Effects of temperature and fertilization on nitrogen cycling and community composition of an urban lawn

NEETA S. BIJOOR*, CLAUDIA I. CZIMCZIK*, DIANE E. PATAKI*† and
SHARON A. BILLINGS‡

*Department of Earth System Science, Croul Hall, University of California, Irvine, CA 92697-3100, USA, †Department of Ecology and Evolutionary Biology, Steinhaus Hall, University of California, Irvine, CA 92697-2525, USA, ‡Department of Ecology & Evolutionary Biology and Kansas Biological Survey, Higuchi Hall, University of Kansas, Lawrence, KS 66047, USA

Abstract

We examined the influence of temperature and management practices on the nitrogen (N) cycling of turfgrass, the largest irrigated crop in the United States. We measured nitrous oxide (N_2O) fluxes, and plant and soil N content and isotopic composition with a manipulative experiment of temperature and fertilizer application. Infrared lamps were used to increase surface temperature by $3.5 \pm 1.3\,^{\circ}C$ on average and control and heated plots were split into high and low fertilizer treatments. The N_2O fluxes increased following fertilizer application and were also directly related to soil moisture. There was a positive effect of warming on N_2O fluxes. Soils in the heated plots were enriched in nitrogen isotope ratio ($\delta^{15}N$) relative to control plots, consistent with greater gaseous losses of N. For all treatments, C_4 plant C/N ratio was negatively correlated with plant $\delta^{15}N$, suggesting that low leaf N was associated with the use of isotopically depleted N sources such as mineralized organic matter. A significant and unexpected result was a large, rapid increase in the proportion of C_4 plants in the heated plots relative to control plots, as measured by the carbon isotope ratio ($\delta^{13}C$) of total harvested aboveground biomass. The C_4 plant biomass was dominated by crabgrass, a common weed in C_3 fescue lawns. Our results suggest that an increase in temperature caused by climate change as well as the urban heat island effect may result in increases in N_2O emissions from fertilized urban lawns. In addition, warming may exacerbate weed invasions, which may require more intensive management, e.g. herbicide application, to manage species composition.

Keywords: C_3 and C_4, C_4 weeds, crabgrass, fescue, nitrous oxide, turfgrass, warming

Received 17 September 2007; revised version received 28 January 2008 and accepted 5 December 2007

Introduction

Turfgrass ecosystems are expanding rapidly in conjunction with urbanization, which is expected to increase 79% in the United States in the next 25 years (Alig *et al.*, 2004). As the largest irrigated crop in the United States, turfgrass currently covers 1.9% of the national surface area (Milesi *et al.*, 2005). Turfgrass land cover can sequester carbon (C) (Qian & Follett, 2002), but can also cause significant emissions of nitrous oxide (N_2O) (Kaye *et al.*, 2005), an important greenhouse gas that

Correspondence: Diane E. Pataki, Department of Earth System Science, Croul Hall, University of California, Irvine, CA 92697-3100, USA, tel. +1 949 824 9411, fax +1 949 824 3874, e-mail: dpataki@uci.edu

has a global warming potential 296 times greater than that of carbon dioxide (CO_2) (IPCC, 2001), and that has been increasing in the troposphere at a rate of about $0.2\%\,yr^{-1}$ (Weiss, 1981; Khalil *et al.*, 2002). It is, therefore, important to understand nitrogen (N) cycling and N_2O emissions from turfgrass, and their potential responses to climate change.

It is difficult to predict how warming caused by climate change and the urban heat island effect (Arnfield, 2003) may influence N cycling of turfgrass. It is also difficult to predict how plant N concentrations will change in response to warming; field experiments have shown that foliar C/N changes can be dynamic or species-specific (Read & Morgan, 1996; Klein *et al.*, 2007). Experimental warming manipulations in various ecosystems have shown that warming may significantly

alter soil N budgets through increased N mineralization (Shaver et al., 2000; Rustad et al., 2001). However, the effects of elevated temperature on nitrification, denitrification, and subsequent N_2O emissions are still unclear due to the small number of field studies (Barnard et al., 2005). The impact of warming on N_2O fluxes is complicated by two possible effects – reduced soil moisture, generally a negative effect, and increased soil temperature, a positive effect (Brumme, 1995; Skiba et al., 1998; Flechard et al., 2007). Studies of fertilized soils are particularly important, as agricultural crops are known to be large sources of N_2O at global, national, and statewide scales (Mosier et al., 1998; Bouwman et al., 2002; Franco, 2002). There have been a surprisingly few direct measurements of N_2O emissions from turfgrass (Maggiotto et al., 2000; Kaye et al., 2004; Bremer, 2006), such that the contributions of managed lawns to local and regional greenhouse gas budgets are largely unconstrained.

We initiated an experimental warming and fertilization manipulation in a fescue-dominated lawn to understand how temperature, fertilizer, and their combination influence N cycling. In addition to measuring soil N_2O fluxes, we examined foliar and soil N content and nitrogen isotope ratios ($\delta^{15}N$) in each treatment in order to determine the effects on plant N availability. $\delta^{15}N$ is a useful indicator of plant N sources and ecosystem N losses, as N derived from mineralized organic matter may undergo microbial fractionation (Shearer et al., 1974; Mariotti et al., 1980; Nadelhoffer & Fry, 1988), and is likely to be lighter than fertilizer-derived N, which is often isotopically enriched due to gaseous N loss (Högberg, 1990, 1997; Robinson, 2001). Our study system was a commonly occurring mixture of C_3 fescue and C_4 crabgrass; therefore, we measured the carbon isotope ratio ($\delta^{13}C$) of each species separately and in total harvested aboveground biomass to quantify the relative abundance of each species in each treatment. The $\delta^{13}C$ of C_4 biomass varies from $-11‰$ to $-15‰$, while the $\delta^{13}C$ of C_3 biomass varies from $-20‰$ to $-35‰$ (Dawson et al., 2002). The isotopic composition of each functional type is isotopically distinct; thus, measurements of $\delta^{13}C$ may be used to determine the proportion of C_3 vs. C_4 plants in each treatment.

As increased temperature is expected to increase the metabolic activity of microbes, we hypothesized that increased temperature as well as fertilization would increase N_2O fluxes in this ecosystem, with a corresponding enrichment of $\delta^{15}N$ in the soil. We also hypothesized that plant C/N would decrease in response to fertilizer addition and then remain relatively constant in response to warming due to the large amounts of fertilizer N applied to this system. While in general, plants utilizing the C_4 photosynthetic path-

way have a competitive advantage over C_3 plants in warmer environments based on the relationship between the quantum yield of photosynthesis and temperature (Ehleringer & Bjorkman, 1977), we did not expect the proportion of C_3 and C_4 biomass to change greatly in our 15 month measurement period. Changes in the proportion of C_3 and C_4 plants may be very important for turfgrass management as C_4 plants are often weeds in C_3 lawns, requiring the application of herbicides to turfgrass in addition to fertilizer. Hence, the impacts of warming and fertilization on N_2O fluxes, ecosystem N cycling, and species composition are highly relevant for turfgrass ecosystem management as well as global change.

Materials and methods

Study site

This study was conducted at the University of California, Irvine, Arboretum (33.7°N 117.7°W, 30 m a.s.l.) on a turfgrass lawn dominated by tall fescue [Schedonorus phoenix (Scop.) Holub], a cool season, C_3 species, and crabgrass (Digitaria Haller), a warm season, C_4 species. The site has a Mediterranean climate, with a mean annual air temperature of 18.6 °C and 352 mm of precipitation, primarily falling between November and April. Tall fescue is a widespread turf species in the United States adapted to cool and humid climates (USDA-NRCS, 2007). Crabgrass is an annual weed that commonly invades domesticated lawn grasses. Bermuda grass (Cynodon dactylon L.), a warm season, C_4 species, was initially present at the site at low abundance. The experimental site was fenced to exclude grazing by wildlife, primarily rabbits. The soil type was alkaline alo clay (USDA-NRCS, 2007). Before the experiment, this site was managed as turfgrass for over 16 years (L. Lyons, personal communication, 2008).

