### **Brief Reviews**

### Brain Cholesterol: Long Secret Life Behind a Barrier

Ingemar Björkhem, Steve Meaney

Abstract—Although an immense knowledge has accumulated concerning regulation of cholesterol homeostasis in the body, this does not include the brain, where details are just emerging. Approximately 25% of the total amount of the cholesterol present in humans is localized to this organ, most of it present in myelin. Almost all brain cholesterol is a product of local synthesis, with the blood—brain barrier efficiently protecting it from exchange with lipoprotein cholesterol in the circulation. Thus, there is a highly efficient apolipoprotein-dependent recycling of cholesterol in the brain, with minimal losses to the circulation. Under steady-state conditions, most of the de novo synthesis of cholesterol in the brain appears to be balanced by excretion of the cytochrome P-450—generated oxysterol 24S-hydroxycholesterol. This oxysterol is capable of escaping the recycling mechanism and traversing the blood—brain barrier. Cholesterol levels and cholesterol turnover are affected in neurodegenerating disorders, and the capacity for cholesterol transport and recycling in the brain seems to be of importance for the development of such diseases. The possibility has been discussed that administration of inhibitors of cholesterol synthesis may reduce the prevalence of Alzheimer disease. No firm conclusions can, however, be drawn from the studies presented thus far. In the present review, the most recent advances in our understanding of cholesterol turnover in the brain is discussed. (Arterioscler Thromb Vasc Biol. 2004;24:806-815.)

Key Words: brain cholesterol ■ blood-brain barrier ■ cholesterol 24S-hydroxylase ■ Alzheimer disease ■ statins

Highly sophisticated regulatory systems have evolved for the maintenance of cholesterol homeostasis in the body. There is a distinct difference between the situation in the central nervous system and that in most other extrahepatic tissues. Outside the brain, the cellular needs for cholesterol is covered by de novo synthesis and by cellular uptake of lipoprotein cholesterol from the circulation. In the brain, the blood—brain barrier effectively prevents uptake from the circulation, and de novo synthesis is responsible for practically all cholesterol present in this organ. The independence of the isolated pool of cholesterol in the brain is likely to reflect a high need for constancy in the cholesterol content of membrane and myelin, a constancy that would be difficult to keep if brain cholesterol had been exchangeable with lipoprotein cholesterol.

The importance of cholesterol in the nervous system was recognized as early as 1834, when Couerbe's observations lead him to regard cholesterol as *un element principal* of the nervous system. Despite concerted efforts in the interim, it is only during the past few decades that the brain has begun to surrender the secrets of the behavior of one of its most abundant lipids.

In the present brief review, we summarize the basal characteristics of brain cholesterol and some recent findings with respect to homeostasis of this compound in the brain. Emphasis is put on the newly described relation between

cholesterol homeostasis and development of Alzheimer disease, and the possible effects of statins in this process. The role of cholesterol in connection with embryological development is not discussed. It should be pointed out that there are several recent excellent reviews discussing different aspects of this field.<sup>2–4</sup>

### **Cholesterol in the Central Nervous System**

In the central nervous system (CNS), essentially all (>99.5%) cholesterol is unesterified, and the majority of cholesterol present in the CNS is believed to reside in 2 different pools: one represented by the myelin sheaths (ie, oligodendroglia) and the other by the plasma membranes of astrocytes and neurons.3 A noteworthy characteristic of myelin is that it contains ≈70% lipid and 30% protein (when related to its dry weight), which is approximately opposite the situation found in most other cell membranes. Although there are no authentic myelin-specific marker lipids, myelin invariably contains cholesterol, phospholipids, and glycosphingolipids (particularly galactocerebroside) in molar ratios of  $\approx$ 4:4:2. It has been estimated that up to 70% of the brain cholesterol is associated with myelin. Because up to half of the white matter may be composed of myelin, it is unsurprising that the brain is the most cholesterol-rich organ in the body. The concentration of cholesterol in the brain, and particularly in myelin, is consistent with an essential function related to its membrane properties.

Received December 3, 2003; revision accepted January 22, 2004.

From the Division of Clinical Chemistry, Karolinska Institute, Huddinge University Hospital, Stockholm, Sweden.

© 2004 American Heart Association, Inc.

Consulting Editor for this article was Alan M. Fogelman, MD, Professor of Medicine and Executive Chair, Departments of Medicine and Cardiology, UCLA School of Medicine, Los Angeles, CA.

Correspondence to Dr Ingemar Björkhem, Division of Clinical Chemistry, Huddinge University Hospital, S-141 86, Sweden. E-mail ingemar.bjorkhem@hs.se

Neurons are composed of a cell body and an axon, and their primary function is the generation and propagation of electrical impulses. The axon is essentially an extended electrical conductor that continually bifurcates and diminishes until it ends at the nerve terminals, the site of neurotransmitter release. Rapid transmission of impulses along the length of the axon is facilitated by the presence of the myelin sheath, which consists of sections of oligodendrocyte plasma membrane repeatedly wrapped around the axon, with the extrusion of almost all of the cytoplasm. Because an individual axon may be ensheathed by myelin from several oligodendrocytes, periodic gaps are present in the sheath. These are termed the nodes of Ranvier and are the sites of propagation of the action potential. Myelin can thus be regarded as a discontinuous insulation that enables the saltatory conduction of the action potential. A detailed description of the neuronal anatomy and function exists elsewhere.3 Cholesterol enrichment of myelin thus leads to reduced permeability to ions (ie, high resistance and low capacitance).3 The net effect is that the current tends to move down the axon rather than diffusing across the membrane.

Cholesterol synthesis in the developing CNS is relatively high, but it declines to a very low level in the adult state. This can be explained by an efficient recycling of brain cholesterol. As a consequence, brain cholesterol has an extremely long half-life; in the adult human brain, the half-life of the bulk of cholesterol has been estimated to be at least 5 years.5 It was recently pointed out that the long half-life of cholesterol in the brain is remarkable in light of the high metabolic rate of this organ.<sup>6</sup> In humans, the organ-specific metabolic rate of the brain is ≈9-fold greater than the average metabolic rate of the individual.

Despite the efficiency of cholesterol reutilization, there appears to be a need for some excretion from the brain to maintain the steady state. Enzymatic conversion of cholesterol into the metabolite 24S-hydroxycholesterol, which is able to traverse the blood-brain barrier, has recently been demonstrated to be the most important excretion mechanism. The presence of a brain-specific oxysterol in the circulation has opened new possibilities to study changes in cholesterol homeostasis in the brain in different physiological, pharmacological, and pathological conditions.

#### **Cholesterol and Membranes**

The appearance of sterols in biological membranes is believed to be a defining step in membrane evolution, and sterols are a major means by which eukaryotic cells can refine the properties of lipid bilayers. By reducing average fluidity and protein free volume, sterols can influence membrane permeability to ions.7

The minimum structural requirements for a compound to be membrane-active include a flat (rigid), fused-ring system, a small polar function at the C-3 position (eg, a hydroxyl group), and a cholesterol-like tail. It has been stated, "Nature has designed the cholesterol side-chain for optimal interaction with phospholipids."8 The structural features contribute to the relatively small area, ie, <40Å per molecule of cholesterol at an air/water interface. Even minor changes in sterol structure lead to a pronounced effect on the membrane activity of the sterol, with deleterious consequences for the organism. This is apparent from the severity of the phenotypes associated with post-squalene defects in cholesterol biosynthesis.

Björkhem and Meaney

In these syndromes, cholesterol is replaced to a varying degree with its precursor, leading to a number of birth defects, with concomitant functional disturbances of the central and peripheral nervous system.9 For example, in the brain of patients with the Smith-Lemli-Opitz syndrome, a potentially fatal condition caused by a defect in the 7-dehydrocholesterol-7-reductase, most of the cholesterol may be replaced with 7-dehydrocholesterol.<sup>9,10</sup> Unsurprisingly, a common feature of these conditions is mental retardation, most likely caused by a deficiency in myelin function. Mouse models have been generated for many of these conditions, and deletion of these genes is associated with a high prenatal or perinatal lethality, except in the case of a recently developed mouse model of desmosterolosis. 11 Disruption of the gene coding for  $3\beta$ hydroxy- $\Delta$ 5-steroid-24-reductase in these mice leads to the replacement of >95% of their cell cholesterol with desmosterol, with resultant neurological and behavioral deficiencies. It is evident that although the introduction of an additional double bond in the cholesterol nucleus or side-chain may be consistent with life, it has serious consequences for the

Although cholesterol therapy is routinely used in these conditions, the unexchangeability of brain and circulating cholesterol precludes its ability to improve the condition within the CNS.

