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PathMolD-AB: Generation of Spatiotemporal Pathways of Protein Folding Using Molecular Dynamics with a Coarse-grained Model

Step-by-step: Generate your first protein folding pathway and analyze it

The purpose of this tutorial is to describe in a simplified way how to generate your first folding trajectory data and visualize the generated folding data for your analysis with PathMolD-AB. The steps required to accomplish this goal are presented below.

step1) Generate your protein AB sequence based on a FASTA file
step2) Set the protein information and the simulation parameters
step3) Compile and execute the protein folding simulation
step4) Realize the protein folding analysis

obs: words highlighted with yellow represent files and red represent line commands.

##
step1) generate your protein AB sequence based on a FASTA file
##
1.1) Download the FASTA file in the PDB database, the square in the figure below highlights the download location.

In this sample, we download the 2GB1 fasta file from https://www.rcsb.org/structure/2GB1

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as presented in the below Figure

1.2) Access the directory SRC, then, execute the command below.
format
$ python AB_sequence.py <pdb id of the fasta file>
example
$ python AB_sequence.py 2GB1

*Make sure that the file fasta is in the INPUT directory.
*The AB_sequence.py was developed in Python 2.7 language.
*Here, we use the Alberts AB classification.

The program will generate the AB sequence file of the 2GB1 protein in the INPUT directory.
example: 2GB1.txt

The content of the 2GB1.txt file is show below.
ABBBAAABABBABABBBBBAABAABABBAABBBABBBAABABABBBBABBBABABB
,where A and B represents the Hydrophobic and Polar amino acids, respectively.

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##
step2) set the protein information and the simulation parameters
##

Insert the information about the protein and the simulation parameters into the input file. The file 2GB1_56.in (in the INPUT directory) is a sample of this input file for the protein 2GB1. The descriptions of each information are present below.

sequence : AB sequence of the protein (use step1 to get this information)
ProtLen : protein size, number of amino acids
LV : length of the 2D or 3D box
stepLimit : max number of steps
temperature : temperature
savepathways : y enable the function that saves amino acids coordinates in text files. This files can be used to show an example of a pathway
pathwaysstep : steps between generations of text coordinate files.

The content of the 2GB1_56.in file is show below.
sequence = ABBBAAABABBABABBBBBAABAABABBAABBBABBBAABABABBBBABBBABABB
ProtLen = 56
LV = 112
stepLimit = 3000000
temperature = 0.1
savepathways = y
pathwaysstep = 3000

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##
step 3) compile and execute the protein folding simulation
##
*The PathMolD-AB program was developed in C language/CUDA.

3.1) ONLY FOR CPU - Access the SRC_CPU directory and execute the command below to compile the MD program.
example
$ gcc -o a.out main_CPU.c func_MD_CPU.c -lm

3.2) Execute the MD program with the executable file (a.out) with the input file (2GB1.in) and a seed number, to generate a random initial structure of the simulation of the protein folding.
format
$ ./executable <input file> <seed>
example
$ ./a.out ../INPUT/2GB1_56.in 0 && mv pathways56_0.txt ../OUTPUT/pathways56_0.txt

after the execution of MD simulation, it will be generated a output file (pathway data) with the information about the protein structure, free energy and radius of gyration along the iterations (in our sample, the output file generated was pathways56_0.txt and we also move this file to the OUTPUT directory).

3.3) ONLY FOR GPU
*parallel PathMolD-AB is written in C and CUDA.

Access the SRC directory and compile the parallel PathMolD-AB program
example
$ nvcc --gpu-architecture = compute_61 --device -c main.c functions.cu
$ nvcc --gpu-architecture = compute_61 main.o functions.o

3.4) execute the parallel PathMolD-AB program
format
$ ./a.out <input file> <output file> <seed> **>** <GPU ID>
example
$ ./a.out ../INPUT/2GB1_56.in ../OUTPUT/pathways56 0 0
<input file> : parameters of the simulation (generate by the step 2)
<output file> : name of the output file/ pathway data
<seed> : seed used to generate the initial structure of the protein

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<GPU ID> : ID of the GPU that will run the simulation. For computers with only one GPU, use the ID as zero.

