

Ionizing Radiation and Autoimmunity

Induction of Autoimmune Disease in Mice by High Dose Fractionated Total Lymphoid Irradiation and Its Prevention by Inoculating Normal T Cells¹

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Ionizing radiation can functionally alter the immune system and break self-tolerance. High dose (42.5 Gy), fractionated (2.5 Gy 17 times) total lymphoid irradiation (TLI) on mice caused various organ-specific autoimmune diseases, such as gastritis, thyroiditis, and orchitis, depending on the radiation dosages, the extent of lymphoid irradiation, and the genetic background of the mouse strains. Radiation-induced tissue damage is not the primary cause of the autoimmune disease because irradiation of the target organs alone failed to elicit the autoimmunity and shielding of the organs from irradiation was unable to prevent it. In contrast, irradiation of both the thymus and the peripheral lymphoid organs/tissues was required for efficient induction of autoimmune disease by TLI. TLI eliminated the majority of mature thymocytes and the peripheral T cells for 1 mo, and inoculation of spleen cell, thymocyte, or bone marrow cell suspensions (prepared from syngeneic nonirradiated mice) within 2 wk after TLI effectively prevented the autoimmune development. Depletion of T cells from the inocula abrogated the preventive activity. CD4⁺ T cells mediated the autoimmune prevention but CD8⁺ T cells did not. CD4⁺ T cells also appeared to mediate the TLI-induced autoimmune disease because CD4⁺ T cells from disease-bearing TLI mice adoptively transferred the autoimmune disease to syngeneic naive mice. Taken together, these results indicate that high dose, fractionated ionizing radiation on the lymphoid organs/tissues can cause autoimmune disease by affecting the T cell immune system, rather than the target self-Ags, presumably by altering T cell-dependent control of self-reactive T cells. *Journal of Immunology*, 1994, 152: 2586.

T cells are key mediators of many organ-specific autoimmune diseases, such as autoimmune thyroiditis, gastritis, and insulinitis in insulin-dependent diabetes mellitus (1–6). To maintain immunologic self-tolerance and prevent autoimmune disease, the pathogenic self-reactive T cells must be eliminated in the thymus or, when produced by the thymus, their expansion/activation must be controlled in the periphery (7–10). Autoimmune disease may develop when exogenous insults affect the

thymus and elicit the production of the pathogenic self-reactive T cells and/or prepare the immunologic conditions favorable to their peripheral expansion/activation (11, 12). Here, we show that ionizing radiation has such an autoimmune-inducing effect.

Biologic effects of ionizing radiation depend on the character of the radiation, the total dose administered, the time when the dose is given, and the site of irradiation (13, 14). High dose irradiation can be achieved by low dose fractionation and by shielding vital organs. One such radiation protocol, called TLI,³ consists of irradiating major lymphoid organs, including the thymus and the spleen, while shielding radiosensitive nonlymphoid organs (e.g., long bones, lung, and skull) with lead plates (15, 16). Total dose of 30 to 40 Gy is achieved within 4 to 5 wk by

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³ Abbreviations used in this paper: TLI, total lymphoid irradiation; Tx, thymectomy; H&E, hematoxylin and eosin; BM, bone marrow; GvHD, graft-versus-host disease; CsA, cyclosporin A.

multiple small fractions of 2 to 2.5 Gy each. TLI has been routinely used for treatment of lymphoid malignancies in humans for the past 30 yr (15). It is an effective treatment for various autoimmune diseases in humans and rodents (17–21). Furthermore, a unique immunologic effect of TLI is that it establishes allograft tolerance in humans and animals when allogeneic bone marrow cells or solid organs are engrafted immediately after TLI (22–26).

In this report, we show that TLI paradoxically breaks self-tolerance in selected strains of normal mice and elicits T cell-mediated organ-specific autoimmune diseases by affecting the T cell immune system not the target self-Ags. Furthermore, the development of autoimmune disease can be prevented by inoculating T cells from syngeneic non-irradiated mice. This is an example of autoimmune disease caused by physical insult to the immune system and would contribute to our understanding of the cause and mechanism of autoimmune disease in humans.

