

Analysis and Prediction of Protein Complexes

Proteomes Interactomes and Biological Networks

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<http://biofold.org/>



**Biomolecules
Folding and
Disease**

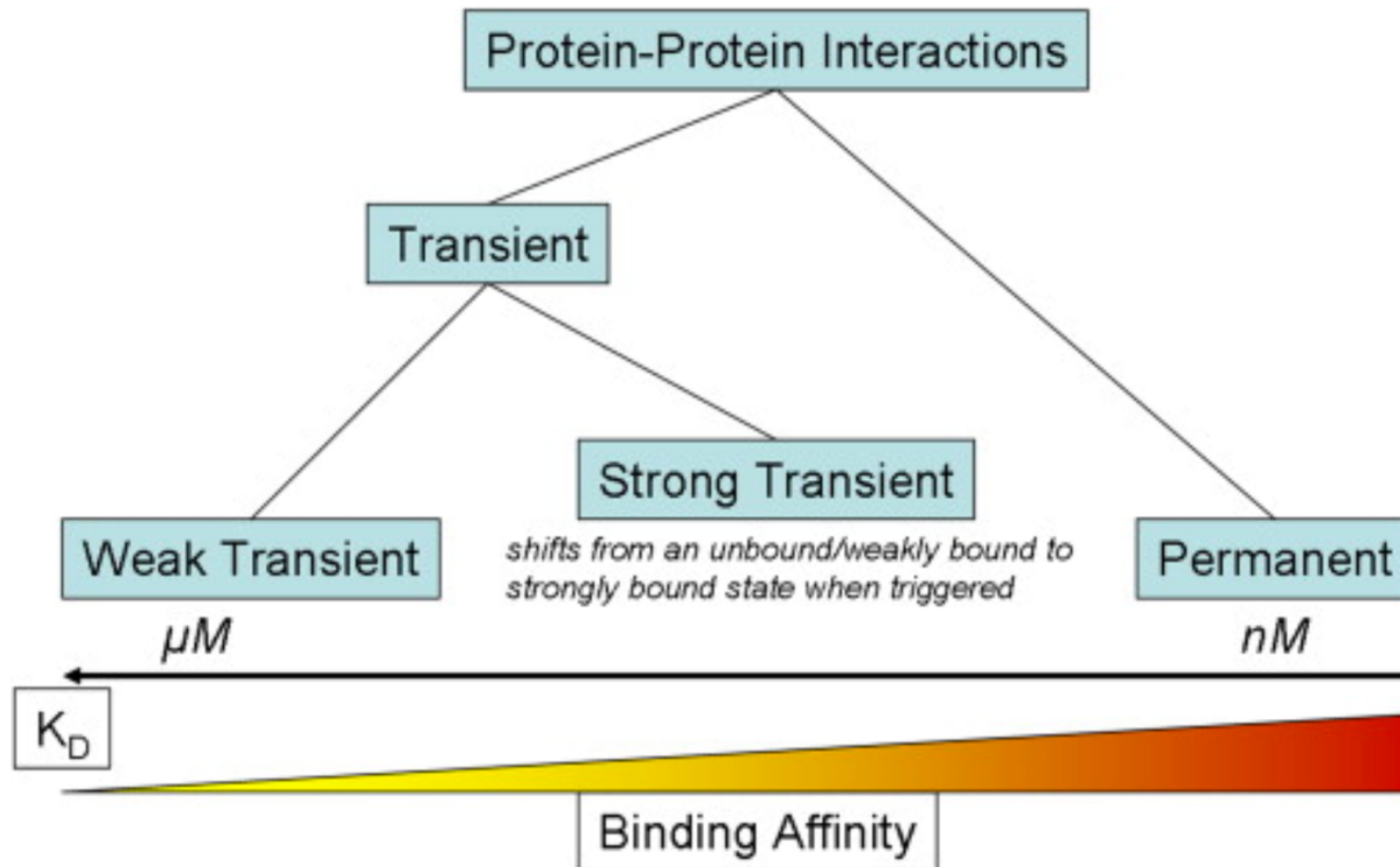
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The Molecular Viewpoint

