The role of oestrogen in the pathogenesis of obesity, type 2 diabetes, breast cancer and prostate disease

Graeme P. Williams

A detailed review of the literature was performed in a bid to identify the presence of a common link between specific hormone interactions and the increasing prevalence of global disease. The synergistic action of unopposed oestrogen and leptin, compounded by increasing insulin. cortisol and xeno-oestrogen exposure directly initiate, promote and exacerbate obesity, type 2 diabetes, uterine overgrowth, prostatic enlargement, prostate cancer and breast cancer. Furthermore these hormones significantly contribute to the incidence and intensity of anxiety and depression, Alzheimer's disease, heart disease and stroke. This review, in collaboration with hundreds of evidencebased clinical researchers, correlates the significant interactions these hormones exert upon the upregulation of p450 aromatase, oestrogen, leptin and insulin receptor function; the normal status quo of their binding globulins; and how adduct formation alters DNA sequencing to ultimately produce an array of metabolic conditions ranging from menopausal symptoms and obesity to Alzheimer's disease and breast and prostate cancer. It reveals the way that poor diet, increased stress, unopposed endogenous oestrogens, exogenous oestrogens, pesticides, xeno-oestrogens and leptin are associated with increased aromatase activity, and how its

products, increased endogenous oestrogen and lowered testosterone, are associated with obesity, type 2 diabetes, Alzheimer's disease and oestrogenic disease. This controversial break-through represents a paradigm shift in medical thinking, which can prevent the raging pandemic of diabetes, obesity and cancer currently sweeping the world, and as such, it will reshape health initiatives, reduce suffering, prevent waste of government expenditure and effectively transform preventative medicine and global health care for decades. *European Journal of Cancer Prevention* 19:256–271 © 2010 Wolters Kluwer Health | Lippincott Williams & Wilkins.

European Journal of Cancer Prevention 2010, 19:256-271

Keywords: benign prostatic hyperplasia, breast cancer, disease prevention, global health initiative, leptin, obesity, oestrogen, prostate cancer, type 2 diabetes mellitus

Medical Practice, Clinical Research, Noosa Heads, Queensland, Australia

Correspondence to Dr Graeme P. Williams, BSc, BAppSci, MBBS (Mon), Medical Practice, Clinical Research, PO box 1574, Noosa Heads, QLD 4567, Australia

Tel: +61417647732; fax: +61754556577; e-mail: graemewilliams8@bigpond.com

Received 13 November 2009 Accepted 17 February 2010

Introduction

Considering that the incidence of obesity, type 2 diabetes, dementia, prostate cancer and breast cancer is now reaching pandemic levels in westernized economies, a detailed review of the literature was carried out in a bid to identify a common link between hormone interactions and the pathogenesis of these prevalent diseases.

In the USA, Canada, Australia and Britain the incidence of obesity is more than 24%, and their combined obesity/ overweight incidence rate has exceeded 60% (World Health Organization, 2004–2006), such that obesity and type 2 diabetes are currently causing more deaths than any recent war or flu epidemic.

By 2025, 380 million people, one in 14, are expected to suffer from type 2 diabetes, the world's fastest growing chronic disease (International Diabetes Foundation, 2007).

Anxiety, depression, hypertension, heart disease, cardiovascular disease, cerebral vascular disease and stressrelated illnesses are growing at a higher rate than most economies. About 80% of women and men suffer from benign uterine and prostate conditions; and breast cancer and prostate cancer, respectively, are now the most common cancers in the world their rates increasing by 31% in each decade.

This detailed analysis reveals an overwhelming plethora of evidence, as to how oestrogen, leptin, insulin, xeno-oestrogens and stress hormones interact to directly and indirectly cause our '21st century pandemic' and its insidious progression from normal function to symptomatic illness, to established disease, and ultimately, to premature death.

Hormone interaction Normal hormone function Oestrogen

Essential for normal fertility, it stimulates endometrial and myometrial growth, increases subcutaneous fat distribution, increases neuromuscular action potentials and muscle excitability, increases ductal breast growth, produces secondary sexual characteristics and increases salt and water retention (Ganong and Lange, 2005).

DOI: 10.1097/CEJ.0b013e328338f7d2

Oestrogen is normally produced by ovarian follicles, adipose cells and breast tissue in women, and by adipose cells and prostate tissue in men.

Leptin

This adipocytokine, a peptide released by adipose cells, is often considered a satiety hormone in excess to restrict eating by reducing neuropeptide Y action on the paraventricular nucleus of the hypothalamus. It probably acts more as an appetite stimulant in humans under conditions of leptin deficiency or leptin nonrecognition, to increase neuropeptide Y release and to stimulate eating.

Insulin

In normality, the consumption of high glycaemic-product carbohydrates, produce increased insulin secretion and sensitivity necessary to convert glucose to glucagon, increase fatty acid oxidation, reduce lipid accumulation in muscle and to assist the entry of glucose into cells.

Stress hormones

The adrenal secretion of corticosteroids, glucocorticoids, cortisol and adrenaline in the short-term normal stress response increases glucose availability to drive the 'fight or flight' mechanism by increasing gluconeogenesis and increasing hepatic glycogenolysis; with protracted action, they increase mobilization of amino acids from skin, bone and muscle and increase visceral adiposity.

Progesterone

Principally produced by the corpus luteum in sufficient quantity at normal ovulation to adequately oppose the growth and irritant properties of endogenous oestrogen. It also inhibits the action of p450 aromatase, the enzyme which catalyses the aromatization of testosterone (and androstenedione, ASD) into endogenous oestradiol (and oestrone) in adipose, breast and prostate tissue. In early reproductive life, late reproductive life and under times of stress, there is often insufficient progesterone production at ovulation to adequately oppose the normal oestrogen present.

During 'ovulopause', the absence of ovulation (whether it be caused by follicle exhaustion, severe stress, contraceptive pill use, the absence of ovaries or being male), no progesterone is produced to offset the normal endogenous production of oestrogen, and so the physiological growth and irritant effects of unopposed oestrogen become evident - the so-called 'symptoms of menopause and andropause'.

The symptomatic progression

Although the rate and degree of metabolic change varies among individuals as a consequence of their in-utero and in-vivo hormone exposures, their genetic backgrounds, predispositions, physical activity and dietary habits, the

process toward symptomatic progression, in general, follows the same path – the cyclic upregulation of p450 aromatase to produce more intracellular oestradiol.

Any compound that upregulates aromatase will increase unopposed intracellular bioactive oestradiol concentrations, and as such will increase the incidence of oestrogen-mediated growth disorders.

The process in brief

The presence of an increasing concentration of unopposed endogenous oestradiol (whether it be associated with the reduced progesterone production of impaired ovulation or by the gradual increase in adipose cell production of oestradiol in men and women) upregulates the production of 11β-hydroxysteroid dehydrogenase 10fold (Dieudonne et al., 2006), (the enzyme that catalyses the conversion of inactive cortisone to active cortisol), which increases intracellular cortisol, and in turn, upregulates aromatase another 9-fold (Thorton et al., 2006).

Although daily stress increases circulating cortisol to accentuate aromatase upregulation, its action in stimulating the increased consumption of palatable carbohydrates (Dallman et al., 2007) is far more significant as it provokes an insulin response which upregulates aromatase a further 6-fold (Samad, 2007).

With further stress and continued eating, the upregulated aromatase creates even more oestradiol, to further upregulate 11β-hydroxysteroid dehydrogenase, which cyclically amplifies the production of more cortisol and further oestradiol to stimulate oestrogen receptors and accentuate growth in oestrogen-sensitive, aromataseactive tissues - fat, breasts, womb and prostate.

The process in detail

Adipocyte-derived leptin and oestradiol both upregulate 11β-hydroxysteroid dehydrogenase (Dieudonne et al., 2006; Paulsen et al., 2007), the enzyme that converts inactive cortisone to bioactive cortisol, to increase the size and number of adipocytes and to increase both intracellular and circulating cortisol. In turn, the cortisol upregulates adipocyte p450 aromatase 9-fold (Thorton et al., 2006), the product of which further upregulates itself (Kinoshita and Chen, 2003) and positively feeds back to cyclically upregulate 11β-hydroxysteroid dehydrogenase 10-fold (Dieudonne et al., 2006), and to stimulate increased leptin production.

The oestrogen (Santner et al., 1997; Dieudonne et al., 2006) and leptin (Daghestani et al., 2007; Geisler et al., 2007), both upregulate p450 aromatase to further increase extra-glandular oestrone and oestradiol production (Catalano et al., 2003; Dundar et al., 2005; Dieudonne et al., 2006) from ASD and testosterone, and potentiate the action of oestradiol on oestrogen receptor- α (ER- α)

(Catalano et al., 2004; Sulkowska et al., 2006; Cirillo et al., 2008) to stimulate breast tissue growth and to further increase subcutaneous fat deposition (Shin et al., 2007) (Fig. 1).

The physical effects of illness and the psychological stress associated with the concern of being loved and accepted by family and peers, compounded by work pressure, financial stress and the perceived urgency of daily living, all contribute to an ever-increasing burden of stressors in the community.

Chronic stress increases palatable food intake (Dallman et al., 2007), including foods that contain 30% sucrose (like many breakfast cereals, cakes, biscuits and confectionary), which increases the output of hypothalamic corticotropin releasing hormone (Foster et al., 2009), pituitary adrenocorticotrophic hormone and adrenal corticosterone to promote further palatable eating and to provoke a normal insulin response, which in turn, increases leptin output (Thomas et al., 2000; Falconnier et al., 2003; Manderson et al., 2003; Lindsay et al., 2004). and upregulates aromatase (Samad, 2007) and oestrogen receptors (Oestrogen Receptors, 1998) to increase subcutaneous fat deposition (Shin et al., 2007), and reduce lipolysis (Pedersen et al., 2004). In effect, this builds the total fat mass to further increase leptin (Castracane et al., 1998) and adipocyte oestrogen output (Vona Davis et al., 2007), which initiates the process of impaired leptin receptor signalling (Kellerer et al., 2001; Strat et al., 2005; Garofolo et al., 2006; Ishida-Takahashi et al., 2006).

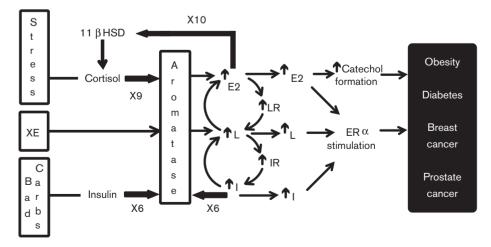
As obesity increases in men and women, there is overexpression and increased activation of 11B-hydroxysteroid dehydrogenase (Paulsen et al., 2007) within local adipose tissue [independent of plasma cortisol levels (Espindola-Antunes and Kater, 2007)], which accentuates intracellular cortisol production and further upregulation of aromatase. In turn, circulating cortisol levels increase, which affects the development of dysmorphic features, central obesity, raised blood pressure, insulin resistance and dyslipidaemia.

The rising intracellular oestrogen concentration stimulates increased leptin production by fat cells (Tanaka et al., 2001), increases adipogenesis by ER-α stimulation and altered Janus kinase (JAK) interactions (Iguchi et al., 2008) and inhibits leptin receptor expression (Ishida-Takahashi et al., 2006; Revillion et al., 2006) by altering downstream signalling and the signal transducer and activator transcription factor (JAK STAT) mechanisms. Moreover, the rising oestrogen inhibits insulin receptor function to directly cause insulin resistance and hyperinsulinaemia (Hilf and Crofton, 1985; Gonzalez et al., 2002; Garcia-Arencibia et al., 2005).

In turn, the increasing insulin level attenuates STAT 3 to create even more leptin resistance (Banks et al., 2000; Kellerer et al., 2001; Fujita et al., 2003; Ishida-Takahashi et al., 2006), and increases arcuate neuropeptide Y release to stimulate more unnecessary eating (Insulin Upregulates Arcuate NPY Release); even to the extent of unpalatable gorging.