Materials and Methods

Mice

BALB/cKa, C57BL/Ka, and C3H/Km mice were obtained from the specific pathogen-free colony of Dr. Robert Kallman, Department of Radiology, Stanford University School of Medicine. Male BALB/c *nu/+* mice and 6-wk-old male athymic BALB/c-*nu/nu* mice were purchased from Life Science Associates, St. Petersburg, FL. SWR/J and A/J mice were purchased from the Jackson Laboratory, Bar Harbor, ME. (C57BL/6J \times BALB/cByJ)F₁ (B6CF1) and DBA/2J mice were provided from the vivarium of the Scripps Research Institute.

TLI

Male mice of 3 to 4 mo of age were used for TLI. Mice were anesthetized with Nembutal (pentobarbital; Abbott Laboratories, Chicago, IL) and positioned in an apparatus designed to irradiate the major lymphoid organs, including the spleen, thymus, and cervical, axillary, mediastinal, inguinal, and mesenteric lymph nodes (22, 23). The skull, lungs, ribs, thoracic spines, tail, and hind legs were shielded with lead. The mice were given 2.5 Gy/day, 5 times/wk to a total accumulative dose of 42.5 Gy over 17 treatments. Irradiation was delivered from a single 250 kV (15A) source (Phillips Medical System Inc., Shelton, CT). The dose was 0.93 Gy/min using a 0.35-mm Cu filter and a 52-cm source-axis distance. A group of mice was irradiated at the Scripps Research Institute from a ¹³⁷Cs source (Gammacell 40 irradiator; Atomic Energy of Canada, Ottawa, Canada) at 0.813 Gy/min with extra lead plates for shielding of vital organs. Neomycin (Pharma-Tek, Inc., Huntington, NY) and polymyxin B (Sigma, St. Louis, MO) were added to the drinking water during TLI and for 1 mo after completion of TLI. For partial irradiation of lymphoid organs, a part of the body was shielded by an additional lead plate.

ELISA

Details of the ELISA (using alkaline phosphatase-conjugated secondary Ab and *p*-nitrophenyl disodium hexahydrate as the substrate) for detecting Abs against the gastric parietal cells, thyroglobulins, ss or dsDNA, or trinitrophenyl haptens and the ELISA for measuring serum immune complexes or IgG concentration were previously described (12, 27). Titers of autoantibodies against the parietal cells or thyroglobulins were defined as the highest dilution at which the absorbance at 405 nm is over 0.1 or 0.05, respectively, when test sera are serially twofold diluted from 1/10 dilution, because the absorbance at 1/10 dilution of 3- to 4-mo-old normal BALB/c mice was 0.07 ± 0.028 ($n = 50$) in the antiparietal cell ELISA and 0.03 ± 0.012 ($n = 20$) in the antithyroglobulin ELISA (27).

Immunohistology

Stomachs were embedded in OCT compound (Miles Inc., Chicago, IL) and instantaneously frozen in liquid nitrogen. Cryostat tissue sections were incubated with monoclonal antiparietal cell Ab (IgG2b) that was prepared from a mouse with autoimmune gastritis induced by neonatal CsA treatment (27, and manuscript in preparation). After washing with PBS, the slides were incubated with peroxidase-conjugated goat anti-mouse IgG (Caltag Laboratories, San Francisco, CA) then washed and incubated with diaminobenzidine and H₂O₂. The slides were then counterstained with hematoxylin.

Cell preparation

Spleen cell suspensions (4×10^7) were treated with anti-Thy-1.2 (mouse IgM) (28), anti-L3T4 (rat IgG2b) (29), or anti-Lyt-2.2 (mouse IgG2a) (30) and nontoxic rabbit C, as previously described (11), and i.v. inoculated into mice. The hybridoma cell lines secreting these mAbs were obtained from American Tissue Culture Collection, Rockville, MD.

Cytofluorometric analysis of T cell subsets

FITC-labeled anti-Thy-1.2 and anti-Lyt-2 mAbs and phycoerythrin-labeled anti-L3T4 mAbs were purchased from Becton Dickinson, San Jose, CA. Biotinylated mAbs specific for TCR V β 11 (RR 3-15) or V β 6 domain (RR 4-7) were provided by Dr. O. Kanagawa, Washington University, St. Louis, MO (31, 32). Anti-V β 3 (KJ23) (33) and anti-V β 8.1 and 8.2 Abs (MR5-2) (34) were purchased from PharMingen, San Diego, CA. Staining procedure was as previously described (27). Stained cells were analyzed by an FACSscan or FACS II flow cytometer (Becton Dickinson).