- The affinity of **PPI varies from millimolar to picomolar**, depending on the type of interaction and signaling needed (Chen et al. Protein Sci. 2013)
- Despite affinity varies over a wide range, **proteins maintain a high degree of specificity** for their partners
- Many **proteins exhibit specificity for multiple partners** (Reichmann et al. Curr. Opin. Struct. Biol. 2007).
- The nature of the **interaction surface** determines how proteins interact
- A detailed knowledge of the **interaction surfaces** of proteins and their energetics is necessary to understand the regulatory **mechanisms of biochemical pathways** (especially to modulate or block these pathways for therapeutic purposes)

Protein-Protein Interactions



Strong transient: This category includes interactions that are triggered/stabilised by an effector molecule or conformational change. An example is given by the Ras proteins, which form tight complexes with their partners when GTP-bound and only weak complexes when GDP-bound.

Surface of Interaction (I)

- The area of PPI interfaces is large (1000 to 4000 Å²)
- **Standard-sized** interfaces are 1200 to 2000 Å²
- **Short-lived and low-stability complexes** -> smaller interfaces (1150–1200 Å²)
- **large surfaces** (2000 to 4600 Å²) ->
 - proteases and particular inhibitors
 - G-proteins and other components of the signal transduction system
- **Protein-small molecule interaction** surfaces have an area of 300 to 1000 Å².

Surface of Interaction (II)

- Surfaces of PPIs are generally **flat** and lack the grooves and pockets that are present at the surfaces of proteins that bind to small molecules.
- PPI **surfaces are generally hydrophobic** in nature.
- Only certain **hydrophobic spots contribute to the free energy** of binding and help to hold the two proteins together.
- Such regions are called **hot spots**.

Hot Spots

- **Hot spots** account for less than **50% of the contact area** of PPI
- A region of protein surface is called a hot spot when **replacement of an amino acid** residue by alanine in that spot **lowers the free energy of binding by at least 2 kcal/mol**
- Analysis of the **amino acid composition of hot spots** shows that some residues are found more frequently in hot spots (Tyr, Trp, and Arg)
- The hot spots are surrounded by energetically less important residues that **separate/prevent bulk water from hot spots**

Analysis of Protein Complex

- identification of **interface residues/hot spots**
- **details** about the interface
solvent accessible surface area, shape, complementarity between surfaces, residue interface propensities, hydrophobicity, segmentation and secondary structure, and conformational changes on complex formation
- assignment of **protein function**
- recognition of **specific residue motifs**

