HOWARD User Guide

1 HOWARD User Guide

HOWARD

Highly Open Workflow for Annotation & Ranking toward genomic variant Discovery

HOWARD annotates and prioritizes genetic variations, calculates and normalizes annotations, translates files in multiple formats (e.g. vcf, tsv, parquet) and generates variants statistics.

HOWARD annotation is mainly based on a build-in Parquet annotation method, and external tools such as BCFTOOLS, ANNOVAR, snpEff, Exomiser and Splice (see docs, automatically downloaded if needed). Parquet annotation uses annotation database in VCF or BED format, in multiple file format: Parquet/duckdb, VCF, BED, TSV, CSV, TBL, JSON.

HOWARD calculation processes variants information to calculate new information, such as: harmonizes allele frequency (VAF), extracts Nomen (transcript, cNomen, pNomen...) from HGVS fields with an optional list of personalized transcripts, generates VaRank format barcode.

HOWARD prioritization algorithm uses profiles to flag variants (as passed or filtered), calculate a prioritization score, and automatically generate a comment for each variants (example: 'polymorphism identified in dbSNP. associated to Lung Cancer. Found in ClinVar database'). Prioritization profiles are defined in a configuration file. A profile is defined as a list of annotation/value, using wildcards and comparison options (contains, lower than, greater than, equal...). Annotations fields may be quality values (usually from callers, such as 'GQ', 'DP') or other annotations fields provided by annotations tools, such as HOWARD itself (example: COSMIC, Clinvar, 1000genomes, PolyPhen, SIFT). Multiple profiles can be used simultaneously, which is useful to define multiple validation/prioritization levels (example: 'standard', 'stringent', 'rare variants', 'low allele frequency').

HOWARD translates VCF format into multiple formats (e.g. VCF, TSV, Parquet), by sorting variants using specific fields (example: 'prioritization score', 'allele frequency', 'gene symbol'), including/excluding annotations/fields, including/excluding variants, adding fixed columns.

HOWARD generates statistics files with a specific algorithm, snpEff and BCFTOOLS.

HOWARD is multithreaded through the number of variants and by database (data-scaling).

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2 Installation

2.1 Python

2.1.1 Quick install

Install HOWARD using Python Pip tool (we strongly suggest to use a virtual environment, such as conda):

```
conda create --name=howard python=3.10 conda activate howard python -m pip install -e .

Run HOWARD for help options:

howard --help
```

2.1.2 Requirements

System packages wget, llvmand libomp are needed.

Even if it is not necessary, we strongly suggest to install aria2 package to speed up databases download.

2.1.3 GUI install

Install HOWARD Graphical User Interface using Python Pip tool with supplementary packages:

```
python -m pip install -r requirements-gui.txt
```

Run HOWARD Graphical User Interface as a tool:

howard gui

HOWARD Graphical User Interface

2.1.4 Configuration

HOWARD Configuration JSON file defined default configuration regarding resources (e.g. threads, memory), settings (e.g. verbosity, temporary files), default folders (e.g. for databases) and paths to external tools.

See HOWARD Configuration JSON for more information.

Configuration file example:

```
{
  "threads": 8,
  "memory": null,
  "verbosity": "warning",
  "folders": {
    "databases": {
      "genomes": "~/howard/databases/genomes/current",
      "annotations": [
        "~/howard/databases/annotations/current",
        "~/howard/databases/dbnsfp/current",
        "~/howard/databases/dbsnp/current"
      ],
      "parquet": [
        "~/howard/databases/annotations/current"
        ],
      "bcftools": [
        "~/howard/databases/annotations/current"
       ],
      "annovar": [
        "~/howard/databases/annovar/current"
      "snpeff": "~/howard/databases/snpeff/current",
      "exomiser": "~/howard/databases/exomiser/current"
    }
  },
  "tools": {
    "bcftools": "bcftools",
    "bgzip": "bgzip",
    "java": "java",
    "snpeff": "~/howard/tools/snpeff/current/bin/snpEff.jar",
    "snpsift": "~/howard/tools/snpeff/current/bin/SnpSift.jar",
    "annovar": "~/howard/tools/annovar/current/bin/table_annovar.pl",
    "exomiser": "~/howard/tools/exomiser/current/bin/exomiser-cli-14.0.0.jar",
    "splice": {
      "docker": {
        "image": "bioinfochrustrasbourg/splice:0.2.1",
        "entrypoint": "/bin/bash",
        "options": null,
        "command": null
      }
    },
    "docker": "docker"
  }
}
```

2.1.5 External tools

In order to use external tools, mainly for annotation (e.g. Annovar, snpEff, Exomiser, Splice), they need to be installed (see doc of each tools). For Splice tool, a Docker image is automatically downloaded using configuration file.

2.2 Docker

2.2.1 Quick Start

In order to build images, launch default setup and create a persitent CLI (Command Line Inferface container), docker-compose command build images and launch services as containers.

docker-compose up -d

The persitent CLI contains external tools, such as:

External tool	Description	
BCFTools	Utilities for variant calling and manipulating VCFs and BCFs	
snpEff	Genomic variant annotations, and functional effect prediction toolbox	
Annovar	Efficient software tool to utilize update-to-date information to functionally annotate genetic variants	
Exomiser	Program that finds potential disease-causing variants from whole-exome or whole-genome sequencing data	
Splice	Image to run SPiP and SpliceAI tools in an nextflow pipeline.	

2.2.2 Setup container

Docker service HOWARD-setup creates HOWARD image and download useful databases to start with HOWARD tools.

docker-compose up -d HOWARD-setup

List of databases downloaded in HOWARD setup for hg19 assembly (see Databases section for more information): - Genome - Annovar - (refGene - COSMIC) - snpEff - refSeq - dbNSFP - AlphaMissense - dnSNP

To avoid databases download (see Databases section to download manually), just start Command Line Interface

2.2.3 Command Line Interface

A Command Line Interface container (HOWARD-CLI) is started with host data and databases folders mounted (by default both in \${HOME}/HOWARD folder). To manually start CLI container:

```
docker-compose up -d HOWARD-CLI
```

To use HOWARD tools within HOWARD-CLI container:

```
docker exec -ti HOWARD-CLI bash
howard --help
```

To run a command into HOWARD-CLI container:

```
docker exec HOWARD-CLI <howard command>
```

Docker HOWARD-CLI container (Command Line Interface) can be used to execute commands.

Example: Query of an existing VCF

```
docker exec HOWARD-CLI \
  howard query \
  --input='/tool/tests/data/example.vcf.gz' \
  --query='SELECT * FROM variants'
```

Example: VCF annotation using HOWARD-CLI (snpEff and ANNOVAR databases will be automatically downloaded), and query list of genes with HGVS

See HOWARD Help for more options.

Let's play within Docker HOWARD-CLI service!