Other methods

When mice were killed, tissues (thyroids, salivary glands, lungs, stomach, pancreas, adrenal glands, liver, kidneys, ovaries, or testes) were fixed in 10% formalin and processed for histologic sectioning and staining with H&E (11, 27).

Results

Male BALB/c mice at 3 mo of age were used in the following experiments unless indicated otherwise.

Development of organ-specific autoimmune disease, especially gastritis, in BALB/c mice after TLI

A total of 55 BALB/c mice were given TLI in the various experiments described below. They were allowed to survive for 6 mo and examined for serologic and/or histologic occurrence of autoimmunity (Table I). Seventy-five percent of the TLI mice developed histologically evident gastritis. Approximately 60% showed achlorhydria (pH 7 to 8 of the gastric contents compared with pH 3 to 4 of control nonirradiated mice) and macroscopically evident giant rugae formation due to destruction of the gastric parietal cells and compensatory hyperplasia of mucous cells (see below). The majority (~90%) developed antiparietal cell autoantibodies (assessed by ELISA) in the circulation (see refs. 11 and 27 for macroscopic view of the giant rugae and antiparietal cell autoantibody demonstrated by indirect immunofluorescence staining).

Autoantibodies against gastric parietal cells reacted with a ~90 kDa gastric membrane component, similar to the H⁺-K⁺-ATPase reported for human autoimmune gastritis/pernicious anemia (6) by Western blotting technique (S. Sakaguchi and N. Sakaguchi, unpublished data). Two

Table 1. Incidence of autoimmune disease in BALB/c male mice given TLI

Gastritis	Thyroiditis	Orchitis/ Epididymitis	Sialoadenitis
41/55 ^a (74.5) (2°; 35 1°; 6) ^b	2/55 (3.6) (1°; 2)	5/55 (9.1) (2°; 2 1°; 3)	3/55 (7.3)

^a Three-month-old male BALB/c mice were given TLI. Incidence of histologically evident autoimmune disease, assessed 5 to 6 mo after TLI, is shown with percentage incidence in parentheses. Thirty-five (63.6%) mice showed achlorhydria (pH 6–8 of gastric contents compared with pH 3–4 of control nonirradiated mice) and macroscopically evident giant rugae formation due to severe destruction of gastric parietal cells and compensatory hyperplasia of mucous cells (grade 2) (see Fig. 3C and ref. 11 for macroscopic view of giant rugae). In six (10.9%) mice, gastritis was histologically observed but not macroscopically evident (grade 1) (see Fig. 3B). Two cases of thyroiditis were histologically evident but showed no goiter formation (grade 1) (11). In two cases of orchitis, both testicles and epididymes were destroyed and aspermatogenic (grade 2); in three cases, epididymitis was histologically evident but orchitis was mild (grade 1) (see ref. 11 for histologic details).

^b 1°, grade 1; 2°, grade 2.

(3.6%) mice histologically developed thyroiditis with serum antithyroglobulin titer of 320 and 640 by ELISA, respectively; ~20% had low but significant titers of antithyroglobulin autoantibody (10 to 40 by ELISA) without histologically evident thyroiditis. Approximately 10% developed epididymitis and/or orchitis; ~7% showed infiltration of inflammatory cells in the salivary gland. Other organs were histologically normal.

Time course of autoimmune development and correlation between histologic severity and autoantibody titer

The antiparietal cell autoantibodies became detectable by ELISA from 1 mo after TLI; the titers steadily rose in approximately 2 mo and stayed constant for the rest of the observation period (Fig. 1). The grade of histologic severity of the gastritis was well correlated with the titer of antiparietal cell autoantibody.

Dose responses

Mice given 4, 8, or 17 TLI treatments were examined 6 mo later for the degree of histologic and serologic gastric autoimmunity (Fig. 2). Histologically evident gastritis developed only in the group given 17 TLI treatments. The eight TLI treatments induced low titers of antiparietal cell autoantibodies in 20% of mice but apparently no histologic changes in the gastric mucosa.