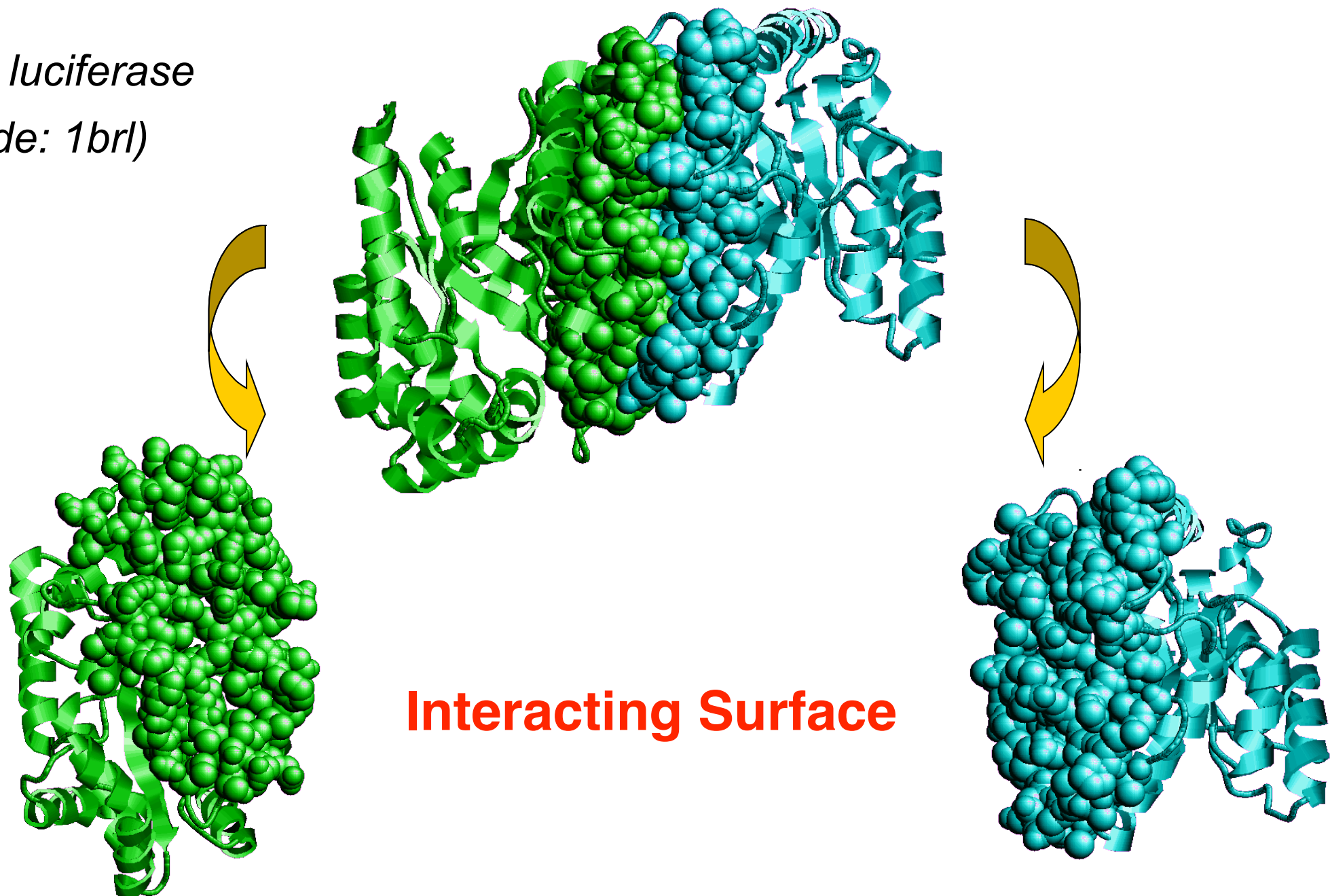
Structure PPI Data

- The most significant contribution to understanding the PPI surface comes from structural biology via **X-ray crystallography** or **NMR** as well as **mutational studies**
- Prediction of interaction/binding sites
- Prediction of protein-protein complexes

Interacting surface

Difference in Accessible Surface Area (ASA) between monomers and complex

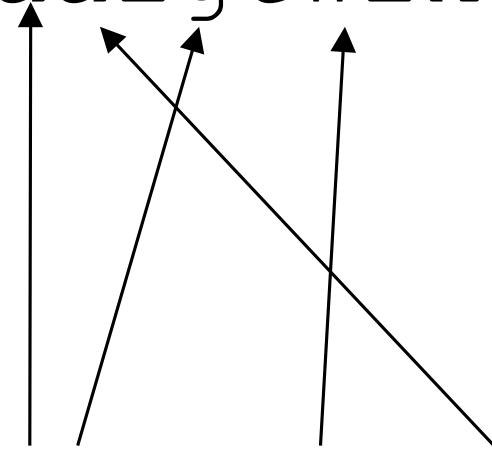
Bacterial luciferase
(PDB code: 1brl)