Experimental design

Six plots of 1.5 m × 2 m were established on June 10, 2005. Pairs of plots were blocked by slope position at three levels on a slight slope. For each pair, plots were randomly assigned to control or heated treatments. Each plot was split into two subplots that were randomly assigned a low or high fertilizer treatment. In March 2006, these subplots were separated with 10 cm deep plastic trenching materials inserted into the soil. Measurements were concentrated in the central 2500 cm^2 of the subplots, while the surrounding area served as a buffer zone. This design resulted in a 2 × 2 factorial experiment of fertilizer and temperature with three replicates (total of 12 plots). In this paper, the treatments

will be referred to as control (C), high temperature × low fertilizer (T), control temperature × high fertilizer (CN), and high temperature × high fertilizer (TN). The plots were watered with domestic water approximately three times a week during summer (May–September) and twice a week during the winter. Starting in July 2006, all plots received exactly equal amounts of water based on recommendations by Hartin *et al.* (2001). The heated subplots were subjected to an average increase in mean daily surface temperature of $3.5 \pm 1.3\,°C$ (average ± SD) by elevating ceramic infrared heaters (250 W, Exo Terra, Rolf C. Hagen Inc., Montreal, Canada) 1 m over the ground. Plots were heated from July 14, 2005 to December 1, 2006. The applied fertilizer was a common commercial formula of 29:3:4 NPK that contained 28% CON_2H_4, 1% NH_4, 3% P_2O_5, 4% K_2O, and 1% Fe (Vigoro Ultra Turf, Spectrum Group, St Louis, MO, USA). The $\delta^{15}N$ of fertilizer was -0.64 ± 0.09‰. Low fertilizer plots received $76.4\,kg\,N\,ha^{-1}\,yr^{-1}$ in two applications in 2006 ($62.3\,kg\,N\,ha^{-1}$ on April 2 and $14.0\,kg\,N\,ha^{-1}$ on July 20) and high fertilizer plots received $118.5\,kg\,N\,ha^{-1}\,yr^{-1}$ in four applications in 2006 ($62.3\,kg\,N\,ha^{-1}$ on April 2, $14.0\,kg\,N\,ha^{-1}$ on May 21, $28.1\,kg\,N\,ha^{-1}$ on July 20, and $14.0\,kg\,N\,ha^{-1}$ on August 31). The treatments were based on the recommended fertilizer application rates for these varieties, which vary approximately from 50 to more than $200\,kg\,N\,ha^{-1}\,yr^{-1}$ divided into two to six applications during the period from March to November (Reynolds & Flint, 2004). These are also plausible fertilization scenarios for householders based on Osmond & Hardy (2004), who found that five North Carolina communities apply 24–$151\,kg\,N\,ha^{-1}$ to turfgrass and had on average between 1.5 and 3 applications per year.

Measurements

Soil surface temperature (T_S) and volumetric soil water content (θ) were measured at the center of each plot. Surface temperature was measured at the mineral soil surface with copper–constantan thermocouples and θ was measured at 5 cm soil depth with water content reflectometers (CS616, Campbell Scientific Inc., Logan, UT, USA). Air temperature and relative humidity were continuously recorded at 1 m above ground in one location at the site starting March 22, 2006 (CS215 shielded by 41303-5a, Campbell Scientific). Before this, meteorological variables were obtained from the California Irrigation Management Information System (www.cimis.ca.gov, Irvine station #75). All environmental data were recorded every 30 min (AM25 and AM16/32 multiplexer, CR10x logger, Campbell Scientific).

The rate of N_2O efflux from the soil surface was measured using a static, polyvinyl chloride (PVC) chamber lid (height 15 cm, inner diameter 26 cm) containing a septum port. The chamber was placed over the soil surface and the lower rim was surrounded by water-filled plastic tubes to prevent the diffusion of ambient air into the chamber. Gas samples were taken using a syringe at four timed intervals over a 15 or 21 min period, and were injected into airtight, pre-evacuated 12 mL vials. The samples were shipped to the University of Kansas where they were analyzed for N_2O on a Varian CP3800 gas-chromatograph fitted with a ^{63}Ni electron capture detector, operated at high temperature (300–400 °C) using N_2 as the carrier gas. Rates of N_2O–N loss were calculated as the rate of N_2O accumulation over time in the chamber. Temperatures inside the N_2O chambers were assumed to be equivalent with ambient air temperatures measured at the site.

Aboveground biomass production was clipped approximately every month to a height of 4 cm at the same time that the area outside the plots was mowed to the same height (clippings removed). The biomass was oven-dried at 70 °C for at least 48 h and weighed. Subsets of well-mixed bulk biomass, C_3 only, and C_4 only biomass material were removed from each harvest for C, N, and stable isotope analysis. Specific leaf area (SLA) was estimated on August 10, September 11, and October 12, 2006 by determining leaf area on a subset of harvested fresh leaves (IMAGEJ software, US National Institute of Health, http://rsb.info.nih.gov/ij/) and dividing leaf area by dry weight. Another subset of samples were ground to a fine powder and analyzed for % C, % N, $\delta^{13}C$, and $\delta^{15}N$ with an elemental analyzer (Carlo Erba NA 1500 NC, Milan, Italy) coupled to an isotope ratio mass spectrometer (Thermofinnigan Delta Plus, San Jose, CA, USA) at the University of California, Irvine, stable isotope facility. Isotope ratios were referenced to the PDB standard for C and the atmospheric standard for N. The precisions of these measurements were 0.14, 0.06, 0.18, and 1.00 (SD) for $\delta^{15}N$, $\delta^{13}C$, % N, and % C, respectively.

Soil samples were collected on July 17, 2005, June 6 and December 1, 2006 at 0–5 and 5–10 cm depths. In 2005, pretreatment samples were collected from the buffer zone of each plot before the plots were split into high and low fertilizer treatments (total of six samples). In June 2006, one sample was taken from the buffer zone of each subplot (total of 12 samples). Samples were collected from the plot buffer zone to minimize disturbance; measurements of the spatial variability of surface temperature in the plots indicated that the soil in the buffer zone was subjected to a similar increase in temperature as the main measurement area. Soil samples from 2005 were oven-dried at 60 °C, and samples from 2006 were freeze-dried. The different processing methods were related to other measurements not reported here and should not affect the determination

of total soil C, N, and isotopic composition. After the removal of roots and litter, subsamples were ground to a fine powder and acid-treated for the removal of inorganic C. Two grams of each subsample were treated with $0.5 M$ HCl and shaken overnight, centrifuged to remove the remaining acid, washed with $20 mL$ of water, centrifuged and decanted, and freeze-dried. The acid-treated soils were analyzed for C and N content and isotope ratios. The precisions of these measurements were 0.20, 0.06, 0.23, and 1.64 (SD) for $\delta^{15}N$, $\delta^{13}C$, % N, and % C, respectively.

Statistical analysis

Data were analyzed using SAS software v. 9.1 (SAS Institute, Cary, NC, USA). Split-plot, repeated measures ANOVAS were used to determine treatment effects, as well as analyses of covariance (ANCOVAS), when volumetric soil water content co-varied with the dependent variable. Paired *t*-tests were used to analyze pre- vs. post-treatment differences, and ANOVAS were used to analyze the differences among treatments for individual sampling periods. For all analyses, *P*-values < 0.05 were considered significant.

