### Before tutorial

Everything, which is written on the green background, is commands, that should be executed in terminal window, string-by-string, each following with the ENTER button.

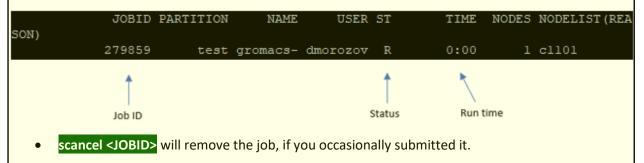
Please note that <...> in the commands means, that everything, including <> symbols, must be replaced with your own specific information. Be careful!

Some exercises will require usage of less Linux tool for looking up into the content of files. In case you are not familiar with it, here is a short list of hotkeys, which could be used inside LESS editor:

- q exit
- / search for a pattern which will take you to the next occurrence.
- n for next match in forward
- N (SHIFT+n) for previous match in backward
- g go to the start of file
- G (SHIFT+g) go to the end of file

All exercises will require you to submit job for computing using sbatch run.sh command. To check status of your job following commands would be useful.

• squeue -u <your username> - checks status of all your jobs. Output will look like that:



#### Setting up tutorial environment

Let's start the tutorial with the following steps:

1) Execute commands in the terminal:

export WRKDIR=/work/ta011/ta011/<your login>

export PATH=/home/ta011/ta011/shared/GRM-CP2K/exe:\$PATH

2) Now go to your work directory and copy tutorial files:

cd \$WRKDIR

cp -r /work/ta011/ta011/shared/tutorial ./

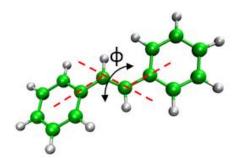
3) Go to the copied tutorial directory

cd tutorial

# Exercise 1: Umbrella sampling simulation with QM/MM

1) Go to stilbene\_vacuum directory:

# cd stilbene\_vacuum



2) Look up in the table and pick-up starting structure and dihedral angle value (anyone you want)

Structure	Dihedral angle, φ
1	-180
2	-173
3	-166
4	-159
5	-152
6	-145
7	-138
8	-131
9	-124
10	-117
11	-110
12	-103
13	-96
14	-89
15	-82
16	-75
17	-68
18	-61
19	-54
20	-47
21	-40
22	-33
23	-26
24	-19
25	-12
26	-5
27	2
28	9
29	16
30	23

3) Copy chosen starting structure:

### cp eq\_gro/md-equilb<your number>.gro ./conf.gro

4) Modify Gromacs input file qmmm\_md\_umbrella.mdp with value of your value of Dihedral angle:

## sed -i "s/@umbr@/<your dihedral angle>/" qmmm\_md\_umbrella.mdp

You can also modify **pull-coord1-init** option in the **qmmm\_md\_umbrella.mdp** file with **vim** or any other editor.

5) Add group of atoms which will be treated with QM to the index file (in that case all atoms are QM):

# gmx\_cp2k make\_ndx -f conf.gro -n index.ndx

> 0

#### > name 7 QMatoms



6) Generate Gromacs-CP2K simulation file:

# gmx\_cp2k grompp -f qmmm\_md\_umbrella.mdp -p topol.top -c conf.gro -n index.ndx -o stilbene.tpr

files stilbene.tpr, stilbene.inp and stilbene.pdb should appear in the directory

7) Run QMMM simulation:

# qsub run<u>.sh</u>

8) While job is running you can check the content of stilbene.inp

#### less stilbene.inp

and of qmmm\_md\_umbrella.mdp

#### less qmmm\_md\_umbrella.mdp

9) At the end of the job inspect **pullx.xvg** file appeared in the directory:

## less pullx.xvg

It contains information about chosen coordinate dynamics over the simulation trajectory. By performing that sampling over the many points along reaction coordinate and gathering all \*.tpr and pull.xvg files you could produce free-energy profile of the reaction with gmx wham tool.

10) Check the free energy profiles generated from 100 steps (100fs) and 10000 steps (10ps) of QMMM MD simulation for each frame from the eq\_gro directory: profile-100fs.xvg and profile-10ps.xvg.

You can open them with Grace or copy data into any other software (i.e. Excel).

Notice, how important is long sampling! For biological systems at least 30 ps of QMMM simulation per frame is recommended.

# Exercise 2: Energy minimization with QM/MM

1) Go to stilbene\_water directory:

## cd ../stilbene\_water

2) Generate QM/MM index file:

gmx\_cp2k make\_ndx -f stilbene-sol.pdb

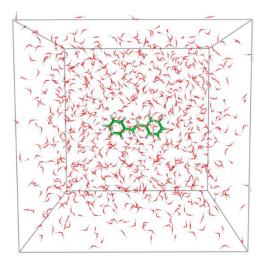


> name 6 QMatoms



file index.ndx should appear in the directory

3) Generate Gromacs-CP2K simulation file:



gmx\_cp2k grompp -f em-qmmm.mdp -p topol.top -c stilbene-sol.pdb -n index.ndx -o stilbene-sol-opt.tpr

files stilbene-sol-opt.tpr, stilbene-sol-opt.inp and stilbene-sol-opt.pdb should appear in the directory