The synergistic action of leptin in the presence of increased insulin directly targets the corpus luteum (Brannian et al., 1999), which impairs fertility, reduces progesterone output, increases relative oestrogen excess and stimulates increased follicle stimulating hormone (FSH) (Arnaoutoglou et al., 2008; Nakamura et al., 2009), which in combination further upregulate p450 aromatase.

Fig. 1



The role of oestradiol, leptin, cortisol and insulin in the pathogenesis of disease. Carbs, refined carbohydrates; E2, estradiol; ER-α, oestrogen receptor; HSD, hydroxysteroid dehydrogenase; I, insulin; IR, insulin resistance; L, leptin; LR, leptin resistance; XE, xeno-oestrogens.

In essence, the complex synergies between oestradiol, leptin, insulin and cortisol are directed towards the cyclic upregulation of p450 aromatase in adipose cells, uterine cells, prostate cells and breast cells, which significantly increases the production of intracellular oestradiol [a measurement that is far more accurate than serum oestradiol concentrations (Lonning and Geisler, 2008). as intracellular oestradiol levels can be 10-50 times higher than serum concentrations (Van Landeghem et al., 1985)] to increase ER-α stimulation (Cohen, 2008), to stimulate cell growth in breast and prostate tissue, to increase leptin levels (Yi et al., 2008) and to increase subcutaneous fat deposition and insulin resistance (Shin et al., 2007).

The rising oestradiol (Revillion et al., 2006) and insulin (Ishida-Takahashi et al., 2006) levels impair leptin receptor function, and in turn, the resultant hyperleptinaemia (Hennige et al., 2006), and unopposed oestradiol (Gonzalez et al., 2002) subsequently impair insulin receptors, which only serves to create more insulin and more leptin, and so on; and so the vicious circles turn faster as their synergistic actions insidiously escalate.

As fat deposition increases – the more leptin and oestradiol concentrations increase - the faster the progression from overweight to obesity. When the fat cells reach maximal size, the oestradiol and leptin stimulate ectopic fat deposition to induce fatty liver and intramuscular lipid storage (Dyck et al., 2006; Daghestani et al., 2007). They increase noradrenaline formation, sympathetic activity (Pausova, 2006), IL-6 and TNF-α to increase vascular resistance, sodium retention and to induce hypertension (Bogaert and Linas, 2009), and they mediate inflammatory reactions to produce hepatic C-reactive protein (Bastard et al., 2006), endothelial inflammation, atherosclerotic change (Knudson, 2007) and heart disease (Martin et al., 2008; Lieb et al., 2009).

Although xeno-oestrogens are relatively weak (Pugh and Moore, 1998), their cumulative exposure in humans from food, water, plastics, pharmaceuticals and cosmetic products synergistically increase p450 aromatase expression (Kinoshita and Chen, 2003; Laville et al., 2006) to act as an initiator of significantly raised intracellular oestrogen production in adipose (Newbold et al., 2008), uterine, prostate and breast tissue. Although oestradiol and DDT [1,1,1,trichloro-2,2-bis[chlorophenyl]ethane] both increase the growth of MCF-7 human breast cancer cells, oestradiol is 200-300 times more potent (Dees et al., 1997).

As such, rather than merely adding to the oestrogenic load, xeno-oestrogens upregulate aromatase to produce much more, highly potent, intracellular oestradiol, which in turn initiates, promotes and exacerbates oestrogenic growth disorders, and when symptoms persist, disease commences.

Oestrogenic disease

Obesity

Obesity is associated with diabetes, heart disease, breast cancer, Alzheimer's disease, hysterectomy frequency and prostate cancer.

Oestrogen increases the deposition of ectopic fat in liver and muscle, visceral fat in and about organs, and subcutaneous fat accumulation especially on the hips, upper thighs and lower abdomen. The amount of abdominal fat is proportional to its amount of aromatase, the intracellular enzyme that converts testosterone (and ASD) to oestradiol (and oestrone), which in essence increases endogenous oestrogen levels at the expense of testosterone levels.

As the increasing fat cells make more oestradiol (Carr and Bradshaw, 2005), they increase their leptin output (Tanaka et al., 2001), which in turn upregulates aromatase, drives insulin and leptin resistance and stimulates the production of more cortisol.

In essence, fat cells make leptin and oestrogen. Leptin, insulin, cortisol and oestrogen all increase aromatase activity. In turn, leptin and oestrogen increase fat deposition and inhibit insulin receptors. So, the body compensates by making more insulin, which in turn increases oestrogen receptor number and activity (Kaaks, 2005).

The increased oestrogen, increased insulin and raised stress hormone cortisol directly cause leptin receptors to fail – in other words, the satiety hormone leptin is not recognized by the receptors, as if no leptin was present so in the absence of satiety, the body is stimulated to eat more high-carbohydrate, low-nutrition, insulin-provoking foods, which deposit even more fat, with even higher aromatase (Catalano et al., 2003) activity, which makes more leptin and more oestrogen to deposit more fat, which makes more oestrogen, which increases leptin, and increases insulin and increases cortisol to make more fat, and so on.

Obesity is associated with increased oestrogen production, and body mass index (BMI) is strongly correlated with each of the oestrogens in obese postmenopausal women (Key et al., 2003).

Obesity and subcutaneous fat deposition are associated with an increased ER- α to ER- β ratio (Shin *et al.*, 2007); in turn, higher fat percentages and increased hip circumferences are strongly correlated with increased leptin levels (Ho et al., 1999), hyperinsulinaemia (Borugian et al., 2003) and breast cancer.

Type 2 diabetes

Endogenous oestrogens, exogenous oestrogens and xenooestrogens, in concert with leptin, impair insulin receptor function to initiate insulin resistance.

This is the missing link between obesity and diabetes. Low-nutrient, high glycaemic-product carbohydrate diets, ongoing stress, reduced exercise and inherited predisposition only compound the problem.

Raised levels of oestradiol, seen in obesity, oral contraceptive use and injudicious hormone replacement therapy (HRT) promote insulin resistance (Livingstone and Collison, 2002), impair insulin sensitivity (Gonzalez et al., 2002), inhibit human insulin receptor gene expression in a dose and time-dependent manner (Garcia-Arencibia et al., 2005), downregulate insulin receptor function and reduce insulin receptor numbers (Hilf and Crofton, 1985) to directly cause insulin resistance and promote type 2 diabetes.

Even triphasic oral contraceptive use increases insulin levels and reduces insulin sensitivity, such that 10% of women develop impaired glucose tolerance after 6 months of use (Petersen, 2002).

In essence, reactive hyperinsulinaemia, subsequent to the consumption of high glycaemic-product foods, upregulates oestrogen receptors, increases leptin levels, accentuates leptin resistance, upregulates 11β-hydroxysteroid dehydrogenase 1 to increase cortisol production, and upregulates aromatase (Samad, 2007) to lower testosterone, increase oestradiol output, increase fat deposition and directly promote type 2 diabetes - a condition closely associated with increased breast cancer, prostate cancer, obesity and heart disease.

Insulin increases serum leptin levels (Thomas et al., 2000; Falconnier et al., 2003; Manderson et al., 2003; Lindsay et al., 2004), inhibits leptin receptor signalling (Kellerer et al., 2001) and promotes leptin resistance (Banks et al., 2000; Fujita et al., 2003) through attenuation of STAT 3 phosphorylation of JAK-2 on the serine 523 (Ser 523) residue (Ishida-Takahashi et al., 2006).

The leptin receptor, LRb, depends upon tyrosine kinases of the JAK family to mediate intracellular signalling, but phosphorylation of Ser 523 on JAK-2 by insulin itself alters downstream signalling, affects nuclear translocation and ultimately ameliorates target gene expression, and hence causes leptin resistance and subsequent hyperleptinaemia.

The excess leptin further downregulates insulin action to inhibit insulin signalling by promoting the phosphorylation of the serine 318 residue in insulin receptor substrate (Hennige et al., 2006), such that in type 2 diabetes, raised serum leptin levels are directly proportional to the degree of insulin resistance, independent of body mass (Kaur and Zhang, 2005).

Alone, the presence of excess insulin increases the synthesis and activity of oestrogen receptors (Oestrogen Receptors, 1998; Kaaks, 2005) to increase the growth and irritant effects of oestradiol. Even insulin at 10⁻⁶ mol/l significantly increases breast cancer cell growth (Garnier et al., 2003).

Alone, oestradiol (Santner et al., 1997; Kinoshita and Chen, 2003) and leptin (Catalano et al., 2003; Cirillo et al., 2008) both upregulate aromatase while insulin alone upregulates aromatase 6-fold (Samad, 2007). Together, oestradiol and leptin are synergistic (Dundar et al., 2005), and in combination with insulin, they increase cortisol production 10-fold in women (Dieudonne et al., 2006). which in turn, as mentioned earlier, acts to further upregulate aromatase another 9-fold (Thorton et al., 2006).

So in combination, the raised levels of oestradiol, leptin, insulin and cortisol produce cyclic amplification and upregulation of cytochrome p450 aromatase activity, which acts in the final stage of the steroid cascade to convert testosterone to oestradiol, to raise oestradiol levels and lower testosterone levels.

This pattern of raised free oestradiol levels (Ding et al., 2006) and low testosterone levels (Dhindsa et al., 2004; Osuna et al., 2006) is commonly seen in type 2 diabetes. Although 64% of men with type 2 diabetes have hypogonadism (Kalyani and Dobs, 2007), men with higher testosterone levels have a 42% lower risk of type 2 diabetes (Ding et al., 2006).

Similarly, women with lower free oestradiol levels have an 80% lower risk of developing type 2 diabetes [relative risk (RR) 0.20], and men with lower free oestradiol levels have a 52% lowered risk (RR 0.48) (Ding et al., 2006).

Reversal of this situation by short-term aromatase inhibition has been shown to essentially double testosterone, halve oestradiol and to reduce both serum insulin and leptin levels (Lapauw et al., 2009); in turn, physiological testosterone supplementation improves glucose homeostasis (Boyanov et al., 2003), and decreases insulin resistance and the risk of developing diabetes (Fukur, 2003).

In essence, the production of excess endogenous oestradiol and leptin from abundant adipose tissue increases cortisol, and stimulates simple carbohydrate consumption to increase insulin production. In combination, they significantly upregulate aromatase to produce even more oestradiol, and together with resultant leptin resistance, insulin resistance and excess cortisol, they promote type 2 diabetes, further obesity, heart disease, and oestrogenic breast, uterine and prostate overgrowth conditions.

Anxiety/depression

Although it is normal for humans to suffer stress from unsatisfactory life experiences, it is the neurogenic effect of unopposed oestrogen that creates the tipping point for progressive anxiety and depression.

The normal stress response triggers the release of adrenaline and cortisol, the short and long-acting adrenal

hormones necessary to activate the 'fight or flight' mechanism; however, it is their combined action with oestradiol, itself a highly neuroactive steroid, which increases the frequency of neuronal action potentials, increases synapse numbers, increases dendritic connections and promotes neuronal irritability and noradrenaline release to stimulate heart rate, sweating and blood pressure (Ganong and Lange, 2005), the hallmark of anxiety and hot-flush symptoms.

Unopposed oestradiol in women, and increasing oestradiol in men, together with increased leptin production, increase noradrenaline release (Leal-Cerro et al., 2001), which encourages carbohydrate consumption (Thomas et al., 2000) and insulin release to promote depression and further fat deposition.

Under prolonged stress, cortisol and oestrogen suppress and activate specific reactions in the normal hormone cascade to preferentially increase the production of even more cortisol and more oestrogen, which only serves to exacerbate symptoms further.