2.2.4 Tests

In order to test HOWARD within Docker, use this command:

```
docker exec -ti HOWARD-CLI bash
cd /tool
# Basic test
coverage run -m pytest .
# Debug test
coverage run -m pytest . -x -v --log-cli-level=DEBUG --capture=tee-sys
```

3 Databases

3.1 Databases tool

Multiple databases can be automatically downloaded with databases tool, such as:

database	description	
Genome	Genome Reference Consortium Human	
Annovar	ANNOVAR is an efficient software tool to utilize	
	update-to-date information to functionally annotate genetic	
	variants detected from diverse genomes	
snpEff	Genetic variant annotation, and functional effect prediction	
	toolbox	
refSeq	A comprehensive, integrated, non-redundant,	
	well-annotated set of reference sequences including genomic,	
	transcript, and protein	
dbSNP	dbSNP contains human single nucleotide variations,	
	microsatellites, and small-scale insertions and deletions	
	along with publication, population frequency, molecular	
	consequence, and genomic and RefSeq mapping information	
	for both common variations and clinical mutations	
dbNSFP	dbNSFP is a database developed for functional prediction	
	and annotation of all potential non-synonymous	
	single-nucleotide variants (nsSNVs) in the human genome	
AlphaMissense	AlphaMissense model implementation	
Exomiser	The Exomiser is a Java program that finds potential	
	disease-causing variants from whole-exome or whole-genome	
	sequencing data	

Example: Download Multiple databases in the same time for assembly 'hg19' (can take a while)

```
howard databases \
   --assembly=hg19 \
   --download-genomes='~/howard/databases/genomes/current' \
```

```
--download-genomes-provider='UCSC'\
--download-genomes-contig-regex='chr[0-9XYM]+$'\
--download-annovar='~/howard/databases/annovar/current'\
--download-annovar-files='refGene,cosmic70,nci60'\
--download-snpeff='~/howard/databases/snpeff/current'\
--download-refseq='~/howard/databases/refseq/current'\
--download-refseq-format-file='ncbiRefSeq.txt'\
--download-dbnsfp='~/howard/databases/dbnsfp/current'\
--download-dbnsfp-release='4.4a'\
--download-dbnsfp-subdatabases\
--download-alphamissense='~/howard/databases/alphamissense/current'\
--download-exomiser='~/howard/databases/exomiser/current'\
--download-dbsnp='~/howard/databases/dbsnp/current'\
--download-dbsnp='~/howard/databases/dbsnp/current'\
--download-dbsnp-vcf\
--threads=8
```

See HOWARD Help Databases tool for more information.

3.2 Home-made Databases

Databases can be generated using an home-made existing annotation file and HOWARD convert tool. The home-made annotation file need to contain specific fields (depending on the annotation type):

- variant annotation: '#CHROM', 'POS', 'ALT', 'REF'
- region annotation: '#CHROM', 'START', 'STOP'

An home-made existing annotation file can be converted into multiple formats (e.g. Parquet, VCF, TSV), but it's strongly suggested to use Parquet format.

After convertion, the database file is associated with a 'header' file ('.hdr'), in VCF header format, to describe annotations within the database. Use the 'header' file to describe annotation fields/columns present in the existing file. An Home-made annotation file in VCF format which is converted in another format will keep all annotation information from the initial VCF header.

Note that a VCF can be directly used as a database (annotation field information within the header of the VCF file). Also, an home-made existing annotation file can be used as a database, but will not be totaly compliant due to the lack of annotation information ('header' will be generated by default).

See HOWARD Help Convert tool for more information.

3.3 Databases from Annovar and Extann

See HOWARD Help Databases tool tool for more information about the tool, and HOWARD Parameters Databases help for generate databases annotation file from Annovar databases (from Annovar, under development) and from Gene annotation file (from Extann).

4 Tools

HOWARD annotates and prioritizes genetic variations, calculates and normalizes annotations, convert on multiple formats, query variations and generates statistics. These tools require options or a Parameters JSON file.

4.1 Parameters

HOWARD Parameters JSON file defined parameters to process annotations, prioritization, calculations, convertions and queries. Use this parameters file to configure tools, instead of options or as a main configuration (options will replace parameters in JSON file).

See HOWARD Parameters JSON for more information.

Example: Use parameters JSON file with query tool

```
howard query \
   --input='tests/data/example.vcf.gz' \
   --param='config/param.json'
  #CHROM
             POS REF ALT
                                             INFO
   chr1
0
           28736 A C
                                CLNSIG=pathogenic
           35144 A C CLNSIG=non-pathogenic
1
   chr1
2
   chr1
            69101 A G
                                            DP=50
3 chr1 768251 A G
                                             None
4 chr1 768252 A G
                                             None
5
   chr1
         768253 A G
                                             None
   chr7 55249063
                    G A
                                           DP=125
Example: Use parameters JSON file with query tool, and add an option to change the query (list of chromosomes)
howard query \
   --input='tests/data/example.vcf.gz' \
   --param='config/param.json' \
   --query="SELECT distinct(\"#CHROM\") as 'chromosomes' FROM variants"
  chromosomes
0
        chr7
1
         chr1
Example: Parameters JSON file with multiple options for tools
  "annotation": {
    "parquet": {
      "annotations": {
        "tests/databases/annotations/current/hg19/avsnp150.parquet": {
          "INFO": null
        },
        "tests/databases/annotations/current/hg19/dbnsfp42a.parquet": {
          "INFO": null
        },
        "tests/databases/annotations/current/hg19/gnomad211_genome.parquet": {
          "INFO": null
        }
      }
    },
    "bcftools": {
      "annotations": {
        "tests/databases/annotations/current/hg19/cosmic70.vcf.gz": {
          "INFO": null
        }
      }
    },
    "snpeff": {
      "options": "-lof -hgvs -oicr -noShiftHgvs -spliceSiteSize 3 "
    },
    "snpsift": {
      "annotations": {
        "tests/databases/annotations/current/hg19/cosmic70.vcf.gz": {
          "INFO": null
        }
      }
    },
    "annovar": {
      "annotations": {
       "refGene": {
          "INFO": null
```

```
}
    },
    "options": {
      "genebase": "-hgvs -splicing_threshold 3 ",
      "intronhgvs": 10
    }
  }
},
"calculation": {
  "calculations": {
    "vartype": null,
    "snpeff_hgvs": null,
    "VAF": "",
    "NOMEN": {
      "options": {
        "hgvs_field": "snpeff_hgvs",
        "transcripts": "tests/data/transcripts.tsv"
      }
    }
  },
  "config/calculations_config.json": "config/calculations_config.json"
},
"prioritization": {
  "profiles": ["default", "GERMLINE"],
  "prioritization_config": "config/prioritization_profiles.json",
  "pzfields": ["PZScore", "PZFlag", "PZComment"],
  "prioritization_score_mode": "VaRank"
},
"hgvs": {
  "full_format": true,
  "use exon": true
},
"stats": {
  "stats_md": null,
  "stats_json": null
},
"query": {
  "query": "SELECT \"#CHROM\", POS, REF, ALT, INFO FROM variants",
  "query_limit": 10,
  "query_print_mode": "default"
},
"explode_infos": {
  "explode_infos": false,
  "explode_infos_prefix": "",
  "explode_infos_fields": null
},
"export": {
  "header_in_output": false,
  "parquet partitions": null,
  "order_by": null
},
"threads": 8
```

Moreover, a transcripts file can be defined, especially to select NOMEN from a list of HGVS annotation (see Calculation and HGVS and NOMEN from snpEff). This file is a tab-delimited with 'transcript' as first column and 'gene' a second column. For a gene, transcripts of reference are ordered (first is priority, e.g. 'NM_001346897' has prior over 'NM_005228').

Example: Transcripts file in tab-delimited format with column 'transcript' and column 'gene'

```
NR_024540 WASH7P
NR_036266 MIR1302-9
NM_001346897 EGFR
NM_005228 EGFR
```

|:----:|

7

Total

4.2 Stats

Generates statistics on genetic variations, such as number of variants, number of samples, statistics by chromosome, genotypes by samples, annotations. These statistics can be applied to VCF files from all database annotation file formats. Statistics can be wrote into files in Markdown and JSON format (resp. --stats_md and --stats_json parameter).