Immunopathology of autoimmune disease

Damage to the gastric parietal cells and infiltration of inflammatory cells became histologically evident 4 to 6 wk after completion of TLI (Fig. 3A). The lesions had progressed in 6 mo to severe infiltration of mononuclear cells, complete loss of parietal cells and chief cells, and hyperplasia of mucous cells (Fig. 3, B and C). Immunohistochemical staining of the gastric mucosa with antiparietal

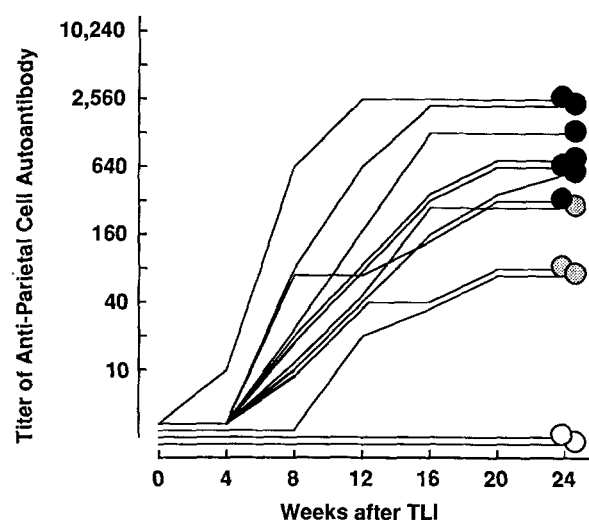


FIGURE 1. Time course of development of gastric autoimmunity after TLI. Male BALB/c mice (3 mo old) were given TLI, checked every 4 wk for serum titers of antiparietal cell autoantibodies by ELISA, and examined for histologic occurrence of gastritis 6 mo after completion of TLI. ○, Histologically intact gastric mucosa; ●, gastritis histologically evident with destruction of parietal cells and cellular infiltration into gastric mucosa as shown in Figure 3B (grade 1); ●, severe destruction of gastric mucosa with formation of giant rugae due to compensatory hyperplasia of mucous cells as shown in Figure 3C (grade 2) (see ref. 11 for giant rugae).

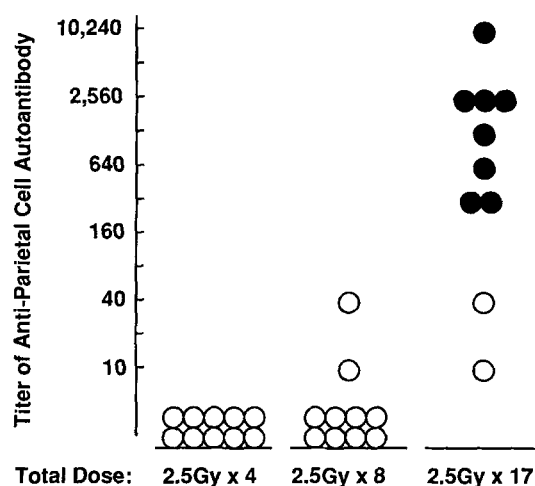


FIGURE 2. Dose effects of TLI on development of gastric autoimmunity. Male BALB/c mice (3 mo old) were given 4, 8, or 17 times of TLI and examined for histology of stomach and titer of antiparietal cell autoantibodies 6 mo after completion of TLI. ○, ●; See Figure 1.

cell mAbs showed no pathologic changes in the parietal cells 1 wk after completion of TLI (Fig. 3D); in contrast, the parietal cells were lost or destroyed in the gastric mucosa 6 mo after TLI (Fig. 3E). The class II MHC Ags were expressed on the gastric epithelium afflicted with histologically evident gastritis but not on the apparently intact