Chronic stress increases glucocorticoid production, which mobilizes peripheral fat stores, increases the production of more oestradiol and leptin (Dallman et al., 2007), reduces the ER-α to ER-β ratio to redirect central fat deposition (Thorton et al., 2006) and stimulates further palatable eating and increased insulin release.

In turn, the increased insulin accentuates ER-α function (Oestrogen Receptors, 1998; Kaaks, 2005) to increase leptin production (Yi et al., 2008) and subcutaneous adipogenesis (Iguchi et al., 2008), and in combination they increase aromatase to create more neurogenic oestradiol and anxiety, and further deplete testosterone levels and energy.

Although the natural process of ageing lowers testosterone levels, any associated aromatase upregulation by oestradiol, xeno-oestrogens, leptin, insulin and cortisol only serves to deplete testosterone levels faster (and to increase oestradiol output), such that women in their 40s have half the testosterone of a woman in her 20s (Goldstat et al., 2003), and 60-year-old obese women have half the testosterone of 'normal sized' 60-year-old women (Padero et al., 2002).

Recently, nearly 4000 men with lower testosterone levels associated with ageing revealed an increased incidence of depression, with those in the lowest quartile having a 300% increased risk of depression (Almeida et al., 2008).

Studies that supplement testosterone to restore waning physiological levels show reduced depression (Margolese, 2000), improved mood (Burris et al., 1992), a significant improvement in Hamilton Depressive ratings (Pope et al., 2003) and a reduction of anxiety in male patients (Cooper, 2000). Similarly, postmenopausal women given judicious physiological testosterone supplementation

showed return of normal physical and psychological function (Padero et al., 2002), and premenopausal women showed improvement of mood, libido and wellbeing, without detrimental side effects (Goldstat et al., 2003).

Heart disease - cerebrovascular disease

Excess oestrogen, reduced testosterone, increased cortisol, raised insulin levels and accentuated adrenergic responses seen in obesity, diabetes and anxiety, all contribute to increase the incidence of hypertension, myocardial infarction and cerebrovascular accidents.

Excess energy intake and subsequent fat storage increases leptin and endogenous oestradiol production, and reduces testosterone levels through aromatase activation to facilitate the formation of large dysfunctional adipoctyes that produce angiotensin, leptin and proinflammatory cytokines, which stimulate ectopic fat deposition in the skeletal muscle and liver sites (Pausova, 2006), activate hepatic C-reactive protein to promote cardiovascular disease (Bastard et al., 2006), activate sympathetic drive, increase vascular resistance and increase sodium retention to produce obesity-related hypertension (Bogaert and Linas, 2009).

Leptin affects endothelial function and coronary circulation (Knudson, 2007) to induce atherosclerotic disease, and as leptin resistance increases, so does the risk of heart failure, coronary heart disease (CHD), cerebrovascular disease and overall mortality (Lieb et al., 2009). Although oestrogen is associated with an improved lipid profile and has been long-considered atheroprotective, increased local oestradiol production and aromatase expression have now been identified in fibroatheromatous placques, adjacent smooth muscle and in thickened human aortic intima (Murakami et al., 2001), which serve to facilitate plaque formation, intimal disruption and to increase thromboembolic risk.

Increased FSH, increased leptin and preexisting unopposed oestradiol also increase CHD risk in surgically induced and physiological menopause (Verhoeven et al., 2009).

Oestrogen has been controversially associated with an increased risk of thromboembolism, progressing to coronary occlusion, cerebrovascular infarcts and pulmonary emboli, for years. The Women's Health Initiative Study of 2002 (Anderson et al., 2002) found long-term combination conjugated equine oestrogen plus medroxyprogesterone acetate HRT use to be associated with a 29% increase in heart attack, a 41% increase in stroke and twice the risk of serious blood clots. Although recent follow-ups suggest that transdermal HRT application is safer than oral dosing, nevertheless, the National Institute of Health and the Food and Drug Administration state that HRT should not be used to prevent CHD, as it does not reduce the incidence of coronary artery

disease or slow the progression of established artery disease (Lauer, 2008), that it increases the risk of stroke at any time, and that oral HRT can double the risk of thromboembolism in the first year of use (Canonico et al., 2008).

Leaving HRT aside, increasing leptin, cortisol, insulin, xeno-oestrogen exposure, and increasing oestradiol itself, maintain ongoing aromatase upregulation to further increase oestradiol and to significantly deplete circulating free testosterone levels.

An independent inverse relationship exists between low levels of testosterone and the presence of atherosclerosis in men (Murakami et al., 2001; Hak et al., 2002) and coronary artery disease in women (Kaczmarec et al., 2003). the effects of which are reversed by physiological testosterone supplementation, which increases coronary flow and produces coronary artery dilatation (Webb et al., 1999), improved angina threshold and exercise tolerance (Channer and Jones, 2003), improved muscle strength, and generally slowed the ageing process (Allen et al., 2007), without deleterious effects.

Dementia and Alzheimer's disease

The incidence of Alzheimer's disease, the most common form of dementia, is currently doubling every 20 years (Ferri et al., 2005).

High serum oestradiol levels in women (Ravaglia et al., 2007) and low testosterone levels in men (Pike et al., 2006) are independent predictors for dementia and Alzheimer's disease.

In men and women, obesity and raised BMI are associated with an increased risk of Alzheimer's disease (Luchsinger and Gustafson, 2009) and an accelerated rate of brain atrophy, such that obese nondementia individuals suffered 8% more brain atrophy over 5 years than lean individuals (Raji et al., 2009).

The raised endogenous oestradiol of obesity and the hyperinsulinaemia of type 2 diabetes are closely linked to a significant increase in the risk of Alzheimer's disease in men and women (Luchsinger and Gustafson, 2009), whereas oestrogen-containing HRT essentially doubled the incidence of dementia in the Women's Health Initiative Memory Study (Craig et al., 2005).

As such, the cumulative effect of upregulated aromatase, increased unopposed endogenous oestradiol and any xeno-oestrogen exposure, together with the consequential reduction of free testosterone associated with aromatase conversion, raised sex hormone binding globulin levels and a naturally lowered output, directly induce progressive β-amyloid accumulation and consequent dementia, in both men and women.

Interestingly, while raised endogenous oestrogen and prescribed HRT oestrogen both increase Alzheimer's

disease, androgen depletion also accelerates the abnormal accumulation of β-amyloid in specific areas of the brain (Pike et al., 2006) to produce Alzheimer's disease-like neuropathy (Rosario et al., 2006). Lower free testosterone levels, a better marker of androgen loss, are associated with reduced processing speed and impaired executive function, whereas higher testosterone levels are associated with improved cognitive function (Muller et al., 2005).

Significantly, the restoration of physiological levels of testosterone in hypogonadal men reversed the degree of dementia while those on placebo continued to deteriorate (Tan and Pu, 2003).

Benign uterine disease, endometrial and ovarian cancer

Oestrogenic compounds promote endometrial proliferation, and promote progression to endometrial hyperplasia, atypia and neoplasia (Vainio et al., 1992).

Endometrial cancer and exogenous oestrogens have been linked since 1951, such that conjugated equine oestrogens and oestradiol both confer an 8-fold increase in the RR of endometrial cancer (Beresford, 1997). Even physiological levels of oestradiol and oestrone, without the opposing effects of progesterone, have been shown to significantly increase the risk (Overall Evaluation of Carcinogenicity, 1987).

Unopposed oestradiol stimulates cyclo-oxygenase-2 to increase the formation of prostaglandin-E2, which promotes aromatase expression in endometrial tissue (normally devoid of aromatase activity), stimulates dysfunctional endometrial growth in endometriosis (Bulun et al., 2005) and increases the production of tumourigenic, hydroxylated oestradiol metabolites, which promote the formation of uterine leiomyomata (Leihr et al., 1995). The 4-hydroxyoestrogens form unstable adducts to induce DNA damage, and transform mutations to cause endometrial and ovarian cancer (Emons et al., 2002), such that ovarian cancer cells display increased cell proliferation and reduced apoptosis, even at physiological concentrations of oestradiol (Seeger et al., 2005). A dosedependent relationship exists between annual exposure to oestrogenic HRT and an increased risk of ovarian cancer (Greiser et al., 2007); similarly, any exposure to oestrogenic HRT increases the risk of ovarian cancer by 50% (Zhou et al., 2008). In fact, the World Health Organization International Agency for Research and Cancer, which included oestrogenic HRT and oestrogenic oral contraception on their 2005 carcinogenic list, added the oestrogenic HRT/ovarian cancer relationship to its known carcinogenic agent list in 2009 (Grosse et al., 2009).

The use of HRT increases the risk of developing and dying of ovarian cancer by 20%, and increases the total incidence of endometrial, breast and ovarian cancer, which represent 40% of all UK cancers, by 63% (Beral, 2007).

Benign prostatic hyperplasia

Oestrogens initiate and promote benign prostatic hyperplasia (BPH) (Hammarsten et al., 2009) through upregulation of prostatic aromatase (Risbridger et al., 2003) to raise intracellular oestradiol levels, increase subsequent 3,4-catechol oestradiol formation (Leihr et al., 1995) and to promote periurethral prostatic adenomatous hyperplasia typically in the transitional zone.

Although BPH has long been attributed to an excess of testosterone, there is no clinical evidence that the incidence of BPH or even prostate cancer increases with testosterone replacement therapy (Morley, 2000); in fact, multiple studies have failed to show any exacerbation of symptoms attributable to BPH during testosterone supplementation (Rhoden and Morgentaler, 2004).

Although it is quite paradoxical to consider oestrogen, and not testosterone, as the cause of prostate disease, it is nevertheless quite understandable, when one considers that prostate disease typically occurs at a stage in a man's life when his testosterone levels are low.

If testosterone is the cause of prostate cancer, then we should have got it when we were young men, and not at a time in our life when our testosterone is at its lowest (Williams, 2006).

Prostate cancer

Prostate cancer, detected in 30% of 50 year-old men in random autopsy studies (Bosland et al., 2002), is now the most common male cancer in many western countries.

Prostate cells, both normal and malignant, contain all the essential enzymes necessary to convert dehydroepiandrosterone to oestradiol, in particular 17β-hydroxysteroid dehydrogenase and aromatase, and they also exhibit ER-α and ER-β, with ER-α being essentially proliferative, and ER-β in the main inhibitory (Carruba, 2007). Testosterone is metabolized by 5α-reductase to form the more potent androgen, dihydroxytestosterone, which in turn activates the androgen receptor.

It seems that the principal role in cell proliferation and metaplastic change rests with the prostatic activation of 17β-hydroxysteroid dehydrogenase, and the expression and upregulation of aromatase, both of which increase intracellular prostatic oestradiol levels (through estrone conversion and androgen aromatization, respectively) to produce divergent activation of the proliferative ER-α more than the inhibitory ER-β (Carruba, 2006).

ER-α activation is essential for BPH, inflammation and prostate cancer to occur while ER-β stimulation reduces prostatic hypertrophy, inflammation and prostate cancer (Ellem and Risbridger, 2007). A reduction in ER-β expression, and hence reduced growth inhibition, has been noted in prostate cancer (Bardin et al., 2004), with some studies revealing that ER-β expression progressively declines in localized prostate cancer, as Gleason scores increase from prostatic intraepithelial neoplasia to advanced prostate cancer (Prins and Korach, 2008).

The days of blaming testosterone alone for prostate cancer have passed. Although it may play a minor role through androgen receptors, it is intracellular oestradiol that directly induces the proliferation of aberrant prostatic basal cell hyperplasia to promote the formation of squamous metaplasia (Risbridger et al., 2001, 2007). The ratio between free testosterone and free oestradiol seems to determine both the extent of aromatase upregulation through nuclear and nonnuclear ERs, and the degree of ER- α stimulation (Prins and Korach, 2008).

The weight of evidence postulates that leptin, xenooestrogens, cortisol and oestradiol all upregulate ER-α, aromatase and 17β-hydroxysteroid dehydrogenase to induce proliferative prostatic metaplasia and neoplasia, just as they do in breast carcinogenesis.

