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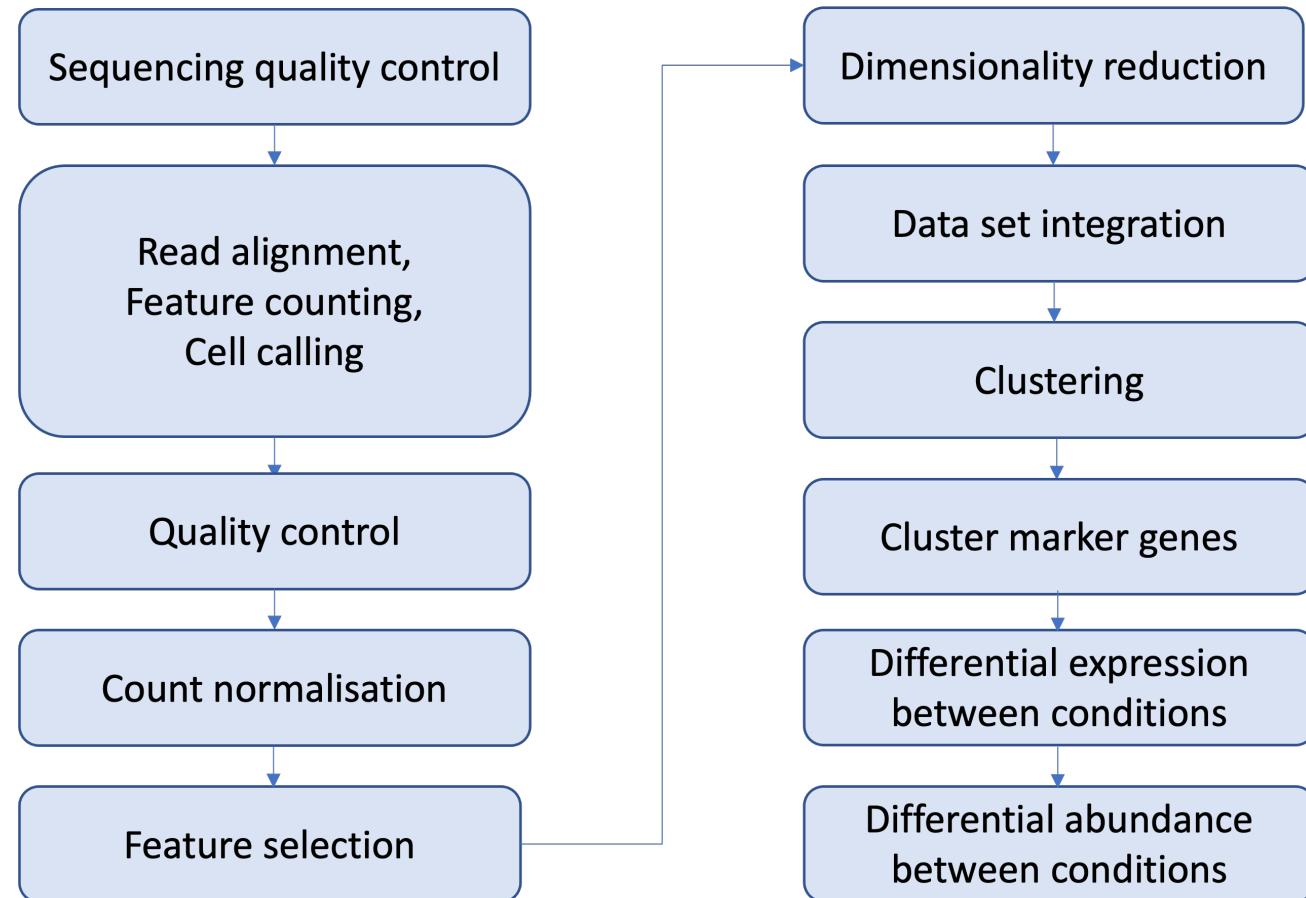
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single-cell RNA-seq Normalisation