*the root directory contains a makefile to run the programs.

```
$ make ab          # convert the AB sequence from the amino acid sequence
$ make gpu         # compile the parallel molecular dynamic version
$ make cpu         # compile the sequential molecular dynamic version
$ make run_gpu     # execute the parallel molecular dynamic version
$ make run_cpu     # execute the sequential molecular dynamic version
$ make vizualize   # generating the folding simulation video
```

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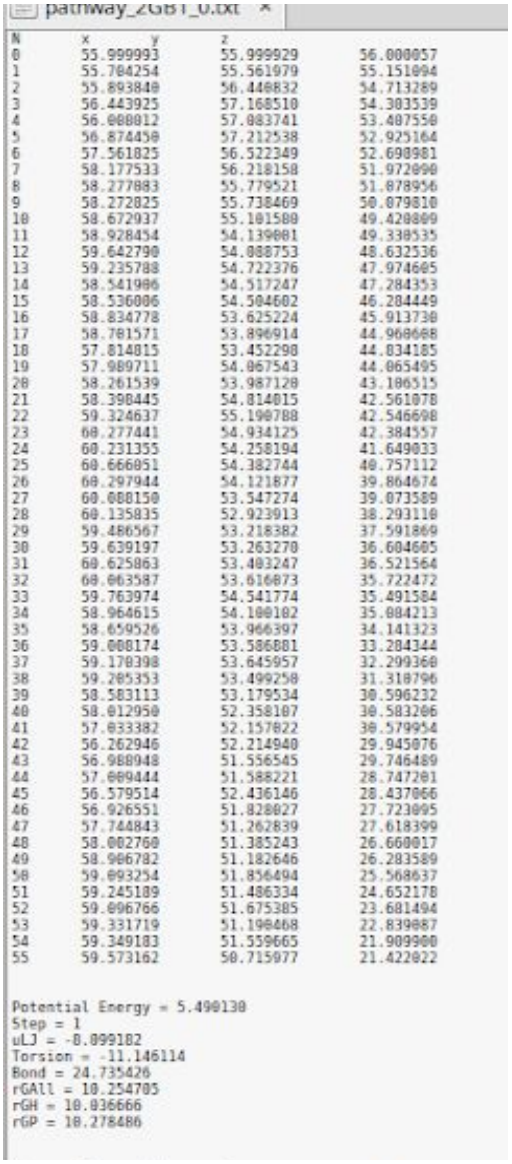
*parallel PathMolD-AB configurations tested (left) a sample of the pathway data generated by PathMolD-AB program (right)

Computing Capability of the PathMolD-AB
The programs were tested in the following configuratic

PathMolD-AB capability (ubuntu 18 LTS)				
	Computing Capability	CUDA7	CUDA8	CUDA9
GTX660	3	NO	NO	NO
K40	3.5	NO	NO	NO
GTX750	5	NO	NO	NO
Titan X	5.2	NO	YES	YES
GTX 1080	6.1	NO	YES	YES
Titan Xp	6.1	NO	YES	YES
GCC/G++		4.8	5.3	6.5
Python		2.7/3.6		