Results

N₂O fluxes

In all treatments, N₂O fluxes were highly variable throughout the year (Fig. 1a). Heated plots had higher fluxes than control plots following the first and second fertilization events on April 3, April 7, and May 22, 2006 ($P = 0.0183$, $P = 0.0360$, and $P < 0.0001$). High N plots had significantly higher fluxes than low N plots on May 22 and July 27, 2006 ($P = 0.0148$ and 0.0327). The highest fluxes were $2.0 mg N m^{-2} day^{-1}$, and were measured in the heated (T and TN) subplots following fertilization in April 2006. The temporal pattern of N₂O fluxes appeared to follow the seasonal pattern in daily average θ in the growing season (Fig. 1b). Before June, when average daily θ was $39.4 \pm 0.42\%$, N₂O fluxes increased immediately following fertilization events, and then decreased (Fig. 1a). Fluxes remained low despite two additional fertilizer applications after June, coincident with a decrease in soil moisture. Although the lawn was watered intensively during summer, soil moisture declined due to unusually high temperatures and high evaporative demand (Czimczik *et al.*, submitted for publication). N₂O fluxes were positively correlated with soil moisture in all four treatments (Fig. 2). An analysis of covariance (ANCOVA) with soil moisture as the covariate showed that there was no difference in the slope of this relationship

among C, CN, and T treatments ($P > 0.05$); however, TN had a greater slope than the other treatments. N₂O fluxes measured from T were higher than C and CN ($P = 0.0391$ and 0.0046, respectively). N₂O fluxes measured from C and CN were not significantly different ($P > 0.05$). For all treatments, there was a decline in N₂O fluxes over time ($P < 0.0001$), indicating that fluxes decreased with the decline in soil moisture in summer, probably caused by an increase in soil temperature during this period (Czimczik *et al.*, submitted for publication). There were interactions of time with temperature on N₂O fluxes before and after the initiation of high fertilizer treatments ($P = 0.0003$ and 0.0002), showing that N₂O fluxes from warmed plots initially increased, but then decreased when soil moisture was low. The integrated flux over the measurement period was $75.2 mg N m^{-2}$ for C and $107.7 mg N m^{-2}$ for T. After the initiation of high fertilizer treatments, the integrated flux was 16.8, 11.4, 32.6, and $16.3 mg N m^{-2}$ from C, CN, T, and TN, respectively.

Plant isotopic and chemical composition

In C₄ plants, leaf C/N increased with time before the fertilization of high N treatments (CN and TN), but then did not change afterwards (Fig. 3a and Table 1; repeated measures ANOVA, $P = 0.0002$ and $P > 0.05$, respectively). The application of fertilizer in the high N treatments decreased leaf C/N of C₄ plants (Table 1 and Fig. 3a; $P = 0.0028$). On October 12, 2006, there was a marginally significant decrease in C/N of C₄ plants in the high N plots ($P = 0.0613$). There was no effect of temperature on C/N in C₄ plants (Table 1; $P > 0.05$), although on October 12, 2006, heated plots had lower C/N ($P = 0.0498$). In C₃ plants, leaf C/N increased with time before the fertilization of high N plots, and then decreased with time afterwards (Fig. 3b and Table 1; repeated measures ANOVA, $P < 0.0001$ and $P = 0.0021$, respectively). At the end of the experiment, C₃ plants in heated (T and TN) plots had lower C/N than in the control (C and CN) plots (Fig. 3b; $P = 0.0410$). After the establishment of CN and TN plots, C/N of C₃ plants were lower than those of C₄ plants (repeated measures ANOVA, $P < 0.0001$). There was a negative correlation between C/N and $\delta^{15}N$ of aboveground biomass for C₄ grasses in each treatment (Fig. 4). This relationship was particularly strong in the C treatment, where leaf $\delta^{15}N$ was also the most isotopically depleted, with a value as low as $-2.9‰$. An analysis of covariance (ANCOVA) with C/N as the covariate showed that there was no difference in the slope among treatments ($P > 0.05$). Heated plots were more enriched in $\delta^{15}N$ ($P < 0.0001$) with no effect of fertilization ($P > 0.05$). For C₃ plants, C/N and $\delta^{15}N$ were correlated only in the
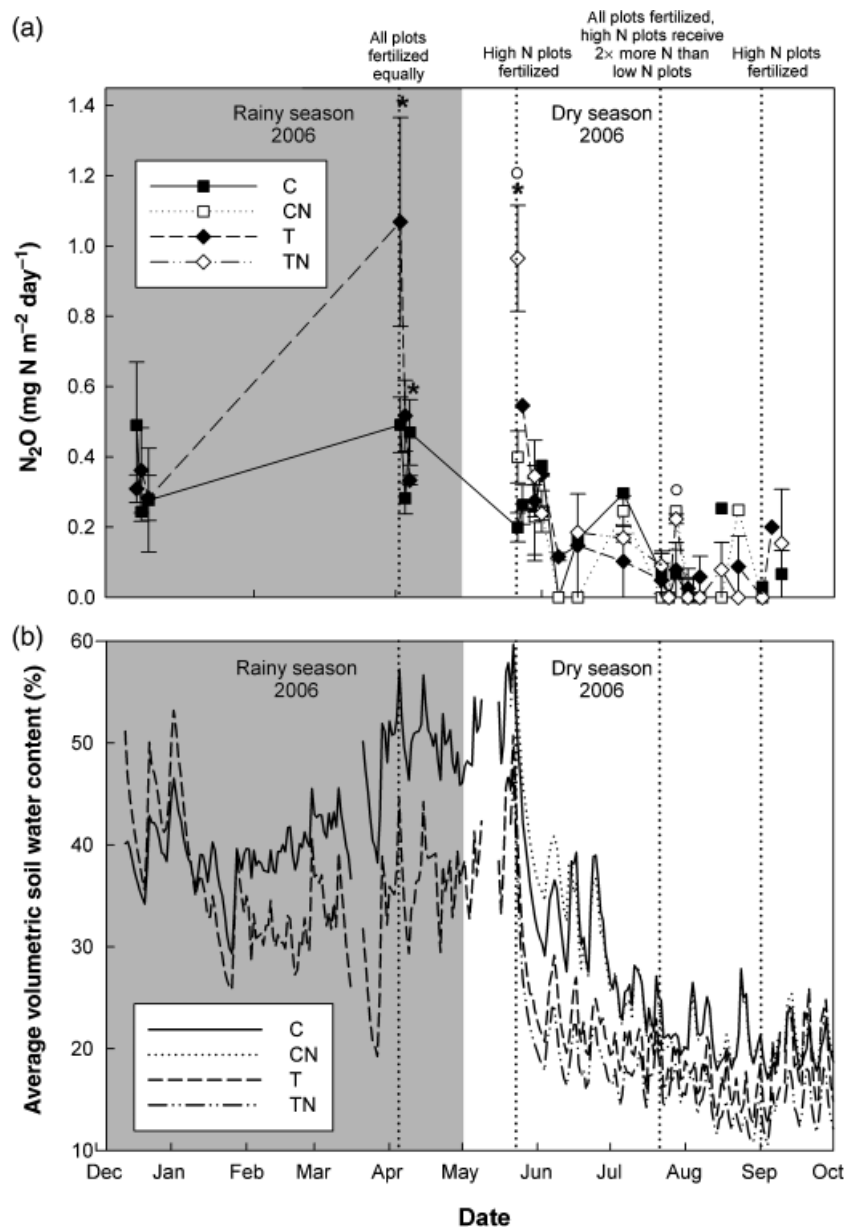
Fig. 1 (a) N_2O fluxes in control (C), high temperature (T), high N (CN), and high N × high temperature (TN) treatments. Dotted lines indicate fertilization events. Warming treatment began on July 14, 2005 (not shown). The asterisk (*) shows a significant treatment difference due to temperature, and circles (○) show treatment differences due to fertilization. Error bars show the standard error. (b) Average volumetric soil water content (%) in each treatment.

T treatment (not shown; $P = 0.0019$). In the C treatment, there was a significant relationship between $\delta^{13}C$ of C_3 plant biomass and soil moisture (Fig. 5a; $P = 0.0021$), and a marginally significant relationship for the CN treatment (Fig. 5b; $P = 0.0507$).

