 2      Austropetalia Tillyard, 1916
## 3      Sogjutella Pritykina, 1980
## 4      Neuragrion Karsch, 1891
## 5      Xamenophlebia Pritykina, 1981
## 6      Lauromacromia Geijskes, 1970
## 7      Sympetrum Newman, 1833
## 8      Corduliochlora Marinov & Seidenbusch, 2007
## 9      Torrenticnemis Lieftinck, 1949
## 10     Cyanallagma Kennedy, 1920
##                                     scientificName    guid  rank
## 1      Gomphomacromia Brauer, 1864 1429753 genus
## 2      Austropetalia Tillyard, 1916 1426725 genus
## 3      Sogjutella Pritykina, 1980 4799335 genus
## 4      Neuragrion Karsch, 1891 4302686 genus
## 5      Xamenophlebia Pritykina, 1981 4799353 genus
## 6      Lauromacromia Geijskes, 1970 1429769 genus
## 7      Sympetrum Newman, 1833 1428195 genus
## 8      Corduliochlora Marinov & Seidenbusch, 2007 4798599 genus
## 9      Torrenticnemis Lieftinck, 1949 1423625 genus
## 10     Cyanallagma Kennedy, 1920 1423468 genus
```

Now we can download the taxonomic data (note that the search is case-sensitive):

```
tx <- taxinfo_download("order_s:Odonata",
  fields = c("guid", "order_s", "genus_s", "specificEpithet_s",
    "scientificName", "canonicalName_s", "rank"),
  verbose = FALSE)
tx <- tx[tx$rank == "species" & tx$genusS != "",] ## restrict to species and not hybrids
```

Now `tx` is our complete species list.

2.2 Get the observations, filter the search, get quality assertions, plotting data on a map and save data

As usual we start by searching for the data resource we are interested in using the function `pick_filter()`. This is an interactive query guiding you through the many resources available to filtering your query (data resources, spatial layers, and curated species lists).

```
library(SBDI4R)
fq_str <- pick_filter("resource")
# follow the instructions
```

Follow the instruction. Your choices here would have been “in3” → “dr5”. Your variable `fq_str` will now contain a string “data_resource_uid:dr5”.

We only need data from 2000 to 2010

```
y1 <- 2000
y2 <- 2010
fq_str <- c(fq_str, paste0("year:[", y1, " TO ", y2, "]"))
# Note the square brackets are hard limits
```

Select data – get records for Southern Sweden (Götaland).

Vector spatial layers (eg. polygons) can be imported in a number of different ways. SBDI APIs take as search input polygons in the s.k. WKT [Well Known Text](#) format. So the first step is to load a vector layer and transform it into a WKT string. You could instead use the data we kindly provided in the SBDI4R package `data("swe")`.

```
data("swe")
wGotaland <- swe$Counties$LnNamn %in% c("Blekinge", "Gotlands", "Hallands", "Jönköpings", "Kalmar", "Kr
```

We could create the WKT string using the `rgeos` library:

```
library(rgeos)
wkt <- writeWKT(gotaland_c)
```

Unfortunately, in this instance this gives a WKT string that is too long and won’t be accepted by the web service. Also, the shapefile we just got is projected in the coordinate system SWEREF99 TM, and the web service only accepts coordinates in a geodesic coordinate system WGS84. Instead, let’s construct the WKT string directly, which gives us a little more control over its format:

```

gotaland_c <- sf::as_Spatial(
  sf::st_transform(
    sf::st_as_sf(gotaland_c),
    crs = sf::st_crs(4326)$wkt )

gotaland <- rgeos::gUnaryUnion(gotaland_c)

# extract the polygons coordinates
nPol <- length(gotaland@polygons[[1]]@Polygons)
lonlat <- list()
for(p in seq(nPol)){
  lonlat[[p]] <- gotaland@polygons[[1]]@Polygons[[p]]@coords
}
lonlat <- do.call(rbind, lonlat)

# create a convex hull of the polygon to reduce the length of the WKT string
gotaland_ch <- chull(lonlat)
lonlat <- lonlat[c(gotaland_ch, gotaland_ch[1]), ]

# create WKT string
# first join each lon-lat coordinate pair
wkt_temp <- apply(lonlat, 1, function(z) paste(round(z,4), collapse=" "))
# now build the WKT string
wkt <- paste("MULTIPOLYGON((" , paste(wkt_temp, collapse=","), "))", sep="")
# NOTE: as of today, the SBDI APIs will only work properly if the polygon is
# submitted as a MULTIPOLYGON

```