Obesity and increased leptin increase the incidence and aggressiveness of prostate cancer (Mistry et al., 2007), and men with a BMI above 30 have a 78% increased risk of high-grade prostate cancer, with a Gleason score above 8 (Gong et al., 2006).

The synergistic action of insulin and leptin, seen in men with higher insulin and waist-hip ratios, have an 8.55-fold increased risk of prostate cancer while a raised insulin alone confers a 2.56-fold increased risk (Hsing et al., 2001).

Raised free oestradiol levels also increase prostate cancer risk (Gann et al., 1996), as do lipophilic, liposoluble, oestrogenic endocrine disruptors, pesticides and environmental carcinogens (Imigaray et al., 2007; Prins, 2008), which include Agent Orange (Chamie et al., 2008), shown to cause prostate cancer sooner in exposed men, with a mean time from exposure to diagnosis of 407 months, and to double the aggressiveness of the cancer, with Gleason scores above 8.

Apart from direct ER activation, the formation of DNA adducts from oxidized electrophilic oestradiol metabolites, in particular estradiol-3,4-quinone, and the generation of excessive reactive oxidizing species, directly induce oxidative modification to DNA bases to form 8-hydroxy-2'-deoxyguanosine and 5-hydroxymethyl-2'deoxyuridine, known prostatic carcinogens (Han et al., 1995; Leihr, 1997).

Low testosterone levels, rather than high testosterone levels, are associated with prostate cancer (Schatzl et al., 2001) with little compelling evidence to suggest that high testosterone, or testosterone supplementation, increases prostate cancer risk (Prehn, 1999; Morley, 2000; Morales, 2002; Rhoden and Morgentaler, 2004).

Lower free testosterone levels are correlated with positive prostate biopsies and high Gleason scores (Gustafsson et al., 1996; Hoffman, 2000; Schatzl et al., 2001), whereas men with higher free testosterone levels have a lower risk of developing prostate cancer (Stattin et al., 2004).

Breast cancer

Anything that upregulates the expression or activity of cytochrome p450 aromatase (or increases the formation of catechol DNA adducts) will increase the risk of breast cancer, whether it be caused by physiological levels of oestradiol, unopposed by progesterone, increased leptin levels, as seen in obesity and leptin resistance, reactive hyperinsulinaemia attributable to injudicious consumption of sugars and high-glycaemic foods, raised cortisol levels associated with chronic stress, or xeno-oestrogen exposure innocently consumed in food, water and skin products or prescribed as medication for humans, stock and farm produce.

Physiological unopposed oestradiol levels and breast cancer: The carcinogenic role of 17β-oestradiol has been confirmed – it induces neoplastic transformation in human breast cancer cells (Russo and Russo, 2006), promotes the growth of breast cancer cells in vivo and in vitro (Dees et al., 1997), 1 nmol/l of 17β-oestradiol induces a 7–13-fold increase in human breast cancer cell numbers (Gupta et al., 1998) and significantly increases the risk of breast cancer (Berrino et al., 1996; Rock et al., 2008), such that unopposed oestradiol alone confers an increased (RR 2.58) risk of breast cancer in post-menopausal women (Key et al., 2002). Even the comparison of women in the upper normal oestradiol range to the lower normal range, revealed an increased RR of 3.3 (Missmer et al., 2004), and in ER-positive breast cancer patients who had never received HRT, those women in the upper quarter of the normal physiological oestradiol range had an almost 5-fold increased risk, compared with those in the lower quarter normal range (Missmer et al., 2004).

Obesity, BMI, percentage fat (Petrelli et al., 2002; Borugian et al., 2003; Key et al., 2003), type 2 diabetes and insulin resistance (Oestrogen Receptors, 1998; Muti et al., 2002; Garnier et al., 2003; Harvard Nurses Health Study, 2003; Kaaks, 2005; Mawson et al., 2005; Ishida-Takahashi et al., 2006), all increase breast cancer risk and mortality. Leptin levels increase remarkably with increasing fat percentage (Daghestani et al., 2007), and so does the ER- α to ER- β ratio (Shin *et al.*, 2007). Leptin interferes with STAT 3 signalling and activates ER-α in malignant breast tissue (Sulkowska et al., 2006); it doubles FSH to upregulate aromatase (Arnaoutoglou et al., 2008), and it interferes with insulin signalling (Kaur and Zhang, 2005) by directly promoting phosphorylation of Ser 318 in insulin receptor substrate to downregulate insulin receptor action (Hennige et al., 2006).

In effect, the increased leptin creates an excess of ineffective insulin, which attenuates STAT 3 phosphorylation, impairs leptin receptor ObR-L function (Banks et al., 2000) and phosphorylates Ser 523 on JAK-2 to inhibit further downstream signalling, nuclear translocation, and receptor gene expression to ultimately amplify the hyperleptinaemia (Ishida-Takahashi et al., 2006), and accentuate the insulin resistance and leptin resistance cycle to increase the risk, incidence and overall mortality of breast cancer.

Oestradiol itself upregulates aromatase activity (Santner et al., 1997), and leptin alone increases aromatase expression (Catalano et al., 2003) to increase intracellular microenvironment oestradiol levels, which induce ER-α (Catalano et al., 2004) and transactivate epidermal growth factor receptors to promote breast cancer invasion and migration (Cirillo et al., 2008). The intracellular levels of aromatase (Lonning and Geisler, 2008) and oestradiol (Van Landeghem et al., 1985) reflect the highly oestrogenic microenvironment of breast cancers better than plasma levels, with intracellular levels being 10–50 times higher than plasma levels.

The cyclic amplification of aromatase by leptin and oestradiol is highlighted by their capacity to increase 11βhydroxysteroid dehydrogenase expression (Catalano et al., 2003), and so increase intracellular cortisol production (Dieudonne et al., 2006), which in turn upregulates aromatase (Thorton et al., 2006), the product of which positively feedsback onto 11β-hydroxysteroid dehydrogenase to complete the cycle, and also increases fat deposition and hence leptin production to upregulate aromatase, increase insulin resistance (Dallman et al., 2007) and stimulate further growth. Leptin and oestradiol combine to potentiate ER-α activation (Catalano et al., 2004) and to enhance cell cycle progression of breast cancer cells (O'Neil et al., 2001; Mawson et al., 2005).

Significantly, unopposed, endogenous, 17β-oestradiol has been shown to induce human breast epithelial cell genomic alterations, at the same locations as other known carcinogens, such as diethylstilboestrol and benzopyrene (Russo et al., 2001). Even at concentrations of 0.1 nmol/l, 17β-oestradiol, 4-hydroxyoestradiol and 16-hydroxyoestradiol display proliferative growth effects in human breast cancer cells (Seeger et al., 2006).

The carcinogenic properties of oestradiol and oestrone are highlighted by their capacity to form catechol oestrogen-3,4-quinones, the ultimate carcinogenic metabolites of excess endogenous and exogenous oestrogens.

Oestrogens can be converted to electrophilic metabolites, particularly the catechol oestrogen-3,4-quinones, oestradiol-3,4-quinone and oestrone-3,4-quinone, which react with DNA to form the depurinating adducts, 4- $OHE_2(E_1)-1-N3$ adenine and $4-OHE_2(E_1)-1-N7$ guanine, which provoke error-prone base excision repair and mutation formation that initiates breast cancer in women and prostate cancer in men from unopposed endogenous oestrogen and synthetic oestrogens (Cavalieri and Rogan, 2006). The 4-hydroxylation pathway producing catechol oestrogen-3,4-quinone is responsible for the genotoxic effects leading to oestrogen-induced cancer (Cavalieri and Rogan, 2004), such that men with prostate cancer and women with breast cancer have increased oestrogenic depurinating adducts in their urine, when compared when healthy men and women (Cavalieri and Rogan, 2006).

Xeno-oestrogens and breast cancer: Pharmacological doses of oestrogen and exposure to environmental oestrogen-like xenoestrogens have been proven to be carcinogenic, teratogenic and embryotoxic (Baan et al., 1999). Environmental oestrogen-like chemicals increase the aromatase sensitivity and expression (Risbridger et al., 2007) of ER-α receptors, supporting the critical role of aromatase upregulation in breast cancer development (Kinoshita and Chen, 2003).

An increased risk of breast cancer has been established with the use of oral contraceptives (Breast Cancer Hormonal Contraceptives, 1996; De Benedetti et al., 1996; Schneider et al., 2005) and HRT (Collaborative Group on Hormonal Factors in Breast Cancer, 1997; Michaeli et al., 2004; Fournier et al., 2005). A 26% increased risk was found in the 18000 postmenopausal women in the Women's Health Initiative prospective study (Anderson et al., 2002), and a 66% increased risk of incident and fatal breast cancer was determined in the Million Women Study (The Million Study Collaborators, 2003). Breast cancer risk has been found to persist for 3 years after cessation of HRT (Heiss et al., 2008).

In June 2005, the WHO International Agency for Research and Cancer classified combination hormone contraception (the oral contraceptive pill) and combined menopausal therapy (HRT) as carcinogenic (Cogliano et al., 2005), and their carcinogenic properties were confirmed by the group in 2009 (Bouvard et al., 2009).

Endocrine disruptors mimic the action of sex hormones (Nilsson, 2000). In particular, the man-made oestrogenic endocrine disruptors, politely referred to as environmental chemicals, exhibit potent, lipophilic, fat-soluble, long half-life, high ER-binding properties that facilitate their spread and residual accumulation through food chains to create an almost irretrievable, global ecotoxic health crisis. This is a human health burden so massive that it significantly contributes to human suffering and loss of productivity worldwide (Earth Summit, 1992). A pandemic so grave and accelerating in nature that it will, if not courageously contained, compromise human fertility and ultimately human survival.

The common commercial chemicals, bisphenol A, butyl benzyl phthalates, dioxin 2,3,7,8-TCDD, cadmium, arsenicals, nonylphenol, octophenol; and the PCB industrial chemicals, endosulfan, lindane, parathion, dieldrin, and DDT insecticides, hexachlorobenzene, maneb and tributyltin fungicides and 2,4,-D, 2,4,5,-T and atrazine herbicides, to name a few, are all endocrine disruptors (Bouvard et al., 2009).

Even ultraviolet filters are a class of endocrine active chemicals, in particular 4-methyl-benzilidine camphor and octyl-methoxycinnamate, shown to display dosedependent oestrogenic action in MCF-7 human breast cancer cells, and to affect oestradiol-regulated genes in the prostate and uterus (Schlumpf et al., 2004).

Although both DDT and endogenous oestradiol (17βoestradiol) promote breast cancer growth in vitro and in vivo (Dees et al., 1997), it should be appreciated that oestradiol is 200-300 times more potent, being capable of inducing change at one femtomol (0.000 000 000 000 001M) (Watson et al., 2008). The extent of human exposure to long-term, low-dose pesticides and their insidious capacity to significantly upregulate intracellular oestradiol production is highlighted by the fact that DDT is applied to crops in litres, and pesticides are applied to edible produce, the skin of animal stock and farm workers, in grams.

In particular, it is not the total accumulated oestrogenic load that has tipped the balance over the last 20 years, it is the intense upregulation of aromatase (through our long-term food and corporate exposure to endocrine disruptors, and the synergistic cyclic amplification of oestradiol, leptin, cortisol and insulin) that has increased intracellular oestradiol production and oestradiol catechol formation, proven to induce cell changes from hypertrophy to neoplasia. Oestrogenic disorders exhibited worldwide today, in the form of obesity, type 2 diabetes, anxiety and depression, cardiovascular disease, benign uterine and prostatic hyperplasia, uterine and prostate cancer and breast cancer (not to mention precocious puberty, genetic malformations and rising infertility), will only increase in incidence, if change is not forthcoming.