All treatments showed increases in C_4 aboveground biomass with time as estimated with $\delta^{13}C$ of bulk harvests (repeated measures ANOVA on $\delta^{13}C$; $P < 0.0001$). Fertilization and its interactions were nonsignificant for

$\delta^{13}C$ ($P > 0.05$), such that fertilization treatments were combined to evaluate the effect of warming (Fig. 6a). Before the warming treatment, the $\delta^{13}C$ of bulk harvests from both control and treatment plots were similar (-25.6 ± 0.7‰ and -24.5 ± 1.2‰, respectively), indicating similar proportions of C_3 vs. C_4 species. In a period of approximately 7 months, the aboveground biomass in heated plots became isotopically enriched by 4.9‰ on average (Fig. 6a), indicating a greater proportion of C_4
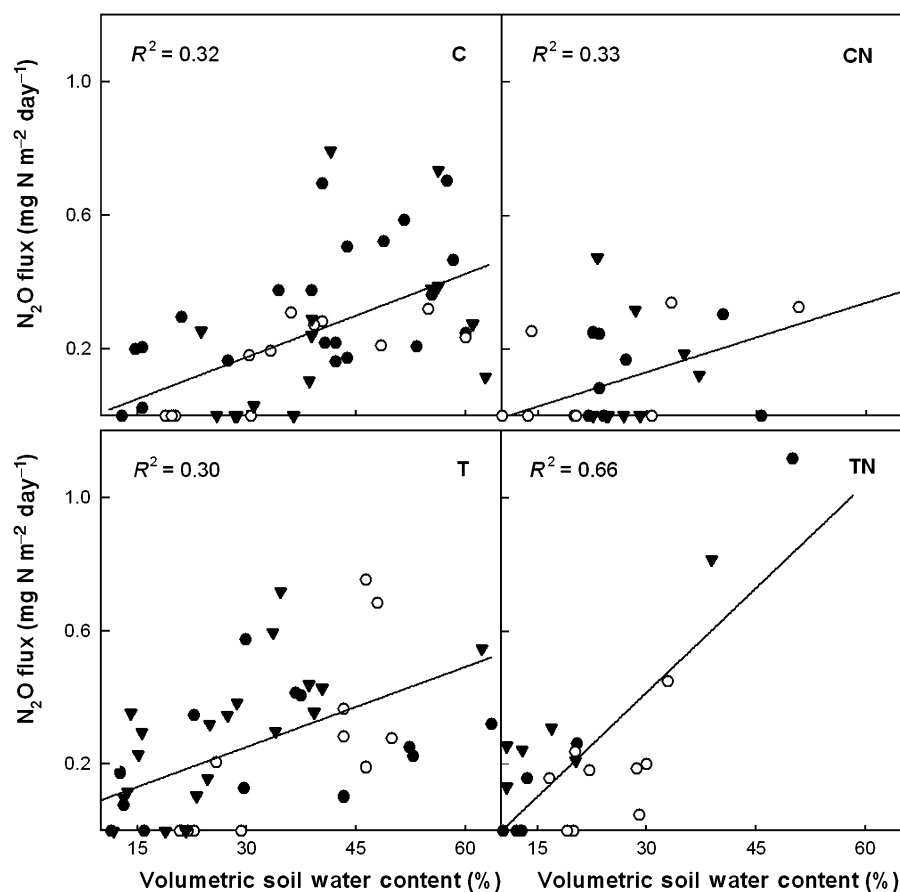
Fig. 2 The relationship between N_2O flux and volumetric soil moisture in each treatment. Treatments are abbreviated as in Fig. 1. Symbols represent different blocks. $P < 0.0001$ for C, T, and TN; $P = 0.003$ for CN; R^2 were 0.32, 0.33, 0.30, and 0.66 for C, CN, T, and TN treatments, respectively.
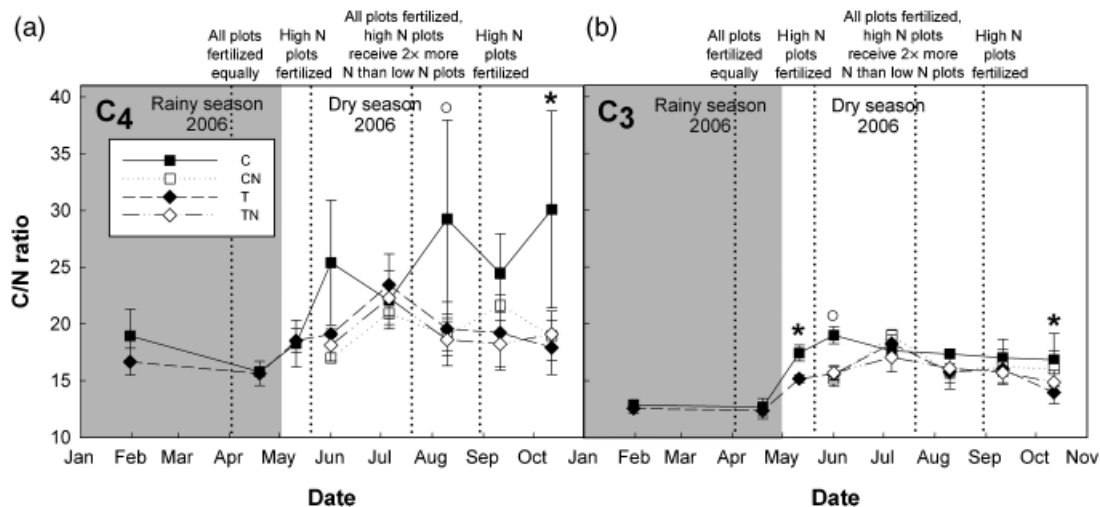

Fig. 3 C/N ratio of (a) C_4 aboveground biomass and (b) C_3 aboveground biomass. Dotted lines indicate fertilization events. Asterisks (*) show significant treatment differences due to temperature, and circles (○) show treatment differences due to fertilization. Treatments are abbreviated as in Fig. 1. Error bars show the standard error.

Table 1 F- and P-values for treatment effects on C_3 and C_4 foliar C/N for time periods before and after fertilization of high N plots (CN and TN)

	C_4				C_3			
	F-value		P-value		F-value		P-value	
	Prefertilization	Postfertilization	Prefertilization	Postfertilization	Prefertilization	Postfertilization	Prefertilization	Postfertilization
Temperature	0.24	3.66	0.7075	0.1959	9.56	5.29	0.0906	0.1481
Fertilizer		10.22		0.0028*		1.46		0.2342
Time	26.69	0.17	0.0002*	0.9506	50.82	5.06	<.0001*	0.0021*
Temperature × fertilizer		7.33		0.0102*		1.79		0.1885
Temperature × time	9.37	1.25	0.0063*	0.3051	3.52	0.53	0.0444*	0.7137
Fertilizer × time		0.62		0.6498		0.99		0.4250

*Significant effects ($P < 0.05$).

plant material. Repeated measures ANOVAs showed no effect of warming on $\delta^{13}C$ before January 31, 2006 ($P > 0.05$), but an effect of warming on $\delta^{13}C$ following January 31, 2006 ($P = 0.0275$). The isotope ratios of each species measured at each harvest were utilized as end-members to calculate the proportion of C_3 vs. C_4 biomass, and confirmed that changes in $\delta^{13}C$ of the bulk harvest were dominated by changes in community composition, rather than changes in $\delta^{13}C$ of C_3 plants (Fig. 6b). The C_4 biomass was $30 \pm 16\%$ greater in the heated plots by October 2006 (Fig. 6b).

Soil chemical and isotopic composition

There was no change in N and $\delta^{13}C$ of soil in response to fertilization, temperature, or time (Fig. 7a and c, $P > 0.05$). However, after the warming treatment was applied, soil $\delta^{15}N$ in heated plots were enriched relative to control plots at the 0–5 cm depth (Fig. 7b, $P = 0.016$ in June 2006) and at the 5–10 cm depth (Fig. 7b, $P = 0.031$ in June 2006 and $P = 0.007$ in December 2006).