### **Proteins**

In the mammalian CNS, proteolipid protein and myelin basic protein account for ≈80% of total myelin proteins. The minor components include cyclic nucleotide phosphodiesterase, myelin-associated glycoprotein, myelin-oligodendrocyte protein, and myelin-associated oligodendrocyte basic protein. Their presence in myelin implies important functional roles for these proteins, an assumption that is supported by a number of recent studies in transgenic animals.3

The lipid and cholesterol composition of membranes are known to affect the activity of some transmembrane proteins. It has been shown that the  $\gamma$ -aminobutyric acid transporter from rat brain has a specific requirement for cholesterol and that some channel and transport proteins require cholesterol for activity in reconstitution experiments.3 Furthermore, the ability of cholesterol to stiffen membranes appears to have a beneficial effect on various aspects of neuronal function, as judged by in vitro experiments.

#### **Isolation of Brain Cholesterol**

Although there has been some controversy in the literature concerning the exchangeability of brain cholesterol, the current consensus is that cholesterol in the brain is insulated from changes in circulating cholesterol. The earliest observations that brain cholesterol was metabolically distinct from that present in the circulation were made in the early 1940s. Waelsch et al demonstrated that after administration of D<sub>2</sub>O to adult rats, there was essentially no incorporation of label into unsaponifiable lipids of the brain. 12 At approximately the same time, Bloch (in an unrelated study on the metabolism of deuterium-labeled cholesterol in the dog) noted, "... the complete absence of deuterio cholesterol in brain and spinal cord." These studies represent the earliest reports of the brains "insulation" from the circulating cholesterol, and they have been confirmed and extended by several investigators. Attacks using a variety of different experimental animals have thus consistently indicated that >95% of the cholesterol content in the brain can be accounted for by de novo synthesis, and there is little if any exchange of plasma and brain cholesterol.

Studies by Chobanian et al in the 1960s clearly demonstrated that the situation is similar in humans. <sup>15</sup> After intravenous injection of a bolus dose of <sup>14</sup>C-labeled cholesterol to terminally ill patients, they observed that the brain contained essentially no label, even after postinjection intervals of up to 7 months. Furthermore, as judged from the extent of incorporation of deuterium in cerebrospinal fluid (CSF) cholesterol after the intravenous infusion of deuterium-labeled cholesterol in a healthy volunteer, CSF cholesterol was found to be predominantly derived from the brain. <sup>16</sup> Finally, this isolation of the brain appears to be present from an early age; after administration of <sup>14</sup>C-cholesterol to pregnant women, there was essentially no label in fetal brain tissue, whereas significant amounts were present in liver and other viscera. <sup>17</sup>

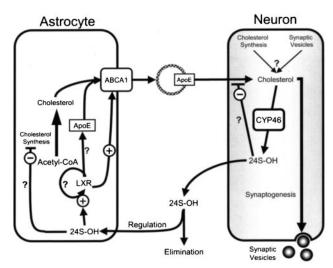
The blood-brain barrier prevents diffusion of large molecules at the level of tight junctional attachments between adjacent capillary endothelial cells. Surprisingly, it has been shown that brain endothelial cells have the potential to take-up low-density lipoprotein (LDL) cholesterol through luminal LDL receptors and translocate this LDL across the cell. 9.20 However, as judged from this isotope experiments, a receptor-mediated uptake of cholesterol from the circulation is of little or no importance under normal conditions.

### Maintenance of the Cholesterol Barrier Function

It is possible that one or more members of the ABC-transporter superfamily may be involved in the exclusion of circulating cholesterol from the brain. It was recently demonstrated that primary porcine brain capillary endothelial cells express mRNA and protein of the cholesterol transporter ABCA1.<sup>21</sup> It was also shown that along with ABCA1 expression, the oxysterol 24S-hydroxycholesterol enhanced apoA1-dependent efflux of cholesterol from cultured brain endothelial cells. Based on results of experiments with an in vitro model system, the possibility was discussed that the ABCA1 transporter and the scavenger receptor SR-B1 may be involved in an autoregulatory mechanism for "backflush" of cholesterol to the brain.

### Mechanism and Site of Synthesis of Brain Cholesterol

According to in vitro studies with cultured cells, astrocytes synthesize at least 2- to 3-fold more cholesterol than neurons and fibroblasts. Evidence has been presented that oligodendrocytes, the cells responsible for myelinization, have an even higher capacity for cholesterol synthesis than astrocytes.<sup>22</sup> Because most studies on neuronal cells have been



**Figure 1.** Interactions of astrocytes and neurons in cholesterol homeostasis according to Pfrieger et al.<sup>24,25</sup> Although neurons are capable of synthesizing cholesterol, it has been suggested that in the adult state, neurons rely on delivery of cholesterol from nearby cells such as astrocytes. This "outsourcing" may allow the neurons to focus on generation of electrical activity and dispense with energetically costly cholesterol synthesis. Cholesterol delivered to neurons in this way may support synaptogenesis and may be incorporated into synaptic vesicles. Generation of 24S-hydroxycholesterol is a strategy to remove excess cholesterol from the neuron. However, because this oxysterol is a potent activator of the LXRs, it may also stimulate efflux of cholesterol from the astrocyte, a mechanism that may involve activation of the cholesterol transporter ABCA1. 24S-Hydroxycholesterol is also an inhibitor of cholesterol synthesis.

made on embryonal cells, there is a possibility that older neurons may lose their capacity to synthesize cholesterol.<sup>23</sup> Pfrieger has recently discussed the possibility that neurons may switch from cholesterol production during embryonic stage to "outsourcing" after birth. He points out that synthesis of the sterol molecule requires a diverse array of enzymes that are distributed in different subcellular organelles and consumes large amounts of energy.<sup>24</sup> Therefore, neurons, which specialize in the generation of electrical activity, may reduce or even abandon cholesterol synthesis. Experimental evidence for a cholesterol shuttle from astrocytes to neurons in an in vitro system has been presented,<sup>25</sup> and cultured neurons from mammalian CNS appear to require glia-derived cholesterol to form numerous and efficient synapses.<sup>26</sup> This is depicted schematically in Figure 1.

There are 2 pathways for the formation of cholesterol in mammals. The major one in most mammalian cells includes 7-dehydrocholesterol as an intermediate, while the alternative route uses 7-dehydrodesmosterol as an intermediate. Oligodendrocytes in CNS appear to preferentially use the 7-dehydrodesmosterol pathway, and disruption of the gene coding for the delta 24-reductase causes accumulation of desmosterol without any accumulation of 7-dehydrodesmosterol. In similarity with most other cells in the body, cultured glial cells have HMG CoA reductase as the major rate-limiting enzyme for their synthesis of cholesterol. Under certain experimental conditions, HMG CoA synthase appears to be rate-limiting in these cells. In similarity with other cells, HMG CoA reductase activity and sterol synthesis are inversely related to the level of

LDL cholesterol in the incubation medium, indicating that glial cells express LDL receptors.<sup>28,29</sup>

In the developing CNS, as well as in the injured CNS, the need for cholesterol synthesis is much higher than in the adult uninjured state. In humans, myelination begins during the second trimester and continues through the second year of life. It is notable that the myelination is preceded by an increased esterification of cholesterol, possibly as a mechanism for accumulation of the required very high amounts of cholesterol. It has been estimated that an active oligodendrocyte may synthesize 3-times its own weight as myelin membranes each day.<sup>30</sup>

### Transport of Cholesterol Within the Brain

Apolipoprotein E (apoE) is one of the major apolipoproteins in plasma. Under normal conditions, this is regarded to be the quantitatively most important transport protein for cholesterol in the brain. Within the CNS, apoE is present in spherical and discoidal particles that are the size of high-density lipoprotein. Astrocytes are believed to have the highest capacity to produce apoE;31 in culture, these cells are able to secrete apoE together with cholesterol into the culture medium.<sup>32</sup> Because apoE and cholesterol-containing lipoproteins can be found in cerebrospinal fluid,33 it is evident that there is some circulation of lipoproteins within the brain.34 In accordance with this, LDL receptor mRNA is present in different structures of the brain, including neuronal cells.35,36 An alternative to uptake by the LDL receptor could be by the multi-ligand receptor LRP (LDL receptor-related protein).<sup>37</sup> Several other lipoproteins, including apoA-I, apo D, and apo J, are known to be synthesized in the brain.<sup>2,4</sup> The brain also expresses other receptors, including the very low-density lipoprotein receptor, apoE receptor 2, and megalin.38 The relative importance of the different lipoproteins and receptors for the transport and recycling of cholesterol within CNS is unknown at present, and it is likely that there is considerable redundancy in these systems. Knocking out the genes coding for the LDL receptor or apoE does not appear to be associated with any major functional abnormalities in the brain of the mouse.<sup>39,40</sup> However, in aged apoE knockout mice, there is a lipid deposition in astrocytes in specific brain areas.<sup>41</sup> Deletion of the genes coding for the very low-density lipoprotein receptor and the apoE receptor 2 results in a highly significant phenotype, with a defect in the neuronal layering,<sup>42</sup> whereas deletion of either of the 2 genes alone has much less effect. Whether the very low-density lipoprotein receptor is involved in cholesterol transport in the brain has not been established, however.