```
sbdi_fields("occurrence")[,1:2]
```

##		name	dataType
## 1	abcd_identification_qualifier		string
## 2	access_rights		string
## 3	assertions		string
## 4	assertions_missing		string
## 5	assertions_passed		string
## 6	assertions_unchecked		string
## 7	basis_of_record		string
## 8	behavior		string
## 9	bibliographic_citation		string
## 10	catalogue_number		string
## 11		cl10038	string
## 12		cl10040	string
## 13		cl10041	string
## 14		cl10042	string
## 15		cl10046	string
## 16		cl10047	string
## 17		cl10048	string
## 18		cl10050	string
## 19		cl10051	string
## 20		cl10052	string
## 21		cl10053	string
## 22		cl10054	string
## 23		cl10055	string

## 24	cl10057	string
## 25	cl10058	string
## 26	cl10059	string
## 27	cl10061	string
## 28	cl10063	string
## 29	cl10064	string
## 30	cl10065	string
## 31	cl10066	string
## 32	cl10067	string
## 33	cl10068	string
## 34	cl10070	string
## 35	cl10071	string
## 36	cl10073	string
## 37	cl10074	string
## 38	cl10082	string
## 39	cl10083	string
## 40	cl10084	string
## 41	cl10087	string
## 42	cl10089	string
## 43	cl10090	string
## 44	cl10097	string
## 45	cl10101	string
## 46	cl10102	string
## 47	cl10104	string
## 48	class	string
## 49	class_id	string
## 50	collection_code	string
## 51	collection_id	string
## 52	collection_name	string
## 53	collection_uid	string
## 54	collector	string
## 55	common_name	string
## 56	common_name_and_lsid	string
## 57	coordinate_uncertainty	tdouble
## 58	country	string
## 59	country_code	string
## 60	county	string
## 61	data_provider	string
## 62	data_provider_uid	string
## 63	data_resource	string
## 64	data_resource_uid	string
## 65	dataset_id	string
## 66	dataset_name	string
## 67	date_precision	string
## 68	datum	string
## 69	day	string
## 70	disposition	string
## 71	dynamic_properties	string
## 72	el10000	tfloat
## 73	el10001	tfloat
## 74	el10002	tfloat
## 75	el10003	tfloat
## 76	el10004	tfloat
## 77	el10005	tfloat

## 78	el10006	tfloat
## 79	el10007	tfloat
## 80	el10008	tfloat
## 81	el10009	tfloat
## 82	el10010	tfloat
## 83	el10011	tfloat
## 84	el10012	tfloat
## 85	el10013	tfloat
## 86	el10014	tfloat
## 87	el10015	tfloat
## 88	el10016	tfloat
## 89	el10017	tfloat
## 90	el10018	tfloat
## 91	el10019	tfloat
## 92	el10020	tfloat
## 93	el10021	tfloat
## 94	el10022	tfloat
## 95	el10023	tfloat
## 96	el10024	tfloat
## 97	el10025	tfloat
## 98	el10026	tfloat
## 99	el10027	tfloat
## 100	el10028	tfloat
## 101	el10029	tfloat
## 102	el10030	tfloat
## 103	el10031	tfloat
## 104	el10032	tfloat
## 105	el10033	tfloat
## 106	el10034	tfloat
## 107	el10035	tfloat
## 108	el10036	tfloat
## 109	el10044	tfloat
## 110	elevation	double
## 111	end_day_of_year	string
## 112	establishment_means	string
## 113	event_id	string
## 114	event_remarks	string
## 115	event_time	string
## 116	family	string
## 117	family_id	string
## 118	field_number	string
## 119	first_loaded_date	tdate
## 120	genus	string
## 121	genus_guid	string
## 122	georeference_remarks	string
## 123	georeferenced_by	string
## 124	georeferenced_date	string
## 125	geospatial_kosher	string
## 126	habitat	string
## 127	higher_classification	string
## 128	higher_geography	string
## 129	id	string
## 130	identification_qualifier	string
## 131	identification_remarks	string

## 132	identified_by	string
## 133	identified_date	tdate
## 134	individual_count	string
## 135	infraspecific_epithet	string
## 136	institution_code	string
## 137	institution_name	string
## 138	institution_uid	string
## 139	island	string
## 140	island_group	string
## 141	kingdom	string
## 142	kingdom_id	string
## 143	language	string
## 144	last_load_date	tdate
## 145	last_processed_date	tdate
## 146	lat_long	string
## 147	latitude	tdouble
## 149	license	string
## 150	location_according_to	string
## 151	location_id	string
## 152	location_remarks	string
## 153	longitude	tdouble
## 154	mappable	string
## 155	max_depth_d	tdouble
## 156	max_elevation_d	tdouble
## 157	min_depth_d	tdouble
## 158	min_elevation_d	tdouble
## 159	min_elevation_d_rng	tdouble
## 160	modified_date	tdate
## 161	month	string
## 162	multimedia	string
## 163	municipality	string
## 164	name_match_metric	string
## 165	name_parse_type	string
## 166	names_and_lsid	string
## 167	occurrence_date	tdate
## 168	occurrence_date_end_dt	tdate
## 169	occurrence_decade_i	tint
## 170	occurrence_id	string
## 171	occurrence_remarks	string
## 172	occurrence_status	string
## 173	occurrence_year	tdate
## 174	order	string
## 175	order_id	string
## 176	organism_quantity	string
## 177	organism_quantity_type	string
## 178	original_name_usage	string
## 179	other_catalog_numbers	string
## 180	outlier_layer_count	tint
## 181	owner_institution_code	string
## 182	phylum	string
## 183	phylum_id	string
## 184	point-0.0001	string
## 185	point-0.001	string
## 186	point-0.01	string

## 187	point-0.02	string
## 188	point-0.1	string
## 189	point-1	string
## 190	preparations	string
## 191	previous_identifications	string
## 192	provenance	string
## 193	rank	string
## 194	rank_id	tint
## 195	raw_associated_references	string
## 196	raw_basis_of_record	string
## 197	raw_class	string
## 198	raw_common_name	string
## 199	raw_continent	string
## 200	raw_coordinate_precision	string
## 201	raw_coordinate_uncertainty	string
## 202	raw_country	string
## 203	raw_datum	string
## 204	raw_day	string
## 205	raw_establishment_means	string
## 206	raw_family	string
## 207	raw_genus	string
## 208	raw_georeference_remarks	string
## 209	raw_georeferenced_by	string
## 210	raw_georeferenced_date	string
## 211	raw_habitat	string
## 212	raw_identification_qualifier	string
## 213	raw_identified_date	string
## 214	raw_information_withheld	string
## 215	raw_institution_id	string
## 216	raw_kingdom	string
## 217	raw_latitude	string
## 218	raw_license	string
## 219	raw_life_stage	string
## 220	raw_locality	string
## 221	raw_longitude	string
## 222	raw_max_depth	string
## 223	raw_max_elevation	string
## 224	raw_min_depth	string
## 225	raw_min_elevation	string
## 226	raw_modified_date	string
## 227	raw_month	string
## 228	raw_name	textgen
## 229	raw_nomenclatural_code	string
## 230	raw_occurrence_date	string
## 231	raw_occurrence_status	string
## 232	raw_occurrence_year	string
## 233	raw_order	string
## 234	raw_phylum	string
## 235	raw_rank	string
## 236	raw_sampling_protocol	string
## 237	raw_sex	string
## 238	raw_state	string
## 239	raw_taxon_name	string
## 240	raw_type_status	string