See HOWARD Help Stats tool for more information.

```
Example: Show example VCF statistics and brief overview
howard stats \
   --input='tests/data/example.vcf.gz'
Example: Show example VCF statistics and generate a file in JSON and Markdown formats (extract)
howard stats \
   --input='tests/data/example.vcf.gz' \
   --stats_json='/tmp/stats.json' \
   --stats_md='/tmp/stats.md'
cat '/tmp/stats.json' '/tmp/stats.md'
{
    "Infos": {
        "Input file": "tests/data/example.vcf.gz",
        "Number of variants": 7,
        "Number of samples": 4,
        "Number of INFO fields": 5,
        "Number of FORMAT fields": 7
    },
    "Variants": {
        "Number of variants by chromosome": {
            "1": {
                "CHROM": "chr1",
                "count": 6,
                "percent": 0.8571428571428571
            },
            "0": {
                "CHROM": "chr7",
                "count": 1,
                "percent": 0.14285714285714285
        }
    }
}
## Variants
### Number of variants by chromosome
| CHROM | count | percent |
|:----:|----:|----:|
                 6 | 0.857143
chr1
         chr7
         1 0.142857
### Counts
Type
       count
```

Example of statistics in Markdown output

• Input file: tests/data/example.vcf.gz

Number of variants: 7
Number of samples: 4
Number of INFO fields: 5
Number of FORMAT fields: 7

CHROM	count	percent
chr1	6	0.857143
chr7	1	0.142857

Type	count
Total	7
SNV	7
MNV	0
InDel	0

4.3 Convert

Convert genetic variations file to another format. Multiple format are available, such as usual and official VCF format, but also other formats such as TSV, CSV, TBL, JSON and Parquet/duckDB. These formats need a header '.hdr' file to take advantage of the power of howard (especially through INFO/tag definition), and using howard convert tool automatically generate header file fo futher use (otherwise, an default '.hdr' file is generated).

Multiple options are available, such as explode VCF INFO/tags (parameter --explode_infos, see HOWARD Help query - Explode infos), order by columns, include header within file (only TSV and CSV format), or use partitioning into multiple files within a folder. See HOWARD Help Convert tool for more information.

4.3.1 CSV, TSV and JSON convert

To convert a file (multiple formats) into another flat file, such as CSV (tab-delimiter) and TSV (comma-delimiter), or JSON format, simply name output file with desired extension. Use .gz extension to compress file.

Example: Convert VCF into TSV and show output file

```
howard convert \
    --input='tests/data/example.vcf.gz' \
    --output='/tmp/example.json.gz'
```

4.3.2 Parquet and duckDB convert

Files can be format into Parquet and duckDB format. For duckDB format, a duckDB database will be created with a variants table.

```
Example: Convert VCF into parquet
howard convert \
    --input='tests/data/example.vcf.gz' \
    --output='/tmp/example.parquet'

Example: Convert VCF into duckDB
howard convert \
    --input='tests/data/example.vcf.gz' \
    --output='/tmp/example.duckdb'
```

4.3.3 BED convert

To convert into BED format, input file needs mandatory columns chrommosome "#CHROM", and positions columns "START"/"END" or "POS" (corresponding to regions with a uniq nucleotide). Header will be automatically included to decribe all columns.

Example: Convert BED in TSV format into BED format

```
howard convert \
    --input='tests/data/example.bed.tsv' \
    --output='/tmp/example.bed'

Example: Convert BED in Parquet format into BED format
howard convert \
    --input='tests/data/example.bed.parquet' \
    --output='/tmp/example.bed'
```

HOWARD input file format does not allow BED format. To read a BED file and export into a CSV or Parquet file format, see Query BED format section.

4.3.4 Partitioning

Partitioning (or Hive partitioning) is a partitioning strategy that is used to split a table into multiple files based on partition keys. The files are organized into folders. Within each folder, the partition key has a value that is determined by the name of the folder (see duckDB hive partitioning).

Simply list columns as keys to process partitioning. Use 'None' (string) for NO partition but split parquet files into a folder. The partitioning is available for all format (e.g. Parquet, TSV, JSON, except duckDB format), by naming output file with desired extension. This option is faster parallel writing, but memory consuming, and also is faster reading.

Example: Convert VCF into partitioned Parquet and show tree structure

```
howard convert \
    --input='tests/data/example.vcf.gz' \
    --output='/tmp/example.partitioned.parquet' \
    --parquet_partitions="#CHROM"

tree '/tmp/example.partitioned.parquet'
/tmp/example.partitioned.parquet
|-- \#CHROM=chr1
|    |-- fe61bf182de640f8840270a527aa1582-0.parquet
|-- #CHROM=chr7
|    |-- fe61bf182de640f8840270a527aa1582-0.parquet
```

Example: Convert VCF into partitioned TSV (compressed) and show tree structure (files are named with .csv extension, but are tab-delimited and compressed)

4.3.5 Explode INFO tags

Use --explode_infos parameter to extract all INFO tags (i.e. annotations) into columns (see HOWARD Help query - Explode infos).

Example: Convert VCF into TSV, export INFO/tags into columns, and show output file

```
howard convert \
   --input='tests/data/example.vcf.gz' \
   --explode_infos \
   --output='/tmp/example.tsv'
cut '/tmp/example.tsv' -f1-4,7,15
#CHROM POS
                   REF
                       ALT FILTER
                                     CLNSIG
        28736
                        C
                             PASS
                                      pathogenic
chr1
        35144
                        С
chr1
                   Α
                             PASS
                                      non-pathogenic
        69101
                   Α
                        G
                             PASS
chr1
chr1
        768251
                   Α
                        G
                             PASS
```

4.4 Query

Query tool provides a simple way to query genetic variations in SQL format. Data are loaded into 'variants' table from various formats (e.g. VCF, TSV, Parquet...). Using --explode_infos allows querying on INFO/tag annotations. SQL query can also use external file within the request, such as a Parquet file(s), or TSV files.

See HOWARD Help Query tool for more information.

4.4.1 Variants file

4.4.1.1 Loading data Query tool is able to read variants (i.e. VCF) or regions files (i.e. BED) files, in various format (e.g. VCF, BED, Parquet, TSV, JSON), using --input parameter. This allows to load data to perfom actions, such as explode VCF INFO/tags (parameter --explode_infos, see HOWARD Help query - Explode infos) in columns to be easier querying. Each columns format (e.g. string, integer) are automatically detected to be used in a SQL query.

Example: Select variants in VCF with REF and POS fields filter

```
howard query \
    --input='tests/data/example.vcf.gz' \
    --query="SELECT * FROM variants WHERE REF = 'A' AND POS < 100000"
Example: Select variants in VCF with INFO Tags criteria filters
howard query \
    --input='tests/data/example.vcf.gz' \
    --explode_infos \</pre>
```

4.4.1.2 External file Variants files can be used directly within the query, espacially if they already contain variants information (e.g. "#CHROM", "POS", "REF", "ALT") and annotations as columns.