Even if apoE does not appear to be obligatory for maintenance of cholesterol homeostasis in the brain of experimental animals, the capacity of the transport function it represents seems to be critical in connection with development of neurodegenerative disorders or recovery from brain injuries. One of the 3 human isoforms of apoE, apoE4, is a well-documented risk factor for late-onset Alzheimer disease. Alzheimer disease. Alzheimer disease associated with poor recovery after head injury. In accordance with this, it has been reported that apoE3 is more effective than apoE4 in delivering cholesterol to neurons. In co-cultures of astrocytes and neurons, apoE3-

containing lipoproteins were found to have a higher stimulatory effect on neurite outgrowth than apoE4.<sup>47</sup>

Björkhem and Meaney

In addition to apoE, ATP-binding cassette (ABC) transporters may be of importance for the shuttling of cholesterol from glial cells to neurons, particularly as the ABCA1 transporter is expressed in astrocytes (Figure 1). It was recently reported that the ABCA2 transporter is predominantly expressed in the brain and in neural tissues. 48,49 Moreover, it has also been shown that the temporal and spatial expression of ABCA2 appears to coincide with the level of myelinization. 50 The available data suggest that ABCA2 is involved in neural lipid redistribution, although direct evidence has not yet been presented. From a regulatory point of view, the promoter region of ABCA2 lacks the conserved regulatory elements for the nuclear receptor LXR.

#### **Efflux of Cholesterol From the Brain**

Despite the efficiency of the cholesterol recycling machinery in the brain, there is a continuing need for export of a small excess of cholesterol into the circulation to maintain steady state. At present, 2 such mechanisms are known. There is some excretion of apoE-bound cholesterol via the CSF.51 Based on the CSF cholesterol content and the rate of CSF renewal, it has been estimated that 1 to 2 mg cholesterol may be eliminated from the brain via the CSF each day. The precise sites and mechanism of the elimination remain to be conclusively established. The other, and quantitatively more important, mechanism involves conversion of cholesterol into 24S-hydroxycholesterol. In contrast to cholesterol, the latter oxysterol is able to traverse the blood-brain barrier. 5,52,53 The reason for the rapid transfer of side-chain oxidized oxysterols over lipophilic membranes is that the introduction of an hydroxyl group in the side chain leads to a local reordering of membrane phospholipids, such that it is energetically more favorable to expel the oxysterol.54 This is outlined schematically in Figure 2 It has been shown that the rate of transfer of side-chain oxidized oxysterols across lipophilic membranes is orders of magnitude greater than that of cholesterol.55,56

Measurements of the arteriovenous concentration difference over the human brain have shown that the flux of 24S-hydroxycholesterol from the brain into the circulation corresponds to ≈6 to 7 mg/24 hours.<sup>5,53</sup> There is a corresponding uptake of the oxysterol in the liver,5 demonstrating that the brain is the exclusive source of 24S-hydroxycholesterol in humans. In mice, there is a considerable production of 24S-hydroxycholesterol, also, in the liver.<sup>57–59</sup> The cytochrome P-450 species responsible for 24S-hydroxylation of cholesterol, CYP46, has been characterized at the molecular level.<sup>57</sup> Interestingly, the enzyme was found to be localized to neuronal cells, indicating that the excretion of the cholesterol from the brain is initiated in the neurons rather than in glial cells. However, in the brain of patients who had died with an advanced Alzheimer disease, some CYP46 immunoreactivity was detected in glial cells.60

By exposing rats to  $^{18}O_2$  in the inhalation air and measuring the incorporation of isotope in brain cholesterol and in brain 24S-hydroxycholesterol, the rate of synthesis of both these compounds can be measured under in vivo conditions.<sup>52</sup> The

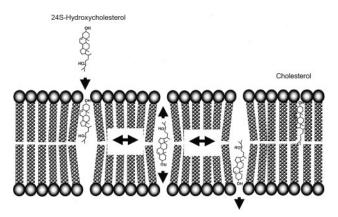


Figure 2. Physiochemical basis for the rapid transfer of sidechain oxidized oxysterols across lipophilic membranes.  $^{55,56}$  In the normal state, cholesterol is oriented approximately perpendicular to the plane of the membrane such that the  $3\beta$ -hydroxyl group can interact with the polar phospholipid head group. Introduction of a hydroxyl group in the side chain of cholesterol leads to local reordering of the phospholipid acyl chains and an increase in the membrane surface area. The oxysterol can rapidly shuttle between the leaflets of the membrane and may easily be extracted by an acceptor (eg, lipoproteins). This property may explain the passage of 24S-hydroxycholesterol across the blood–brain barrier while cholesterol is retained. The direction of the net flux across the membrane is most probably dependent on the concentration gradient of the oxysterol.

rate of conversion of cholesterol into 24S-hydroxycholesterol was found to be  $\approx$  two-thirds that of cholesterol synthesis. Very recently, this was confirmed in mice with a disruption of the gene coding for cyp46.<sup>6,59</sup> These mice had a reduction of the sterol export from the brain by  $\approx$ 64%. The results of both studies suggest that additional mechanisms are likely to exist for removal of cholesterol from the brain. Interestingly, the absence of the CYP46-mediated mechanism for sterol efflux was associated with 40% reduction in cholesterol synthesis,<sup>59</sup> suggesting a close relation between synthesis and metabolism of cholesterol in the brain.

In view of the fact that almost all 24S-hydroxycholesterol present in human circulation is of cerebral origin, it may be used as a surrogate marker for brain cholesterol homeostasis. It has been shown that the levels of this oxysterol in the circulation reflects the balance between cerebral production and hepatic metabolism.61 The cerebral production appears to be relatively constant, although the hepatic metabolism varies with age. As a consequence, there is an inverse relationship between plasma levels of 24S-hydroxycholesterol and body surface. Levels of 24S-hydroxycholesterol are therefore high during the first decade of life but relatively constant after the second decade. There is a tendency toward slightly increased levels of the oxysterol in the circulation after the sixth decade. It has also been shown that different neurological diseases affect the levels of 24S-hydroxycholesterol in the circulation.62-64 It should be emphasized that 24S-hydroxycholesterol is transported in the circulation by the same lipoproteins as cholesterol,65 and changes in lipoprotein content may therefore affect the levels of 24S-hydroxycholesterol.<sup>61</sup> This relationship must always be taken into account when using 24S-hydroxycholesterol as a marker for brain cholesterol homeostasis. Under conditions with changes in plasma cholesterol levels, the ratio between 24S-hydroxycholesterol and cholesterol may therefore better-reflect brain cholesterol homeostasis than the absolute levels.

## Possible Roles of Nuclear Receptors and Their Target Genes in the Brain

The liver-X receptors (LXR) LXR $\alpha$  and LXR $\beta$  are important proteins in the regulation of cholesterol homeostasis in the body. <sup>66,67</sup> The LXRs are expressed in most tissues and organs and are activated by a number of oxysterols, including 24S-hydroxycholesterol. <sup>68,69</sup>

The LXRs regulate their target genes in the form of heterodimers with the 9-cis-retinoic acid receptors, which bind to LXR-response elements in the regulatory regions. The target genes include the ATP-binding cassette transporters ABCA1 and ABCG1, which mediate efflux of phospholipids and cholesterol from a number of cells, including astrocytes.70 It was recently reported that LXR agonists have marked effects on gene expression in murine brain tissues in vitro and in vivo.70 In primary astrocyte cultures, LXR agonists regulated several established LXR target genes, including ABCA1, and led to enhanced cholesterol efflux. In contrast, a much lower effect on gene expression or cholesterol efflux was observed in primary neuronal cultures. These findings are in consonance with the hypothesis of a net export of cholesterol from astrocytes to neurons that is feedbackregulated with participation of LXR $\beta$ , ABCA1, CYP46, and 24S-hydroxycholesterol.<sup>24</sup> A mechanistic model for this has been suggested by Pfrieger and is shown in Figure 1.