```

## 241         raw_verbatim_depth    string
## 242         raw_verbatim_elevation string
## 243         record_number          string
## 244         rightsholder            string
## 245         sampling_effort         string
## 246         scientific_name_authorship string
## 247         scientific_name_id      string
## 248         sensitive               string
## 249         species                 string
## 250         species_group           string
## 251         species_guid            string
## 252         species_subgroup        string
## 253         specific_epithet        string
## 254         start_day_of_year       string
## 255         state                   string
## 256         subgenus                string
## 257         subspecies              string
## 258         subspecies_guid         string
## 259         subspecies_id           string
## 260         subspecies_name         string
## 261         suitable_modelling       string
## 262         system_assertions        string
## 263         taxon_concept_lsid       string
## 264         taxon_id                string
## 265         taxon_name              string
## 266         taxon_remarks           string
## 267         taxonomic_issue          string
## 268         taxonomic_kosher        string
## 269         type_status              string
## 270         verbatim_coordinate_system string
## 271         verbatim_coordinates     string
## 272         verbatim_event_date      string
## 273         verbatim_latitude        string
## 274         verbatim_locality        string
## 275         verbatim_longitude       string
## 276         verbatim_srs            string
## 277         verbatim_taxon_rank      string
## 278         water_body              string
## 279         year                     int
## 280

```

```

xf <- SBDI4R::occurrences(taxon = "order:Odonata",
                           fq = fq_str,
                           wkt = wkt,
                           extra = "collector",
                           email = "sbdi4r-test@biodiversitydata.se",
                           download_reason_id = 10)

```

```
xf$meta
```

```

##      UID                                     Name
## 1 dp0                                     IPT GBIF Sweden
## 2 co3 Artportalen - The Swedish Species Observation System
## 3 dr5      Artportalen (Swedish Species Observation System)
## 4 in3      The Swedish University of Agricultural Sciences

```