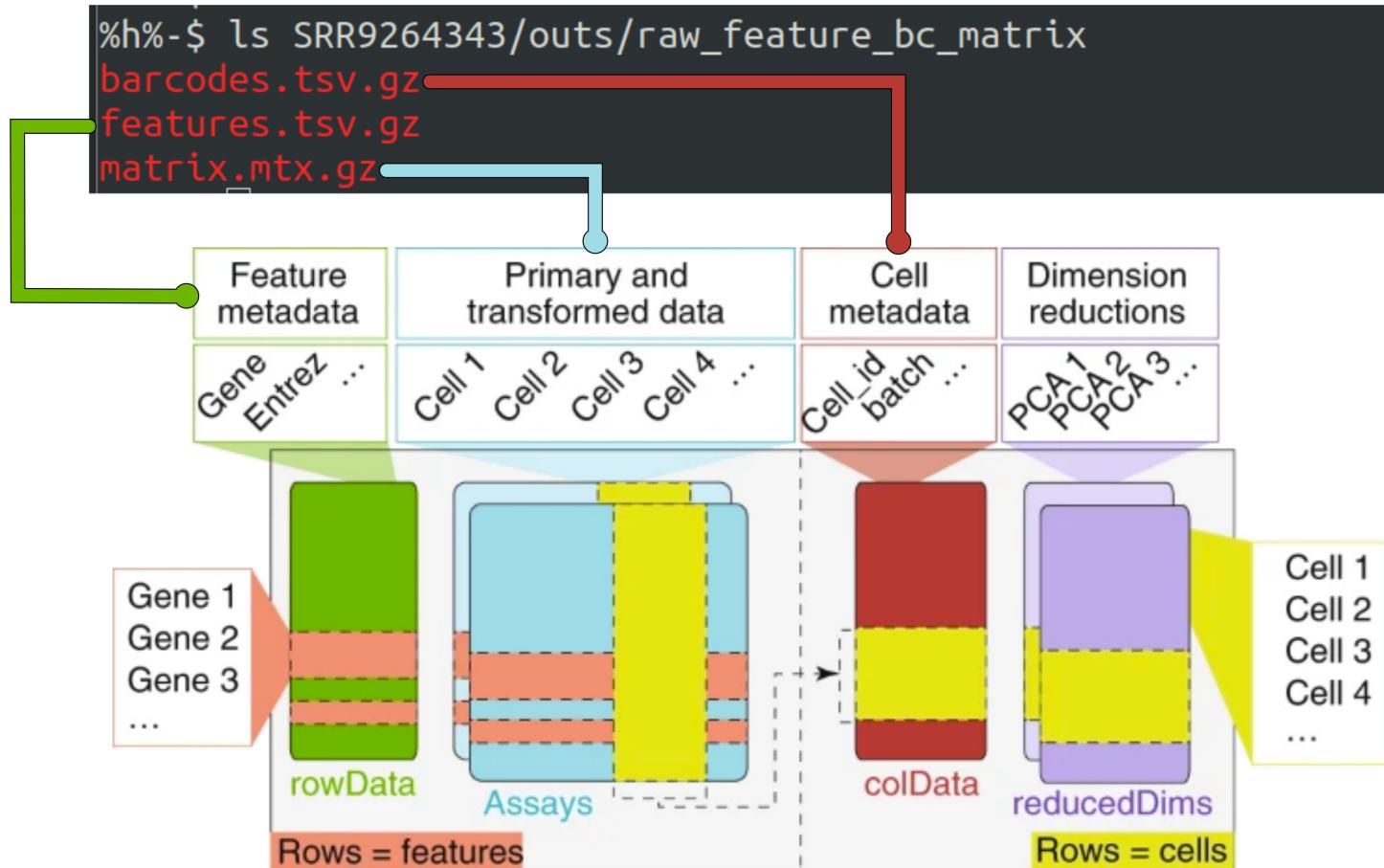
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Workflow



Quality Control (Recap)