Plant canopy properties

Specific leaf area averaged $181.9 \pm 11.1 \, cm^2 \, g^{-1}$ (mean $\pm$ SE) in C_3 plants and 217.8 ± 5.6 (mean $\pm$ SE) in C_4 plants. There were no significant effects of temperature, fertilizer, time, or their interactions on SLA for C_3 or C_4 plants (repeated measures ANOVA, $P > 0.05$). Leaf area index (LAI) changed significantly over time for C_3 and C_4 plants ($P = 0.0367$ and 0.0006 respectively), but was not affected by fertilization ($P > 0.05$). C_3 plants in heated plots had lower LAI than control plots on two sampling dates (August 10 and September 11, $P = 0.0088$ and 0.0422, respectively). The LAI of C_4 plants was unaffected by heating. Average total plot LAI in 2006 ranged from $0.60 \pm 0.08 \, m^2 \, m^{-2}$ (mean $\pm$ SE) in August–September to $0.80 \pm 0.11 \, m^2 \, m^{-2}$ (mean $\pm$ SE) in October.

Discussion

Measured N_2O fluxes were within the reported range for turfgrass (Maggiotto et al., 2000; Kaye et al., 2004; Bremer, 2006). As hypothesized, fertilizer application caused an increase in N_2O fluxes, although the effects were significant only on two sampling dates. Maggiotto et al. (2000) and Bremer (2006) also reported increased N_2O emissions from fertilized turfgrass plots relative to control plots. The positive effect of warming on N_2O fluxes was consistent with our hypothesis. We are not aware of prior warming manipulations in turfgrass, and field warming studies in other ecosystems have shown conflicting effects on N_2O fluxes. Kamp et al. (1998),
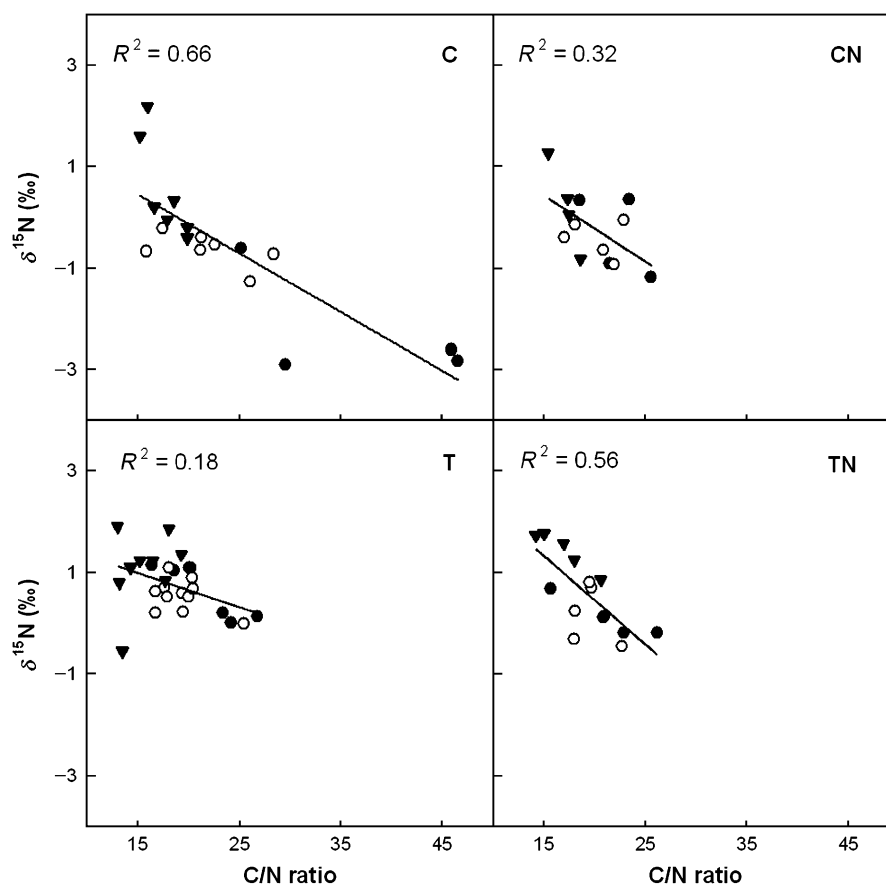
Fig. 4 The relationship between $\delta^{15}N$ and C/N ratio of C_4 aboveground biomass in each treatment. Treatments are abbreviated as in Fig. 1. Symbols represent different blocks. $P < 0.0001$, 0.0446, 0.0334, 0.0013 for C, CN, T, TN, respectively; R^2 were 0.66, 0.32, 0.18, 0.56 for C, CN, T, and TN treatments, respectively.
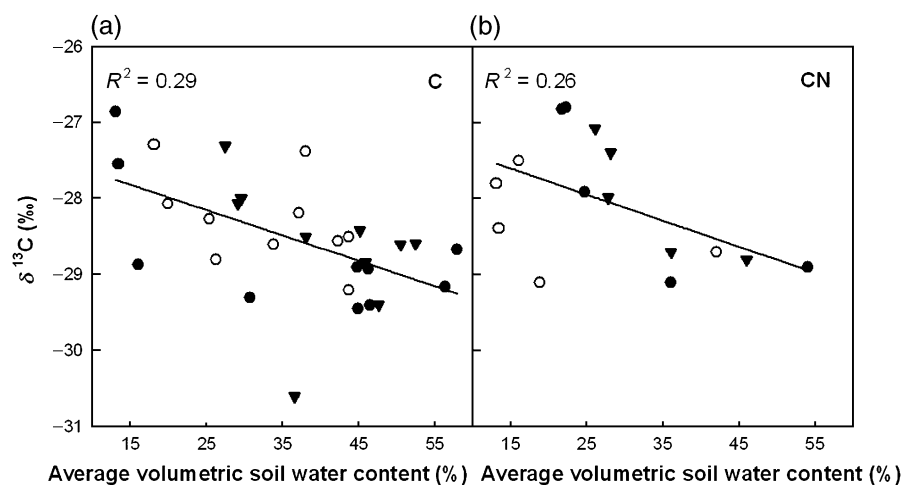