```

##                               DOI
## 1
## 2
## 3 doi.org/10.15468/kllkyl
## 4
##
## 1                               Records provided by IPT GBIF Sweden, a
## 2                               Records provided by Artportalen - The Swedish Species Observation System, a
## 3 Artportalen (Swedish Species Observation System). ArtDatabanken. Dataset/Occurrence. http://www.gb
## 4                               Records provided by The Swedish University of Agricultural Sciences, a
##
## 1
## 2
## 3 Public Domain (CC0) To the extent possible under law, the publisher has waived all rights to these
## 4
##                               More.Information
## 1 For more information: https://collections.biodiversitydata.se/public/show/dp0
## 2 For more information: https://collections.biodiversitydata.se/public/show/co3
## 3 For more information: https://collections.biodiversitydata.se/public/show/dr5
## 4 For more information: https://collections.biodiversitydata.se/public/show/in3
##   Data.generalisations Information.withheld Download.limit
## 1           NA           NA           NA
## 2           NA           NA           NA
## 3           NA           NA           NA
## 4           NA           NA           NA
##   Number.of.Records.in.Download
## 1           27290
## 2           26744
## 3           31779
## 4           26744

```

but before we can use the observation records we need to know how the observation effort has varied over time and in space. For this we define field visits i.e. occasions at which an observer has sampled observations – if we have information on observer id, location id and date we can aggregate data into “field visits”. We do this using BIRDS, and 25km grid:

```
library(BIRDS)
```

```
## Warning: package 'BIRDS' was built under R version 4.0.5
```

```
##
```

```
## Attaching package: 'BIRDS'
```

```
## The following object is masked _by_ '.GlobalEnv':
```

```
##
```

```
##      gotaland
```

```
OB <- organiseBirds(xf$data, sppCol = "species" ,
                    taxonRankCol = "rank", taxonRank = "species",
                    idCols = c("locality", "collector"),
                    timeCols = c("year", "month", "day"),
                    xyCols = c("longitude", "latitude") )
```

```
## 252 observations did not match with the specified taxon rank and were removed.
```

```
gotaland_grid25 <- raster::intersect(gotaland, Sweden_Grid_25km_Wgs84)

# gotaland_grid25 <- gIntersection(spTransform(Sweden_Grid_25km_Wgs84,
#                                             CRSobj = CRS(sf::st_crs(4326)$wkt)),
#                                gotaland,
#                                byid = TRUE)

SB <- summariseBirds(OB, grid = gotaland_grid25, spillOver = "unique")
```

```
## 1664 observations did not overlap with the grid and will be discarded.
```

```
## 0.009 % of the visits spill over neighbouring grid cells.
```

```
maxC <- max(SB$spatial@data$nObs, na.rm = TRUE)
palBW <- leaflet::colorNumeric(c("white", "navyblue"),
                              c(0, maxC),
                              na.color = "transparent")

oldpar <- par()
par(mar = c(4,0,4,0), mfrow=c(1,3))
plot(SB$spatial, col=palBW(SB$spatial@data$nObs),
     border = "grey", main="All years") ## with palette
legend("topleft", inset = c(0,0.05),
      legend = round(seq(0, maxC, length.out = 5)),
      col = palBW(seq(0, maxC, length.out = 5)),
      title = "Number of \nobservations", pch = 15, bty="n")

## or export other combinations, e.g. one map per observed year
yearlySp <- exportBirds(SB,
                       dimension = "spatial",
                       timeRes = "yearly",
                       variable = "nObs",
                       method = "sum")

maxC <- max(yearlySp@data$'2005', na.rm = TRUE)
palBW <- leaflet::colorNumeric(c("white", "navyblue"),
                              c(0, maxC),
                              na.color = "transparent")