In the model shown in Figure 1, there is an activation of LXRβ by 24S-hydroxycholesterol. Whether 24Shydroxycholesterol and other oxysterols really are able to bind to LXR in the presence of the great excess of cholesterol present in most biological systems has been questioned.<sup>71</sup> In the circulation and in most tissues, the excess of cholesterol in relation to any oxysterol is >10 000-fold. In the brain, however, the ratio between 24S-hydroxycholesterol and cholesterol varies between 1/500 and 1/1000.53 In preliminary experiments, it was recently shown that at >500-fold excess, cholesterol can suppress the binding of 24S-hydroxycholesterol to a recombinant LXR $\alpha$  ligand-binding domain.<sup>72</sup> Thus, it appears that at least some activation of LXR by 24Shydroxycholesterol may occur in the brain, and that this may be of regulatory importance. Of interest in this connection is the fact that the expression patterns of CYP46 and LXR $\beta$  in the brain are very similar. 57,73 Moreover, in the rat, the expression of LXR $\beta$  is ontogenically regulated, suggesting that these receptors are involved in maintenance of brain sterol homeostasis even at early developmental stages.<sup>74</sup>

The importance of the nuclear receptors LXR $\alpha$  and LXR $\beta$  in the brain was recently elucidated in a mouse model in which both genes had been knocked out.<sup>75</sup> While young, such mice had normal brain histology whereas aged mice had closed lateral ventricles lined with lipid-laden cells. Other features of the aged brain were excessive lipid deposits, proliferation of astrocytes, loss of neurons, and disorganized myelin sheaths. Evidence was presented that loss of apoE could not explain the phenotype of the LXR-deficient mice. It was emphasized that the findings are consistent with a role of LXR-dependent ABC transporters in

efflux of cholesterol from astroglial cells in CNS, particularly from perivascular astrocytes.

### **Cholesterol Homeostasis in Neurodegenerative Conditions**

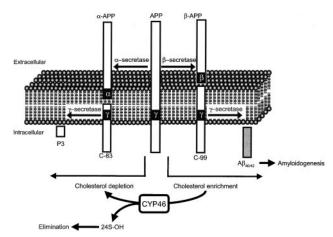
In the past decade, evidence of a possible link between cholesterol and neurodegeneration has accumulated. Some of the earliest observations of this link were the recognition of  $\epsilon 4$  isotype of apoE as an important risk factor for late-onset Alzheimer disease<sup>76</sup> and that hypercholesterolemia was associated with increased brain A $\beta$  immunoreactivity in rabbits.<sup>77</sup> Alzheimer disease and Niemann-Pick disease type C are the best-studied of the clinical conditions and represent prototypic models of the role of cholesterol homeostasis in neurodegeneration.

The initial epidemiological studies implicating apoE polymorphisms with the risk of Alzheimer disease have been repeatedly extended and confirmed. In a recent meta-analysis,78 it was calculated that the odds ratio for late-onset Alzheimer disease associated with the carriers of the  $\epsilon 4/\epsilon 4$ compared with  $\epsilon 3/\epsilon 3$  was 11.6. Moreover, the authors estimated that ≈60% of late-onset Alzheimer disease was attributable to presence of the  $\epsilon 4$  isotype. A role for apoE in cholesterol recycling during peripheral nerve regeneration has long been recognized, although its function in the CNS has remained more elusive. While apoE is undoubtedly involved in CNS cholesterol homeostasis, several nonlipid activities have also been attributed to it, including protein chaperoning<sup>79</sup> and signal transduction.<sup>80</sup> However, the physiological relevance of these latter activities remain controversial.

The second important observation was that experimentally induced hypercholesterolemia in rabbits lead to accumulation of pathogenic amyloid beta peptide ( $A\beta$ ) in brain.<sup>77</sup> As the blood–brain barrier efficiently prevents the passage of peripheral cholesterol into the brain, the mechanism behind this accumulation remains obscure. The possibility has to be considered that the effect of cholesterol is mediated by effects at the level of the cerebral microcirculation rather than at the level of glial cells or neurons.

Under in vitro conditions, cholesterol has been shown to influence a number of processes involved in the generation of the neuritic plaques (which predominantly consist of  $A\beta$  peptides of 40 or 42 residues) and neurofibrillary tangles. The generation of  $A\beta$  peptides from the amyloid precursor protein (APP) is perhaps the best-studied of these processes. APP is a transmembrane protein with a large extracellular domain and one transmembrane region. APP can undergo an initial  $\alpha$  cleavage or  $\beta$  cleavage, followed by  $\gamma$  cleavage, to yield the  $\alpha$ -APP and P3 peptides or the  $\beta$ -APP and A $\beta$  peptides, respectively. This metabolism of APP is outlined in Figure 3.

Numerous studies using immortalized and primary cell lines  $^{83-85}$  have demonstrated that increased cholesterol levels can increase the activity of the  $\beta$ -secretase pathway, leading to an accumulation of  $A\beta_{1-40}$  and  $A\beta_{1-42}$  peptides, with a resulting increase in formation of extracellular amyloid deposits. Moreover, it has recently been demonstrated that cholesterol is capable of modulating the production of mature



Björkhem and Meaney

Figure 3. Metabolism of the APP. APP is a single transmembrane protein that undergoes regulated intramembrane cleavage to give rise to a variety of products. Two major pathways have been defined for the metabolism of this protein. An initial cleavage by the  $\alpha$ -secretase leads to the formation of a soluble product ( $\alpha$ -APP), with further processing by  $\gamma$ -secretase leading to the generation of nontoxic C-terminal fragments (P3 in the figure). Because the site of  $\alpha$  cleavage is within the A $\beta$  region, processing of APP via this pathway does not generate amyloidogenic peptides. However, when an initial  $\beta$  cleavage is followed by  $\gamma$  cleavage, A $\beta$  peptides are generated. In recent years, in vitro experiments have implicated cholesterol as a modulator of APP processing, with increased cholesterol levels associated with increased formation of  $A\beta$  peptides. Conversely, reduction in cholesterol levels, either by direct cholesterol extraction from the membrane or by inhibition of synthesis (eg, via statins), appears to drive processing through the  $\alpha$ -secretase pathway. CYP46 represents the major pathway for removal of cholesterol from the brain, and induction of this enzyme is likely to have cholesterol-lowering effects. CYP46 thus represents a valid target for the modulation of brain cholesterol levels.

glycosylated APP.<sup>86</sup> Cholesterol has also been suggested to alter the conformation of  $A\beta$ , leading to the generation of "amyloid seeds," regarded to promote the formation of amyloid fibrils from  $A\beta$ . The possibility has also been discussed that  $A\beta$  itself can influence cholesterol homeostasis, leading to a decrease in intracellular free cholesterol.<sup>87</sup> Based on in vitro work, it has been suggested.<sup>88</sup> that the ratio between free and total cholesterol is a primary regulator of APP processing, and that ACAT is fundamentally important in this regard. However, because of the extremely high ratio between free and esterified cholesterol in the adult human brain, it is difficult to understand how ACAT modulates APP processing in vivo.

However, this hypothesis may be relevant to Niemann-Pick disease. This is a fatal neurovisceral cholesterol storage disease caused by mutations in the ubiquitously expressed NPC-1 or NPC-2 gene products. Although these proteins are structurally unrelated, they are both involved in the intracellular trafficking of cholesterol, and mutations lead to accumulation in the endo-lysosomal system. In the past, it was generally accepted that this accumulation was not present in the CNS. Recent studies indicate, however, that cholesterol accumulation does indeed occur in NPC neurons. Based on studies using a mouse model of the disease, it has been suggested that the turnover of cholesterol is also increased. The net effect is that neurons are unable to adequately

### Summary of Statin Effects on Biomarkers for the Development of Alzheimer Disease

Statin	Dose	N	Subject Characteristics	Age (range or mean)	Effect	Ref
Simvastatin	80 mg/d	18	Hypercholesterolemic	21–70	Decreased plasma 24S-OH/chol*	106
Pravastatin	10 mg/d	46	MMSE >25	33-83	No effect on plasma $A\beta_{1-40}$ or $A\beta_{1-42}$	107
Simvastatin	20 mg/d	19	AD patients	64-84	No effect on plasma $A\beta_{1-42}$	108
			MMSE 22±4		Decreased CSF $\alpha$ APP $_{\rm sol}$ and $\beta$ APP $_{\rm sol}$	
Lovastatin	10-60 mg/d	94	Hypercholesterolemic	29–70	Dose related decrease in serum $Aeta$	109
Simvastatin	40 or 80 mg/d	20/24	AD patients	68	No effect on CSFA $eta_{ ext{1-40}}$ or A $eta_{ ext{1-42}}$	110
			MMSE 12-26		No change in CSF 24S-0H/chol*	
Statin†	40 mg/d	31	AD patients‡	65	No change in plasma 24S-OH/chol*	111
Statin†	Variable, see reference	150	See reference	See reference	No change in 24S-0H/chol* <i>vs</i> a normocholesterolemic group	112

<sup>\*24</sup>S-OH/chol indicates 24S-hydroxycholesterol/cholesterol.

respond to increased cholesterol levels via storage or excretion, with the consequence of vastly accelerated development of neurofibrillary tangles. These are essentially identical to those observed in Alzheimer disease.