Prediction features

Protein Sequence

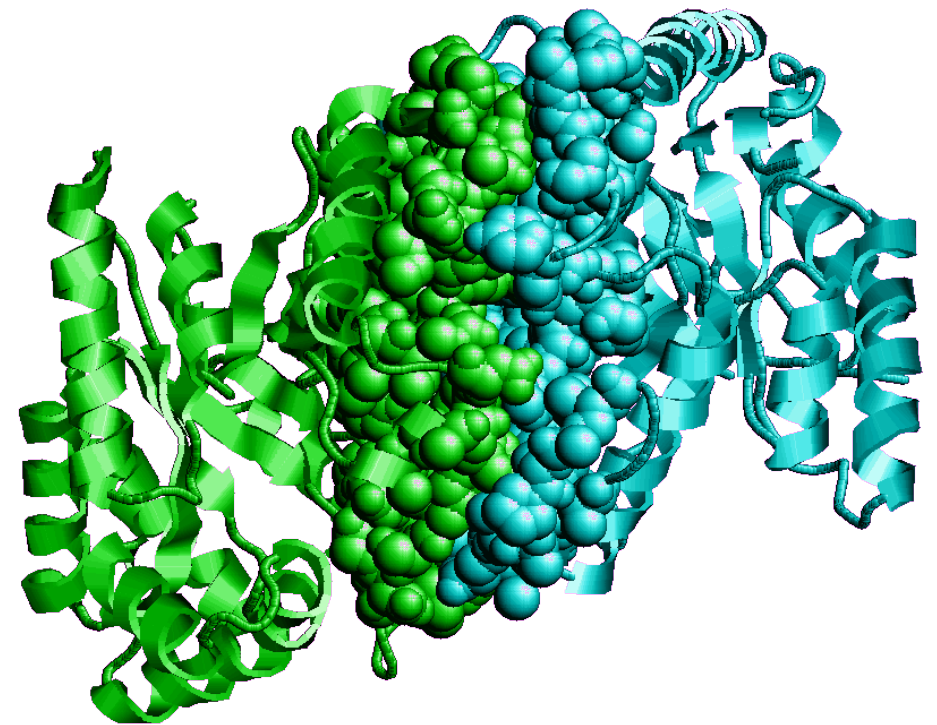
...aalgtwlkts.....
...stwlgtaal kts.....



+ Whole genome computation

- No exact location, No atomic description

Protein Structure



+ Exact location Atomic description

- Availability of the 3D coordinates

Three major problems

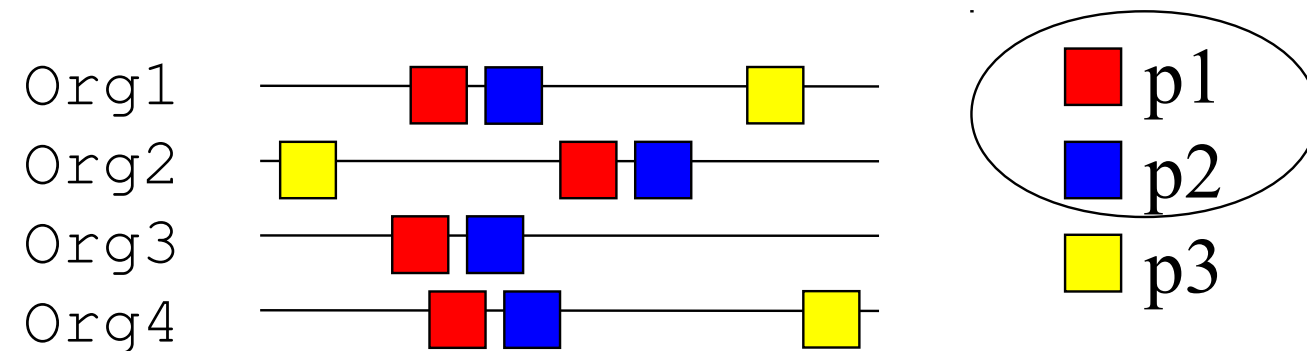
- **Protein-Protein interaction networks:** given a set of proteins, predict the possible partners
- **Docking:** given a pairs of proteins, known to interact, predict the geometry of the complex
- **Protein-interaction sites:** given a single protein, predict possible interacting regions

