## Genetic induction and upregulation of cyclooxygenase (COX) and aromatase (CYP19): an extension of the dietary fat hypothesis of breast cancer

## R. E. Harris, F. M. Robertson, H. M. Abou-Issa, W. B. Farrar, R. Brueggemeier

The Ohio State University Comprehensive Cancer Center, Columbus, Ohio, USA

**Summary** A novel model of mammary carcinogenesis is proposed involving sequential induction and upregulation of cyclooxygenase and aromatase genes by essential fatty acids prominent in the US diet. The basic carcinogenic processes are: (1) constitutive prostaglandin biosynthesis and formation of mutagenic oxygen and nitrogen free radicals responsible for tumor initiation; (2) PGE-2-induced expression of aromatase and constitutive estrogen biosynthesis which sustains mitogenesis and tumor promotion; and (3) PGE-2-induced expression of vascular endothelial growth factor which stimulates angiogenesis and tumor metastasis.

Epidemiologic investigations suggest that nonsteroidal anti-inflammatory drugs (NSAIDs) have chemopreventive potential against human breast cancer (1–4). This effect apparently stems from the blockade of the prostaglandin cascade by inhibition of its rate limiting enzyme, cyclooxygenase (COX).

Two primary genes are responsible for the genetic control of COX, a constitutive gene (COX-1) and an inducible isoform (COX-2) (5). Recent molecular studies of breast cancer tissues indicate that COX-2 is inappropriately induced and that both COX-2 and COX-1 are upregulated in malignant cells (6).

Prostaglandin (PG) biosynthesis is the key component of the human response to immune stimulation (7). However, inappropriate activation and upregulation of this immunoreactive pathway may lead to untoward results; namely, the intermediate genesis of free radicals (FRs) which are mutagenic (8).

Since mammary carcinogenesis is undoubtedly a func-

Received 19 November 1997 Accepted 2 July 1998

Correspondence to: Randall E. Harris MD, PhD, The Ohio State University, Comprehensive Cancer Center, M-116 Starling Loving Hall, 320 West 10th Avenue, Columbus, OH 43210-1240, USA

tion of deregulation of estrogen biosynthesis and metabolism, the above components of PG deregulation are not by themselves sufficient to constitute an important model of mammary carcinogenesis. However, it has recently been discovered that PGE-2 effectively and specifically induces the promoter II region of the cytochrome P-450 gene (CYP-19) which is transcribed and translated into aromatase, the chief enzyme in the biosynthesis of estrogen (9). Thus, the PG model of mammary carcinogenesis involves a mutagenic autocrine function of mammary epithelial cells (production of PG and FRs), and a paracrine function in contiguous adipocytes and fibroblasts (induction of CYP-19 with subsequent biosynthesis of estrogen by aromatase). It is also noteworthy that the model potentiates angiogenesis in developing tumors, since PGE-2 induces the expression of vascular endothelial growth factor (VEGF) which stimulates neovascularization (10). Figure 1 depicts the basic features of the model.

The key feature of the model is elucidation of a common mechanism by which inappropriate induction and upregulation of estrogen biosynthesis occurs with regularity in the ductal epithelium of the mammary gland. Since estrogen is under tight homeostatic regulation, it is hypothesized that the loss of control is facilitated primarily

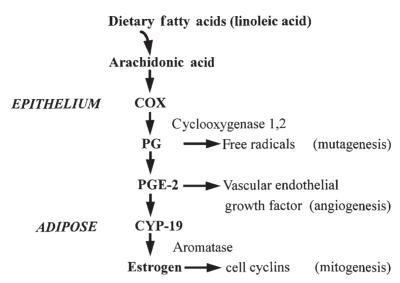


Fig. 1 Model of mammary carcinogenesis.