Taken together, the aforementioned studies implicate cholesterol homeostasis as a central component of neurodegenerative cascades, which can influence several aspects of disease development. Modulation of cholesterol levels thus becomes a potential target for the prevention or treatment of neurodegenerative conditions.

Although brain cholesterol levels may be altered in neurodegenerative conditions, it is impossible to make analyses on brain cholesterol content during the process of neurodegeneration. Theoretically, the recognition that almost all of the 24S-hydroxycholesterol present in the circulation originates from the brain should provide a window into this otherwise analytically inaccessible compartment. Many studies based on the measurement of this sterol (either in the circulation or in the CSF) have appeared over the past few years. Initial studies reported an increase in plasma 24S-hydroxycholesterol during the early stages of Alzheimer disease or vasculardemented patients,64 possibly reflecting ongoing demyelinization. A similar slight increase may occur in connection with active periods of multiple sclerosis.62 In the advanced Alzheimer disease, as well as in chronic stages of multiple sclerosis, however, the levels of 24S-hydroxycholesterol are reduced,61,62,91 probably as a consequence of the loss of neuronal cells containing CYP46. In accordance with this, the levels of 24S-hydroxycholesterol were decreased in brain samples from patients who had died from Alzheimer disease.92 Interestingly, there was a close correlation between cholesterol synthesis (as estimated by lathosterol levels) and degradation (ie, formation of 24S-hydroxycholesterol) in the frontal cortex of Alzheimer disease patients, but not in controls. This finding is consistent with an increased turnover of cholesterol in these subjects. Similar changes were observed in the brain of mice carrying a mutation in the APP gene.93

# Are Statins Effective in the Treatment of Neurodegenerative Disorders?

The possibility that cholesterol influences the progression or development of neurodegenerative diseases has obvious im-

plications for therapy. HMG CoA reductase inhibitors are one of the most widely prescribed and well-tolerated lipidlowering drugs in use today. According to several observational studies,94-100 statins may have a protective effect on the development of dementia. The limitations of these studies have been extensively discussed, particularly in relation to indication bias, and scenarios have been proposed that account for the observed protective effect.<sup>101</sup> Some studies fail to take into account the well-documented influence of gender on the development of dementia. 102 Despite these objections, a recent meta-analysis of the previously listed observational studies concluded that statins significantly lowered the risk of Alzheimer disease (RR: 0.43; 95% CI: 0.31 to 0.62).103 In contrast, 2 large, randomized, controlled trials 104,105 concluded that there was no effect of statins on the prevalence of dementia. It is obvious that new prospective trials are imperative to evaluate the usefulness of statins for neurodegenerative disorders.

Because the blood-brain barrier would be expected to block the entrance of most statins to the brain, at least if the statin is hydrophilic, the protective effect is unexpected. According to the data reported thus far, there is no difference between the protective effect of a hydrophobic and a hydrophilic statin. If statin effects on the brain can be confirmed, then they are likely to be of indirect nature and mediated by changes in nitric oxide levels and the microcirculation in the brain.

To date, several studies have investigated the effects of different statins on biomarkers used for the evaluation of the development of Alzheimer disease. The typical endpoints are the changes in plasma or CSF levels of  $\beta$ -amyloid or 24S-hydroxycholesterol. Regardless of the specific endpoint, the results of these studies are thus quite inconsistent. Most or all of the apparent decreases in 24S-hydroxycholesterol measured in these studies may be a consequence of the statin-induced cholesterol lowering. With one exception,  $^{106}$  the different studies have failed to demonstrate a reduction in the ratio between 24S-hydroxycholesterol and cholesterol, as a consequence of statin treatment. The results of these investigations are summarized in Table.

In addition to these clinical investigations, several studies have been performed aimed at characterizing the effects of

<sup>†</sup>Mixed group with either lovastatin, simvastatin, pravastatin, fluvastatin, cerivastatin.

<sup>‡</sup>See reference for a complete description of the patient groups.

statins on the metabolism of APP in different in vitro and in vivo systems. Invariably, inhibition of cholesterol synthesis results in a decrease in the  $\beta$ -secretase pathway in vitro. These studies are echoed by studies in laboratory animals, which report a statin-induced decrease in the amyloidogenic pathways. The levels of statin used in these studies in general are, however, ≈2 orders of magnitude higher than the normal dose in humans, and the results observed may be complicated by effects at the level of the vascular wall (eg, increased nitric oxide production) and alteration of the inflammatory response (eg, abrogation of LFA-1-mediated adhesion and activation of lymphocytes). A further complicating factor in the context of neurodegeneration is that secretion of apoE is dependent on prenylation, and production of prenyl groups may be reduced by the supraphysiological doses used in these studies. Because of the extremely high concentrations used in these studies, it is likely that the data presented are confounded by effects unrelated to lipid lowering.

A general concern with the aforementioned animal studies is that they are usually performed with male animals, and there are some indications that there may be gender difference in the production of  $A\beta$ . It has been reported that female mice expressing mutant APP have increased rather than reduced deposition of amyloid as a result of lovastatin treatment. 113

### **General Conclusion**

During the past decade, there has been a significant increase of knowledge about cholesterol homeostasis in the brain, and powerful new experimental tools have been introduced in this field of research. Overall, the current data provide tantalizing indications that modulation of intracerebral cholesterol levels may be a possible strategy for protection against Alzheimer disease and possibly also other forms of neurodegeneration. At present, it is impossible to draw a firm conclusion about the value of using statins to prevent (or delay) the onset of clinical dementia. In addition to HMG CoA reductase, there are many other possible targets for modulation of cholesterol homeostasis in the brain, and new therapeutic strategies are likely to be tested in the near future. The brain has only begun to reveal the secrets of its cholesterol homeostasis, and many more are likely to be uncovered.

### Acknowledgments

The studies from the authors' laboratory have been supported by grants from the Swedish Medical Research Council and the Swedish Heart-Lung Foundation.

#### References

- 1. Couerbe JP. Du Cerveau, considéré sous le point du vue chimique et physiologique. Annales de Chimie et de Physique. 1834;56:160-193.
- 2. Raffai RL, Weisgraber KH. Cholesterol: from heart attacks to Alzheimer's disease. J Lipid Res. 2003:44:1423-1430.
- 3. Snipes G, Suter U. Cholesterol and Myelin. In: Bittman R, ed. Cholesterol. New York: Plenum Press: 1998.
- 4. Dietschy JM, Turley SD. Cholesterol metabolism in the brain. Curr Opin Lipidol. 2001;12:105-112.
- 5. Björkhem I, Lutjohann D, Diczfalusy U, Stahle L, Ahlborg G, Wahren J. Cholesterol homeostasis in human brain: turnover of 24S-hydroxycholesterol and evidence for a cerebral origin of most of this oxysterol in the circulation. J Lipid Res. 1998;39:1594-1600.
- 6. Xie C, Lund EG, Turley SD, Russell DW, Dietschy JM. Quantitation of two pathways for cholesterol excretion from the brain in normal mice and mice with neurodegeneration. J Lipid Res. 2003;44:1780-1789.

7. Barenholz Y. Cholesterol and other membrane active sterols: from membrane evolution to "rafts." Prog Lipid Res. 2002;41:1-5.

