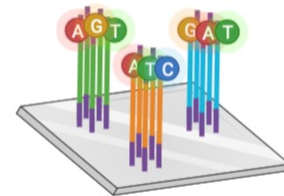


Quality control – ChIP–Seq Data

Junfan Huang

MRC Cancer Unit
University of Cambridge

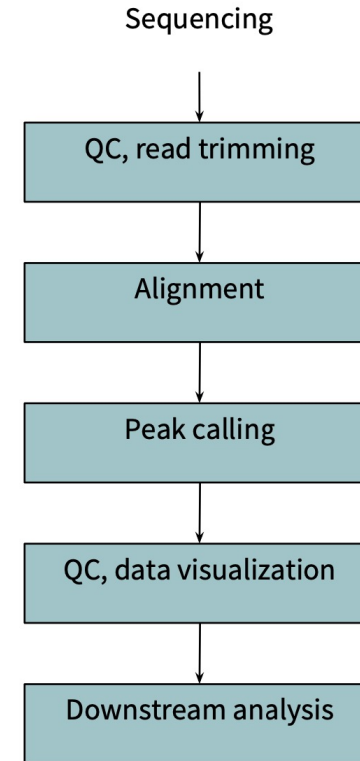
CRUK Bioinformatics Summer School 2021
27th July 2021



Quality control – ChIP-Seq data

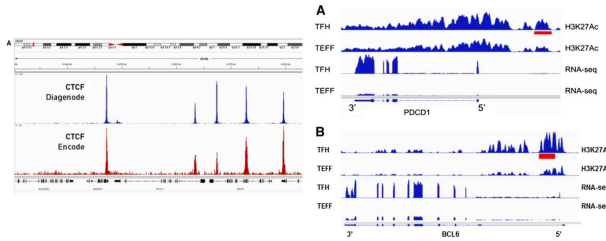
Visually

Computationally

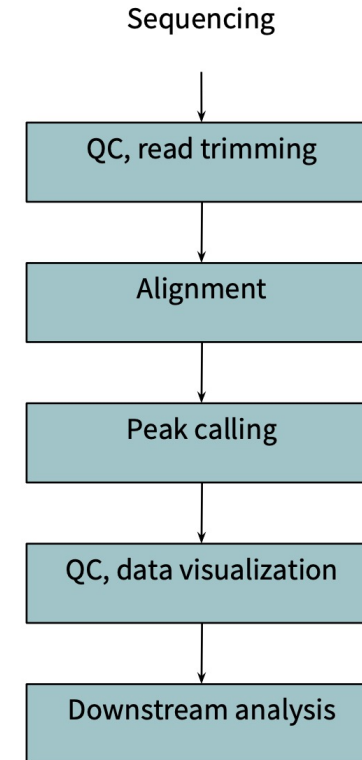


Quality control – ChIP–Seq data

Visually

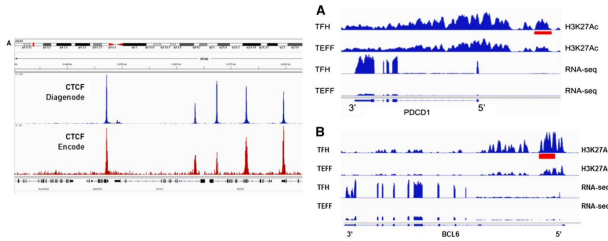


Computationally

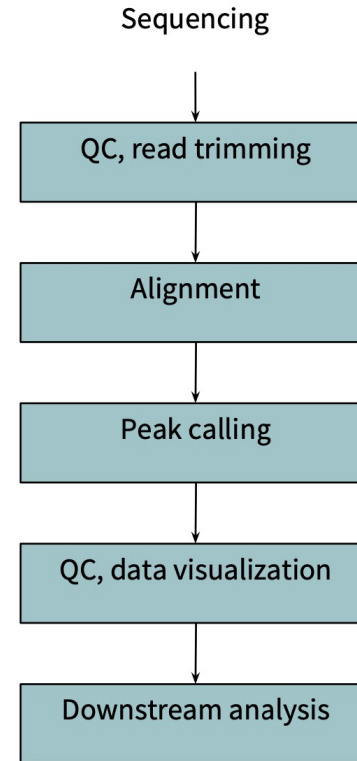


Quality control – ChIP–Seq data

Visually (IGV or USCS genome browser)