Symptom control and disease prevention

In the final stage of the steroid cascade, cytochrome p450 aromatase converts testosterone to oestradiol. As such, any compound that upregulates aromatase will increase intracellular bioactive oestradiol concentrations, and increase the incidence of oestrogen-mediated growth disorders, including obesity, diabetes, uterine overgrowth, BPH, breast cancer and prostate cancer.

Conversely, aromatase inhibition has been shown to lower oestradiol levels by 62%, and to reduce fasting insulin levels by 37% and circulating leptin levels by 24% (Lapauw et al., 2009).

The application of dosed quantities of transdermal progesterone is a cost-effective means of aromatase

inhibition, readily available in most economic regions. Furthermore, it attenuates the deleterious effects of unopposed oestrogen, improves leptin and insulin receptor function, downregulates ER-α action, reduces the incidence of breast cancer and provides a substrate for testosterone synthesis, necessary to naturally restore the waning levels seen in osteoporosis, Alzheimer's disease, depression and type 2 diabetes.

Although oestradiol (Dieudonne et al., 2006), leptin (Dundar et al., 2005; Dieudonne et al., 2006), insulin (Samad, 2007), cortisol (Brueggemeier et al., 2001), FSH (Arnaoutoglou et al., 2008) and xeno-oestrogens (Laville et al., 2006; Nakanishi, 2008; Prins 2008) all upregulate aromatase, physiological levels of progesterone downregulate aromatase (Kinoshita and Chen, 2003), decrease leptin secretion (Cova et al., 2005) and improve leptin receptor expression and signalling (Revillion et al., 2006).

Oestradiol upregulates ER-α, whereas progesterone reduces ER number 5-fold (Iosif and Batra, 1994), and reduces nuclear ER count and activity (Fuentes et al., 1990).

Normal progesterone levels reduce breast cancer risk by 88% (RR 0.12) in premenopause (Kaaks *et al.*, 2005), and halve the risk (RR 0.5) in postmenopausal women (Michaeli et al., 2004). Progesterone also inhibits human breast cancer cell growth by upregulation of p27 gene (Gizard et al., 2005); it downregulates breast cancer IGF-1 growth factor (Yamada et al., 2004), which has been shown to act with leptin to increase invasion and migration of breast cancer (Saxena et al., 2008); and progesterone action can improve the prognosis of those with primary breast cancer (Revillion et al., 2006).

Synthetic progestins increase breast cancer risk (Michaeli et al., 2004) and upregulate aromatase (Xu et al., 2007), and increase leptin (Sagsoz et al., 2009) while natural progesterone does not (Michaeli et al., 2004). Never confuse natural human hormones with synthetic, similarshaped, patented hormones, which have different affinities for receptors and binding globulins, and rarely any capacity for natural metabolic action.

Increased intracellular oestradiol and reduced free testosterone levels are the result of upregulated aromatase. Low testosterone levels are closely related to type 2 diabetes, obesity, prostate cancer, osteoporosis, anxiety and depression, heart disease and accelerated ageing.

The restoration of physiological levels of testosterone (with the appropriate aromatase blockade) increases muscle size, reduces abdominal fat and slows the ageing process (Allen et al., 2007). Although oestradiol increases the progression of Alzheimer's disease (Craig et al., 2005; Ravaglia et al., 2007), testosterone has been shown to improve cognitive performance (Muller et al., 2005) and to reverse its effects (Tan and Pu, 2003). Testosterone reduces the incidence of insulin resistance and metabolic syndrome (Fukur, 2003), and improves leptin function (Söderberg et al., 2001; Horenburg et al., 2008).

Bearing in mind the inverse relationship between low testosterone levels and atherosclerosis in men (Hak et al., 2002) and women (Kaczmarec et al., 2003), testosterone supplementation increases coronary blood flow and coronary artery dilatation (Webb et al., 1999), increases exercise tolerance and reduces angina (Channer and Jones, 2003).

Psychologically, low testosterone has been shown to triple depression in men (Almeida et al., 2008). Physiological testosterone supplementation in men improves mood (Cooper, 2000; Margolese, 2000), increases Hamilton depressive rating scores (Pope et al., 2003) and improves libido and erection quality (Burris et al., 1992), without any adverse association with BPH or prostate cancer (Morley, 2000). In fact, higher free testosterone levels are associated with reduced prostate cancer incidence and aggressiveness (Schatzl et al., 2001; Stattin et al., 2004).

Women aged in their 40s have half the testosterone of those in their 20s (Goldstat et al., 2003). Testosterone supplementation restores physical and psychological function (Padero et al., 2002), and improves mood and wellbeing (Goldstat et al., 2003), without adverse effects.

The restoration of physiological hormone levels requires accurate dosing, whether it be insulin, thyroxine, progesterone or testosterone. If a particular human hormone is deficient, use the very same hormone at an exact dose necessary to restore the natural physiological level.

Unfortunately, the use of vitamins, herbs and massage to 'balance hormones' is unacceptable, as they will never restore natural hormone levels; similarly, the prescription of a 'one dose fits all' hormone is medically inappropriate, bearing in mind the widespread individual variation in body fat percentages, bioavailable oestrogen levels, current medications and coexisting conditions in the community. However, the prescription of an appropriate dose of progesterone, together with dietary modification to avoid the consumption of high glycaemic product, high insulin-producing foods, and some objective exercise advice is easily achievable by most caring medical practitioners – a cost-effective, high-yield, preventative measure that will reduce wasted expenditure and improve global health.

Moderate medium exercise improves insulin action (Park et al., 2005; Franks et al., 2007), and in leptin resistance, it improves both insulin and leptin function through improved signal transduction (Dyck, 2005; Flores et al., 2006), such that even short-term exercise and diet programmes have been shown to improve hormone function (Miller et al., 2008).

The importance of a balanced regimen incorporating dietary modification, moderate aerobic exercise, avoiding exposure to xeno-oestrogens, and the restoration of physiological progesterone (and in some cases testosterone) levels to counteract the excesses of oestrogen, leptin and aromatase cannot be overemphasized.

Discussion

At a time when governments worldwide are struggling to meet the financial demands of burgeoning health systems, a credible opportunity to resoundingly reduce the incidence of obesity, diabetes, depression, breast cancer and prostate cancer is at hand – an opportunity to improve the diminishing health of nations and to significantly reduce government health expenditure.

The upregulation of aromatase in adipose tissue, breast cells and prostate cells by xeno-oestrogens, the stress hormone cortisol, and insulin from injudicious carbohydrate consumption results in increased intracellular oestradiol and leptin concentrations, both of which further upregulate aromatase to create even more intracellular oestradiol through cyclic amplification.

The peer-reviewed evidence of hundreds of clinical researchers has been integrated to reveal, for the first time, the common link between aromatase upregulation by the synergistic actions of oestrogen, leptin, insulin and cortisol, and the pathogenesis of obesity, diabetes, breast cancer and prostate cancer - the 21st century epidemic and how simple preventative measures, including combined aromatase blockade, dietary modification and exercise, can reduce the global incidence of these diseases.

The significance of how stress, poor diet and xenooestrogen exposure combine to significantly increase intracellular oestradiol levels by cyclically upregulating aromatase cannot be underestimated, for it initiates fat deposition, further oestradiol and leptin production, increased anxiety, insulin and leptin resistance, type 2 diabetes and overgrowth conditions of oestrogenic tissues from obesity and uterine/prostate hypertrophy to prostate cancer and breast cancer. This revelation alone will reshape preventative medicine and global health for decades.

Although it is now possible to ameliorate the symptoms of menopause and andropause, and to reduce the risk of individuals developing oestrogenic disease, it is critical that governments acknowledge the deleterious effects that xeno-oestrogen exposure and ultimately cumulative oestrogen levels have upon global health, if we are to prevent the progressive wave of oestrogen-related disease that threatens the future health of nations.

Acknowledgements

There are no grants, fundings, and disclaimers for this paper. Dr Graeme P. Williams is the sole author of this paper. No medical writers or editors have contributed to

this study. Dr Graeme P. Williams confirms that he has total access to all aspects of this research paper, and that he holds total responsibility for the decision to submit this paper for publication. Conflict of interest statement: Dr Graeme P. Williams has no conflict of interest associated with the contents or publication of this paper. All research and production was performed by, and selffunded by, Dr Graeme P. Williams.

References

- Allen C, Strauss B, Burger H, Forbes E, McLachlan R (2007). Testosterone therapy prevents gain in visceral adipose tissue and loss of skeletal muscle in non-obese aging men. J Clin Endocrinol Metab 93:139-146.
- Almeida O, Yeap B, Hankey GJ, Jamrozik K, Flicker L (2008). Low free testosterone concentration as a potentially treatable cause of depressive symptoms in older men. Arch Gen Psychiatry 65:283-289.
- Anderson GI, Limacher M, Assaf AR, Bassford T, Beresford SA, Black H, et al. (2002). Effects of conjugated equine estrogen in post-menopausal women: principal results from the WHI randomised controlled trial. JAMA 288:321-333.
- Arnaoutoglou C, Keivanidou A, Arnaoutoglou M, Kastrouni E, Samaras V, Kaiki-Astara A, et al. (2008). The effect of leptin on the tonic secretion of gonadotropins in female rats. Neuro Endocrinol Lett 29:999-1006.
- Axelson O, Bannasch P, Betazzi P, Blair A, Brown A, Bykorez A, et al. WHO/IARC Working Group (1987). Overall Evaluation of Carcinogenicity: an updating of IARC Monographs Evaluation of the carcinogenic risk to humans; Lyon, WHO Press, pp. 272-310.
- Baan R, Barrett-Connor E, Beral V, Bosland MC, Cook L, Franceschi S, et al.; WHO/IARC Working Group (1999). Hormonal contraception and postmenopausal hormone therapy. IARC monographs on the evaluation of carcinogenic risk to humans. Vol. 72. Lyon (France): WHO Press.
- Banks A, Davis S, Bates S, Myers M Jr (2000). Activation of downstream signals by the long form of the leptin receptor. J Biol Chem **275**:14563-14577.
- Bardin A, Boulle N, Lazennec G, Vignon F, Pujol P (2004). Loss of ERb expression as a common step in estrogen-dependent tumor progression. Endocr Relat Cancer 11:537-551.
- Bastard JP, Maachi M, Lagathu C, Kim M, Caron M, Vidal H, et al. (2006). Recent advances in the relationship between obesity, inflammation and insulin resistance. Eur Cytokin Netw 17:4-12.
- Beral V (2007). Ovarian cancer and HRT in the million women study. Lancet 369:1707-1710.
- Beresford SA (1997). Risk of endometrial cancer in relation to estrogen and combined cyclic progestagen therapy. Lancet 349:458-461.
- Berrino F, Muti P, Micheli A, Bolelli G, Krough V, Sciajno R, et al. (1996). Serum sex hormone levels after menopause and subsequent breast cancer. J Natl Cancer Inst 88:291-296.
- Bogaert YC, Linas S (2009). The role of obesity in the pathogenesis of hypertension, Nat Clin Pract Nephrol 5:101-111.
- Borugian MJ, Sheps SB, Kim-Sing C, Olivotto IA, Van Patten C, Dunn BP, et al. (2003). Waist to hip ratio directly related to breast cancer mortality. Am J Fnidemiol 158:963-968.
- Bosland MC, McCormick DL, Melamed J, Walden PD, Zeleniuch-Jacquotte A, Lumey LH (2002). Chemoprevention strategies for prostate cancer. Eur J Cancer Prev 11 (Suppl 2):S18-S27.
- Bouvard V, Baan R, Straif K, Grosse Y, Secretan B, Ghissassi F, et al. (2009). A review of human carcinogens - Part A: pharmaceuticals. Lancet Oncol
- Boyanov MA, Boneva Z, Christov V (2003). Testosterone supplementation in men with type 2 diabetes, visceral obesity and partial androgen deficiency. Aging Male 6:1-7.
- Brannian J, Zhao Y, McElroy M (1999). Leptin inhibits progesterone production by luteinised granulosa cells only in the presence of increased insulin. Human Reprod 6:1445-1448.
- Breast Cancer Hormonal Contraceptives (1996). Further results. Contraception 54 (Suppl 3):1S-106S.
- Brueggemeier RW, Richards JA, Joomprabutra S, Bhat AS, Whetstone JL (2001). Molecular pharmacology of aromatase and its regulation by endogenous and exogenous agents. J Steroid Biochem Mol Biol 79:75-84.
- Bulun SE, Imir G, Utsunomiya H, Thung S, Gurates B, Tamuraa M, Lin Z (2005). Aromatase in endometriosis and leiomyomata. J Ster Biochem Mol Biol **95**:57-62.