through a second biosynthetic pathway inextricably linked to estrogen biosynthesis. The PG cascade is sufficient for this purpose. It is ubiquitous in all cells including mammary epithelium and its controlling genes (especially COX-2) are readily induced and upregulated by a number of intra- and intercellular effector molecules including viral and bacterial antigens, growth regulatory factor and, most importantly, arachidonic acid, which serves as the pathway's primary substrate (11). In US women, the sustained presence of excess arachidonic acid results from excess consumption of red meat and certain vegetable oils rich in the essential polyunsaturated fatty acid, linoleic acid (12,13). Upon entering adipose and muscle cells, linoleic acid is converted to arachidonic acid which, in turn, activates constitutive transcription and translation of COX genes in the mammary epithelium, thereby leading to autocrine and paracrine effects of mutagenesis (tumor initiation), mitogenesis (tumor promotion), and angiogenesis (tumor metastasis). The model is therefore an extension of the dietary fat hypothesis of breast cancer and, since NSAIDs selectively inhibit cyclooxygenase, it portends an important new area of research in breast cancer chemoprevention (14,15).

## REFERENCES

- 1. Harris R. E., Namboodiri K. K., Stellman S. D., Wynder E. L. Breast cancer and NSAID use: heterogeneity of effect in a casecontrol study. Prev Med 1995; 24: 119-120.
- 2. Harris R. E., Namboodiri K. K., Farrar W. B. Epidemiologic study of non-steroidal anti-inflammatory drugs and breast cancer. Oncol Reports 1995; 2: 591-592.
- 3. Harris R. E., Namboodiri K. K., Farrar W. B. Nonsteroidal antiinflammatory drugs and breast cancer. Epidemiology 1996;
- 4. Schreinemachers D. M., Everson R. B. Aspirin use and lung,

- colon, and breast cancer incidence in a prospective study. Epidemiology 1994; 5: 138-146.
- 5. Herschman H. R. Regulation of prostaglandin synthase-1 and prostaglandin synthase-2. Cancer Metas Rev 1994; 13: 241-256
- 6. Parrett M. L., Harris R. E., Joarder F. S., Ross M. S., Clausen K. P., Robertson F. M. Cyclooxygenase-2 gene expression in human breast cancer. Int J Oncol 1997; 10: 503-507.
- 7. Vane J. R. Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. Nature (New Biol) 1971; 231: 232-235.
- 8. Marnett L. J. Aspirin and the potential role of prostaglandins in colon cancer. Cancer Res 1992; 52: 5575-5589.
- 9. Zhao Y., Agarwal V. R., Mendelson C. R., Simpson E. R. Estrogen biosynthesis proximal to a breast tumor is stimulated by PGE2 via cyclic AMP, leading to activation of promoter II of the CYP19 (aromatase) gene. Endocrinology 1996; 137(12): 5739-5742.
- 10. Ben-Av P., Crofford L. J. Wilder R. L., Hla T. Induction of vascular endothelial growth factor expression in synovial fibroblasts by Prostaglandin E and interleukin-1: a potential mechanism for inflammatory angiogenesis. FEBS Lett 1995; **372**(1): 83-87.
- 11. Wu K. K. Cyclooxygenase 2 induction: molecular mechanism and pathophysiologic roles [Review]. J Lab Clin Med 1996; **128**: 242-245.
- 12. Karmali R. A., Adams L., Trout J. R. Plant and marine n-3 fatty acids inhibit experimental metastasis of rat mammary adenocarinoma cells. Prostaglandins Leukot Essent Fatty Acids 1993; 48(4): 309-314.
- 13. Ramchurren N., Karmali R. A. Effects of gamma-linolenic and dihomo-gamma-linolenic fatty acids on 7.12 dimethylbenz(alpha)anthracene mammary tumors in rats. Prostaglandins Leukot Essent Fatty Acids 1995; 53(2): 95-101.
- 14. Mitchell J. A., Akarasereenont P., Thiemermann C., Flower R. J., Vane J. R. Selectivity of nonsteroidal antiinflammatory drugs as inhibitors of constitutive and inducible cyclooxygenase. Proc Natl Acad Sci USA 1994; 90: 11693-11697.
- 15. Harris R. E., Kasbari S., Farrar W. B. Prospective study of nonsteroidal anti-inflammatory drugs and breast cancer. Oncol Reports 1999; 6: 71-73.