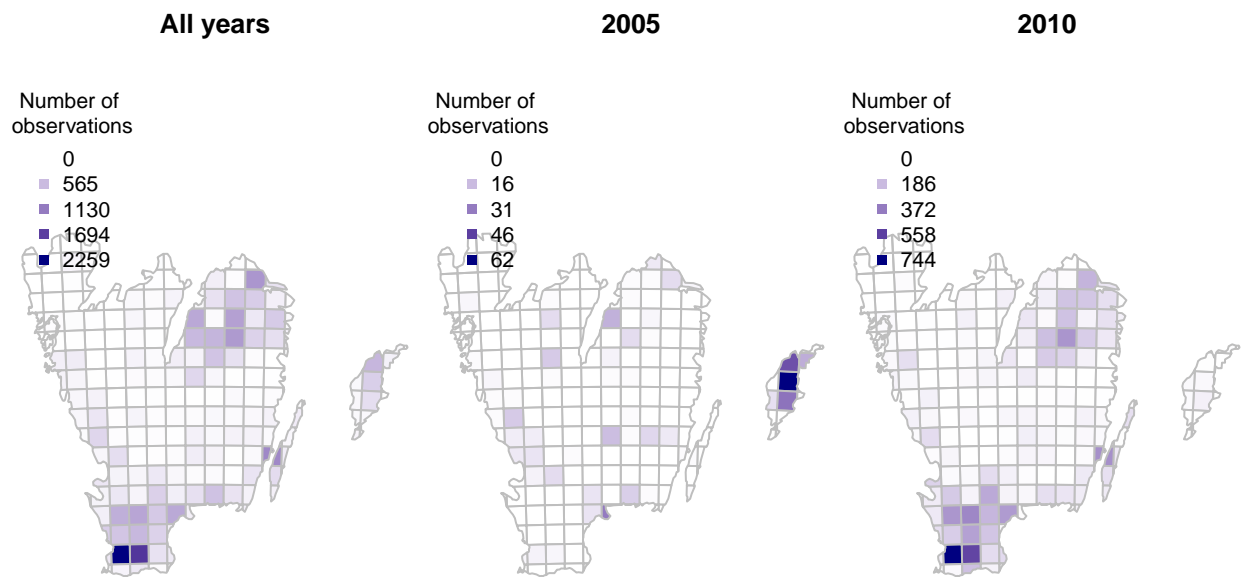
plot(yearlySp["2005"], col=palBW(yearlySp@data$'2005'),
     border = "grey", main="2005")
legend("topleft", inset = c(0,0.05),
      legend = round(seq(0, maxC, length.out = 5)),
      col = palBW(seq(0, maxC, length.out = 5)),
      border = "grey",
      title = "Number of \nobservations", pch = 15, bty="n")

maxC <- max(yearlySp@data$'2010', na.rm = TRUE)
palBW <- leaflet::colorNumeric(c("white", "navyblue"),
                              c(0, maxC),
                              na.color = "transparent")
```

```

plot(yearlySp["2010"], col=palBW(yearlySp@data$'2010'),
     border = "grey",main="2010")
legend("topleft", inset = c(0,0.05),
     legend = round(seq(0, maxC, length.out = 5)),
     col = palBW(seq(0, maxC, length.out = 5)),
     border = "grey",
     title = "Number of \nobservations", pch = 15, bty="n")

```



```
suppressWarnings(par(oldpar))
```

```
library(sf)
```

```
## Warning: package 'sf' was built under R version 4.0.5
```

```
## Linking to GEOS 3.9.0, GDAL 3.2.1, PROJ 7.2.1
```

```
library(cowplot)
```

```
## Warning: package 'cowplot' was built under R version 4.0.3
```

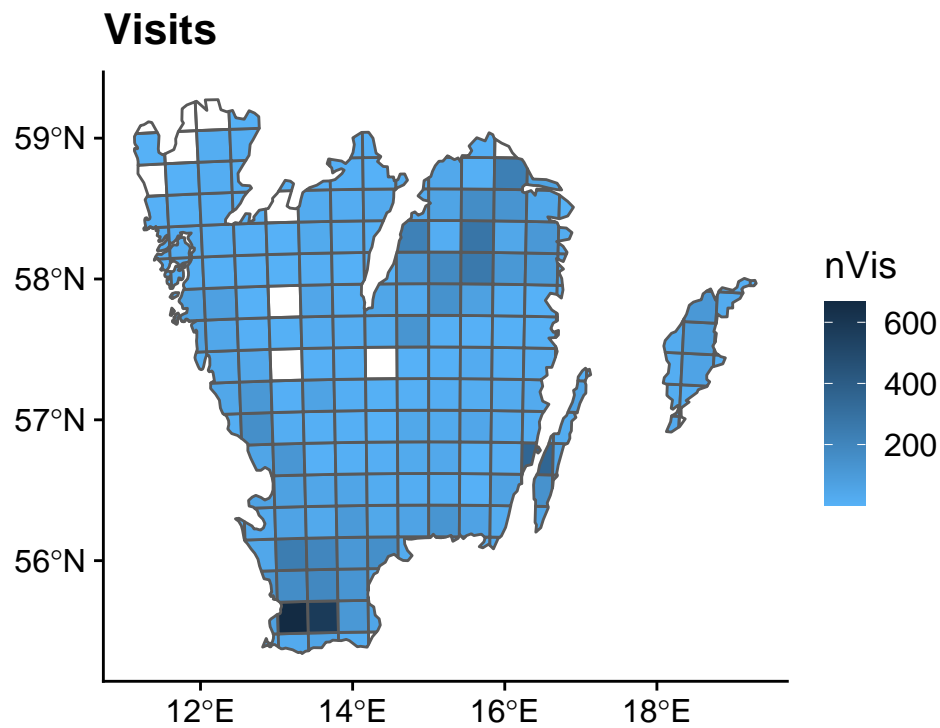
```
library(ggplot2)
library(colorRamps)
```