- Burris A, Banks S, Carter C, Davidson J, Sherins R (1992). A long-term prospective study of the physiological and behavioural effects of testosterone replacement therapy in untreated hypogonadal men. Androl 13:297-304
- Canonico M, Plu-Bureau G, Lowe G, Scarabin P-Y (2008). HRT and risk of venous thrombo-embolism in post-menopausal women: systemic review and meta-analysis. BMJ 336:1227-1231.
- Carr B, Bradshaw K (2005). Disorders of the ovary and the female reproductive tract. In: Kasper D, Sauci A, Longo D, Braunwald E, Hauser S, Jameson J, editors. Harrison's Principles of Internal Medicine. 16th ed. New York: McGraw Hill Medical Publishing Div. pp. 2201. Chap 326.
- Carruba G (2006). Estrogens and mechanisms of prostate cancer progression. Ann N Y Acad Sci 1089:201-217.
- Carruba G (2007). Estrogen and prostate cancer: an eclipsed truth in an androgen-dominated scenario. J Cell Biochem 102:899-911.
- Castracane VD, Kraemer RR, Franken MA, Giupel TL (1998). Serum leptin concentration in women: the effect of age, obesity and estrogen administration. Fertil Steril 70:472-477.
- Catalano S, Marsico S, Giordano C, Mauro L, Rizza P, Panno ML, Andro S (2003), Leptin enhances aromatase content, mRNA expression aromatase enzymic activity and upregulates p450 aromatase gene expression in epithelial breast cancer cells. J Biol Chem 278:2866-2867.
- Catalano S, Marsico S, Giordano C, Mauro L, Rizza P, Panno ML, Andro S (2004). Leptin induces via ERK1/ERK2 signal, functional activation of estrogen receptor-α in MCF-7 cells. J Biol Chem 279:19908-19915.
- Cavalieri EL, Rogan EG (2004). A unifying mechanism in the initiation of cancer and other diseases by catechol quinones. Ann N Y Acad Sci 1028:247-252.
- Cavalieri E, Rogan E (2006). Catechol guinones of estrogens in the initiation of breast cancer, prostate cancer and other human cancers. Ann N Y Acad Sci 1089:286-301
- Chamie K, DeVere-White RW, Lee D, Ok JH, Ellison LM (2008). Agent orange exposure, Vietnam war veterans, and the risk of prostate cancer. Cancer **113**:2464-2470.
- Channer KS, Jones TH (2003). Cardiovascular effects of testosterone: implications of the male menopause. Heart 89:121-122.
- Cirillo D, Rachiglio AM, la Montagna R, Giordano A, Normanno N (2008). Leptin signalling in breast cancer: an overview. J Cell Biochem 105:956-964.
- Cogliano V. Grosse Y. Baan R. Straif K. Secretan B. El Ghissassi F: WHO (IARC) Monograph Working Group (2005). Carcinogenicity of combined oestrogenprogestogen contraceptives and menopausal treatment. Lancet Oncol 6:552-553
- Cohen PG (2008). Obesity in men: the hypogonadal-estrogen receptor relationship and its effect on glucose homeostasis. Med Hypotheses 70:358-360.
- Collaborative Group on Hormonal Factors in Breast Cancer (1997). Breast cancer and HRT: collaborative re-analysis of data from 51 epidemiological studies of 52 705 women with breast cancer and 108 411 women without breast cancer. Lancet 350:1047-1059.
- Cooper MA (2000). Testosterone replacement therapy for anxiety. Am J Psychiatry **157**:1884.
- Coya R, Martul P, Algorta J, Aniel-Quiroga MA, Busturia MA, Señarís R (2005). Progesterone and hPL inhibit leptin secretion. Gynecol Endocrinol 21:27-32.
- Craig MC, Maki PM, Murphy DG (2005). The Women's Health Initiative Memory study: findings and implications of treatment. Lancet Neurol 4:190-194.
- Daghestani MH, Ozand PT, Al-Himadi AR, Al-Odaib AN (2007). Hormonal levels of leptin, insulin, ghrelin, and neuropeptide Y in lean, overweight and obese Saudi females. Saudi Med J 28:1191-1197.
- Dallman MF, Akana SF, Pecoraro N, Warne J, la Fleur S, Foster M (2007). Corticosteroids, the etiology of obesity and the metabolic syndrome. Curr Alzheimer Res 4:199-204.
- De Benedetti V, Welsh J, Yu M, Bennett W (1996). p53 mutations in carcinoma related to oral contraceptive use. Carcinogenesis 17:145-149.
- Dees C, Askari M, Foster JS, Ahamed S, Wimalasena J (1997). DDT mimics oestrogen stimulation of breast cancer cells. Mol Carcinog 18:107-114.
- Dhindsa S, Prabhakar S, Sethi M, Bandyopadhyay A, Chaudhuri A, Dandona P (2004). Frequent occurrence of hypogonadotrophic hypogonadism in type 2 diabetes. J Clin Endocrinol Metab 89:5462-5468.
- Dieudonne MN, Sammari A, Dos Santos E, Leneveu M-C, Giudicelli Y, Pecquery R (2006). Sex steroids and leptin regulate 11β-hydroxysteroid dehydrogenase 1 and p450 aromatase expressions in human preadipocytes: sex specificities. J Steroid Biochem Mol Biol 99:189-196.
- Ding EL, Song Y, Malik VS, Liu S (2006). Sex differences of endogenous sex hormones and risk of type 2 diabetes - a systematic review and metaanalysis. JAMA 295:1288-1299.

- Dundar B, Dundar M, Erci T, Buber E, Buyukgebiz A (2005). Leptin levels in boys with pubertal gynecomastia. J Paed Endocrinol Metab 18:929-934.
- Dyck D (2005). Leptin sensitivity in skeletal muscle is modulated by diet and exercise. Exerc Sport Sci Rev 33:189-194.
- Dyck DJ, Heigenhauser GJ, Bruce CR (2006). Role of adipokines as regulators of skeletal muscle fatty acid metabolism and insulin sensitivity. ACTA Physiol (Oxf) 186:5-16.
- Earth Summit: United Nations Conference on Environment and Development (UNCED), Rio de Janeiro: World Scientists' Warning To Humanity, November 18 1992
- Ellem SJ, Risbridger GP (2007). Treating prostate cancer: a rationale for targeting local oestrogens. Nat Rev Cancer 7:621-627.
- Emons G, Grundker C, Hanf V (2002). 4-hydroxyestrogens induce DNA damage and transforming mutations to cause endometrial cancer and ovarian cancer. Zentralbl Gynacol 124:559-565.
- Espindola-Antunes D, Kater CE (2007). Adipose tissue expression of 11betahydroxysteroid dehydroganase type 1 in Cushing's syndrome and obesity. Arg Bras Endocrinol Metab 51:1397-1403.
- Falconnier Y, Delavaud C, Chillard Y (2003). Insulin and dexamethasone effects on leptin production and metabolic activities of adipose tissue. Reprod Nutr Devel 43:237-250.
- Ferri CP, Prince M, Brayne C, Brodaty H, Fratiglioni L, Ganguli M (2005). Global prevalence of dementia: a Delphi consensus study. Lancet 366:2112-2117.
- Flores M, Fernandes M, Ropelle E, Faria M, Ueno M, Velloso L, et al. (2006). Exercise improves insulin and leptin sensitivity. Diabetes 55:2554-2561.
- Foster MT, Warne JP, Ginsberg AB, Horneman H, Pecoraro C, Akana S, Dallman M (2009). Palatable foods, stress, and energy stores sculpt corticotropin-releasing factor, adrenocorticotropin and corticosterone concentrations after restraint. Endocrinology 150:2325-2333.
- Fournier A, Berrino F, Riboli E, Avenel V, Clavel-Chapelon F (2005). Breast cancer risk in relation to different types of HRT in the E3N-EPIC cohort. Int J Cancer 114:448-454.
- Franks PW, Loos RJ, Brage S, O'Rahilly S, Wareham NJ, Ekelund U (2007). Physical activity energy expenditure may mediate the relationship between plasma leptin levels and worsening insulin resistance independently of adiposity. J Appl Physiol 102:1921-1926.
- Fuentes M, Muldoon TG, Mahesh VB (1990). Inhibitory effect of progesterone on occupied oestrogen receptors of anterior pituitary and uterus in adult rats. J Neuroendocrinol 2:517-522.
- Fujita N, Sakamaki H, Uotani S, Takahashi R, Kuwahara H, Kita A, et al. (2003). Intracerebroventricular administration of insulin inhibits anorectic action of leptin in rats. Exp Biol Med 228:1156-1161.
- Fukur M (2003). Supraphysiological testosterone dosages will increase insulin resistance. Diabetes Care 26:1869-1873.
- Gann PH, Hennekens CH, Ma J, Longcope C, Stampfer M (1996). Prospective study of sex hormone levels and risk of prostate cancer. J Nat Cancer Inst 88:1118-1126.
- Ganong W, Lange JD (2005). The gonads development and function of the reproductive system. In: Ganong W, editor. Review of medical physiology. 22nd ed. New York: Lange Medical Books/McGraw-Hill Medical Publishing. pp. 441-442. Chap 23.
- Garcia-Arencibia M, Molero S, Davila N, Carranza M, Calle C (2005). 17B estradiol transcriptionally represses human insulin receptor gene expression causing cellular insulin resistance. Leukemia Res 29:79-87.
- Garnier M, Gianarchi C, Delrieu I, Rio MC, Chinestra P, Bayard F, et al. (2003). Insulin, estradiol receptor ligand influence FGF-2 activities in MCF-7 cells. Biochem Pharmacol 65:629-636.
- Garofolo C, Koda M, Cascio S, Sulkowska M, Kanczuga-Koda L, Golaszewska J, et al. (2006). Increased expression of leptin and leptin receptors as a marker of breast cancer progression - the possible role of obesity-associated stimuli. Clin Cancer Res 12:1447-1453.
- Geisler J, Haines B, Eske D, Dowsett M, Lonning PE (2007). Total body aromatization in postmenopausal breast cancer patients is closely correlated to plasma leptin levels, J Steroid Biochem Mol Biol 104:27-34.
- Gizard F, Robillard F, Gevois P, Faucompre A, Revillion F, Peyrat J-P, et al. (2005). Progesterone inhibits human breast cancer cell growth through transcriptional upregulation of the cyclin-dependent kinase inhibitor p27Kip1 gene. Federation of European Biochemical **579**:5535-5541.
- Goldstat R, Briganti E, Tran J, Wolfe R, Davis SR (2003). Transdermal testosterone application in women improves mood, well-being and sexual function in premenopausal women. Menopause 10:390-398.
- Gong Z. Neuhouser ML. Goodman P. Albanes D. Chi C. Hsing A. et al. (2006). Obesity, diabetes and risk of prostate cancer: results from the prostate cancer prevention trial. Cancer Epidemiol Biomarkers Prev 15:1977-1983.