Sequence-based methods

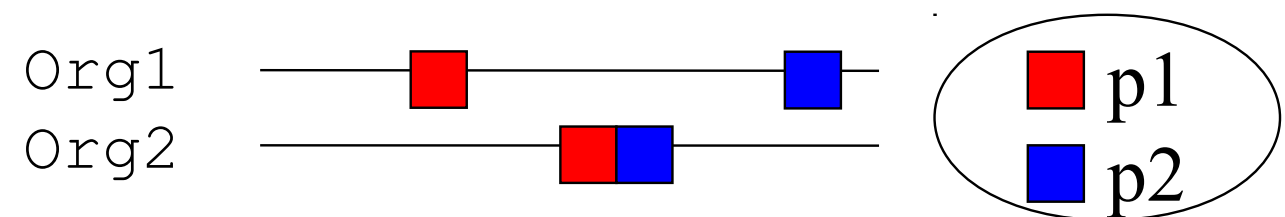
Phylogenetic Profiling: interacting proteins should co-evolve and should have orthologs in closely related species.

	p1	p2	p3	p4
Org1	1	1	1	1
Org2	0	1	0	1
Org3	1	0	1	0
Org4	1	0	1	1

Gene Neighborhood: interacting proteins and co-evolving homologs tend to have close genomic locations.