```
## Warning: package 'colorRamps' was built under R version 4.0.3
```

```
spatial_sf <- st_as_sf(SB$spatial)

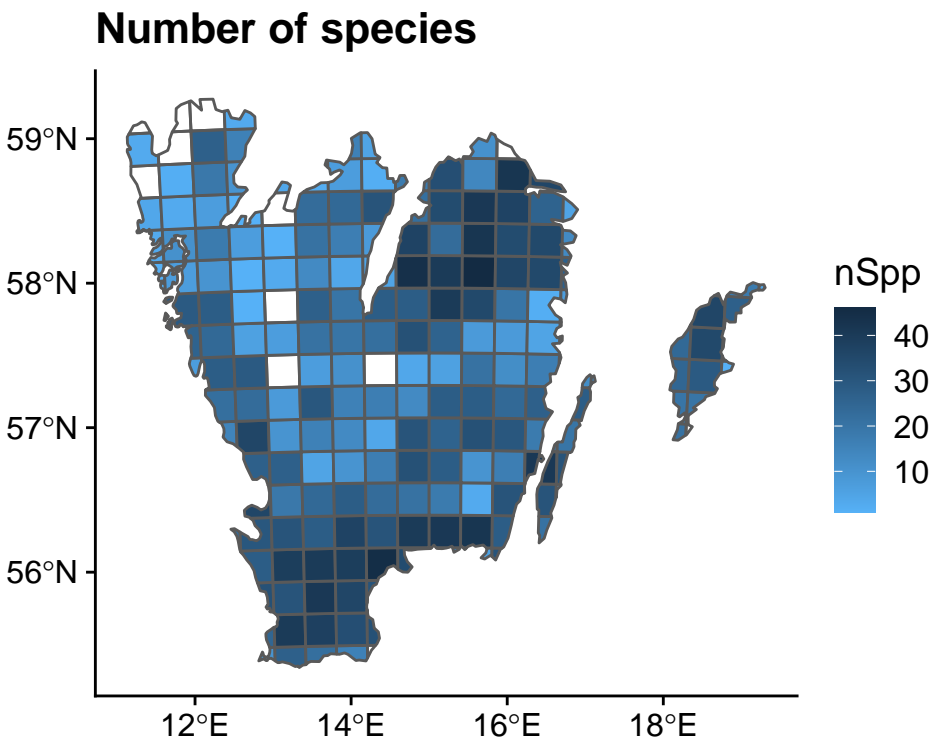
obs <- ggplot(data = spatial_sf, aes( fill = nVis))+
  geom_sf()+
  ggtitle("Visits")+
  scale_fill_gradient(low = "#56B1F7",
                     high = "#132B43",
                     na.value = NA) +
  theme_cowplot()
```

obs



```
spp <- ggplot(data = spatial_sf ,aes( fill = nSpp))+
  geom_sf()+
  ggtitle("Number of species")+
  scale_fill_gradient(low = "#56B1F7",
                     high = "#132B43",
                     na.value = NA) +
  theme_cowplot()
```

spp



How has observation effort (frequency of visits) varied over time and space? – 1) show maps as in Example 7 (all years, year 2000, 2002, 2004, 2006, 2008, 2010), 2) make also a time line plot with no. visits against years, no. of gridcells with visits against years.

we see that SB contains an element called `SB$temporal` that contains a daily time series with time specific rows when there is information. `xts` also supports time, but dating below day resolution is not yet implemented in the BIRDS package.

```
sb.xts <- SB$temporal
head(sb.xts)
```

```
##           nObs nVis nSpp
## 2000-03-24     1     1     1
## 2000-04-05     4     3     3
## 2000-04-06    11     6     3
## 2000-04-10     1     1     1
## 2000-04-12     3     3     1
## 2000-04-13     8     5     2
```

```
dim(sb.xts)
```

```
## [1] 1118    3
```

Sub-setting is convenient in `xts` as you can do it with its dates and with a `/` for a range of dates.