- Gonzalez C. Alonso A. Grueso N. Diaz F. Estaban M. Fernandez S. Patterson A. (2002). The role of 17β-estradiol administration on insulin sensitivity and its implications for the insulin receptor. Steroids 67:993-1005.
- Greiser CM, Greiser EM, DoANren M (2007), Menopausal hormone therapy and risk of ovarian cancer: systematic review and meta-analysis. Hum Reprod Update 13:453-463.
- Grosse Y. Baan R. Straif K. Secretan B. El Ghissasi F. Bouvard V. et al.: IARC Working Group (2009). Special report human carcinogens: pharmaceuticals. Lancet Oncol 10:13-14.
- Gupta M, McDougal A, Safe S (1998). Estrogenic activities of 16-OHE2 and 17β-estradiol in MC-7 and T47D human breast cancer cells. J Steroid Biochem Mol Biol 67:413-419.
- Gustafsson O, Norming U, Gustafsson S, Eneroth P, Aström G, Nyman CR (1996). DHT and testosterone levels in men screened for prostate cancer. Br J Urol 77:433-440.
- Hak E, Witteman C, de Jong F, Geerlings M, Hofman A, Pols H (2002). Low levels of endogenous androgens and raised oestrogen levels increase risk of atherosclerosis in elderly men. J Clin Endocrinol Metab 87:3632-3639
- Hammarsten J, Damber JE, Karlsson M, Knutson T, Ljunggren O, Ohlsson C, et al. (2009), Insulin and free oestradiol are independent risk factors for benign prostatic hyperplasia. Prostate Cancer Prostatic Dis 12:160-165.
- Han X, Liehr J, Bosland M (1995). Induction of a DNA adduct detectable by 32Ppostlabelling in the dorsolateral prostate of rats treated with 178-estradiol and testosterone. Carcinogenesis 16:951-954.
- Harvard Nurses Health Study (2003). Type 2 diabetes and subsequent incidence of breast cancer in the Nurses Health Study. Diabetes Care 26:1752-1758.
- Heiss G, Wallace R, Anderson G, Aragaki A, Beresford S, Brzyski R, et al. (2008). Health risks and benefits 3 years after stopping randomized treatment with estrogen and progestin, JAMA 299:1036-1045.
- Hennige AM, Stefan M, Kapp K, Lehmann R, Weigert C, Beck A, et al. (2006). Leptin down-regulates insulin action through phosphorylation of Ser-318 in insulin receptor substrate 1. FASEB J 20:1206-1208.
- Hilf R, Crofton DH (1985). Effects of estradiol on insulin receptor distribution in primary cultures of R3230AC mammary adenocarcinoma of the rat. Endocrinology 116:154-163.
- Ho SC, Tai ES, Eng PH, Ramli A, Tan CE, Fok AC (1999). A study of the relationships between leptin, insulin, and body fat in Asian subjects. Int J Obes Relat Metab Disord 23:246-252.
- Hoffman MA (2000). Is a low serum free testosterone a marker for high-grade prostate cancer? J Urol 163:824-827.
- Horenburg S, Fischer-Posovszky P, Debatin KM, Wabitsch M (2008). Influence of sex hormones on adiponectin expression by human adipocytes. Horm Metab Res 40:779-786.
- Hsing AW, Chau S Jr, Gao YT, Gentzschein E, Chang L, Deng J, Stanczyk FZ (2001). Prostate cancer risk and serum levels of insulin and leptin: a population-based study. J Nat Cancer Inst 93:783-789.
- Iguchi T, Wantanabe H, Ohta Y, Blumberg B (2008). Developmental effect: oestrogen induced vaginal changes and adipogenesis. Int Jour Andrology
- Imigaray P, Newby JA, Lacomme S, Belpomme D (2007). Overweight/obesity and cancer genesis: more than a biological link. Biomed Pharmacother 61:665-678.
- Insulin upregulates arcuate NPY release. www.nature.com/neuro/journal/voru8/ n5/full/nn1455.html
- International Diabetes Foundation: 2007.
- losif CS, Batra S (1994). Changes in the density of nuclear estrogen receptors in urogenital tissue during pseudopregnancy in the rabbit. Internat Urogynecol J 4:264-268.
- Ishida-Takahashi R, Rosario F, Gong Y, Kopp K, Stancheva Z, Chen X, et al. (2006). Phosphorylation of Jak-2 on Ser 523 inhibits Jak-2 dependent leptin receptor signalling. Mol Cell Biol 26:4063-4073.
- Kaaks R (2005), Nutrition, insulin, IGF-1 metabolism and cancer risk; a summary of epidemiological evidence. In: Bock G, Goode J, editors. Biology of IGF-1. Novartis Foundation Symposium 262. London: Novartis Foundation; pp. 247-264.
- Kaaks R, Berrino F, Key T, Rinaldi S, Dossus L, Biessy C, et al. (2005). Serum sex steroids in pre-menopausal women and breast cancer risk within the European Prospective Investigation into Cancer and Nutrition. J Natl Cancer Inst 97:755-765.
- Kaczmarec A, Reczuch K, Madja J, Banaslak W, Ponikowski P (2003). The association of lower testosterone levels with coronary artery disease in postmenopausal women. Int J Cardiol 87:53-57.
- Kalyani RR, Dobs AS (2007). Androgen deficiency, diabetes and the metabolic syndrome in men. Curr Opin Endocrinol Diabetes Obes 14:226-234.
- Kaur T, Zhang ZF (2005). Obesity, breast cancer, and the role of adipocytokines. Asian Pac J Cancer Prev 6:547-552.

- Kellerer M. Lammers R. Fritsche A. Strack V. Machicao F. Borboni P. et al. (2001). Insulin inhibits leptin receptor signalling at the JK-2 level: a possible mechanism for hyperinsulinemia associated leptin resistance. Diabetologica 9:1125-1132
- Key TJ, Appleby P, Barnes I, Reeves G (2002). Endogenous sex hormones and breast cancer in post menopausal women: reanalysis of nine prospective studies. J Natl Cancer Inst 94:606-616.
- Key TJ, Appleby PN, Reeves GK, Roddam A, Dorgan JF, Longcope C, et al.; The Endogenous Hormones and Breast Cancer Collaborative Group (2003). BMI, serum sex hormones and breast cancer risk in post-menopausal women. J Natl Cancer Inst 95:1218-1226.
- Kinoshita Y, Chen S (2003). Induction of aromatase expression in breast cancer cells. Cancer Res 63:3546-3555.
- Knudson JD (2007). Adipokines and coronary vasomotor dysfunction. Exp Biol Med 232:727-736.
- Lapauw B, T'sjoen G, Mahmoud A, Kaufman JM, Ruigeet JB (2009). Short term aromatase inhibition: effects on glucose metabolism, and serum leptin levels in young and elderly men. Eur J Endocrinol 160:397.
- Lauer M (2008). Women's Health Initiative Follow-up Study. NIH and NHLBI Public Release 4 March 2008.
- Laville N, Balaquer P, Brion F, Hinfray N, Casellas C, Porcher J, Ait-Aissa S (2006). Modulation of aromatase activity and mRNA by various selected pesticides in human choriocarcinoma TEG-3 cell line. Toxicology 228:98-10.
- Leal-Cerro A, Soto A, Martinez MA, Dieguez C, Casaneva FF (2001). Influence of cortisol status on leptin secretion. Pituitary 4:111-116.
- Leihr J (1997). Hormone associated cancer, Mechanistic similarities between human breast cancer and estrogen induced carcinogenesis. Environ Health Perspect 105 (Suppl 3):565-569.
- Leihr J, Ricci M, Jeffcoate C, Hannigan E, Hokanson J, Zhu B (1995). 4-Hydroxylation of estradiol by human uterine myometrium and myoma microsomes: implications for the mechanism of uterine tumorigenesis. Proc Natl Acad Sci U S A 92:9220-9224.
- Lieb W, Sullivan LM, Harris TB, Roubenoff R, Benjamin E, Levy D, et al. (2009). (Framingham Heart Study). Plasma leptin levels and incidence of heart failure, cardiovascular disease and total mortality in the elderly. Diabetes Care 32:612-616.
- Lindsay RS, Hamilton BA, Calder AA, Johnstone FD, Walker TD (2004). The relation of insulin, leptin, and IGF-1 to birth weight in offspring of women with diabetes. Clin Endocrinol 61:353-359.
- Livingstone C, Collison M (2002). Sex steroids and insulin resistance. Clin Sci 102:151-166
- Lonning PE, Geisler J (2008). Aromatase inhibitors assessment of biochemical efficacy measured by total body aromatase inhibition and tissue suppression. J Ster Biochem Mol Biol 108:196-202.
- Luchsinger JA, Gustafson DR (2009). Adiposity and Alzheimer's disease. Curr Opin Clin Nutr Care 12:15-21.
- Manderson JG, Patterson CC, Hadden D, Traub Al, Leslie H, McCance D (2003). Leptin concentration in maternal blood in diabetic and non-diabetic pregnancy. Am Jour Obstet Gynecol 188:1326-1332.
- Margolese HC (2000). The male menopause and mood: testosterone decline and depression in the ageing male - is there a link? J Geriatr Psych Neurol **13**:93-101.
- Martin SS, Qasim A, Reilly MP (2008). Leptin resistance: a possible interface of inflammation and metabolism in obesity-related heart disease. J Amer Coll Cardiol 52:1201-1210.
- Mawson A, Lai A, Carroll JS, Sergio CM, Mitchell CJ, Sarcevic B (2005). Oestrogen and insulin/IGF-1 cooperatively increase cell cycle progression in MCF-7 breast cancer cells. Mol Cell Endocrinol **229**:161-173.
- Michaeli A, Muti P, Secreto G, Krogh V, Meneghini E, Venturelli E, et al. (2004). Endogenous sex hormones and subsequent breast cancer in pre-menopausal women. Int J Cancer 112:312-318.
- Miller PB, Forstein DA, Styles S (2008). Effect of short-term diet and exercise on hormonal levels and menses in obese infertile women. J Reprod Med **53**:315-319.
- Missmer S, Eliassen AH, Barberi R, Hankinson S (2004). Endogenous estrogen, androgen, and progesterone concentrations and breast cancer risk amongst post-menopausal women. J Nat Cancer Instit 96:1856-1865.
- Mistry T, Digby JE, Desai KM, Randeva HS (2007). Obesity and prostate cancer: a role for adipokines. Eur Urol 52:46-53.
- Morales A (2002). Androgen replacement therapy and prostate safety. Eur Urol 41:113-120.
- Morley JE (2000). Testosterone replacement and the physiological aspects of ageing in men. Mayo Clin Proc 75:83-87.