Fig. 5 Average volumetric soil water content before harvest vs. $\delta^{13}C$ of C_3 aboveground biomass in (a) control (C) and (b) high N (CN) treatments. Symbols distinguish different blocks.
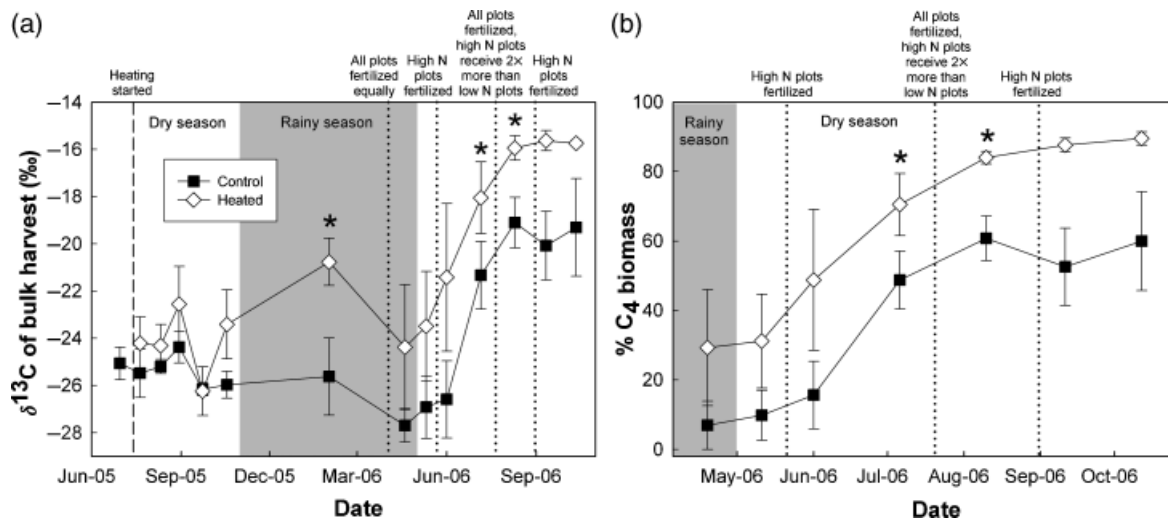
Fig. 6 (a) The $\delta^{13}C$ of bulk turfgrass aboveground biomass. (b) The percentage of total aboveground biomass comprised of C_4 plants as calculated from the isotopic composition of C_3 biomass, C_4 biomass, and the bulk harvest. The dashed line indicates the beginning of the warming treatment, and dotted lines indicate fertilization events. High and low N treatments are combined as they were not statistically different. Asterisks (*) show significant treatment differences due to temperature. Error bars show the standard error.
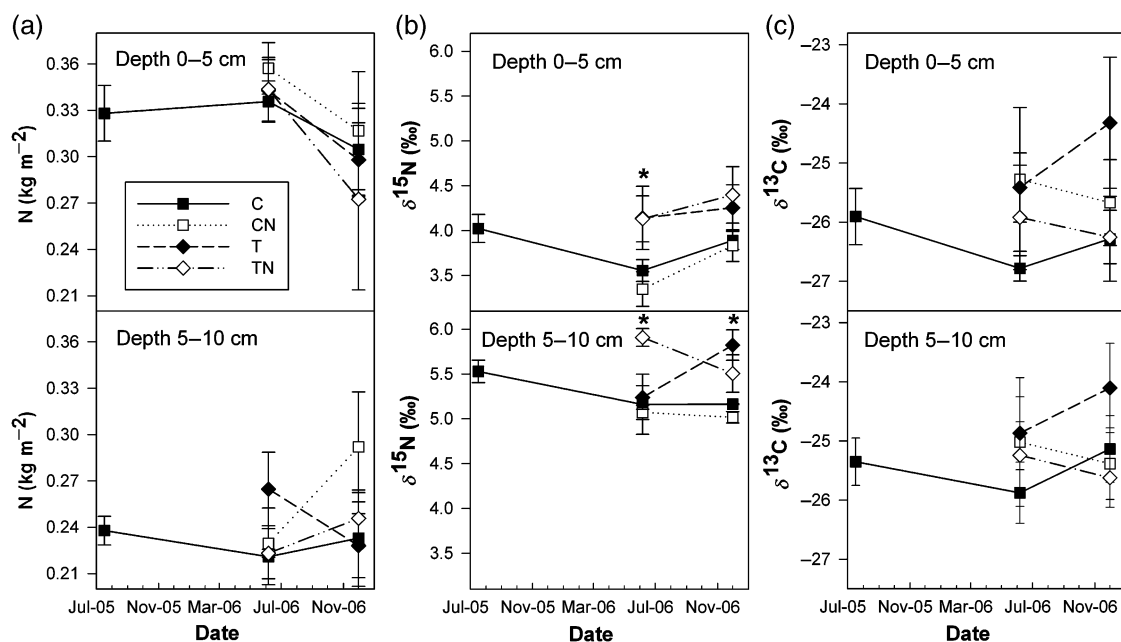

Fig. 7 (a) N (kg m^{-2}), (b) $\delta^{15}N$, and (c) $\delta^{13}C$ of soil measured at two depths at the beginning of the warming treatment (July 2005), following fertilization (June 2006), and at the end of the experiment (December 2006). Treatments are abbreviated as in Fig. 1. Asterisks (*) show significant treatment differences due to temperature. Error bars show the standard error.

who applied a 3 °C warming to wheat and fallow fields, did not find differences in cumulative emissions of N_2O, but did find that heated fallow plot emissions were three times higher than control plots during summer. Conversely, Hantschel *et al.* (1995) found lower N_2O fluxes in 3 °C heated wheat fields during winter in Germany. Peterjohn *et al.* (1994) did not find an effect of a 5 °C warming on N_2O fluxes in a deciduous hardwood forest. McHale *et al.* (1998) did not find a strong response of heating on N_2O fluxes in plots heated to 2.5, 5.0, or 7.5 °C above ambient in a northern hardwood forest. In a companion study, we found increases in ecosystem respiration in heated plots relative to control plots in winter when aboveground biomass production was low (Czimczik *et al.*, submitted for publication). It is possible that heating increased heterotrophic respira-

tion and N-mineralization. Higher rates of N-mineralization in heated plots may have increased available ammonium pools, causing the positive response of N_2O fluxes to heating. In summer, N_2O fluxes declined, perhaps because low soil moisture suppressed the hydrolysis of applied urea fertilizer to ammonium, the substrate for N_2O production via nitrification. The activity of urease, the enzyme responsible for urea hydrolysis, peaks near field capacity and declines with decreasing soil moisture (Vlek & Carter, 1983; Sahrawat, 1984). In addition, urea hydrolysis strongly depends on the incorporation of urea into soil through diffusion of dissolved ammonia (Sadeghi *et al.*, 1989). Thus, it is possible that fertilizer was unavailable for microbial processes during the period of low soil moisture in summer, resulting in low N_2O fluxes (Fig. 1). A reduction in soil denitrification, which requires anoxic conditions associated with soil moisture, may also explain the decline of N_2O fluxes when soil moisture was low.

Leaf C/N of C_3 plants decreased in the period following fertilization of high N plots, supporting our hypothesis (Fig. 3b). However, C/N of C_4 plants did not change over time in response to fertilization, contrary to our hypothesis (Fig. 3a), indicating plant N-limitation despite the additional application of fertilizer. The negative correlation between leaf C/N of C_4 grasses and leaf $\delta^{15}N$ (Fig. 4) suggests the greater use of isotopically light forms of N when N was limiting. The labile soil organic matter that is fractionated by mineralization (Shearer *et al.*, 1974; Mariotti *et al.*, 1980; Nadelhoffer & Fry, 1988) is likely to be isotopically lighter than fertilizer N, which can undergo isotopic enrichment due to rapid gaseous loss through volatilization (Högberg, 1990, 1997; Robinson, 2001). As the soils in this study were alkaline (pH ranging from 7 to 9), ammonia volatilization may have been an important pathway for gaseous N loss (Kirchmann & Witter, 1989). Thus, the correlation between C/N of C_4 grasses and $\delta^{15}N$ in all treatments suggests greater use of fertilizer-derived ammonium during wetter conditions earlier in the year vs. uptake of N mineralized from organic matter late in the year when applied fertilizer was not biologically available. Additional measurements of inorganic N forms and their isotopic composition would be required to validate this interpretation, as many processes may cause fractionation of plant and soil N (Evans, 2001; Robinson, 2001). Plants in the high N treatments showed greater isotopic enrichment and weaker correlations between $\delta^{15}N$ and C/N than the C treatment, possibly because of increased fertilizer uptake. The T treatment also showed a weaker trend and greater isotopic enrichment than the C treatment, possibly because of greater N_2O and other gaseous losses such as NH_3, NO, and N_2 (Fig. 1). This is supported by the soil data, which shows enrichment of $\delta^{15}N$ in heated plots (Fig. 7b).

The $\delta^{13}C$ of leaves is determined by c_i/c_a, the ratio of CO_2 inside and outside of the leaf, which is in turn determined by the balance between photosynthesis and stomatal conductance (Farquhar *et al.*, 1989). Previous studies have reported more positive $\delta^{13}C$ values in response to reduced soil moisture in C_3 grasses, including tall fescue turfgrass (Johnson & Bassett, 1991; Ebdon *et al.*, 1998; Johnson & Li, 1999). In the C and CN treatments, we found a direct correlation between $\delta^{13}C$ and soil moisture in the C_3 plants (Fig. 5), which suggests stomatal closure in response to declining soil moisture. In the T and TN treatments, there was no correlation with variations in soil moisture. However, the $\delta^{13}C$ of C_3 plants was significantly more enriched in the two high temperature treatments than the C and CN treatments (ANOVA, $P = 0.0385$), suggesting that plants in the T and TN treatments were generally more water-stressed, consistent with the lower soil moisture (Fig. 1b).