**Björkhem and Meaney** 

- 8. Bittman R. Has nature designed the cholesterol side chain for optimal interaction with phospholipids? Subcell Biochem. 1997;28:145-171.
- 9. Tint G. Batta A. Xu G. Shefer S. Honda A. Irons M. Elias E. Salen G. The Smith-Lemli-Opitz syndrome: A potentially fatal birth defect caused by a block in the last enzymatic step in cholesterol biosynthesis. In: Bittman R, ed. Cholesterol. New York: Plenum Press; 1998.
- 10. Jira PE, Waterham HR, Wanders RJ, Smeitink JA, Sengers RC, Wevers RA. Smith-Lemli-Opitz syndrome and the DHCR7 gene. Ann Hum Genet. 2003:67:269-280.
- 11. Wechsler R, Brafman A, Shafir M, Heverin M, Gottlieb H, Damari G, Gozlan-Kelner S, Spikvak I, Moshkin O, Fridman E, Becker Y, Skaliter R, Einat P, Faerman A, Björkhem I, Feinstein E. Generation of viable cholesterol-free mouse. Science. 2003;302:2087.
- 12. Waelsch H, Sperry W, Stoyanoff VA. A study of the synthesis and deposition of lipids in brain and other tissues with deuterium as an indicator. J Biol Chem. 1940;135:291-296.
- 13. Bloch K, Berg B, Rittenberg D. The biological conversion of cholesterol to cholic acid. J Biol Chem. 1943;149:511-517.
- 14. Jurevics H, Morell P. Cholesterol for synthesis of myelin is made locally, not imported into brain. J Neurochem. 1995;64:895-901.
- 15. Chobanian A, Hollander W. Body cholesterol metabolism in man. I The equilibration of serum and tissue cholesterol. J Clin Invest. 1962;41:
- 16. Meaney S, Hassan M, Sakinis A, Lutjohann D, von Bergmann K, Wennmalm A, Diczfalusy U, Björkhem I. Evidence that the major oxysterols in human circulation originate from distinct pools of cholesterol: a stable isotope study. J Lipid Res. 2001;42:70-78.
- 17. Plotz E, Kabara J, Davis M, LeRoy G, Gould RG. Studies on the synthesis of cholesterol in the brain of the human fetus. Am J Obstet Gynecol. 1968:101:534-538
- 18. Reese TS, Karnovsky MJ. Fine structural localization of a blood-brain barrier to exogenous peroxidase. J Cell Biol. 1967;34:207-217.
- 19. Dehouck B, Dehouck MP, Fruchart JC, Cecchelli R. Upregulation of the low density lipoprotein receptor at the blood-brain barrier: intercommunications between brain capillary endothelial cells and astrocytes. J Cell Biol. 1994;126:465-473.
- 20. Dehouck B, Fenart L, Dehouck MP, Pierce A, Torpier G, Cecchelli R. A new function for the LDL receptor: transcytosis of LDL across the blood-brain barrier. J Cell Biol. 1997;138:877-889.
- 21. Panzenboeck U, Balazs Z, Sovic A, Hrzenjak A, Levak-Frank S, Wintersperger A, Malle E, Sattler W. ABCA1 and scavenger receptor class B, type I, are modulators of reverse sterol transport at an in vitro blood-brain barrier constituted of porcine brain capillary endothelial cells. J Biol Chem. 2002; 277:42781-42789.
- 22. Saito M, Benson EP, Rosenberg A. Metabolism of cholesterol and triacylglycerol in cultured chick neuronal cells, glial cells, and fibroblasts: accumulation of esterified cholesterol in serum-free culture. J Neurosci Res. 1987:18:319-325.
- 23. Koper JW, Zeinstra EC, Lopes-Cardozo M, van Golde LM. Acetoacetate and glucose as substrates for lipid synthesis by rat brain oligodendrocytes and astrocytes in serum-free culture. Biochim Biophys Acta. 1984;796: 20 - 26.
- 24. Pfrieger FW. Outsourcing in the brain: do neurons depend on cholesterol delivery by astrocytes? Bioessays. 2003;25:72-78.
- 25. Mauch DH, Nagler K, Schumacher S, Goritz C, Muller EC, Otto A, Pfrieger FW. CNS synaptogenesis promoted by glia-derived cholesterol. Science. 2001;294:1354-1357.
- 26. Pfrieger FW. Role of cholesterol in synapse formation and function. Biochim Biophys Acta. 2003;1610:271-280.
- 27. Volpe JJ, Obert KA. Coordinate regulation of cholesterol synthesis and 3-hydroxy-3-methylglutaryl coenzyme A synthase but not 3-hydroxy-3methylglutaryl coenzyme A reductase in C-6 glia. Arch Biochem Biophys. 1981:212:88-97.
- 28. Birkenmeier G, Grosche J, Reichenbach A. Immunocytochemical demonstration of alpha 2-M-R/LRP on Muller (glial) cells isolated from rabbit and human retina. Neuroreport. 1996;8:149-151.
- 29. Jung-Testas I, Weintraub H, Dupuis D, Eychenne B, Baulieu EE, Robel P. Low density lipoprotein-receptors in primary cultures of rat glial cells. J Steroid Biochem Mol Biol. 1992;42:597-605.
- 30. Norton W. Isolation and characterisation of myelin. In: Bittman R, ed. Cholesterol. New York: Plenum Press: 1998.
- 31. Boyles JK, Pitas RE, Wilson E, Mahley RW, Taylor JM. Apolipoprotein E associated with astrocytic glia of the central nervous system and with

- nonmyelinating glia of the peripheral nervous system. *J Clin Invest.* 1985; 76:1501–1513.
- Shanmugaratnam J, Berg E, Kimerer L, Johnson RJ, Amaratunga A, Schreiber BM, Fine RE. Retinal Muller glia secrete apolipoproteins E and J which are efficiently assembled into lipoprotein particles. *Brain Res Mol Brain Res*. 1997;50:113–120.
- 33. Pitas RE, Boyles JK, Lee SH, Hui D, Weisgraber KH. Lipoproteins and their receptors in the central nervous system. Characterization of the lipoproteins in cerebrospinal fluid and identification of apolipoprotein B,E(LDL) receptors in the brain. *J Biol Chem.* 1987;262:14352–14360.
- Danik M, Champagne D, Petit-Turcotte C, Beffert U, Poirier J. Brain lipoprotein metabolism and its relation to neurodegenerative disease. *Crit Rev Neurobiol*. 1999;13:357–407.
- Hofmann SL, Russell DW, Goldstein JL, Brown MS. mRNA for low density lipoprotein receptor in brain and spinal cord of immature and mature rabbits. Proc Natl Acad Sci U S A. 1987;84:6312–6316.
- Swanson LW, Simmons DM, Hofmann SL, Goldstein JL, Brown MS. Localization of mRNA for low density lipoprotein receptor and a cholesterol synthetic enzyme in rabbit nervous system by in situ hybridization. *Proc Natl Acad Sci U S A*. 1988;85:9821–9825.
- Bu G, Maksymovitch EA, Nerbonne JM, Schwartz AL. Expression and function of the low density lipoprotein receptor-related protein (LRP) in mammalian central neurons. J Biol Chem. 1994;269:18521–18528.
- Herz J, Bock HH. Lipoprotein receptors in the nervous system. Annu Rev Biochem. 2002;71:405–434.
- Osono Y, Woollett LA, Herz J, Dietschy JM. Role of the low density lipoprotein receptor in the flux of cholesterol through the plasma and across the tissues of the mouse. *J Clin Invest*. 1995;95:1124–1132.
- Masliah E, Mallory M, Ge N, Alford M, Veinbergs I, Roses AD. Neurodegeneration in the central nervous system of apoE-deficient mice. *Exp Neurol.* 1995;136:107–122.
- Mato M, Ookawara S, Mashiko T, Sakamoto A, Mato TK, Maeda N, Kodama T. Regional difference of lipid distribution in brain of apolipoprotein E deficient mice. *Anat Rec.* 1999;256:165–176.
- Trommsdorff M, Gotthardt M, Hiesberger T, Shelton J, Stockinger W, Nimpf J, Hammer RE, Richardson JA, Herz J. Reeler/Disabled-like disruption of neuronal migration in knockout mice lacking the VLDL receptor and ApoE receptor 2. Cell. 1999;97:689–701.
- 43. Saunders AM, Strittmatter WJ, Schmechel D, George-Hyslop PH, Pericak-Vance MA, Joo SH, Rosi BL, Gusella JF, Crapper-MacLachlan DR, Alberts MJ, et al. Association of apolipoprotein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. *Neurology*. 1993;43: 1467–1472.
- Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, Roses AD, Haines JL, Pericak-Vance MA. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science*. 1993;261:921–923.
- Teasdale GM, Nicoll JA, Murray G, Fiddes M. Association of apolipoprotein E polymorphism with outcome after head injury. *Lancet*. 1997; 350:1069–1071.
- Gong JS, Kobayashi M, Hayashi H, Zou K, Sawamura N, Fujita SC, Yanagisawa K, Michikawa M. Apolipoprotein E (ApoE) isoform-dependent lipid release from astrocytes prepared from human ApoE3 and ApoE4 knock-in mice. *J Biol Chem.* 2002;277:29919–29926.
- Sun Y, Wu S, Bu G, Onifade MK, Patel SN, LaDu MJ, Fagan AM, Holtzman DM. Glial fibrillary acidic protein-apolipoprotein E (apoE) transgenic mice: astrocyte-specific expression and differing biological effects of astrocyte-secreted apoE3 and apoE4 lipoproteins. *J Neurosci*. 1998;18:3261–3272.
- Schmitz G, Kaminski WE. ABCA2: a candidate regulator of neural transmembrane lipid transport. Cell Mol Life Sci. 2002;59:1285–1295.
- Zhou C, Zhao L, Inagaki N, Guan J, Nakajo S, Hirabayashi T, Kikuyama S, Shioda S. Atp-binding cassette transporter ABC2/ABCA2 in the rat brain: a novel mammalian lysosome-associated membrane protein and a specific marker for oligodendrocytes but not for myelin sheaths. *J Neurosci.* 2001; 21:849–857.
- Tanaka Y, Yamada K, Zhou CJ, Ban N, Shioda S, Inagaki N. Temporal and spatial profiles of ABCA2-expressing oligodendrocytes in the developing rat brain. *J Comp Neurol.* 2003;455:353–367.
- Pitas RE, Boyles JK, Lee SH, Foss D, Mahley RW. Astrocytes synthesize apolipoprotein E and metabolize apolipoprotein E-containing lipoproteins. *Biochim Biophys Acta*. 1987;917:148–161.
- Björkhem I, Lutjohann D, Breuer O, Sakinis A, Wennmalm A. Importance of a novel oxidative mechanism for elimination of brain cholesterol. Turnover of cholesterol and 24(S)-hydroxycholesterol in rat brain as