```
sb.xts["2010-09"] #a specific month
```

##		nObs	nVis	nSpp
##	2010-09-01	38	15	14
##	2010-09-02	26	12	12
##	2010-09-03	20	9	10
##	2010-09-04	63	19	18
##	2010-09-05	71	25	12
##	2010-09-06	16	4	9
##	2010-09-07	9	7	5
##	2010-09-08	13	6	8
##	2010-09-09	32	12	14
##	2010-09-10	1	1	1
##	2010-09-11	15	8	8
##	2010-09-12	15	7	8
##	2010-09-13	14	5	9
##	2010-09-14	1	1	1
##	2010-09-15	3	3	2
##	2010-09-17	3	2	3
##	2010-09-18	9	5	5
##	2010-09-19	12	7	5
##	2010-09-21	3	2	3
##	2010-09-22	4	4	2
##	2010-09-23	3	3	2
##	2010-09-24	10	5	5
##	2010-09-25	6	3	6
##	2010-09-26	7	6	2
##	2010-09-28	2	2	2
##	2010-09-29	5	3	4
##	2010-09-30	2	2	2

```
sb.xts["2010-09-07"] #a specific day
```

##		nObs	nVis	nSpp
##	2010-09-07	9	7	5

```
sb.xts["2007-01-01/2007-05-01"] #for a period
```

##		nObs	nVis	nSpp
##	2007-03-05	1	1	1
##	2007-03-14	1	1	1
##	2007-03-20	4	4	4
##	2007-04-02	7	4	4
##	2007-04-11	14	7	3
##	2007-04-12	8	6	4
##	2007-04-13	1	1	1
##	2007-04-15	1	1	1
##	2007-04-17	6	4	3
##	2007-04-18	1	1	1
##	2007-04-21	1	1	1
##	2007-04-23	1	1	1

```
## 2007-04-27 11 6 4
## 2007-04-28 4 4 3
## 2007-04-30 2 2 2
```

The package `xts` has several tools for converting to different periods. Here we will use `to.monthly`. This provides, the first, min, max, and last of the data. We can plot the daily maximum number of observations. The plot command with an `xts` object provides a TON of features. This makes it fairly easy to customize your plots. Read more in `?plot.xts`.

```
library(xts)
obs.m <- to.monthly(sb.xts$nObs)
obs.m["2007-04"]
```

```
##          sb.xts$nObs.Open sb.xts$nObs.High sb.xts$nObs.Low sb.xts$nObs.Close
## Apr 2007                7              14                1                2
```

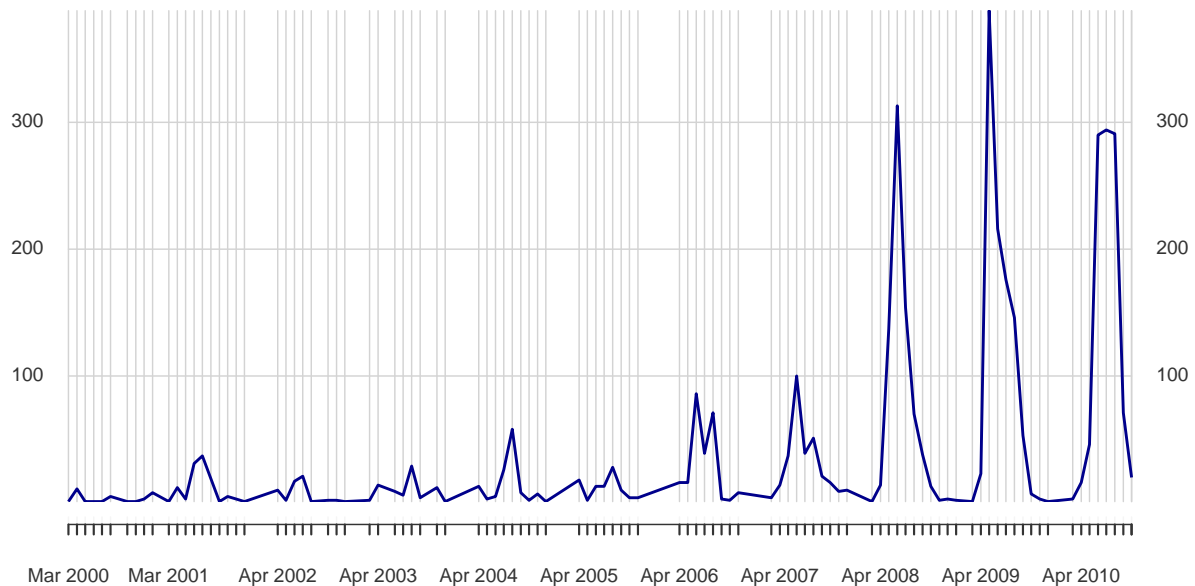
```
sb.xts["2007-04"]
```