One of the most significant results of this study was unexpected: the temperature treatments facilitated a rapid ($\sim$7 months) shift in C_4 weed biomass relative to C_3 plant biomass, as shown by the time series of $\delta^{13}C$ of total biomass (Fig. 6a). C_4 biomass was $30 \pm 16\%$ greater in the heated plots at the end of the experiment in October 2006 (Fig. 6b). Immediately following the first spring fertilizer application, the $\delta^{13}C$ of total biomass in the control and high temperature plots had become more similar and were not significantly different, possibly because fertilization enabled the C_3 plants to compete more successfully with the C_4 species, which have higher N-use efficiency (Sage & Pearcy, 1987). A greater ability to take up isotopically heavy N early in the growing season may also have contributed to the competitive advantage of C_4 species in all plots, and particularly in heated plots, and may explain why C_4 C/N did not change over time. Because C_3 and C_4 plants can have different patterns of N uptake and allocation, changes in C_3/C_4 community composition could ultimately influence other aspects of N cycling such as N_2O fluxes, as both microbes and plants share the same soil N pool.

Plants utilizing the C_4 photosynthetic pathway have a competitive advantage over C_3 plants in warmer environments (Ehleringer & Bjorkman, 1977). Because most weeds are C_4 plants (Holm, 1977), there is great concern about the possibility of more widespread weed invasions as a result of global climate change (Patterson, 1995; Dukes & Mooney, 1999; Sage & Kubien, 2003; Schmitz, 2006). However, field experiments often fail to confirm simple predictions based on physiological

principles. For example, C_4 plants were predicted to be relatively unresponsive to elevated atmospheric CO_2, but this has been contradicted by experimental evidence (Dukes & Mooney, 1999). As most field warming experiments have been conducted in high latitude ecosystems where C_4 plants are largely absent, there have been few of these studies on C_3 vs. C_4 plants (White *et al.*, 2000, 2001; Wan *et al.*, 2005; Luo, 2007), particularly in agricultural and highly managed ecosystems, e.g. White *et al.*, 2000, 2001; Ziska, 2000, 2003; Derner *et al.*, 2003; Fuhrer, 2003. Weed expansion under higher temperatures is particularly relevant because herbicide is commonly applied to fescue lawns to control the invasion of C_4 weeds such as crabgrass, often at higher application rates than in other types of agriculture (Templeton *et al.*, 1998).

Conclusions

This study showed reduced N_2O fluxes and plant N limitation with declining soil moisture and increases in N_2O fluxes with warming. Negative correlations between C/N of C_4 plants and plant $\delta^{15}N$ suggest that N limitation was associated with the increased use of isotopically depleted N sources, such as mineralized organic matter. The increases in N_2O fluxes with warming suggest that soil N_2O fluxes could serve as a positive feedback to global warming in turfgrass. The strong influence of soil moisture on N_2O fluxes suggests that best management practices for turfgrass should optimize the tradeoff between soil moisture enhancement of urea fertilizer hydrolysis and gaseous N emissions. That is, soil moisture should be regulated so that plant use of fertilizer is maximized while gaseous N loss is minimized (e.g. Matson *et al.*, 1998).

These results also provide *in situ* evidence for rapid, warming-induced C_4 weed expansion in turfgrass and suggest that other managed systems, such as agricultural crops, may experience rapid weed invasions or changes in community composition in response to warming. Because turfgrass is often associated with urban and suburban land cover, the urban heat island effect as well as climate change may exacerbate weed invasions, which would require more intensive management, e.g. herbicide application, to manage species composition.

Acknowledgements

References

Alig RJ, Kline JD, Lichtenstein M (2004) Urbanization on the US landscape: looking ahead in the 21st century. *Landscape and Urban Planning*, **69**, 219–234.

Arnfield AJ (2003) Two decades of urban climate research: a review of turbulence, exchanges of energy and water, and the urban heat island. *International Journal of Climatology*, **23**, 1–26.

Barnard R, Leadley PW, Hungate BA (2005) Global change, nitrification, and denitrification: a review. *Global Biogeochemical Cycles*, **19**, GB1007.

Bouwman AF, Boumans LJM, Batjes NH (2002) Emissions of N_2O and NO from fertilized fields: summary of available measurement data. *Global Biogeochemical Cycles*, **16**, 1058–1070.

Bremer DJ (2006) Nitrous oxide fluxes in turfgrass: effects of nitrogen fertilization rates and types. *Journal of Environmental Quality*, **35**, 1678–1685.

Brumme R (1995) Mechanisms of carbon and nutrient release and retention in beech forest gaps: 3. Environmental-regulation of soil respiration and nitrous-oxide emissions along a microclimatic gradient. *Plant and Soil*, **168/169**, 593–600.

Czimczik CI, Bijoor NS, Pataki DE. The effects of temperature and fertilization on the carbon budget of an urban lawn in Southern California, submitted for publication.

Dawson TE, Mambelli S, Plamboeck AH, Templer PH, Tu KP (2002) Stable isotopes in plant ecology. *Annual Review of Ecology and Systematics*, **33**, 507–559.

Derner JD, Johnson HB, Kimball BA *et al.* (2003) Above- and below-ground responses of C-3-C-4 species mixtures to elevated CO_2 and soil water availability. *Global Change Biology*, **9**, 452–460.

Dukes JS, Mooney HA (1999) Does global change increase the success of biological invaders? *Trends in Ecology & Evolution*, **14**, 135–139.

Ebdon JS, Petrovic AM, Dawson TE (1998) Relationship between carbon isotope discrimination, water use efficiency, and evapotranspiration in Kentucky bluegrass. *Crop Science*, **38**, 157–162.

Ehleringer J, Bjorkman O (1977) Quantum yields for CO_2 uptake in C3 and C4 plants – dependence on temperature, CO_2, and O_2 concentration. *Plant Physiology*, **59**, 86–90.

Evans RD (2001) Physiological mechanisms influencing plant nitrogen isotope composition. *Trends in Plant Science*, **6**, 121–126.

Farquhar GD, Ehleringer JR, Hubick KT (1989) Carbon isotope discrimination and photosynthesis. *Annual Review of Plant Physiology and Plant Molecular Biology*, **40**, 503–537.

Flechard CR, Ambus P, Skiba U *et al.* (2007) Effects of climate and management intensity on nitrous oxide emissions in grassland systems across Europe. *Agriculture Ecosystems & Environment*, **121**, 135–152.

Franco G (2002) *Inventory of California greenhouse gas emissions and sinks: 1990–1999.* Report No. 600-02-001F.

Fuhrer J (2003) Agroecosystem responses to combinations of elevated CO_2, ozone, and global climate change. *Agriculture Ecosystems & Environment*, **97**, 1–20.

Hantschel RE, Kamp T, Beese F (1995) Increasing the soil temperature to study global warming effects on the soil nitrogen cycle in agroecosystems. *Journal of Biogeography*, **22**, 375–380.

Hartin J, Geisel PM, Unruh CL (2001) *Lawn watering guide for California*. Publication 8044. University of California, Agriculture and Natural Resources, Davis.

Högberg P (1990) Forests losing large quantities of nitrogen have elevated $^{15}N/^{14}N$ ratios. *Oecologia*, **84**, 229–231.

Högberg P (1997) Tansley review no 95 – ^{15}N natural abundance in soil–plant systems. *New Phytologist*, **137**, 179–203.

Holm L (1977) Weeds and water in world food production. *Weed Science*, **25**, 338–342.

IPCC (2001) In: *Climate Change 2001: the Scientific Basis – Contribution of Working Group I to the Third Assessment Report of the Intergovernmental Panel on Climate Change (IPCC)* (eds Houghton JT, Ding Y, Griggs DJ, Naguer M, van der Linden PJ, Dai X, Maskell K, Johnson CA), p. 388. Cambridge University Press, Cambridge, UK.

Johnson RC, Bassett LM (1991) Carbon isotope discrimination and water-use efficiency in 4 cool-season grasses. *Crop Science*, **31**, 157–162.