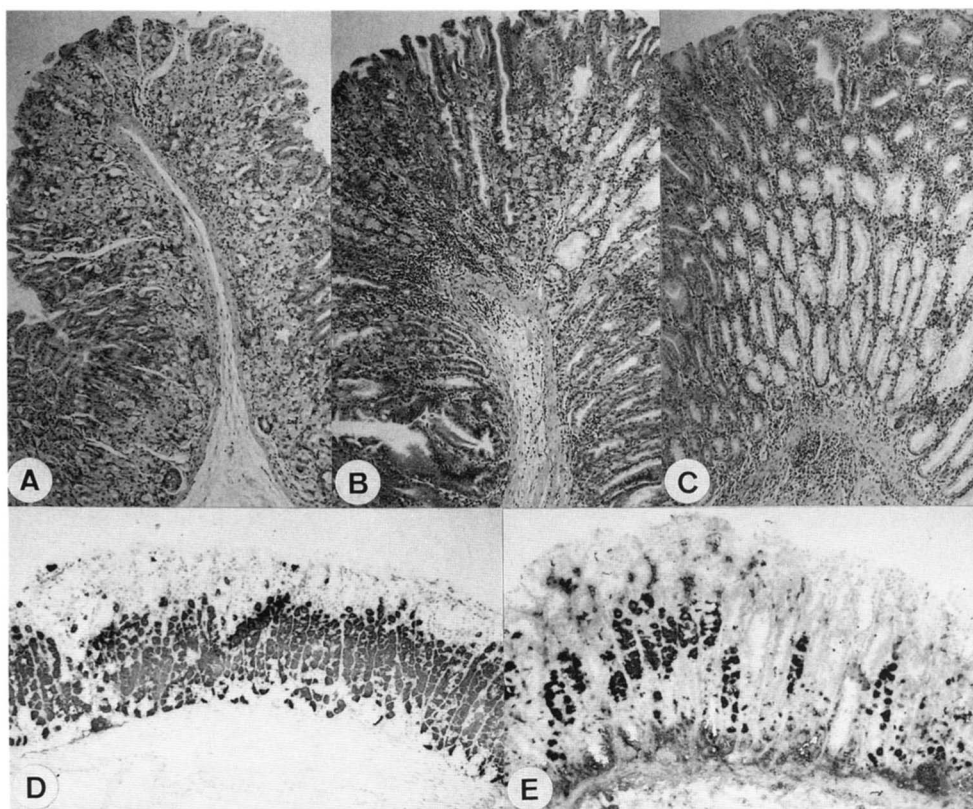


FIGURE 3. Immunopathology of autoimmune gastritis. Gastric mucosa of mice 1 mo (A), 2 mo (B), or 6 mo (C) after TLI (H&E staining). Gastritis progressed from mild infiltration of mononuclear cells (A) to severe infiltration and damage of parietal cells (grade 1) (B), and to complete loss of parietal cells and chief cells with replacement by mucous cells (grade 2) (C). Immunoperoxidase staining with parietal cell-specific mAbs shows intact parietal cells in gastric mucosa 1 wk after completion of TLI (D) but loss and damage of parietal cells in gastric mucosa 6 mo after TLI (E). A to E; $\times 100$.

epithelium immediately after TLI (data not shown). Immunopathology of thyroiditis, orchitis/epididymitis, and sialoadenitis was similar to that described elsewhere (11, 27).

Adoptive transfer of autoimmune disease by T cells

TLI was given to 12 BALB/c *nu/+* mice (3 mo of age) whose testes were carefully shielded by additional lead plates during TLI. Nine (75%) mice developed histologically severe gastritis 4 mo after TLI; two of them also suffered from epididymitis and low grade orchitis (see ref. 11 for immunopathology of epididymitis/orchitis). Spleen and lymph node cell suspensions (4×10^7) prepared from individual mice were i.v. inoculated into 6-wk-old syngeneic BALB/c *nu/nu* mice; the *nu/nu* mice were histologically and serologically examined 2 mo later.

Table II shows that the lymphocytes could adoptively transfer the autoimmune disease, and removal of CD4⁺ (L3T4⁺) cells eliminated the transfer activity of the lymphocytes, whereas removal of CD8⁺ (Lyt-2⁺) cells did not. The result indicates that CD4⁺ cells can transfer the disease in a disease-specific manner by helping B cells to form organ-specific autoantibodies and, presumably, by eliciting cell-mediated immune responses to the target

Table II. Adoptive transfer of autoimmune disease to syngeneic *nu/nu* mice by CD4⁺ T cells

Autoimmune Disease in Donor TLI Mice ^a	Treatment of Lymphocytes ^b	Autoimmune Disease in Recipient <i>nu/nu</i> Mice ^c	
		Gastritis	Orchitis
Gastritis (7)	C	3/3 (640, 320, 320)	0/3
	Anti-Thy-1.2+C	0/2 (10, <10)	0/2
	Anti-Lyt-2.2+C	3/3 (640, 640, 320)	0/3
	Anti-L3T4+C	0/3 (10, <10, <10)	0/3
Gastritis and Orchitis (2)	C	1/1 (320)	1/1
	Anti-Thy-1.2+C	0/1 (<10)	0/1