```
##          nObs nVis nSpp
## 2007-04-02   7    4    4
## 2007-04-11  14    7    3
## 2007-04-12   8    6    4
## 2007-04-13   1    1    1
## 2007-04-15   1    1    1
## 2007-04-17   6    4    3
## 2007-04-18   1    1    1
## 2007-04-21   1    1    1
## 2007-04-23   1    1    1
## 2007-04-27  11    6    4
## 2007-04-28   4    4    3
## 2007-04-30   2    2    2
```

```
plot(obs.m["2000/2010",2], col = "darkblue", grid.ticks.on = "month",
     major.ticks = "month", grid.col = "lightgrey",
     main = "Maximum number of daily observations per month")
```

Maximum number of daily observations per month

Mar 2000 / Oct 2010



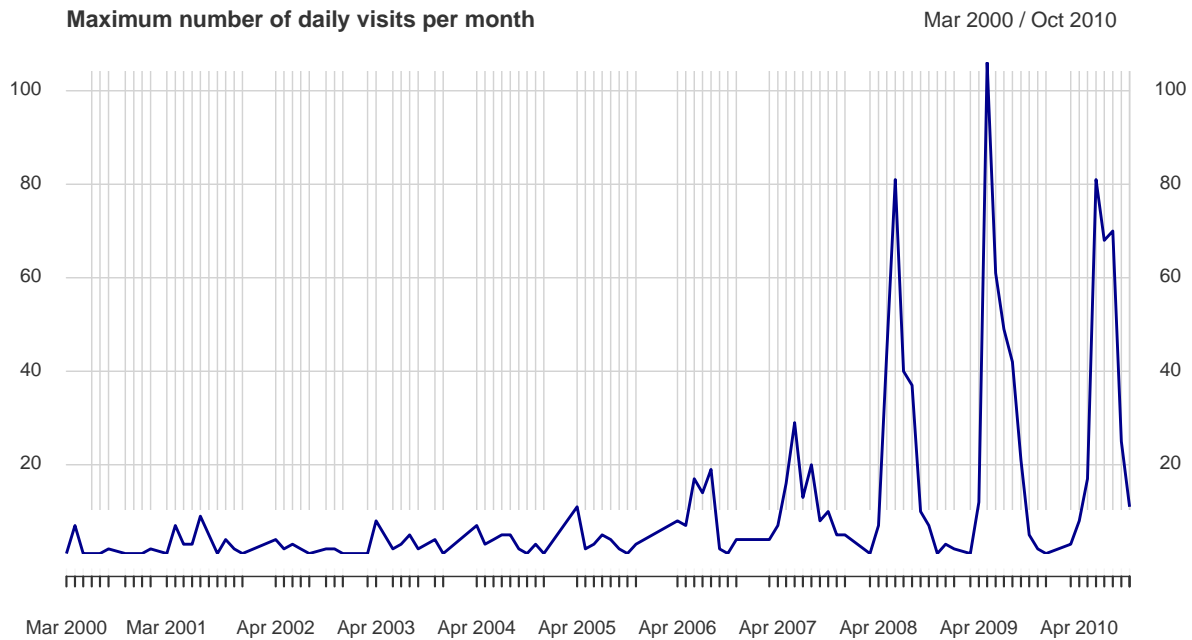
```
vis.m <- to.monthly(sb.xts$nVis)
vis.m["2007-04"]
```

```
##          sb.xts$nVis.Open sb.xts$nVis.High sb.xts$nVis.Low sb.xts$nVis.Close
## Apr 2007                4                7                1                2
```

```
sb.xts["2007-04"]
```

```
##          nObs nVis nSpp
## 2007-04-02    7    4    4
## 2007-04-11   14    7    3
## 2007-04-12    8    6    4
## 2007-04-13    1    1    1
## 2007-04-15    1    1    1
## 2007-04-17    6    4    3
## 2007-04-18    1    1    1
## 2007-04-21    1    1    1
## 2007-04-23    1    1    1
## 2007-04-27   11    6    4
## 2007-04-28    4    4    3
## 2007-04-30    2    2    2
```

```
plot(vis.m["2000/2010",2], col = "darkblue", grid.ticks.on = "month",
     major.ticks = "month", grid.col = "lightgrey",
     main = "Maximum number of daily visits per month")
```



We can now look at some particular species and ask whether this has changed in occurrence over time: Plot no. records of species x and no. visits all species over years (we simply explore by comparing records for a species with no visits, can assume that species has increased of stronger positive trend than for no. visits)

Plot no. gridcells with visits for species x and no. gridcells with visits for all species over years (we simply explore by comparing records for a species with no visits, can assume that species has increased of stronger positive trend than for no. visits) (species x: Tvåfläckad trollslända *Epitheca bimaculata*)