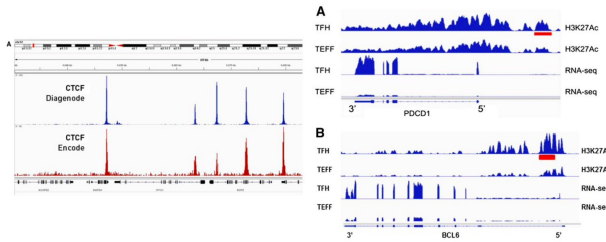


Computationally

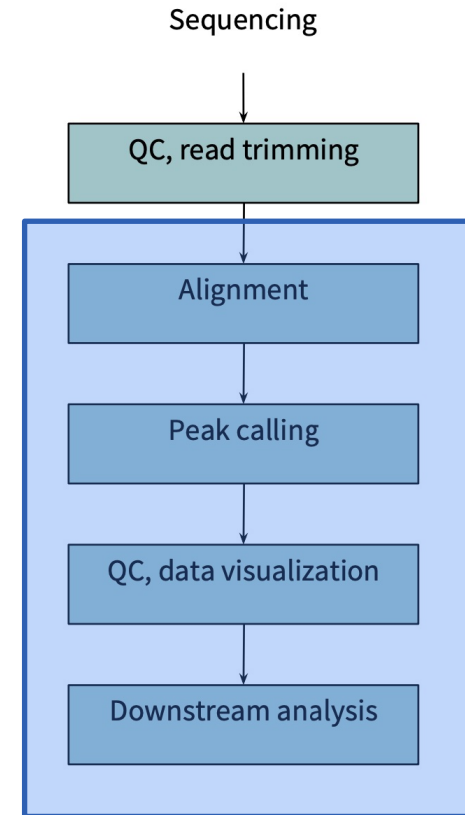


Quality control – ChIP–Seq data

Visually



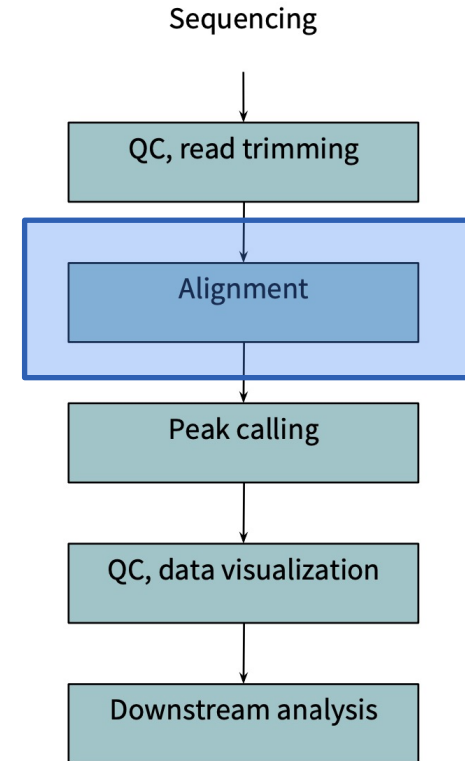
Computationally



Quality control – ChIP-seq data

Visually (Alignment)

- Relative Enrichment in genomic intervals (REGI)

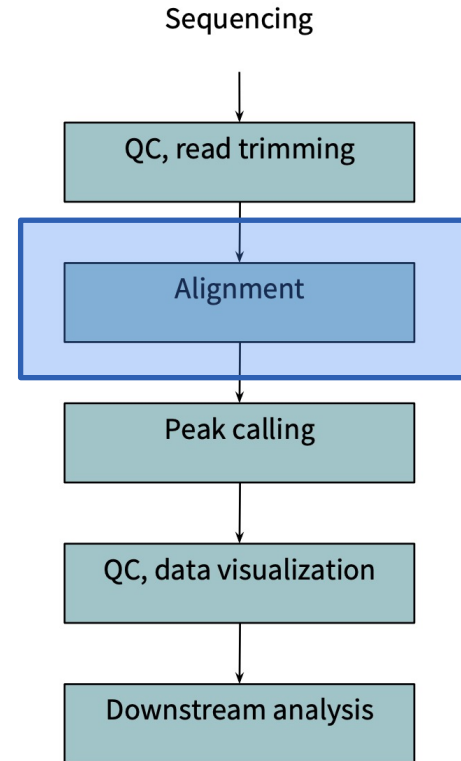


Computationally

Quality control – ChIP-seq data

Visually (Alignment)

- Relative Enrichment in genomic intervals (REGI)
 - Proteins might have a high expected enrichment in certain genomic regions



Quality control – ChIP-seq data

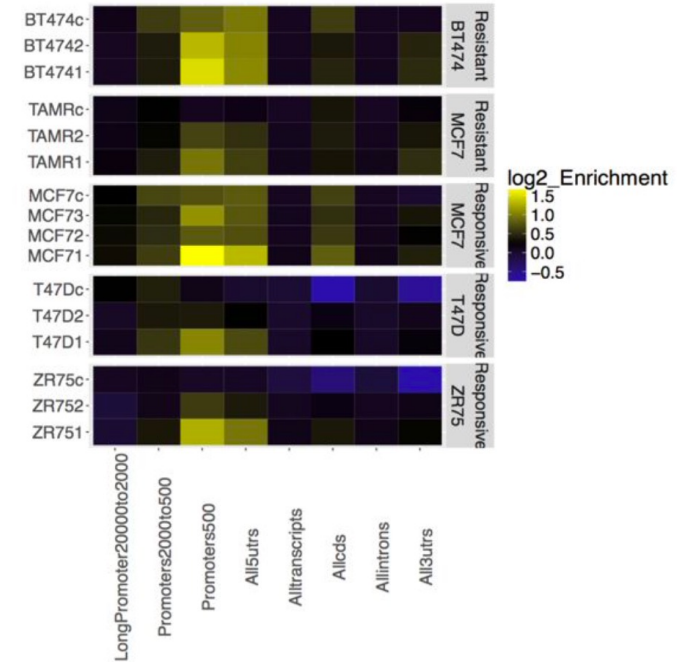
Visually (Alignment)