Example: Query a Parquet file with specific columns (e.g. from VCF convertion to Parquet)

```
howard query \
    --query="SELECT * \
         FROM 'tests/databases/annotations/current/hg19/dbnsfp42a.parquet' \
         WHERE \"INFO/Interpro_domain\" NOT NULL \
         ORDER BY \"INFO/SiPhy_29way_logOdds_rankscore\" DESC"
```

Example: Query multiple Parquet files, merge INFO columns, and extract as TSV (in VCF format)

```
howard query \
    --query="
    SELECT \
    \"#CHROM\" AS \"#CHROM\", \
    POS AS POS, \
    '' AS ID, \
    REF AS REF, \
    ALT AS ALT, \
    '' AS QUAL, \
    '' AS FILTER, \
    STRING_AGG(INFO, ';') AS INFO \
    FROM 'tests/databases/annotations/current/hg19/*.parquet' \
    GROUP BY \"#CHROM\", POS, REF, ALT" \
    --output='/tmp/full_annotation.tsv' \
    --include_header
```

4.4.2 Other files

Whatever the external file format, if it is compatible with duckDB, query tool is able to query data (see duckDB Select Statement).

4.4.2.1 Parquet format Simply use Parquet file path within the query (as descibe above).

Example: Query a Parquet file with specific columns (e.g. from VCF convertion to Parquet)

```
howard query \
    --query="SELECT * \
        FROM 'tests/databases/annotations/current/hg19/dbnsfp42a.parquet' \
        WHERE \"INFO/Interpro_domain\" NOT NULL \
        ORDER BY \"INFO/SiPhy_29way_logOdds_rankscore\" DESC"
```

4.4.2.2 CSV format Use duckDB function **read_csv_auto** to read a TSV file format as a table. See duckDB CSV import for more information.

Example: Query a TSV file

4.4.2.3 BED format In order to read a BED file, create a query (using appropriate columns), and export file into a desired format.

Example: Read a BED file and export in Parquet format

```
howard query \
   --query="SELECT * FROM read_csv_auto('tests/data/example.bed',
                    columns={'#CHROM': 'VARCHAR', 'START': 'INTEGER', 'END': 'INTEGER'})" \
   --output='/tmp/example.bed.parquet'
Example: Convert a BED file in a Parquet format into a BED file format
howard convert \
   --input='tests/data/example.bed.parquet' \
   --output='/tmp/example.bed'
cat '/tmp/example.bed'
          START
#CHR.OM
                  END
chr1 28735
            69101
chr1 768250 768253
chr7 55249060
                  55249069
```

A BED file can be filtered using positions or other columns such as gene names, or transcripts.

Example: Filter a BED using positions

```
howard query \
   --input='tests/data/example.bed.parquet' \
   --query="SELECT * \
            FROM variants \
            WHERE \
               \" \# CHROM \" = 'chr1' and \
                  (START>28000 and \"END\"<70000) or \
                  (START>760000 and \"END\"<770000)
               ) "
 #CHROM
          START
                     END
           28735
                   69101
0
    chr1
    chr1 768250 768253
```

4.4.3 Extract variants

In order to extract variants from a VCF file, without annotations and samples, use a query to construct the VCF.

Example: Extract variants onnly

4.5 Annotation

Annotation is mainly based on a build-in Parquet annotation method, using annotation database file (in multiple format such as Parquet, duckdb, VCF, BED, TSV, JSON).

See HOWARD Help Annotation tool for more information.

These annotation databases can be automatically downloaded using HOWARD Databases tool and manually generated using existing annotation files and HOWARD Convert tool. Annotation databases need a header file (.hdr) to describe each annotation in the database. However, a default header will be generated if no header file is associated to the annotation database file.

Moreover, some external annotation tools are integrated into HOWARD to extend annotation with their own options and databases.

HOWARD annotation tool can use annotation databases files in 2 differents ways: Quick annotation and Annotation Parameters JSON file.

4.5.1 Quick annotation

4.5.1.1 Parquet annotation method Quick annotation allows to annotates by simply listing annotation databases (in multiple format).

4.5.1.1.1 Parquet annotation with path These annotation databases are defined with their full path (e.g. /full/path/to/my.database.parquet) or relative path (e.g. databases/my.database.parquet). A list (separator : or +) of annotation databases files can be used.

Example: VCF annotation with full path Parquet databases

```
howard annotation \
   --input='tests/data/example.vcf.gz' \
   --annotations=$(pwd)'/tests/databases/annotations/current/hg19/dbnsfp42a.parquet'
   --output='/tmp/example.howard.vcf.gz'
Example: VCF annotation with relative path VCF databases
howard annotation \
   --input='tests/data/example.vcf.gz' \
   --annotations='tests/databases/annotations/current/hg19/cosmic70.vcf.gz'
   --output='/tmp/example.howard.vcf.gz'
Example: VCF annotation with relative path BED databases
howard annotation \
   --input='tests/data/example.vcf.gz' \
   --annotations='tests/databases/annotations/current/hg19/refGene.bed.gz' \
   --output='/tmp/example.howard.vcf.gz'
Example: VCF annotation with 3 annotation databases files
howard annotation \
   --input='tests/data/example.vcf.gz' \
   --annotations=$(pwd)'/tests/databases/annotations/current/hg19/dbnsfp42a.parquet,
       tests/databases/annotations/current/hg19/cosmic70.vcf.gz,
       tests/databases/annotations/current/hg19/refGene.bed.gz' \
   --output='/tmp/example.howard.vcf.gz'
```

4.5.1.1.2 Parquet annotation with annotation folder If annotation folder is configured in HOWARD Configuration JSON, just mention the annotation database basename file. Annotation database file will be found (depending of the assembly).

Example: VCF annotation with Parquet and VCF databases, with annotation database defined in JSON configuration (as a string)

```
howard annotation \
    --input='tests/data/example.vcf.gz' \
```

```
--annotations='dbnsfp42a.parquet,cosmic70.vcf.gz' \
--config='{"folders": {"databases": {"annotations": ["tests/databases/annotations/current"]}}}' \
--output='/tmp/example.howard.vcf.gz'
```

4.5.1.1.3 Full annotation In order to annotate with all available annotation databases, the keyword ALL will autodetect files in the databases annotation folder. The option format (defaut parquet) can filter annotation databases by listing (separator +) desired formats (such as parquet, vcf). The option release (default current) is able to scan annotation databases in one or more specific releases in a list (separator +). See HOWARD Configuration JSON - Folders - Databases for more information about databases structure.

Example: VCF annotation with all available database annotation files in Parquet format (within the database annotation folder in configuration):

```
howard annotation \
    --input='tests/data/example.vcf.gz' \
    --assembly='hg19' \
    --annotations='ALL:format=parquet+vcf:release=current' \
    --config='{"folders": {"databases": {"annotations": ["tests/databases/annotations/current"]}}}' \
    --output='/tmp/example.howard.tsv'
```

- **4.5.1.2** External tools annotation External annotation tools are also available, such as BCFTOOLS, Annovar, snpEff, Exomiser and Splice. Annovar, snpEff and Exomiser databases are automatically downloaded (see HOWARD Help Databases tool). Quick annotation allows to annotates by simply defining external tools keywords.
- **4.5.1.2.1 BCFTools annotation** For BCFTools, use HOWARD keyword bcftools and list (separator : or +) annotation databases with format such as VCF or BED (compressed). More options are available using HOWARD Parameters JSON file.

Example: VCF annotation with Cosmic VCF databases and refGene BED database

```
howard annotation \
--input='tests/data/example.vcf.gz' \
--annotations='bcftools:tests/databases/annotations/current/hg19/cosmic70.vcf.gz,
tests/databases/annotations/current/hg19/refGene.bed.gz' \
--output='/tmp/example.howard.vcf.gz'
```

4.5.1.2.2 Annovar annotation For Annovar tool, use HOWARD keyword annovar and mention specific Annovar database keywords (separator :). More options are available using HOWARD Parameters JSON file.