- measured with 18O2 techniques in vivo and in vitro. J Biol Chem. 1997; 272:30178-30184.
- Lutjohann D, Breuer O, Ahlborg G, Nennesmo I, Siden A, Diczfalusy U, Björkhem I. Cholesterol homeostasis in human brain: evidence for an age-dependent flux of 24S-hydroxycholesterol from the brain into the circulation. *Proc Natl Acad Sci U S A*. 1996;93:9799–9804.
- Kessel A, Ben-Tal N, May S. Interactions of cholesterol with lipid bilayers: the preferred configuration and fluctuations. *Biophys J.* 2001;81:643–658.
- Meaney S, Bodin K, Diczfalusy U, Björkhem I. On the rate of translocation in vitro and kinetics in vivo of the major oxysterols in human circulation: critical importance of the position of the oxygen function. *J Lipid Res*. 2002;43:2130–2135.
- Lange Y, Ye J, Strebel F. Movement of 25-hydroxycholesterol from the plasma membrane to the rough endoplasmic reticulum in cultured hepatoma cells. J Lipid Res. 1995;36:1092–1097.
- Lund EG, Guileyardo JM, Russell DW. cDNA cloning of cholesterol 24-hydroxylase, a mediator of cholesterol homeostasis in the brain. *Proc Natl Acad Sci U S A*. 1999;96:7238–7243.
- Meaney S, Lutjohann D, Diczfalusy U, Björkhem I. Formation of oxysterols from different pools of cholesterol as studied by stable isotope technique: cerebral origin of most circulating 24S-hydroxycholesterol in rats, but not in mice. *Biochim Biophys Acta*. 2000;1486:293–298.
- Lund EG, Xie C, Kotti T, Turley SD, Dietschy JM, Russell DW. Knockout of the cholesterol 24-hydroxylase gene in mice reveals a brain-specific mechanism of cholesterol turnover. *J Biol Chem.* 2003;278:22980–22988.
- Bogdanovic N, Bretillon L, Lund EG, Diczfalusy U, Lannfelt L, Winblad B, Russell DW, Björkhem I. On the turnover of brain cholesterol in patients with Alzheimer's disease. Abnormal induction of the cholesterol-catabolic enzyme CYP46 in glial cells. *Neurosci Lett.* 2001;314:45–48.
- Bretillon L, Lutjohann D, Stahle L, Widhe T, Bindl L, Eggertsen G, Diczfalusy U, Björkhem I. Plasma levels of 24S-hydroxycholesterol reflect the balance between cerebral production and hepatic metabolism and are inversely related to body surface. *J Lipid Res.* 2000;41:840–845.
- Leoni V, Masterman T, Diczfalusy U, De Luca G, Hillert J, Björkhem I. Changes in human plasma levels of the brain specific oxysterol 24Shydroxycholesterol during progression of multiple sclerosis. *Neurosci Lett.* 2002;331:163–166.
- Bretillon L, Siden A, Wahlund LO, Lutjohann D, Minthon L, Crisby M, Hillert J, Groth CG, Diczfalusy U, Björkhem I. Plasma levels of 24Shydroxycholesterol in patients with neurological diseases. *Neurosci Lett.* 2000;293:87–90.
- 64. Lutjohann D, Papassotiropoulos A, Björkhem I, Locatelli S, Bagli M, Oehring RD, Schlegel U, Jessen F, Rao ML, von Bergmann K, Heun R. Plasma 24S-hydroxycholesterol (cerebrosterol) is increased in Alzheimer and vascular demented patients. *J Lipid Res.* 2000;41:195–198.
- Babiker A, Diczfalusy U. Transport of side-chain oxidized oxysterols in the human circulation. *Biochim Biophys Acta*. 1998;1392:333–339.
- Edwards PA, Kennedy MA, Mak PA. LXRs; oxysterol-activated nuclear receptors that regulate genes controlling lipid homeostasis. *Vascul Pharmacol*. 2002;38:249–256.
- Repa JJ, Mangelsdorf DJ. The role of orphan nuclear receptors in the regulation of cholesterol homeostasis. Annu Rev Cell Dev Biol. 2000;16: 459–481
- Janowski BA, Willy PJ, Devi TR, Falck JR, Mangelsdorf DJ. An oxysterol signalling pathway mediated by the nuclear receptor LXR alpha. *Nature*. 1996;383:728–731.
- Lehmann JM, Kliewer SA, Moore LB, Smith-Oliver TA, Oliver BB, Su JL, Sundseth SS, Winegar DA, Blanchard DE, Spencer TA, Willson TM. Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway. *J Biol Chem.* 1997;272:3137–3140.
- Whitney KD, Watson MA, Collins JL, Benson WG, Stone TM, Numerick MJ, Tippin TK, Wilson JG, Winegar DA, Kliewer SA. Regulation of cholesterol homeostasis by the liver X receptors in the central nervous system. *Mol Endocrinol*. 2002;16:1378–1385.
- Björkhem I. Do oxysterols control cholesterol homeostasis? J Clin Invest. 2002;110:725–730.
- Meaney S. Studies on Oxysterols: Origins. Properties and Roles *Laboratory Medicine*. Stockholm; 2003.
- Teboul M, Enmark E, Li Q, Wikstrom AC, Pelto-Huikko M, Gustafsson JA.
   OR-1, a member of the nuclear receptor superfamily that interacts with the 9-cis-retinoic acid receptor. *Proc Natl Acad Sci U S A*. 1995;92:2096–2100.
- Kainu T, Kononen J, Enmark E, Gustafsson JA, Pelto-Huikko M. Localization and ontogeny of the orphan receptor OR-1 in the rat brain. *J Mol Neurosci.* 1996;7:29–39.

**Björkhem and Meaney** 

- 75. Wang L, Schuster GU, Hultenby K, Zhang Q, Andersson S, Gustafsson JA. Liver X receptors in the central nervous system: from lipid homeostasis to neuronal degeneration. Proc Natl Acad Sci USA. 2002;99:13878-13883.
- 76. Burns M, Duff K. Cholesterol in Alzheimer's disease and autopathy. Ann N Y Acad Sci. 2002;977:367-375.
- 77. Sparks DL, Scheff SW, Hunsaker JC, 3rd, Liu H, Landers T, Gross DR. Induction of Alzheimer-like beta-amyloid immunoreactivity in the brains of rabbits with dietary cholesterol. Exp Neurol. 1994;126:88-94.
- 78. Rubinstein DC, Easton DF. Apolipoprotein E genetic variation and Alzheimer's disease. a meta-analysis. Dement Geriatr Cogn Disord. 1999;10:
- 79. Wisniewski T, Frangione B. Apolipoprotein E: a pathological chaperone protein in patients with cerebral and systemic amyloid. Neurosci Lett. 1992;135:235-238.
- 80. Strittmatter WJ. Apolipoprotein E and Alzheimer's disease: signal transduction mechanisms. Biochem Soc Symp. 2001;67:101-109.
- 81. Kirsch C, Eckert GP, Koudinov AR, Muller WE. Brain cholesterol, statins and Alzheimer's disease. Pharmacopsychiatry. 2003;36:113-119.
- 82. Simons M, Keller P, Dichgans J, Schulz JB. Cholesterol and Alzheimer's disease: is there a link? Neurology. 2001;57:1089-1093.
- 83. Bodovitz S, Klein WL. Cholesterol modulates alpha-secretase cleavage of amyloid precursor protein. J Biol Chem. 1996;271:4436-4440.
- 84. Frears ER, Stephens DJ, Walters CE, Davies H, Austen BM. The role of cholesterol in the biosynthesis of beta-amyloid. Neuroreport. 1999;10: 1699 - 1705
- 85. Howland DS, Trusko SP, Savage MJ, Reaume AG, Lang DM, Hirsch JD, Maeda N, Siman R, Greenberg BD, Scott RW, Flood DG. Modulation of secreted beta-amyloid precursor protein and amyloid beta-peptide in brain by cholesterol. J Biol Chem. 1998;273:16576-16582.
- 86. Galbete JL, Martin TR, Peressini E, Modena P, Bianchi R, Forloni G. Cholesterol decreases secretion of the secreted form of amyloid precursor protein by interfering with glycosylation in the protein secretory pathway. Biochem J. 2000;348:307-313.
- 87. Liu Y, Peterson DA, Schubert D. Amyloid beta peptide alters intracellular vesicle trafficking and cholesterol homeostasis. Proc Natl Acad Sci USA. 1998;95:13266-13271.
- 88. Puglielli L, Konopka G, Pack-Chung E, Ingano LA, Berezovska O, Hyman BT, Chang TY, Tanzi RE, Kovacs DM. Acyl-coenzyme A: cholesterol acyltransferase modulates the generation of the amyloid beta-peptide. Nat Cell Biol. 2001:3:905-912.
- 89. Treiber-Held S, Distl R, Meske V, Albert F, Ohm TG. Spatial and temporal distribution of intracellular free cholesterol in brains of a Niemann-Pick type C mouse model showing hyperphosphorylated tau protein. Implications for Alzheimer's disease. J Pathol. 2003;200:95-103.
- 90. Xie C, Burns DK, Turley SD, Dietschy JM. Cholesterol is sequestered in the brains of mice with Niemann-Pick type C disease but turnover is increased. J Neuropathol Exp Neurol. 2000;59:1106–1117.
- 91. Papassotiropoulos A, Lutjohann D, Bagli M, Locatelli S, Jessen F, Rao ML, Maier W, Björkhem I, von Bergmann K, Heun R. Plasma 24S-hydroxycholesterol: a peripheral indicator of neuronal degeneration and potential state marker for Alzheimer's disease. Neuroreport. 2000;11:1959–1962.
- 92. Heverin M, Bogdanovic N, Lutjohann D, Bayer T, Pikuleva I, Bretillon L, Diczfalusy U, Winblad B, Björkhem I. Changes in the levels of cerebral and extracerebral sterols in the brain of patients with Alzheimer's disease. J Lipid Res. 2003.
- 93. Lutjohann D, Brzezinka A, Barth E, Abramowski D, Staufenbiel M, von Bergmann K, Beyreuther K, Multhaup G, Bayer TA. Profile of cholesterol-related sterols in aged amyloid precursor protein transgenic mouse brain. J Lipid Res. 2002;43:1078-1085.
- 94. Green RC, Jayakumar P, Benke K, Farrer L. Statin use is associated with reduced risk of Alzheimer's disease. Neurobiol Aging. 2002;23:273-274.
- 95. Hajjar I, Schumpert J, Hirth V, Wieland D, Eleazer GP. The impact of the use of statins on the prevalence of dementia and the progression of cognitive impairment. J Gerontol A Biol Sci Med Sci. 2002;57:M414-M418.
- 96. Jick H, Zornberg GL, Jick SS, Seshadri S, Drachman DA. Statins and the risk of dementia. Lancet. 2000;356:1627-1631.