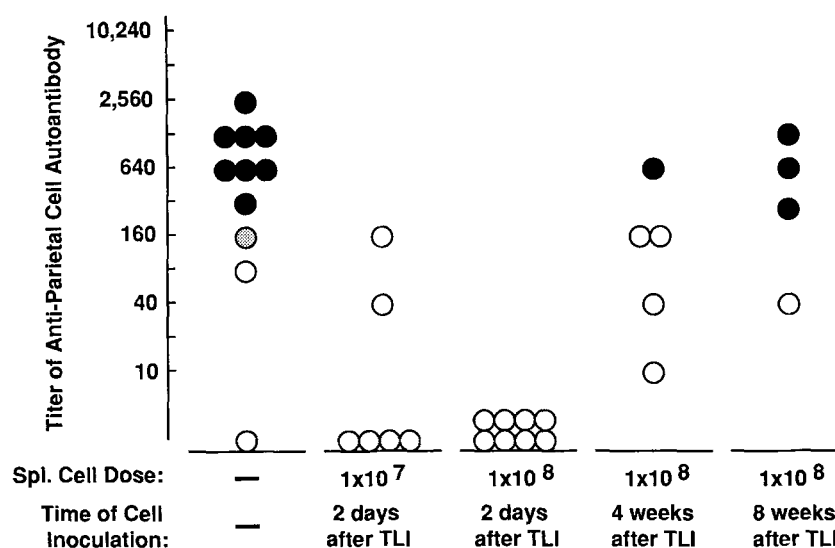
^a Male BALB/c *nu/+* mice (3 mo old) were given TLI with careful shielding of testes with extra lead plate. Spleen and lymph node cell suspensions prepared from the *nu/+* mice with respective autoimmune disease(s) at 4 mo after TLI were i.v. transferred to 6- to 8-wk-old BALB/c *nu/nu* mice. Number of mice used as donors is shown in parentheses.

^b Spleen and lymph node cell suspensions (4×10^7) were treated with antisera and rabbit complement (C) before transfer.

^c Incidence of histologically evident autoimmune diseases in *nu/nu* mice that survived 2 mo after cell transfer. Gastritis in *nu/nu* recipients was all grade 2. Titers of antiparietal cell autoantibodies assessed by ELISA for individual *nu/nu* mice are shown in parentheses.

self-Ags (35). In these transfers, however, approximately 50% of the recipient *nu/nu* mice (10 of 22 mice), especially those transferred with lymphocytes treated with C

FIGURE 6. Prevention of autoimmune disease by inoculating spleen cells from syngeneic nonirradiated mice. Male BALB/c mice (3 mo old) received TLI and inoculation of normal BALB/c spleen cell suspensions (1×10^7 or 1×10^8) 2 days, 4 wk, or 8 wk after completion of TLI. The mice were histologically and serologically examined 6 mo after TLI. ○, ●, ■; See Figure 1.



might contribute to development of organ-specific autoimmunities; e.g., V β 11⁺ cells, the majority of which are deleted in normal BALB/c mice, were found to increase in neonatally thymectomized BALB/c mice, some of which spontaneously developed autoimmune gastritis similar to the one after TLI (36–39). Compared with the post-thymectomy autoimmune model, there was not a marked difference between TLI BALB/c mice 3 mo after TLI and age-matched control BALB/c mice in percentage of composition of lymph node α/β TCR⁺ cells expressing the following TCR V β domains: V β 3 (0.09 ± 0.05 in TLI mice vs 0.06 ± 0.05 in control mice); V β 8.1, 8.2 (23.36 ± 1.23 vs 25.91 ± 1.34); V β 11 (2.04 ± 0.96 vs 1.73 ± 0.50) (the mean \pm SD, $n = 3$).

Prevention of autoimmune disease by inoculating lymphocytes from syngeneic nonirradiated mice

To determine whether restoration of the TLI-induced lymphocyte depletion can prevent or ameliorate autoimmune disease, spleen cell suspensions (1×10^7 or 1×10^8) prepared from syngeneic nonirradiated mice were i.v. inoculated (Fig. 6). The inoculation of 1×10^7 cells 2 days after TLI completely prevented the occurrence of gastritis; however, the cell inoculation 4 or 8 wk after TLI was less effective and ineffective, respectively, even with the dose of 1×10^8 cells.