Example: VCF annotation with Annovar refGene and cosmic70

```
howard annotation \
    --input='tests/data/example.vcf.gz' \
    --annotations='annovar:refGene:cosmic70' \
    --output='/tmp/example.howard.tsv'
```

4.5.1.2.3 snpEff annotation For snpEff tool, use HOWARD keyword **snpeff**. Options are available for quick annotation with snpEff, see HOWARD Parameters JSON - snpEff for more options.

Example: VCF annotation with snpEff

```
howard annotation \
    --input='tests/data/example.vcf.gz' \
    --annotations='snpeff' \
    --output='/tmp/example.howard.tsv'
```

4.5.1.2.4 Exomiser Annotation For Exomiser tool, use HOWARD keyword exomiser. A list of options can be provided as key-value format, such as exomiser release, a preset (pre-configured options), source of transcripts (e.g. 'refseq', 'ucsc'), a list of HPO terms (do not use ':' separator, e.g. '0001156', 'HP0001156', 'hpo0001156'). More options are available using HOWARD Parameters JSON file.

Example: VCF annotation with Exomiser (exome preset, list of HPO terms, transcript as refseq and release 2109)

4.5.1.2.5 Splice Annotation For Splice tool, use HOWARD keyword splice. A list of options can be provided as key-value format, such as split mode, spliceAI distance, spliceAI mask. More options are available using HOWARD Parameters JSON file.

Example: VCF annotation with Splice (split mode, spliceAI distance, spliceAI mask)

```
howard annotation \
    --input='tests/data/example.vcf.gz' \
    --annotations='splice:split_mode=one:spliceai_distance=500:spliceai_mask=1' \
    --output='/tmp/example.howard.tsv'
```

4.5.1.3 Annotation combination Quick annotation allows to combine annotations, from build-in Parquet method and external tools. Simply use a list with a comma separator.

Example: VCF annotation with build-in Parquet method and external tools (Annovar, snpEff and Exomiser)

See HOWARD Help Annotation tool tool for more information.

4.5.2 Annotation parameters

All annotation parameters can be defined in HOWARD Parameters JSON file. All annotations can be combined (bild-in parquet method and external tools annotation), and options can be detailed in a full JSON file format, including selection of annotation database columns (and rename them) and specific options for external tools.

4.6 Calculation

Calculation processes variants annotations to generate new annotation, such as: identify variation type (VarType), harmonizes allele frequency (VAF) and calculate statistics (VAF_stats), extracts Nomen (transcript, cNomen, pNomen...) from an HGVS field (e.g. snpEff, Annovar) with an optional list of personalized transcripts, generates VaRank format barcode, identify trio inheritance. These calculations are based on existing annotations of variants (and genotypes).

Calculations are either provided by HOWARD within code, or configured into a JSON file. Calculations are either an inner HOWARD Python code, or a SQL query.

See HOWARD Help Calculation tool tool for more information.

To process a calculation, use is keyword with the --calculations parameter.

Example: calculation of the variant type with vartype keyword

```
howard calculation \
    --input='tests/data/example.full.vcf' \
    --calculations='vartype' \
    --output='/tmp/example.calculation.tsv'
```

4.6.1 Available calculations

To list all available calculations, from HOWARD default configuration or with a homemade Calculation configuration JSON file, use the --show_calculations parameter.

Example: List of build-in calculation

```
howard calculation \
   --show calculations
#[INFO] Start
#[INFO] Available calculation operations:
#[INFO]
           BARCODE: BARCODE as VaRank tool
           BARCODEFAMILY: BARCODEFAMILY as VaRank tool
#[INFO]
#[INFO]
           DP STATS: Depth (DP) statistics
           FINDBYPIPELINE: Number of pipeline that identify the variant (for multi pipeline VCF)
#[INFO]
#[INFO]
           FINDBYSAMPLE: Number of sample that have a genotype for the variant (for multi sample VCF)
           GENOTYPECONCORDANCE: Concordance of genotype for multi caller VCF
#[INFO]
           NOMEN: NOMEN information (e.g. NOMEN, CNOMEN, PNOMEN...) from HGVS nomenclature field
#[INFO]
           SNPEFF ANN EXPLODE: Explode snpEff annotations
#[INFO]
#[INFO]
           SNPEFF_ANN_EXPLODE_JSON: Explode snpEff annotations in JSON format
#[INFO]
           SNPEFF_ANN_EXPLODE_UNIQUIFY: Explode snpEff annotations with uniquify values
#[INFO]
           SNPEFF_HGVS: HGVS nomenclatures from snpEff annotation
           TRANSCRIPTS_ANN: Add transcripts annotations in structured format (field 'transcripts_ann')
#[INFO]
#[INFO]
           TRANSCRIPTS_ANNOTATIONS: Add transcripts annotations in JSON and/or structured format (see pa
#[INFO]
           TRANSCRIPTS JSON: Add transcripts annotations in JSON format (field 'transcripts json')
           TRANSCRIPTS_PRIORITIZATION: Prioritize transcripts with a prioritization profile (using param
#[INFO]
#[INFO]
           TRIO: Inheritance for a trio family
           VAF: Variant Allele Frequency (VAF) harmonization
#[INFO]
#[INFO]
           VAF_STATS: Variant Allele Frequency (VAF) statistics
           VARIANT_ID: Variant ID generated from variant position and type
#[INFO]
#[INFO]
           VARTYPE: Variant type (e.g. SNV, INDEL, MNV, BND...)
#[INFO] End
```

4.6.2 Calculation configuration JSON file

All calculations are configured in a JSON file. A default configuration is provided with default calculations.

Basically, a calculation is defined by: - Type: either 'sql' for a SQL query or 'python' for a Python function - Name/keyword: a keyword that is used with --show_calculations parameter (case unsensitive) - Description: a description of the calculation - Output column information: Name, type and decription of the new annotation calculated - Query and fields: an SQL query (for 'sql' type) with parameters such as mandatory INFO fields - Function name and parameters: a existing Python function and parameters (for 'python' type)

Example: Configuration with calculation of variant type using an SQL query and calculation of variant id using an existing Python function calculation_variant_id

```
{
 "VARTYPE": {
   "type": "sql",
   "name": "VARTYPE",
   "description": "Variant type (e.g. SNV, INDEL, MNV, BND...)",
   "available": true,
   "output_column_name": "VARTYPE",
   "output_column_type": "String",
   "output_column_description": "Variant type: SNV, MOSAIC or INDEL",
   "operation_query": [
      "CASE",
      "WHEN \"SVTYPE\" NOT NULL THEN \"SVTYPE\"",
     "WHEN LENGTH(REF) = 1 AND LENGTH(ALT) = 1 THEN 'SNV'",
     "WHEN REF LIKE '%,%' OR ALT LIKE '%,%' THEN 'MOSAIC'",
     "WHEN LENGTH(REF) == LENGTH(ALT) AND LENGTH(REF) > 1 THEN 'MNV'",
      "WHEN LENGTH(REF) <> LENGTH(ALT) THEN 'INDEL'",
     "ELSE 'UNDEFINED'",
     "END"
   "info_fields": ["SVTYPE"],
```

```
"operation_info": true
},

"variant_id": {
    "type": "python",
    "name": "variant_id",
    "description": "Variant ID generated from variant position and type",
    "available": true,
    "function_name": "calculation_variant_id",
    "function_params": []
}
```

See Calculation configuration JSON file example.

See Calculation configuration JSON file for more information.