Johnson RC, Li YY (1999) Water relations, forage production, and photosynthesis in tall fescue divergently selected for carbon isotope discrimination. *Crop Science*, **39**, 1663–1670.

Kamp T, Steindl H, Hantschel RE, Beese F, Munch JC (1998) Nitrous oxide emissions from a fallow and wheat field as affected by increased soil temperatures. *Biology and Fertility of Soils*, **27**, 307–314.

Kaye JP, Burke IC, Mosier AR, Guerschman JP (2004) Methane and nitrous oxide fluxes from urban soils to the atmosphere. *Ecological Applications*, **14**, 975–981.

Kaye JP, McCulley RL, Burke IC (2005) Carbon fluxes, nitrogen cycling, and soil microbial communities in adjacent urban, native and agricultural ecosystems. *Global Change Biology*, **11**, 575–587.

Khalil MAK, Rasmussen RA, Shearer MJ (2002) Atmospheric nitrous oxide: patterns of global change during recent decades and centuries. *Chemosphere*, **47**, 807–821.

Kirchmann H, Witter E (1989) Ammonia volatilization during aerobic and anaerobic manure decomposition. *Plant and Soil*, **115**, 35–41.

Klein JA, Harte J, Zhao XQ (2007) Experimental warming, not grazing, decreases rangeland quality on the Tibetan Plateau. *Ecological Applications*, **17**, 541–557.

Luo Y (2007) Terrestrial carbon-cycle feedback to climate warming. *Annual Review of Ecology and Systematics*, **38**, 683–712.

Maggiotto SR, Webb JA, Wagner-Riddle C, Thurtell GW (2000) Nitrous and nitrogen oxide emissions from turfgrass receiving different forms of nitrogen fertilizer. *Journal of Environmental Quality*, **29**, 621–630.

Mariotti A, Pierre D, Vedy JC, Bruckert S, Guillemot J (1980) The abundance of natural nitrogen-15 in the organic matter of soils along an altitudinal gradient. *Catena*, **7**, 293–300.

Matson PA, Naylor R, Ortiz-Monasterio I (1998) Integration of environmental, agronomic, and economic aspects of fertilizer management. *Science*, **280**, 112–115.

McHale PJ, Mitchell MJ, Bowles FP (1998) Soil warming in a northern hardwood forest: trace gas fluxes and leaf litter decomposition. *Canadian Journal of Forest Research – Revue Canadienne De Recherche Forestiere*, **28**, 1365–1372.

Milesi C, Running SW, Elvidge CD, Dietz JB, Tuttle BT, Nemani RR (2005) Mapping and modeling the biogeochemical cycling of turf grasses in the United States. *Environmental Management*, **36**, 426–438.

Mosier A, Kroeze C, Nevison C, Oenema O, Seitzinger S, van Cleemput O (1998) Closing the global N_2O budget: nitrous oxide emissions through the agricultural nitrogen cycle – OECD/IPCC/IEA phase II development of IPCC guidelines for national greenhouse gas inventory methodology. *Nutrient Cycling in Agroecosystems*, **52**, 225–248.

Nadelhoffer KF, Fry B (1988) Controls on natural ^{15}N and ^{13}C abundances in forest soil organic-matter. *Soil Science Society of America Journal*, **52**, 1633–1640.

Osmond DL, Hardy DH (2004) Characterization of turf practices in five North Carolina communities. *Journal of Environmental Quality*, **33**, 565–575.

Patterson DT (1995) Weeds in a changing climate. *Weed Science*, **43**, 685–700.

Peterjohn WT, Melillo JM, Steudler PA, Newkirk KM, Bowles FP, Aber JD (1994) Responses of trace gas fluxes and N availability to experimentally elevated soil temperatures. *Ecological Applications*, **4**, 617–625.

Qian YL, Follett RF (2002) Assessing soil carbon sequestration in turfgrass systems using long-term soil testing data. *Agronomy Journal*, **94**, 930–935.

Read JJ, Morgan JA (1996) Growth and partitioning in *Pascopyrum smithii* (C_3) and *Bouteloua gracilis* (C_4) as influenced by carbon dioxide and temperature. *Annals of Botany*, **77**, 487–496.

Reynolds CA, Flint ML (2004) *The UC guide to healthy lawns*. University of California Statewide Integrated Pest Management Program, Davis, CA.

Robinson D (2001) $\delta^{15}N$ as an integrator of the nitrogen cycle. *Trends in Ecology & Evolution*, **16**, 153–162.

Rustad LE, Campbell JL, Marion GM *et al.* (2001) A meta-analysis of the response of soil respiration, net nitrogen mineralization, and aboveground plant growth to experimental ecosystem warming. *Oecologia*, **126**, 543–562.

Sadeghi AM, Kissel DE, Cabrera ML (1989) Estimating molecular-diffusion coefficients of urea in unsaturated soil. *Soil Science Society of America Journal*, **53**, 15–18.

Sage RF, Kubien DS (2003) Quo vadis C_4? An ecophysiological perspective on global change and the future of C_4 plants. *Photosynthesis Research*, **77**, 209–225.

Sage RF, Pearcy RW (1987) The nitrogen use efficiency of C_3 and C_4 plants: 2. Leaf nitrogen effects on the gas-exchange characteristics of *Chenopodium album* (L) and Amaranthusretroflexus (L). *Plant Physiology*, **84**, 959–963.

Sahrawat KL (1984) Effects of temperature and moisture on urease activity in semi-arid tropical soils. *Plant and Soil*, **78**, 401–408.

Schmitz U (2006) Increase of alien and C_4 plant species in annual river bank vegetation of the River Rhine. *Phytocoenologia*, **36**, 393–402.

Shaver GR, Canadell J, Chapin FS *et al*. (2000) Global warming and terrestrial ecosystems: a conceptual framework for analysis. *Bioscience*, **50**, 871–882.

Shearer G, Duffy J, Kohl DH, Commoner B (1974) Steady-state model of isotopic fractionation accompanying nitrogen transformations in soil. *Soil Science Society of America Journal*, **38**, 315–322.

Skiba UM, Sheppard LJ, MacDonald J, Fowler D (1998) Some key environmental variables controlling nitrous oxide emissions from agricultural and semi-natural soils in Scotland. *Atmospheric Environment*, **32**, 3311–3320.

Templeton SR, Zilberman D, Yoo SJ (1998) An economic perspective on outdoor residential pesticide use. *Environmental Science & Technology*, **32**, 416A–423A.

USDA-NRCS (2007) *National Soil Survey Characterization Data (http://soils.usda.gov/, 11 July 2007)*. National Soil Survey Center, Lincoln, NE.

USDA-NRCS (2007) *The PLANTS Database (http://plants.usda.gov, 10 July 2007)*. National Plant Data Center, Baton Rouge, LA, USA.

Vlek PLG, Carter MF (1983) The effect of soil environment and fertilizer modifications on the rate of urea hydrolysis. *Soil Science*, **136**, 56–63.

Wan SQ, Hui DF, Wallace L, Luo YQ (2005) Direct and indirect effects of experimental warming on ecosystem carbon processes in a tallgrass prairie. *Global Biogeochemical Cycles*, **19**, doi: 10.1029/2004GB002315.

Weiss RF (1981) The temporal and spatial distribution of tropospheric nitrous oxide. *Journal of Geophysical Research – Oceans and Atmospheres*, **86**, 7185–7195.

White TA, Campbell BD, Kemp PD, Hunt CL (2000) Sensitivity of three grassland communities to simulated extreme temperature and rainfall events. *Global Change Biology*, **6**, 671–684.

White TA, Campbell BD, Kemp PD, Hunt CL (2001) Impacts of extreme climatic events on competition during grassland invasions. *Global Change Biology*, **7**, 1–13.

Ziska LH (2000) The impact of elevated CO on yield loss from a C_3 and C_4 weed in field-grown soybean. *Global Change Biology*, **6**, 899–905.

Ziska LH (2003) Evaluation of yield loss in field sorghum from a C_3 and C_4 weed with increasing CO_2. *Weed Science*, **51**, 914–918.