Inoculation of thymocytes (1×10^8) or BM cells (3×10^7) immediately after TLI significantly reduced the incidence/severity of gastritis, although less effectively than the spleen cell inoculation (Fig. 7). Depletion of T cells from the splenic or BM cell inocula abrogated the autoimmune preventive activity.

To determine which splenic T cell subset, CD4⁺ or CD8⁺ population, mediates the prevention, the equivalent numbers of CD4⁺ or CD8⁺ splenic T cells were inoculated 2 days after TLI (Fig. 8). CD4⁺ cells completely prevented the disease development, whereas CD8⁺ cells did not.

Strain difference in the susceptibility to TLI-induced autoimmune disease

TLI elicited gastritis in the BALB/c (H-2^d), A (H-2^a), and SWR (H-2^q) strains of mice at high histologic incidences (70 to 90%) with high titers of antiparietal cell autoantibodies (Fig. 9). In contrast to these three strains, C57BL/6 (H-2^b), DBA/2 (H-2^d), and C3H (H-2^k) were resistant to the autoimmune gastritis, although the C3H strain developed low titers of antiparietal cell autoantibodies. The histologic incidence of gastritis in (C57BL/6 \times BALB/c)_F₁ mice was intermediate between the incidences in C57BL/6 and BALB/c. Mild epididymitis/orchitis was observed in 30% of SWR mice and 10 to 20% of A and BALB/c mice. One A mouse developed thyroiditis with thyroglobulin autoantibodies (x320 by ELISA); 15 to 20% of A and BALB/c and 50% of SWR developed significant titers of antithyroglobulin autoantibodies (80 to 160 by ELISA). Significantly high titers (i.e., higher than the 95th percentile of the titers in normal nonirradiated mice) of IgG autoantibodies against ssDNA developed in ~50, ~60, and ~80% of BALB/c, A, and SWR strains, respectively, after TLI. Titers of IgG autoantibodies specific for dsDNA were not significantly high in these strains. Serum concentration of immune complexes was significantly high (i.e., higher than the 95th percentile of the concentrations in normal nonirradiated mice) in 40 to 50% of the A and SWR strains given TLI but with no histologic evidence of immune complex glomerulonephritis or vasculitis.

Discussion

Inflammatory lesions observed after TLI in the gastric mucosa, thyroid glands, and testes were T cell-mediated autoimmune diseases, because CD4⁺ T cells could adoptively transfer the lesions to syngeneic athymic nude mice

FIGURE 7. Prevention of autoimmune disease by inoculating spleen cell, thymocyte, or BM cell suspensions from syngeneic nonirradiated mice. Male BALB/c mice (3 mo old) were given TLI and inoculated 2 days later with normal BALB/c spleen cells (1×10^8), spleen cells treated with anti-Thy-1 plus complement ($\sim 6 \times 10^7$), thymocytes (1×10^8), BM cells (3×10^7), or BM cells treated with anti-Thy-1 plus complement ($2.5-3 \times 10^7$). The mice were histologically and serologically examined 4 mo later. ○, ●, ●; See Figure 1.

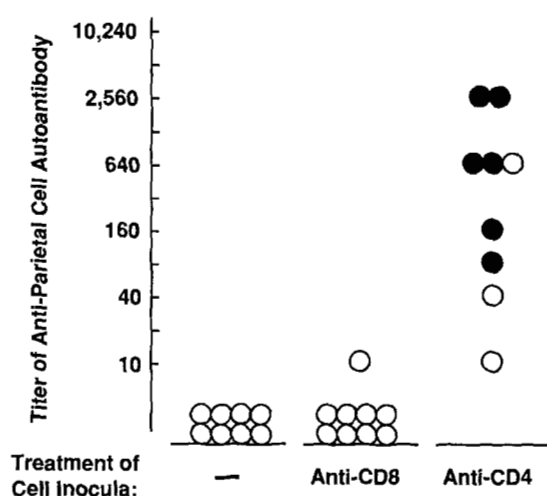
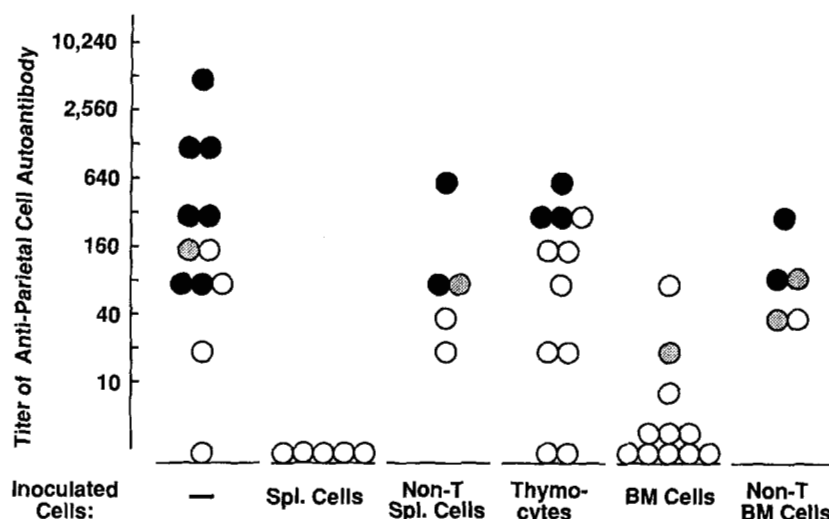