- Muller M, Aleman A, Grobbee DE, de Haan EH, van der Schouw YT (2005). Endogenous sex hormone levels and cognitive function in aging men: is there an optimal level? Neurology 64:866-871.
- Murakami H, Harada N, Sasano H (2001). Aromatase in atherosclerotic lesions of human aorta. J Ster Biochem Mol Biol 79:67-74.
- Muti P, Quattrin T, Grant B, Krogh V, Micheli A, Holger J, et al. (2002). Fasting glucose is a risk factor for breast cancer. Cancer Epidemiology, Biomarkers and Prevention 11:1361-1368.
- Nakamura S, Nishii N, Yamanaka A, Kitagawa H, Asano M, Tsubota T, Suzuki M (2009). Leptin receptor (Ob-R) expression in the ovary and uterus of wild Japanese black bears. J Reprod Dev 55:110-115.
- Nakanishi T (2008). Endocrine disruption induced by organotin compounds: organotins function as a powerful agonist for nuclear receptors rather than being an aromatase inhibitor. J Toxicol Sci 33:269-276.
- Newbold R, Padilla-Banks E, Jefferson W, Heindel J (2008). Effects of endocrine disruptors on obesity. Int J Andrology 31:201-208.
- Nilsson R (2000). Endocrine modulators in the food chain. Toxicol Pathol 28:420-431.
- O'Neil JS, Burow ME, Green AE, Henson MC (2001). Effects of estrogen on leptin gene promoter activation in MCF-7 breast cancer cells. Mol Cell Endocrinol 176:67-75.
- Oestrogen receptors are increased twelvefold in the presence of insulin on the surface of human breast cancer cells. Experimental Biology Conference 1998, San Francisco, April 21,
- Osuna JA, Gomez-Perez R, Arata-Bellabarba G, Villaroel V (2006). Relationship between BMI, total testosterone, sex hormone binding globulin, leptin, insulin and insulin resistance in obese men. Arch Andriol 52:355-361.
- Padero MC, Bhasin S, Friedman TC (2002). Androgen supplementation in older women. J Am Geriat Soc 50:1131-1140.
- Park S, Jang J, Jun DW, Hong SM (2005). Exercise enhances insulin and leptin signaling in the hypothalamus during dexamethasone induced stress in diabetic rats. Neuroendocrinol 82:282-293.
- Paulsen SK, Pedersen SB, Fisker S, Richelsen B (2007). 11β-HSD type 1 expression in human adipose tissue: impact of gender, obesity, and fat localization. Obesity (Silver Spring) 15:1954-1960.
- Pausova Z (2006). From big fat cells to high blood pressure: a pathway to obesity-associated hypertension. Curr Opin Nephrol Hypertens 15:173-178.
- Pedersen SB, Kristensen K, Hermann PA, Katzenellenbogen JA, Richelsen B (2004). Estrogen controls lipolysis by upregulating alpha2A-adrenergic receptors directly in human adipose tissue through estrogen receptor-α. Implications for the female fat distribution. J Clin Endocrinol Metab 89:1869-1878.
- Petersen KR (2002). Pharmacodynamic effects of oral contraceptive steroids on biochemical markers for arterial thombosis. Studies in non-diabetic women and in women with insulin dependent diabetes mellitus. Dan Med Bull
- Petrelli J, Calle E, Rodriguiz C, Thun M (2002). BMI, height and post menopausal breast cancer mortality in a prospective cohort of US women. Cancer Causes Control 13:325-332.
- Pike CJ, Rosario ER, Nguyen TV (2006). Androgens, aging and Alzheimer's disease. Endocrine 29:233-241.
- Pope HG Jr, Cohane GH, Kanayama G, Siegel AJ, Hudson JI (2003). Testosterone gel supplementation for men with refractive depression: a random placebo controlled study. Am J Psychiatry 160:105-111.
- Prehn RT (1999). The prevention and therapy of prostate cancer by androgen administration. Cancer Res 59:4161-4164.
- Prins G (2008). Endocrine disrupters and prostate cancer risk. Endocr Relat Cancer 15:649-656.
- Prins G, Korach K (2008). The role of estrogens and estrogen receptors in normal prostate growth and disease. Steroids 73:233-244.
- Pugh J, Moore M. National Research Centre for Environmental Toxicology: national science and industry forum report, 1998.
- Raji C, Ho A, Pariksak N, Becker J, Lopez O, Kuller L, et al. (2010). Brain structure and obesity. Human Brain Mapping 31:353-364.
- Ravaglia G, Forti P, Maioli F, Bastagli L, Montesi F, Pisacana N, et al. (2007). Endogenous sex hormones as risk factors for dementia in elderly men and women. J Gerontol A Biol Sci Med Sci 62:1035-1041.
- Revillion F, Charlier M, L'hotellier V, Hornez L, Giard S, Baranzelli MC, et al. (2006), Messenger RNA expression of leptin and leptin receptors, and their prognostic value in 322 human primary breast cancers. Clin Cancer Res
- Rhoden EL, Morgentaler A (2004). Risks of testosterone-replacement therapy and recommendations for monitoring. New Eng J Med 350:440-442.
- Risbridger G, Wang H, Young P, Kurita T, Wang YZ, Lubahn D, et al. (2001). Evidence that epithelial and mesenchymal estrogen receptor-a mediates effects of estrogen on prostatic epithelium. Dev Biol 229:432-442.

- Risbridger GP Bianco II Fllem SI McPherson SI (2003). Oestrogens and prostate cancer. Endocr Relat Cancer 10:187-191.
- Risbridger GP, Ellem SJ, McPherson SJ (2007). Estrogen action on the prostate gland: a critical mix of endocrine and paracrine signaling. J Mol Endocrinol 39:183-188.
- Rock CL, Flatt SW, Laughlin GA, Gold EB, Thomson CA, Natarajan L, et al. (2008). Reproductive steroid hormones and recurrence-free survival in women with a history of breast cancer. Cancer Epidem Biomarkers and Prevention 17:204-211.
- Rosario E, Carroll J, Oddo S, Laferla F, Pike C (2006). Androgens regulate the development of neuropathology in a triple transgenic mouse model of Alzheimer's disease. J Neurosci 26:13384-13389.
- Russo J, Russo IH (2006). The role of estrogen in the initiation of breast cancer. J Steroid Biochem Mol Biol 102:89-96.
- Russo J, Hu YF, Tahin Q, Mihaila D, Slater C, Lareef M, et al. (2001). Carcinogenicity of estrogens in human breast epithelial cells. APMIS 109:39-52.
- Sagsoz N, Orbak Z, Noyan V, Yücel A, Uçar B, Yildiz L (2009). Effects of oral contraceptives including low dose estrogen and drospirenone on leptin and ghrehlin in PCOS. Fertil Sterility 92:660-666.
- Samad F. Adipose estrogen and increased breast cancer risk in obesity: Regulation by leptin and insulin. La Jolla Institute for Molecular Medicine, SanDiego. CA 92121/US Army Med Research Command, Sept 2007.
- Santner S, Pauley R, Tait L, Kaseta J, Santen R (1997). Aromatase activity and expression in breast cancer and benign breast tissue stromal cells. J Clin Endocrinol Metab 82:200-208.
- Saxena NK, Taliaferro-Smith L, Knight B, Merlin D, Anania F, O'Regan R, et al. (2008). Bi-directional crosstalk between leptin and insulin-like growth factor 1 signaling, promotes invasion and migration of breast cancer via transactivation of epidermal growth factor receptor. Cancer Res 68:9712-9722.
- Schatzl G, Madersbacher S, Thurridl T, Waldmüller J, Kramer G, Haitel A, Marberger M (2001). High-grade prostate cancer associated with low serum testosterone levels. Prostate 47:52-58.
- Schlumpf M, Schmid P, Durrer S, Conscience M, Maerkel K, Henseler M, et al. (2004). Endocrine activity and developmental toxicity of cosmetic UV filters an update. Toxicology 205:113-122.
- Schneider HP, Mueck AO, Kuhl H (2005). IARC monograph on carcinogenicity of combined hormonal contraceptives and menopausal therapy. Climacteric 8:311-316
- Seeger H, Wallwiener D, Kraemer E, Mueck AO (2005). Estradiol metabolites are potent mitogenic substances for human ovarian cancer cells. Eur J Gynaecol Oncol 26:383-385.
- Seeger H, Wallwiener D, Kraemer E, Mueck AO (2006). Comparison of possible carcinogenic estradiol metabolites: effects on proliferation, apoptosis, and metastasis of human breast cancer cells. Maturitas 54:72-77.
- Shin JH, Hur JY, Seo HS, Jeong YA, Lee JK, Oh MJ, et al. (2007). The ratio of estrogen receptor-α to estrogen receptor-β in adipose tissue is associated with leptin production and obesity. Steroids 72:592–599.
- Söderberg S, Olsson T, Eliasson M, Johnson O, Brismar K, Carlström K, Ahrén B (2001). A strong inverse association between biologically active testosterone and leptin in non-obese men and women is lost with increasing central adiposity. Internat J Obesity 25:98-105.
- Stattin P, Lumme S, Tenkanen L, Alfthan H, Jellum E, Hallmans G, et al. (2004). High levels of circulating testosterone are not associated with increased prostate cancer risk - a pooled prospective study. Int J Cancer **108**:418-424.
- Strat A, Kokta T, Dodson M, Gertler A, Wu Z, Hill R (2005). Early signalling interactions between insulin and leptin pathways in bovine myogenic cells. Biochimica et Biophysica: Molecular Cell Research 1744:164-175.
- Sulkowska M, Golaszewska J, Wincewicz A, Koda M, Baltaziak M, Sulkowski S (2006). Leptin - from regulation of fat metabolism to stimulation of breast cancer growth, Pathol Oncol Res 12:69-72.
- Tan RS, Pu SJ (2003). A pilot study on the effects of testosterone in hypogonadal aging male patients with Alzheimer's disease. Aging Male 6:13-17.
- Tanaka M, Nakaya S, Kumai T, Watanabe M, Tateishi T, Shimizu S (2001). Effects of estradiol on serum leptin levels and leptin mRNA synthesis in rat adipose tissue. Horm Res 56:98-104.
- The Million Study Collaborators (2003). Breast cancer and hormone replacement therapy in the Million Women Study. Lancet 362:419-427.
- Thomas T, Burguera B, Melton LJ, Atkinson EJ, O'Fallon WM, Riggs LJ, Khosla S (2000). Relationship of serum leptin levels with body composition and sex steroid and insulin levels in men and women. Metabolism 49:1278-1284.
- Thorton MJ, Nelson L, Taylor AH, Birch MP, Laing I, Messenger AG (2006). The modulation of aromatase and ER-α in cultured human dermal papilla. J Investigat Dermatol 126:2010-2018.

- Vainio H, Magee PN, McGregor DB, McMichel AJ (1992). Sex hormones and cancer. In: Vainio H, Magee P, McGregor D, McMichel A, editors. Mechanisms of carcinogenesis in risk identification. WHO/IARC Scientific Publication No. 116. Lyon: IARC Scientific Publishing. pp. 225-269.
- Van Landeghem A, Poortman J, Nabuurs M, Thijssen J (1985). Endogenous concentrations and subcellular distribution of estrogens in normal and malignant breast tissue. Cancer Res 45:2900-2906.
- Verhoeven MO, van der Mooren MJ, Teerlink T, Verheijen RHM, Scheffer PG, Kenemans P (2009). The influence of physiological and surgical menopause on coronary heart disease risk markers. Menopause 16:37-49.
- Vona Davis L, Howard-McNatt M, Rose DP (2007). Adiposity, type two diabetes and the metabolic syndrome in breast cancer. Obes Rev
- Watson C, Jeng Y-J, Kochukov M (2008). Non-genomic actions of estradiol compared to estrone and estriol in pituitary tumor cells significantly increases proliferation. FASEB 22:3328-3336.

- Webb CM, McNeill JG, Hayward CS, de Zeigler D, Collins P (1999). Effects of testosterone on coronary vasomotor regulation in men with CHD. Circulation **100**:1690-1696.
- Williams GP (2006). Unlock your hormones. Melbourne: Griffin Press. World Health Organization Global Infobase: 2004-2006.
- Xu B, Kitawaki J, Koshiba H, Ishihara H, Kiyomizu M, Teramoto M, et al. (2007). Differential effects of progestogens, by type and regimen, on estrogenmetabolizing enzymes in human breast cancer cells. Maturitas 56:142-152.
- Yamada T, Natagos S, Kurachi O, Wang J, Takekida S, Matsuo H, Maruo T (2004). Progesterone down-regulates insulin-like growth factor-I expression in cultured human uterine leiomyoma cells. Human Reprod 19:815-821.
- Yi KW, Shin JH, Seo HS, Lee JK, Oh MJ, Kim T, et al. (2008). Role of estrogen receptor alpha and beta in regulating leptin expression in 3T3-L1 adipocytes. Obesity (Silver Spring) 16:2393-2399.
- Zhou B, Sun Q, Cong R, Gu H, Tang N, Yang L, et al. (2008). Hormone replacement therapy and ovarian cancer risk: a meta-analysis. Gynecol Oncol **108**:641-651.