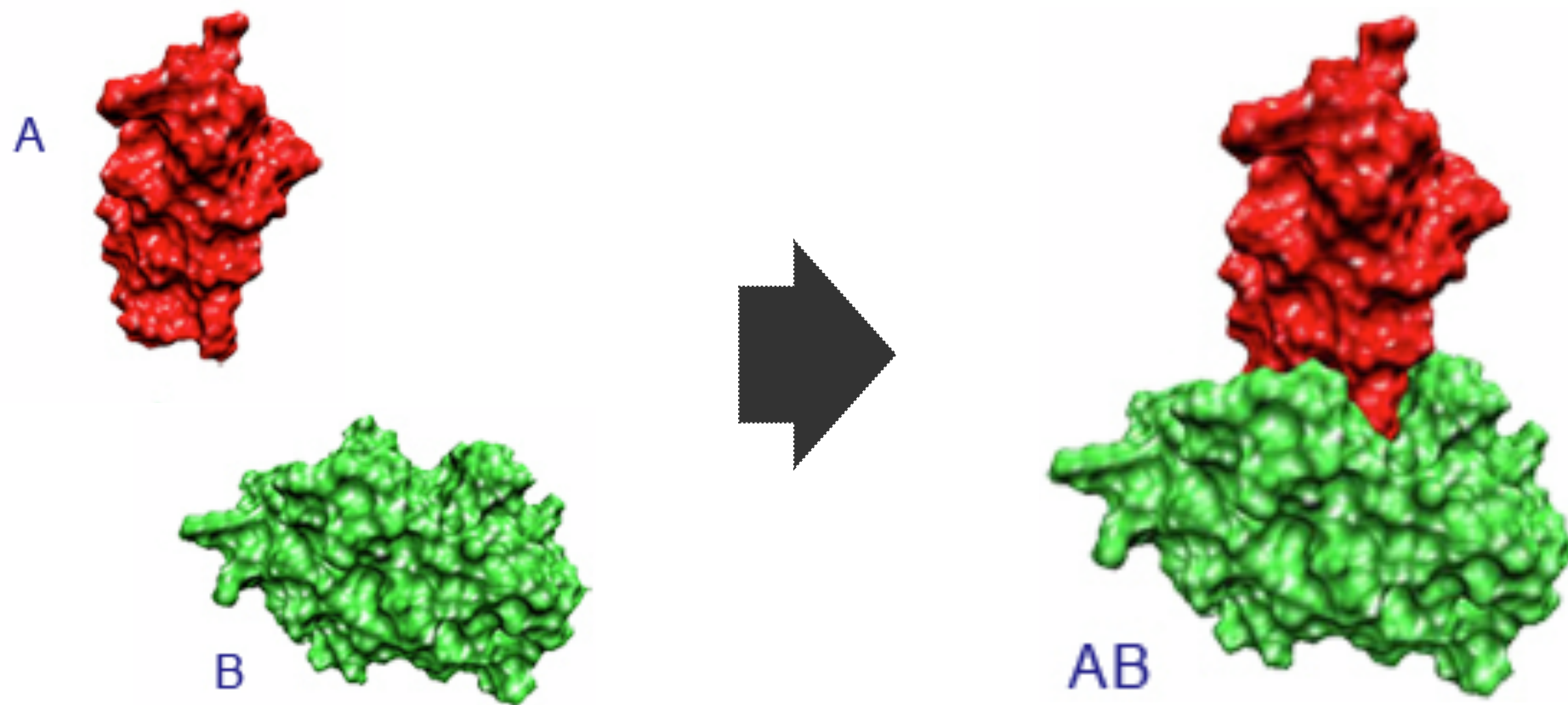


Gene Fusion: two proteins that interact may have homologs in other genomes that are fused into a unique protein



Protein Docking

- Computational schemes that aims to **find the “best” matching between two molecules**, a **receptor** and a **ligand**
- The molecular docking problem can be defined as follows: **given the atomic coordinates of two molecules, predict their “correct” bound association**



Some methods

- **HADDOCK** (software/web server).
<http://haddock.chem.uu.nl>
- **CLUSPRO** (software/web server)
<http://cluspro.bu.edu>
- **ICM-pro** (desktop-modeling environment)
http://www.molsoft.com/protein_protein_docking.html
- **ROSETTADOCK** (software/web server)
<http://graylab.jhu.edu/docking/rosetta/>
- <http://rosettadock.graylab.jhu.edu/submit>
- **GRAMM-X** (web server)
<http://vakser.bioinformatics.ku.edu/resources/gramm/grammx>
- **PATCHDOCK/FIREDOCK** (software/web server)
<http://bioinfo3d.cs.tau.ac.il/PatchDock/>
- **HEX** (software/web server)
<http://hexserver.loria.fr>

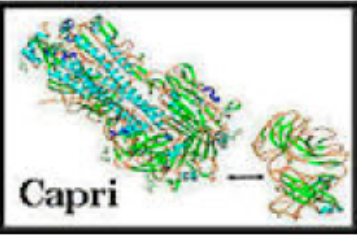
Docking limitations

Although the docking algorithms shows good performances in the prediction of **enzyme-inhibitor and antigen-antibody complexes**. The molecular docking problem **is far from being solved**

- It is difficult to find very specific properties of protein-protein interfaces
- Results are generally **poor with weakly interacting proteins**
- Proteins are flexible and may undergo even **large conformational changes upon binding**

CAPRI Experiments

- CAPRI is a community-wide experiment in modelling the molecular structure of protein complexes
- CAPRI is a **blind prediction experiment** aimed at testing the performance of protein docking methods
- Rounds take place about every six months
- Each round contains between one and six target protein–protein complexes whose structures have been recently determined experimentally
- Targets are unpublished crystal or NMR structures of complexes, whose coordinates are held privately by the assessors, with the co-operation of the structural biologists who determined them
- The atomic coordinates of the two proteins are given to groups for prediction



Capri

PDB idcodes for past targets

Home > Databases > PDBe > Services > Capri-Home

CAPRI: Critical Assessment of PRediction of Interactions

CAPRI communitywide experiment on the comparative evaluation of protein-protein docking for structure prediction

Hosted By EMBL/EBI-PDBe Group

Exercise

Download the DSSP file of the **Bacterial luciferase** (*Vibrio harveyi*) from the PDB (code: 1BRL)

- Generate the **DSSP** file for the protein complex and the isolated chains A and B
- Calculate the total **solvent accessible area** of the complex and isolated chains and calculate the surface of interaction for both chains.
- Given the size of the binding surface **what kind of protein interaction** it is expected?
- Find the **residue at the interface** and calculate the **variation of relative solvent accessible area**. Which residue are buried in the interacting surface?

Chain = col 12, AA = col 14, SS = col 17, Acc: cols 36-38, Phi: cols 104-109, Psi: cols 110-115