- 97. Rockwood K, Kirkland S, Hogan DB, MacKnight C, Merry H, Verreault R, Wolfson C, McDowell I. Use of lipid-lowering agents, indication bias, and the risk of dementia in community-dwelling elderly people. Arch Neurol. 2002;59:223-227.
- 98. Rodriguez EG, Dodge HH, Birzescu MA, Stoehr GP, Ganguli M. Use of lipid-lowering drugs in older adults with and without dementia: a community-based epidemiological study. J Am Geriatr Soc. 2002;50: 1852-1856.
- 99. Wolozin B, Kellman W, Ruosseau P, Celesia GG, Siegel G. Decreased prevalence of Alzheimer disease associated with 3-hydroxy-3-methyglutaryl coenzyme A reductase inhibitors. Arch Neurol. 2000;57:1439-1443.
- 100. Yaffe K, Barrett-Connor E, Lin F, Grady D. Serum lipoprotein levels, statin use, and cognitive function in older women. Arch Neurol. 2002; 59:378-384
- 101. Haley RW, Dietschy JM. Is there a connection between the concentration of cholesterol circulating in plasma and the rate of neuritic plaque formation in Alzheimer disease? Arch Neurol. 2000;57:1410-1412.
- 102. Green RC, Cupples LA, Go R, Benke KS, Edeki T, Griffith PA, Williams M, Hipps Y, Graff-Radford N, Bachman D, Farrer LA. Risk of dementia among white and African Am relatives of patients with Alzheimer disease. JAMA. 2002;287:329-336.
- 103. Etminan M, Gill S, Samii A. The role of lipid-lowering drugs in cognitive function: a meta-analysis of observational studies. Pharmacotherapy. 2003;23:726-730.
- 104. MRC/BHF Heart Protection Study of cholesterol lowering with simvastatin in 20,536 high-risk individuals: a randomised placebo-controlled trial. Lancet. 2002;360:7-22.
- 105. Shepherd J, Blauw GJ, Murphy MB, Bollen EL, Buckley BM, Cobbe SM, Ford I, Gaw A, Hyland M, Jukema JW, Kamper AM, Macfarlane PW, Meinders AE, Norrie J, Packard CJ, Perry IJ, Stott DJ, Sweeney BJ, Twomey C, Westendorp RG. Pravastatin in elderly individuals at risk of vascular disease (PROSPER): a randomised controlled trial. Lancet. 2002;360:1623-1630.
- 106. Locatelli S, Lutjohann D, Schmidt HH, Otto C, Beisiegel U, von Bergmann K. Reduction of plasma 24S-hydroxycholesterol (cerebrosterol) levels using high-dosage simvastatin in patients with hypercholesterolemia: evidence that simvastatin affects cholesterol metabolism in the human brain. Arch Neurol. 2002;59:213-216.
- 107. Ishii K, Tokuda T, Matsushima T, Miya F, Shoji S, Ikeda S, Tamaoka A. Pravastatin at 10 mg/day does not decrease plasma levels of either amyloid-beta (Abeta) 40 or Abeta 42 in humans. Neurosci Lett. 2003; 350:161-164
- 108. Sjogren M, Gustafsson K, Syversen S, Olsson A, Edman A, Davidsson P. Wallin A. Blennow K. Treatment with Simvastatin in Patients with Alzheimer's Disease Lowers Both alpha- and beta-Cleaved Amyloid Precursor Protein. Dement Geriatr Cogn Disord. 2003;16:25-30.
- 109. Buxbaum JD, Cullen EI, Friedhoff LT. Pharmacological concentrations of the HMG-CoA reductase inhibitor lovastatin decrease the formation of the Alzheimer beta-amyloid peptide in vitro and in patients. Front Biosci. 2002;7:a50-a59.
- 110. Simons M, Schwarzler F, Lutjohann D, von Bergmann K, Beyreuther K, Dichgans J, Wormstall H, Hartmann T, Schulz JB. Treatment with simvastatin in normocholesterolemic patients with Alzheimer's disease: A 26-week randomized, placebo-controlled, double-blind trial. Ann Neurol. 2002;52:346-350.
- 111. Vega GL, Weiner MF, Lipton AM, Von Bergmann K, Lutjohann D, Moore C, Svetlik D. Reduction in levels of 24S-hydroxycholesterol by statin treatment in patients with Alzheimer disease. Arch Neurol. 2003; 60:510-515.
- 112. Fassbender K, Stroick M, Bertsch T, Ragoschke A, Kuehl S, Walter S, Walter J, Brechtel K, Muehlhauser F, Von Bergmann K, Lutjohann D. Effects of statins on human cerebral cholesterol metabolism and secretion of Alzheimer amyloid peptide. Neurology. 2002;59: 1257-1258.
- 113. Park IH, Hwang EM, Hong HS, Boo JH, Oh SS, Lee J, Jung MW, Bang OY, Kim SU, Mook-Jung I. Lovastatin enhances Abeta production and senile plaque deposition in female Tg2576 mice. Neurobiol Aging. 2003;24:637-643.

# Arteriosclerosis, Thrombosis, and Vascular Biology



JOURNAL OF THE AMERICAN HEART ASSOCIATION

### Brain Cholesterol: Long Secret Life Behind a Barrier

Ingemar Björkhem and Steve Meaney

Arterioscler Thromb Vasc Biol. 2004;24:806-815; originally published online February 5, 2004; doi: 10.1161/01.ATV.0000120374.59826.1b

Arteriosclerosis, Thrombosis, and Vascular Biology is published by the American Heart Association, 7272
Greenville Avenue, Dallas, TX 75231
Copyright © 2004 American Heart Association, Inc. All rights reserved.

Print ISSN: 1079-5642. Online ISSN: 1524-4636

The online version of this article, along with updated information and services, is located on the World Wide Web at:

http://atvb.ahajournals.org/content/24/5/806

**Permissions:** Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Arteriosclerosis*, *Thrombosis*, *and Vascular Biology* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

**Reprints:** Information about reprints can be found online at: http://www.lww.com/reprints

**Subscriptions:** Information about subscribing to *Arteriosclerosis*, *Thrombosis*, *and Vascular Biology* is online at:

http://atvb.ahajournals.org//subscriptions/