4.6.3 Build-in calculations examples

4.6.3.1 Variant type Variant type calculation vartype detect the type of variant (e.g. SNV, INDEL, MNV). Variant type are claculated with these criteria: SNV if X>Y, MOSAIC if X>Y,Z or X,Y>Z, INDEL if XY>Z or X>YZ.

Example: Identify variant types and generate a table of variant type count

```
howard calculation \
   --input='tests/data/example.full.vcf' \
   --calculations='vartype' \
   --output='/tmp/example.calculation.tsv'
howard query \
   --input='/tmp/example.calculation.tsv' \
   --explode_infos \
   --query='SELECT
                "VARTYPE" AS 'VariantType',
               count(*) AS 'Count'
            FROM variants
            GROUP BY "VARTYPE"
            ORDER BY count DESC'
  VariantType Count
          BND
                    7
          DUP
1
                    6
2
          INS
                    5
3
          SNV
                    4
          CNV
                    3
4
5
          DEL
6
          INV
7
       MOSAIC
                    2
                    2
8
        INDEL
9
          MNV
                    1
```

4.6.3.2 HGVS and NOMEN from snpEff NOMEN can be extracted from snpEff annotation (see HOWARD Parameters JSON - snpEff). The first calculation extract list of HGVS annotations from snpEff annotation (snpeff_hgvs keyword), the second calculation choose the NOMEN from snpEff HGVS annotations using a list of reference transcripts (NOMEN keyword, --hgvs_field and --transcripts parameters). More options are available (see HOWARD Parameters JSON). See Parameters for more information about list of reference transcripts

Example: Calculate NOMEN by extracting hgvs from snpEff annotation and identifying transcripts from a list

```
howard calculation \
    --input='tests/data/example.ann.vcf.gz' \
    --calculations='snpeff_hgvs,NOMEN' \
    --hgvs_field='snpeff_hgvs' \
    --transcripts='tests/data/transcripts.tsv' \
```

4.7 Prioritization

See HOWARD Help Prioritization tool tool for more information.

Prioritization algorithm uses profiles to flag variants (as passed or filtered), calculate a prioritization score, and automatically generate a comment for each variants (example: 'polymorphism identified in dbSNP. associated to Lung Cancer. Found in ClinVar database'). Prioritization profiles are defined in a configuration file in JSON format. A profile is defined as a list of annotation/value, using wildcards and comparison options (contains, lower than, greater than, equal...). Annotations fields may be quality values (usually from callers, such as 'DP') or other annotations fields provided by annotations tools, such as HOWARD itself (example: COSMIC, Clinvar, 1000genomes, PolyPhen, SIFT).

4.7.1 Prioritization options

Multiple profiles can be used simultaneously (--prioritizations option), which is useful to define multiple validation/prioritization levels (e.g. 'standard', 'stringent', 'rare variants', 'low allele frequency', 'GERMLINE'). By default, all profiles will be processed. A default profile can be defined with --default_profile option (by default, the first profile in list of profiles is selected).

Prioritization score can be calculated following multiple mode. The HOWARD mode will increment scores of all passing filters (default). The VaRank mode will select the maximum score from all passing filters.

Prioritization fields can be selected from: - PZScore: calculated score from all passing filters, depending of the mode - PZFlag: final flag ('PASS' or 'FILTERED'), with strategy that consider a variant is filtered as soon as at least one filter do not pass. By default, the variant is considered as 'PASS' (no filter pass) - PZComment: concatenation of all passing filter comments - PZ-Tags: combinason of score, flags and comments in a tags format (e.g. 'PZFlag#PASS|PZScore#15|PZComment#Described on ...') - PZInfos: information about passing filter criteria

Example: Prioritize variants from criteria on INFO annotations for profiles 'default' and 'GERMLINE' (from 'prioritization_profiles.json' profiles configuration), export prioritization tags, and query variants by ordering with flags and scores

```
howard prioritization \
   --input='tests/data/example.vcf.gz' \
   --prioritizations='default,GERMLINE' \
   --prioritization_config='tests/data/prioritization_profiles.json' \
   --default_profile='default' \
   --pzfields='PZFlag,PZScore,PZClass,PZComment,PZTags,PZInfos' \
   --prioritization_score_mode='HOWARD' \
   --output='/tmp/example.prioritized.vcf.gz'
howard query \
   --input='/tmp/example.prioritized.vcf.gz' \
   --explode_infos \
   --query="SELECT \"#CHROM\", POS, ALT, REF, PZFlag, PZScore, DP, CLNSIG \
            FROM variants \
            ORDER BY PZFlag DESC, PZScore DESC"
                               PZFlag
  #CHROM
               POS ALT REF
                                      PZScore
                                                    DP
                                                                CLNSIG
0
    chr1
             69101
                      G
                          Α
                                 PASS
                                            105
                                                  50.0
                                                                  None
          55249063
                          G
                                 PASS
                                                 125.0
    chr7
                     Α
                                            105
                                                                  None
1
2
                     С
    chr1
             28736
                          A
                                 PASS
                                            15
                                                   NaN
                                                            pathogenic
```

```
3
    chr1
              768251
                        G
                             Α
                                     PASS
                                                    0
                                                          NaN
                                                                            None
4
              768252
                        G
                                     PASS
                                                    0
                                                                           None
    chr1
                             Α
                                                          NaN
5
    chr1
              768253
                        G
                             Α
                                     PASS
                                                    0
                                                          NaN
                                                                           None
                                                 -85
6
    chr1
               35144
                        C
                             A FILTERED
                                                          \mathtt{NaN}
                                                              non-pathogenic
```

4.7.2 Prioritization query

Prioritization fields can be then easily querying, by filtering on fields and order by fields.

Example: Query variants using prioritization fields

```
howard query \
   --input='/tmp/example.prioritized.vcf.gz' \
   --explode_infos \
   --query="SELECT \"#CHROM\", POS, ALT, REF, PZFlag, PZScore, DP, CLNSIG \
            FROM variants \
            WHERE PZScore > 0 \
               AND PZFlag == 'PASS' \
            ORDER BY PZFlag DESC, PZScore DESC"
  #CHROM
               POS ALT REF PZFlag PZScore
                                                 DP
                                                          CLNSIG
0
    chr1
             69101
                      G
                          Α
                              PASS
                                         105
                                               50.0
                                                            None
1
    chr7
          55249063
                      Α
                          G
                              PASS
                                         105
                                              125.0
                                                            None
2
    chr1
             28736
                      С
                          Α
                              PASS
                                          15
                                                NaN
                                                     pathogenic
Example: Query variants with different prioritization flag between profiles
howard query \
   --input='/tmp/example.prioritized.vcf.gz' \
   --explode_infos \
   --query="SELECT \"#CHROM\", POS, ALT, REF, PZFlag_default, PZFlag_GERMLINE \
            FROM variants \
            WHERE PZFlag_default != PZFlag_GERMLINE \
            ORDER BY PZFlag DESC, PZScore DESC"
  #CHROM
            POS ALT REF PZFlag_default PZFlag_GERMLINE
         35144
                               FILTERED
                                                     PASS
Example: Showing propritization comments of variants, with flags and scores
howard query \
   --input='/tmp/example.prioritized.vcf.gz' \
   --explode_infos \
   --query="SELECT \"#CHROM\", POS, ALT, REF, PZComment, PZFlag \
            FROM variants WHERE PZComment IS NOT NULL \
            ORDER BY PZFlag DESC, PZScore DESC"
  #CHROM
               POS ALT REF
                                                                        PZComment
                                                                                     PZFlag
    chr1
             69101
                      G
                          Α
                                                DP, Variant probably pathogenic
                                                                                        PASS
                          G
1
          55249063
                                                DP, Variant probably pathogenic
                                                                                        PASS
    chr7
                      Α
2
                      С
                                                  Described on CLINVAR database
                                                                                        PASS
             28736
                          Α
    chr1
3
             35144
                      C
                          Α
                             Described on CLINVAR database, Described on CL...
                                                                                   FILTERED
    chr1
```

Prioritization profiles are defined in a JSON configuration file. Each profiles are defined as a list of annotation fields with associated filters (type of comparison and threshold, with related score, flag and comment).