N	x	y	z
0	55.999993	55.999929	56.000057
1	55.704254	55.561979	55.151094
2	55.893840	56.446832	54.713289
3	56.443925	57.168510	54.303539
4	56.008012	57.083741	53.407550
5	56.874450	57.212538	52.925164
6	57.561825	56.522349	52.698981
7	58.177533	56.218158	51.972090
8	58.277883	55.779521	51.078956
9	58.272825	55.738469	50.079810
10	58.672937	55.101580	49.420809
11	58.928454	54.139081	49.338535
12	59.642790	54.088753	48.632536
13	59.235788	54.722376	47.974605
14	58.541986	54.517247	47.284353
15	58.536006	54.504602	46.284449
16	58.834778	53.625224	45.913730
17	58.781571	53.896914	44.960668
18	57.814815	53.452298	44.834185
19	57.989711	54.067543	44.065495
20	58.261539	53.987128	43.106515
21	58.398445	54.814015	42.561078
22	59.324637	55.190788	42.546698
23	60.277441	54.934125	42.384557
24	60.231355	54.258194	41.649033
25	60.666851	54.382744	40.757112
26	60.297944	54.121877	39.864674
27	60.088150	53.547274	39.073589
28	60.135835	52.923913	38.293110
29	59.486567	53.218382	37.591869
30	59.639197	53.263270	36.604605
31	60.625863	53.403247	36.521564
32	60.063587	53.616873	35.722472
33	59.763974	54.541774	35.491584
34	58.964615	54.100102	35.084213
35	58.659526	53.966397	34.141323
36	59.008174	53.586881	33.284344
37	59.170398	53.645957	32.299360
38	59.285353	53.499258	31.318796
39	58.583113	53.179534	30.596232
40	58.012950	52.358107	30.583206
41	57.033382	52.157022	30.579954
42	56.262946	52.214940	29.945076
43	56.988948	51.556545	29.746489
44	57.009444	51.588221	28.747201
45	56.579514	52.436146	28.437066
46	56.926551	51.828027	27.723095
47	57.744843	51.262839	27.618399
48	58.002760	51.385243	26.660017
49	58.906782	51.182646	26.283589
50	59.093254	51.856494	25.568637
51	59.245189	51.486334	24.652178
52	59.096766	51.675385	23.681494
53	59.331719	51.190468	22.839087
54	59.349183	51.559665	21.909900
55	59.573162	50.715977	21.422022

Potential Energy = 5.490130
Step = 1
uLJ = -8.099182
Torsion = -11.146114
Bond = 24.735426
rGAll = 10.254705
rGH = 10.036666
rGP = 10.278486

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step 4) realize the protein folding analysis

##

*The pathway_print_multi-subplot.py script was developed in Python 2.7/3.6 language.

With the output file generated by the PathMolD-AB program (step 3), you can generate images of the protein folding trajectory with the program pathway_print_multi-subplot.py.

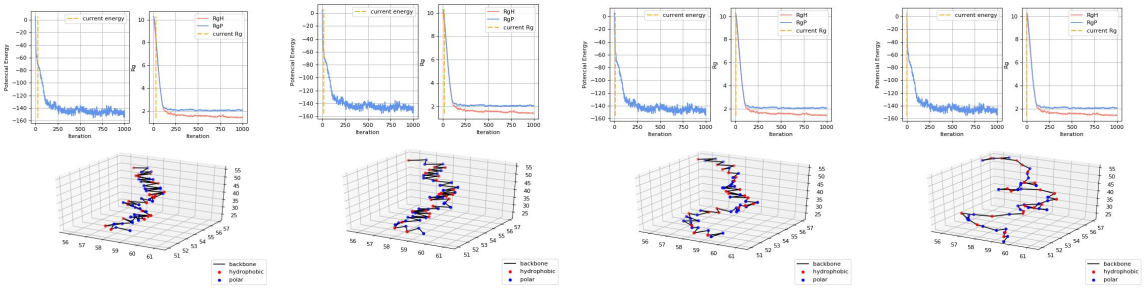
4.1) Before executing pathway_print_multi-subplot.py, you need to set the parameters to read and save the files. Then, open pathway_print_multi-subplot.py file and change the variables *path_pathways*, *filename*, *filesequence* and *path_save*.

path_pathways : folder of the protein pathway data
filename : name of the pathway data (generated by step 3)
filesequence : file containing the AB sequence of the protein (generated by step 1)
path_save :folder where the images will be saved

4.2)execute the command

```
$ python pathway_print_multi-subplot.py
```

Then, the program will produces images of the folding information at each iteration. The samples of the frame 0, 9, 18, and 27 are described below.



The image files of the folding process will be saved in the 'img' folder in this tutorial, and a video of protein protein folding will be generated called folding.mp4.

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Thanks for using PathMolD-AB!!

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for any doubt send a email to leandrotakeshihattori@gmail.com

Link for the Dataset of Spatiotemporal Pathways of Protein Folding:
https://mega.nz/#F!C5QkHQ6A!Ng2xowc2hVPoHHiSB7ww-w