4) Run QMMM minimization

#### sbatch run.sh

5) While job is running you can check the content of stilbene-sol-opt.inp

## less stilbene-sol-opt.inp

and of em-qmmm.mdp

## less em-qmmm.mdp

6) At the end of the job you can print out energy at each step with the following command:

### gmx\_cp2k energy

6

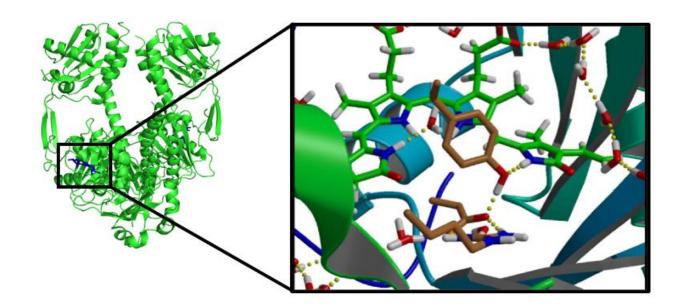
## <!Press ENTER button second time!>

energy.xvg file should appear in the directory.

It contains data with Potential energy (kJ/mol) against optimization step. You can open it in Grace or copy data into any other software (i.e. Excel).

7) You can also check geometry changes by downloading **stilbene-sol.pdb** (coordinates) and **traj.trr** (trajectory) and open them with PyMOL or VMD

Exercise 3: Large protein system setup with QM/MM



1) Go to phytochrome directory:

# cd ../phytochrome

2) Generate Gromacs-CP2K simulation file:

gmx\_cp2k grompp -f md-qmmm.mdp -p topol-sol.top -c lumi-R-conf.gro -n index.ndx -o phytochrome.tpr files phytochrome.tpr, phytochrome.inp and phytochrome.pdb should appear in the directory

3) Run QMMM MD simulation:

# sbatch run.sh

4) While job is running you can check the content of **phytochrome.inp** 

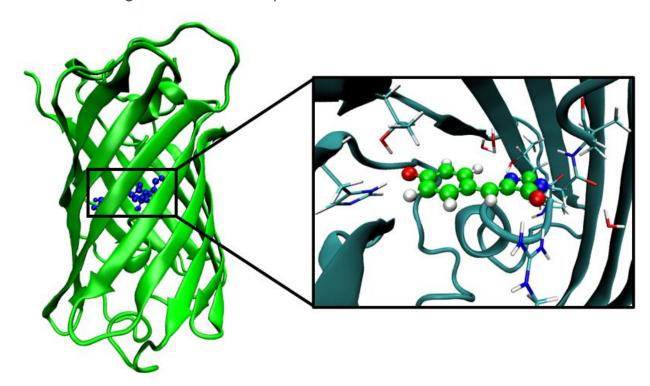
# less phytochrome.inp

and of md-qmmm.mdp

## less md-qmmm.mdp

5) Notice how CP2K input file has changed. Four **&LINK** sections appeared at the end of **&QMMM** part. They are notifying CP2K that broken chemical bond should be treated between MM and QM atoms. From the side of QM part that that result in adding "fake" Hydrogen Link-atom between QM and MM atoms. This is automatically done inside the CP2K.

Exercise 4: Using user-defined QM input file



1) Go to egfp\_spectra directory:

# cd ../egfp\_spectra

2) Generate Gromacs-CP2K simulation file:

## gmx\_cp2k grompp -f md-qmmm.mdp -p topol.top -c conf.gro -n index.ndx -o egfp-spec.tpr

file **egfp-spec.tpr** should appear in the directory, while **egfp-spec.inp** and **egfp-spec.pdb** was already there before tpr file generation.

3) Run QMMM MD simulation:

#### sbatch run.sh

4) While job is running you can check the content of egfp-spec.inp

## less egfp-spec.inp

and of md-qmmm.mdp

# less md-qmmm.mdp

5) Notice that in the md-qmmm.mdp file:

qmmm-qmmethod = INPUT

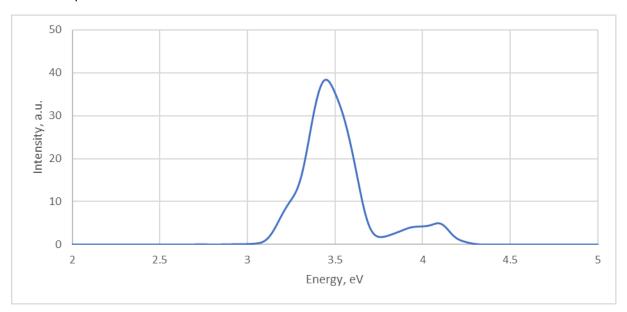
qmmm-qminputfile = egfp-spec.inp

And CP2K input file **egfp-spec.inp** has and additional section **&PROPERTIES.** It requests CP2K to perform additional TDDFT calculation for the lowest 5 excitations at each MD step.

6) After job is finished, we need to gather information about excitation energies over the calculated trajectory:

grep " TDDFPT|" egfp-spec.out | awk '{ print \$3 " " \$7 }' > excitations

The excitations file should appear in the directory, it will consist out of two columns. First column is an excitation energy in eV and second is an oscillator strength in a.u. for each excitation computed by CP2K. Final absorption spectra could be convolved by representing each excitation with gaussian function and summed up over all of them:



# 7) Spectra collected over 3 ps (3000 MD steps will look like that):