Example: Profiles with 2 filters on annotation field 'DP' (threashold 50), 2 filters on annotation field 'CLNSIG' ("pathogenic" or "non-pathogenic"), and 1 filter combining 'DP' and 'CLNSIG' (with SQL)

```
"score": 5,
            "flag": "PASS",
            "comment": [
                "DP higher than 50"
            ]
        },
            "type": "lt",
            "value": "50",
            "score": 0,
            "flag": "FILTERED",
            "comment": [
                "DP lower than 50"
        }
    ],
    "CLNSIG": [
            "type": "equals",
            "value": "pathogenic",
            "score": 15,
            "flag": "PASS",
            "comment": [
                "Described on CLINVAR database as pathogenic"
       },
        {
            "type": "equals",
            "value": "non-pathogenic",
            "score": -100,
            "flag": "FILTERED",
            "comment": [
                "Described on CLINVAR database as non-pathogenic"
            ]
        }
    ],
    "CLNSIG and DP filter": [
          "sql": " DP >= 50 OR regexp_matches(CLNSIG, 'Pathogenic') ",
          "fields": ["DP", "CLNSIG"],
          "score": 100,
          "flag": "PASS",
          "comment": ["Variant probably pathogenic"]
        }
      ]
}
```

See HOWARD Help Prioritization Profiles for more options.

4.8 HGVS annotation

}

HOWARD annotates variants with HGVS annotation using HUGO HGVS internation Sequence Variant Nomenclature (http://varnomen.hgvs.org/). Annotation refere to refGene and genome to generate HGVS nomenclature for all available transcripts. This annotation add 'hgvs' field into VCF INFO column of a VCF file.

To enhance the functionality of HGVS tool, several options available. The --use_gene option enables the utilization of gene information for the generation of HGVS annotation, providing a concise representation such as 'NM_152232(TAS1R2):c.231T>C'. Alternatively, the --use_exon option incorporates exon details into the annotation, as seen in the example 'NM_152232(exon2):c.231T>C', but this is only applicable if 'use_gene' is not enabled. For a

more detailed HGVS annotation, activate the <code>--full_format</code> option, which generates a comprehensive HGVS annotation, including all available information, such as 'TAS1R2:NM_152232:NP_689418:c.231T>C:p.Cys77Arg'. This is non-standard format that ensures exhaustive data representation. Furthermore, specify the format of amino acid codons with the <code>--codon_type</code> option, choosing between single-character ('1'), three-character ('3'), or full-name formats ('FULL'). Additionally, protein-level information can be integrated using the <code>--use_protein</code> option, enabling annotations like 'NP_689418:p.Cys77Arg', which can be combined with DNA-level annotations using the <code>--add_protein</code> option (e.g. 'NM_152232:c.231T>C,NP_689418:p.Cys77Arg').

All these options can be combined into one quick option --hgvs. Simply concatenate options (separator ',') with there value (e.g --hgvs=use_gene:True or --hgvs=use_gene:False,codon_type:FULL). By default, value option is 'True' (e.g. --hgvs=use_gene is equal to --hgvs=use_gene:True)

For database management, specify refSeq annotation files with --refgene and --refseqlink options (or set custom folders with --refseq-folder), and genomes folder using and --genomes-folder (folders are set by default, see HOWARD Configuration JSON file).

See HOWARD Help Prioritization tool for more options.

```
Example: HGVS annotation with quick options
howard hgvs \
   --input='tests/data/example.vcf.gz' \
   --output='/tmp/example.process.tsv' \
   --hgvs='full_format,use_exon'
howard query \
   --input='/tmp/example.process.tsv' \
   --explode_infos \
   --query="SELECT hgvs \
            FROM variants "
                                                 hgvs
0
                      WASH7P:NR 024540.1:n.50+585T>G
1
      FAM138A:NR_026818.1:exon3:n.597T>G:p.Tyr199Asp
2
  OR4F5:NM_001005484.2:NP_001005484.2:exon3:c.74...
  LINCO1128:NR_047526.1:n.287+3767A>G,LINCO1128:...
  LINCO1128:NR_047526.1:n.287+3768A>G,LINCO1128:...
  LINCO1128:NR_047526.1:n.287+3769A>G,LINCO1128:...
  EGFR: NM_001346897.2: NP_001333826.1: exon19: c.22...
Example: HGVS annotation with separated options
howard hgvs \
   --input='tests/data/example.vcf.gz' \
   --output='/tmp/example.process.tsv' \
   --full_format \
   --use exon
howard query \
   --input='/tmp/example.process.tsv' \
   --explode_infos \
   --query="SELECT hgvs \
            FROM variants "
                                                 hgvs
\cap
                      WASH7P: NR_024540.1:n.50+585T>G
      FAM138A:NR_026818.1:exon3:n.597T>G:p.Tyr199Asp
1
2
  OR4F5:NM_001005484.2:NP_001005484.2:exon3:c.74...
3
  LINCO1128:NR 047526.1:n.287+3767A>G,LINCO1128:...
  LINCO1128:NR_047526.1:n.287+3768A>G,LINCO1128:...
4
5
  LINCO1128:NR_047526.1:n.287+3769A>G,LINCO1128:...
  EGFR: NM_001346897.2: NP_001333826.1: exon19: c.22...
```

4.9 Process

HOWARD process tool manage genetic variations to:

- annotates genetic variants with multiple annotation databases/files and tools
- calculates and normalizes annotations
- prioritizes variants with profiles (list of citeria) to calculate scores and flags
- annotates genetic variants with HGVS nomenclature
- translates into various formats
- query genetic variants and annotations
- generates variants statistics

This process tool combines all other tools to pipe them in a uniq command, through available options or a parameters file in JSON format (see HOWARD Parameters JSON file).

See HOWARD Help Process tool tool for more information (under development).

Process with options

Process tool uses quick options for annotation, calculation and prioritization to enrich variant file, and to query variants annotations.