FIGURE 8. Prevention of autoimmune disease by inoculating splenic CD4⁺ cells from syngeneic nonirradiated mice. Male BALB/c mice (3 mo old) given TLI were inoculated with normal BALB/c spleen cell suspensions treated with anti-CD4 or anti-CD8 Ab and complement then examined 4 mo later for histologic and/or serologic development of autoimmune gastritis. To inoculate an equivalent number of T cells, 1×10^8 or 5×10^7 spleen cells were treated with anti-L3T4 or anti-Lyt-2, respectively, because the number of CD4⁺ cells in normal spleen is roughly twice the number of CD8⁺ cells in BALB/c mice. As control, 5×10^7 cells were treated with complement only and i.v. inoculated. ○, ●; See Figure 1.

with resulting histologic damage of the corresponding organs and appearance of specific autoantibodies in the circulation. These autoimmune diseases were similar in immunopathology to human organ-specific autoimmune diseases, such as autoimmune gastritis with pernicious anemia, Hashimoto's autoimmune thyroiditis, or autoimmune orchitis in male infertility (1).

For the following reasons, TLI appears to affect primarily the T cell immune system not the target organs in eliciting autoimmune disease; i.e., 1) shielding the target organs (e.g., testes) during TLI did not prevent autoimmune disease, and irradiation of the target organ alone (e.g., stomach) failed to induce autoimmunity; 2) irradiation of both the thymus and the peripheral lymphoid tissues was required for efficient induction of autoimmune disease; 3) TLI depleted mature T cells from the thymus (40) and the periphery, and inoculation of normal mature thymocytes/T cells effectively prevented the autoimmune disease; and 4) similar organ-specific autoimmune diseases including gastritis, thyroiditis, and orchitis can be induced in normal mice by preparing a similar immunologic condition as in TLI without manipulating the target self-Ags, e.g., transplanting x-irradiated thymi into syngeneic athymic nude mice or mice depleted of T cells by Tx and x-irradiation at a T cell-depleting dose (12).

The finding that Tx and subsequent TLI could produce autoimmune disease indicates that TLI elicits pathogenic self-reactive T cells from the peripheral T cell pool. Penhale et al. (41, 42) reported a similar finding with rats: Tx in young adults and subsequent four times of fractionated whole body irradiation (2.5 Gy each every two weeks) induced chronic autoimmune thyroiditis and pancreatic insulinitis/insulin-dependent diabetes mellitus in selected strains of rats. The irradiation on non-Tx rats elicited marginal pathologic changes in the thyroid (41). In our experiments, the incidence/severity of autoimmune disease in Tx-TLI mice was significantly lower than non-Tx mice given TLI (Fig. 4). In the non-Tx TLI mice, the T cells regenerating from the irradiated thymus may predominantly contain the pathogenic self-reactive T cells due to insufficient clonal deletion or defective production of mature CD4⁺ cells with autoimmune preventive activity (see below) or both (12); the self-reactive T cells thus produced

PVG strain) even with shielding of the thyroid gland from irradiation (N. Sakaguchi and S. Sakaguchi, unpublished data). Further immunologic and genetic study of this radiation-induced autoimmune disease in humans (61, 62) and animals would contribute to our understanding of the roles of environmental, immunologic, and genetic factors in autoimmune pathogenesis.

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