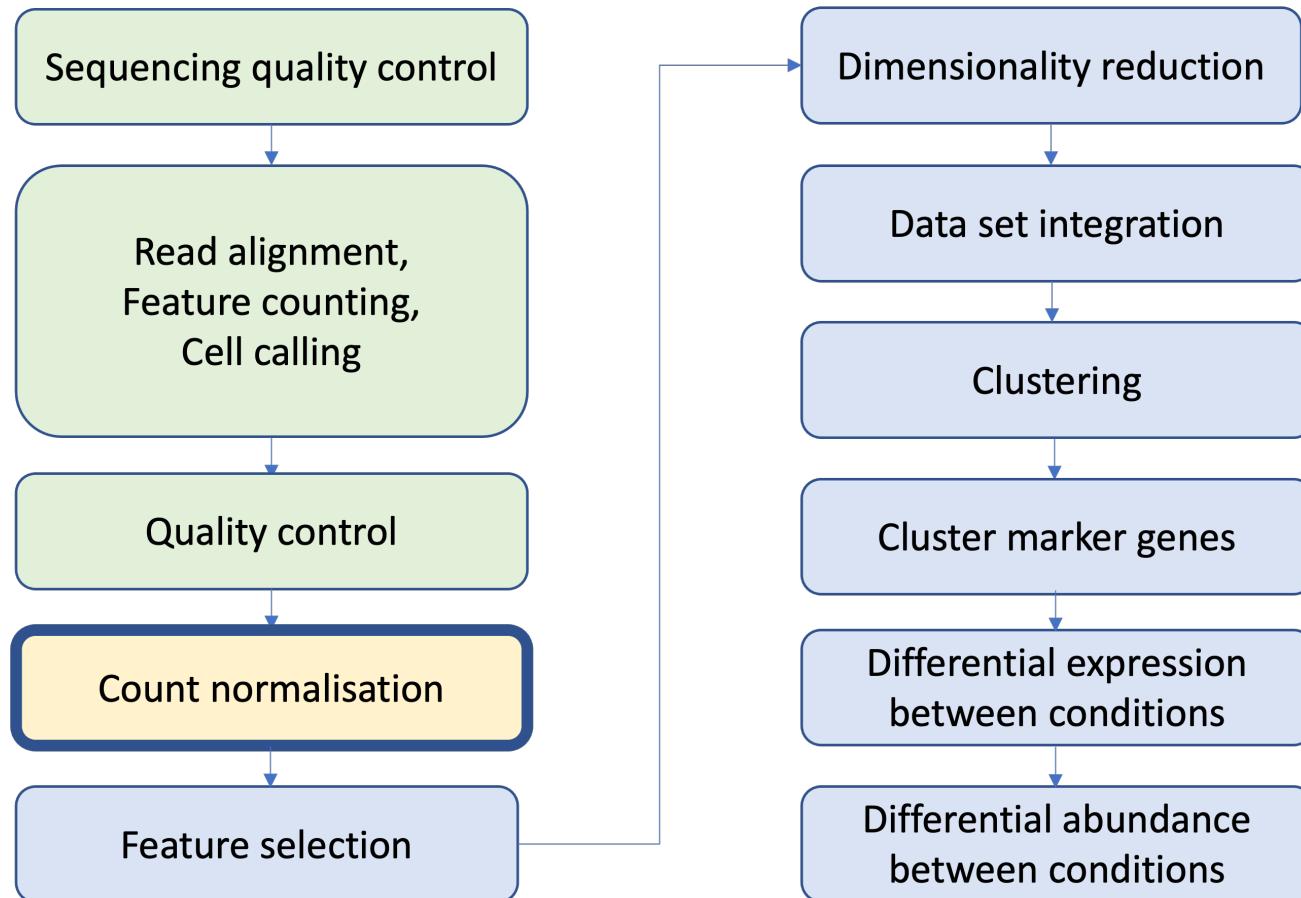


SingleCellExperiment Object

Quality Control (Recap)

- Remove undetected genes
- Empty = no cell in droplet : low gene count
- Doublet/multiplet = more than one cell in droplet : large gene count
- Broken/dead cell in droplet : high proportion of UMIs mapping to mitochondrial genes

Workflow



Why Normalize?

Systematic differences in sequencing coverage between libraries occur because of:

- ✓ low input material
- ✓ differences in mRNA capture
- ✓ differences in PCR amplification

Scaling Normalization

- ✓ In scaling normalization, the “normalization factor” is an estimate of the library size relative to the other cells.
- ✓ Steps usually include:
 - Computation of a cell-specific 'scaling' or 'size' factor
 - division of all counts for the cell by that factor to remove that bias.
- ✓ Assumption: any cell specific bias will affect genes the same way.

Library Size Normalization

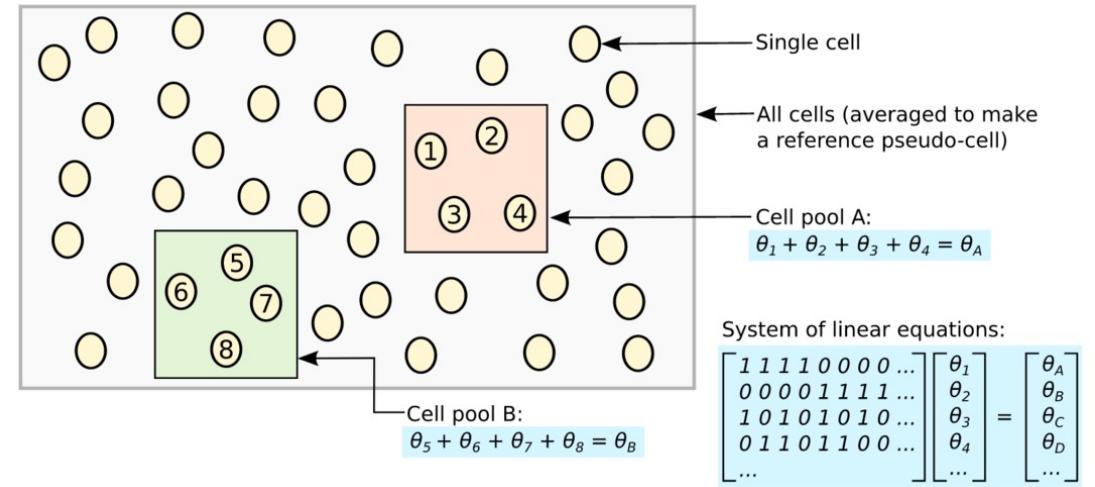
- ✓ The library size factor is proportional to the library size such that the average size factor across cell is one.
- ✓ Divide gene's UMI count in a cell by total number of UMIs in the cell.
- ✓ Multiply the ratio by a scale factor (10,000 by default).
- ✓ Transform the result by taking natural log.

Deconvolution

Deconvolution strategy [Lun et al 2016](#)

The deconvolution method consists of several key steps:

- Defining a pool of cells
- Summing expression values across all cells in the pool
- Normalizing the cell pool against an average reference, using the summed expression values
- Repeating this for many different pools of cells to construct a linear system
- Deconvolving the pool-based size factors to their cell-based counterparts (Fig. 3)



Steps :

- *compute Scaling Factor*
- *apply scaling Factor*