Example: Process command with options (HGVS, annotation, calculation)

```
howard process \
   --input='tests/data/example.vcf.gz' \
   --output='/tmp/example.process.tsv' \
   --hgvs='full_format,use_exon' \
   --annotations='tests/databases/annotations/current/hg19/avsnp150.parquet,
                  tests/databases/annotations/current/hg19/dbnsfp42a.parquet,
                  tests/databases/annotations/current/hg19/gnomad211_genome.parquet' \
   --calculations='NOMEN' \
   --prioritizations='default' \
   --prioritization_config='tests/data/prioritization_profiles.json' \
   --explode_infos \
   --query="SELECT NOMEN,
                   PZFlag,
                   avsnp150 AS 'snpID',
                   SIFT score AS 'SIFT',
                   AF AS 'gnomAD',
                   ClinPred_pred AS 'ClinPred' \
                FROM variants"
                                             NOMEN
                                                                           SIFT
                                                                                 gnomAD ClinPred
                                                      PZFlag
                                                                   snpID
                     WASH7P: NR_024540: n.50+585T>G
                                                        PASS
                                                                    None
                                                                           None
                                                                                   None
                                                                                             None
     FAM138A:NR_026818:exon3:n.597T>G:p.Tyr199Asp
                                                    FILTERED
                                                                    None
                                                                           None
                                                                                   None
                                                                                             None
      OR4F5:NM_001005484:exon3:c.74A>G:p.Glu25Gly
                                                        PASS
                                                                    None
                                                                          0.005
                                                                                   None
                                                                                                D
                LINCO1128:NR_047526:n.287+3767A>G
                                                        PASS
                                                                    None
                                                                           None
                                                                                   None
                                                                                             None
                LINCO1128:NR_047526:n.287+3768A>G
                                                        PASS
                                                                    None
                                                                           None
                                                                                   None
                                                                                             None
                LINCO1128:NR 047526:n.287+3769A>G
                                                        PASS
                                                                    None
                                                                           None
                                                                                   None
                                                                                             None
  EGFR:NM 001346897:exon19:c.2226G>A:p.Gln742Gln
                                                        PASS
                                                              rs1050171
                                                                           None
                                                                                0.5029
                                                                                             None
```

Example: Full process command with options (HGVS, annotation parquet, annotation beftools, snpEff and Annovar, calculation and prioritization, and query)

```
howard process \
```

0

1

2

3

4

5

```
--input='tests/data/example.vcf.gz' \
--output='/tmp/example.process.tsv' \
--hgvs='full_format,use_exon' \
--annotations='tests/databases/annotations/current/hg19/avsnp150.parquet,
               tests/databases/annotations/current/hg19/dbnsfp42a.parquet,
               tests/databases/annotations/current/hg19/gnomad211_genome.parquet,
               bcftools:tests/databases/annotations/current/hg19/cosmic70.vcf.gz,
               snpeff,
```

```
annovar:refGene' \
   --calculations='vartype, snpeff hgvs, VAF, NOMEN' \
   --prioritizations='default' \
   --prioritization_config='config/prioritization_profiles.json' \
   --explode_infos \
   --query="SELECT string_split(snpeff_hgvs, ',')[1] AS 'HGVS',
                   PZScore,
                   Func_refGene AS 'Location',
                   string_split(string_split(cosmic70, ',')[2], '=')[2] AS 'COSMIC' \
            FROM variants \
            ORDER BY PZScore DESC"
                                             HGVS
                                                  PZScore
                                                                   Location
                                                                                          COSMIC
  EGFR: NM_005228.5 : exon20 : c.2361_{G} > A : p.Gln787Gln
0
                                                        105
                                                                            1(large_intestine)
                                                                     exonic
1
                MIR1302-2:NR 036051.1:n.-1630A>C
                                                        15
                                                             ncRNA intronic
                                                                                            None
2
    OR4F5:NM_001005484.1:exon1:c.11A>G:p.Glu4Gly
                                                         5
                                                                                            None
                                                                     exonic
                                                          0 ncRNA_intronic
                                                                                            None
3
       LINCO1128:NR_047519.1:exon2:n.287+3767A>G
4
       LINCO1128: NR_047519.1: exon2:n.287+3768A>G
                                                          0 ncRNA_intronic
                                                                                            None
5
       LINCO1128:NR_047519.1:exon2:n.287+3769A>G
                                                          0
                                                             ncRNA_intronic
                                                                                            None
6
                MIR1302-2:NR_036051.1:n.*4641A>C
                                                       -100
                                                               ncRNA exonic
                                                                                            None
```

4.9.2 Process with parameters JSON file

In order to fine tune process, all tools can be defined in a HOWARD Parameters JSON. This allows to add specific options, such as selecting specific fields (and rename them) for annotation, defining options for external tools, specifying a list of transcripts of preference for NOMEN calculation. This Parameters JSON file can be combine with options.

Example: Full process command with Parameters JSON file example and a query as option

```
howard process \
   --input='tests/data/example.vcf.gz' \
   --output='/tmp/example.process.tsv' \
   --param='config/param.json' \
   --explode_infos \
   --query="SELECT NOMEN, PZComment \
            FROM variants \
            ORDER BY PZScore DESC"
                                                                                              PZComment
                                             NOMEN
   EGFR:NM 001346897:exon19:c.2226G>A:p.Gln742Gln
                                                     DP higher than 50, Described on CLINVAR databa...
0
1
               WASH7P: NR_024540: exon1: n.50+585T>G
                                                           Described on CLINVAR database as pathogenic
2
       OR4F5:NM_001005484:exon1:c.11A>G:p.Glu4Gly
                                                                                      DP higher than 50
3
          LINCO1128:NR 047519:exon2:n.287+3767A>G
                                                                                                    None
4
          LINC01128: NR_047519: exon2: n.287+3768A>G
                                                                                                    None
5
          LINCO1128: NR_047519: exon2: n.287+3769A>G
                                                                                                    None
                   MIR1302-9:NR_036266:n.*4641A>C
                                                       Described on CLINVAR database as non-pathogenic
cat '/tmp/example.process.tsv' | cut -c 1-80
NOMEN
        PZComment
EGFR:NM_001346897:exon19:c.2226G>A:p.Gln742Gln DP higher than 50, Described on C
WASH7P:NR_024540:exon1:n.50+585T>G
                                         Described on CLINVAR database as pathogenic
OR4F5:NM_001005484:exon1:c.11A>G:p.Glu4Gly
                                                  DP higher than 50
LINC01128:NR_047519:exon2:n.287+3767A>G
LINCO1128:NR_047519:exon2:n.287+3768A>G
LINCO1128:NR 047519:exon2:n.287+3769A>G
MIR1302-9:NR_036266:n.*4641A>C Described on CLINVAR database as non-pathogenic
Example: Full process command with Parameters JSON file example and switch off query (configured in param
JSON file) and generate a VCF file
howard process \
   --input='tests/data/example.vcf.gz' \
```

```
--output='/tmp/example.process.vcf' \
  --param='config/param.json' \
   --explode infos \
  --query=""
howard query \
  --input='/tmp/example.process.vcf' \
  --explode_infos \
  --query="SELECT \"#CHROM\", POS, ALT, REF, NOMEN, PZFlag, PZScore \
          FROM variants \
          ORDER BY PZScore DESC"
             POS ALT REF
 #CHROM
                                                               NOMEN
                                                                       PZFlag PZScore
0
  chr7 55249063 A G EGFR:NM_001346897:exon19:c.2226G>A:p.Gln742Gln
                                                                         PASS
                                                                               100.0
         28736 C A
                                    WASH7P:NR 024540:exon1:n.50+585T>G
                                                                         PASS
                                                                                 15.0
1 chr1
2 chr1
          69101 G A
                             OR4F5:NM_001005484:exon1:c.11A>G:p.Glu4Gly
                                                                         PASS
                                                                                  5.0
3 chr1 768251 G A
                                LINCO1128:NR_047519:exon2:n.287+3767A>G
                                                                         PASS
                                                                                  0.0
4 chr1 768252 G A
                                LINC01128:NR_047519:exon2:n.287+3768A>G
                                                                         PASS
                                                                                  0.0
5 chr1 768253 G A
                                LINCO1128: NR_047519: exon2: n.287+3769A>G
                                                                         PASS
                                                                                  0.0
6 chr1 35144 C A
                                        MIR1302-9:NR_036266:n.*4641A>C FILTERED
                                                                                  NaN
```