- Relative Enrichment in genomic intervals (REGI)
 - Proteins might have a high expected enrichment in certain genomic regions

Promoter region

UTRs

introns

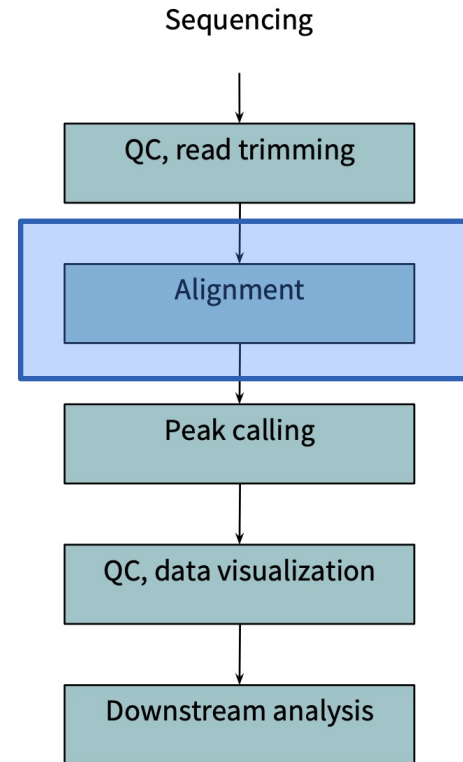


Quality control – ALL NGS data

Visually

Computationally

- Read Mapping% (**Higher the better**)

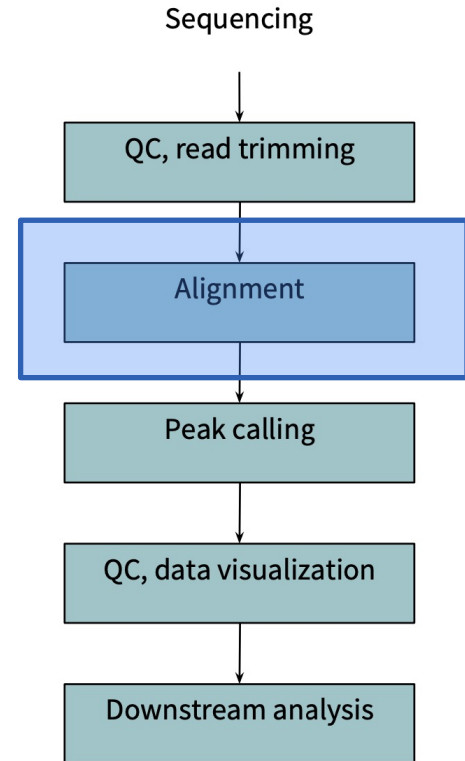


Quality control – ChIP-seq data

Visually

Computationally (Alignment)

- Remove Blacklisted regions
- Strand cross-correlation
- PCR Bottleneck coefficient (PBC)

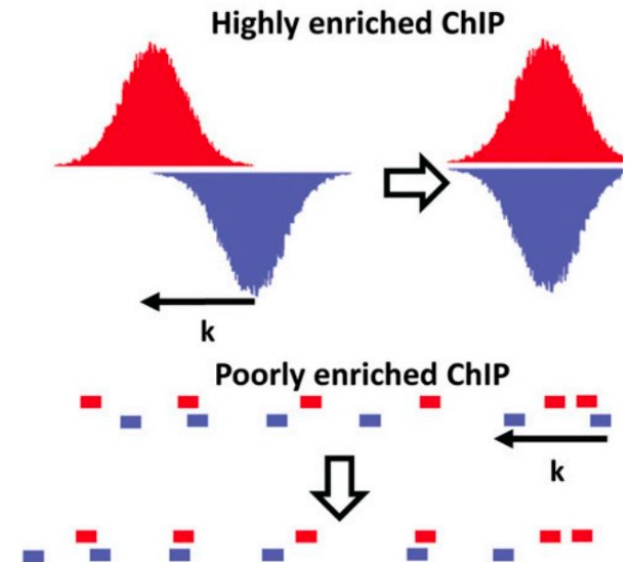


Quality control – ChIP-seq data

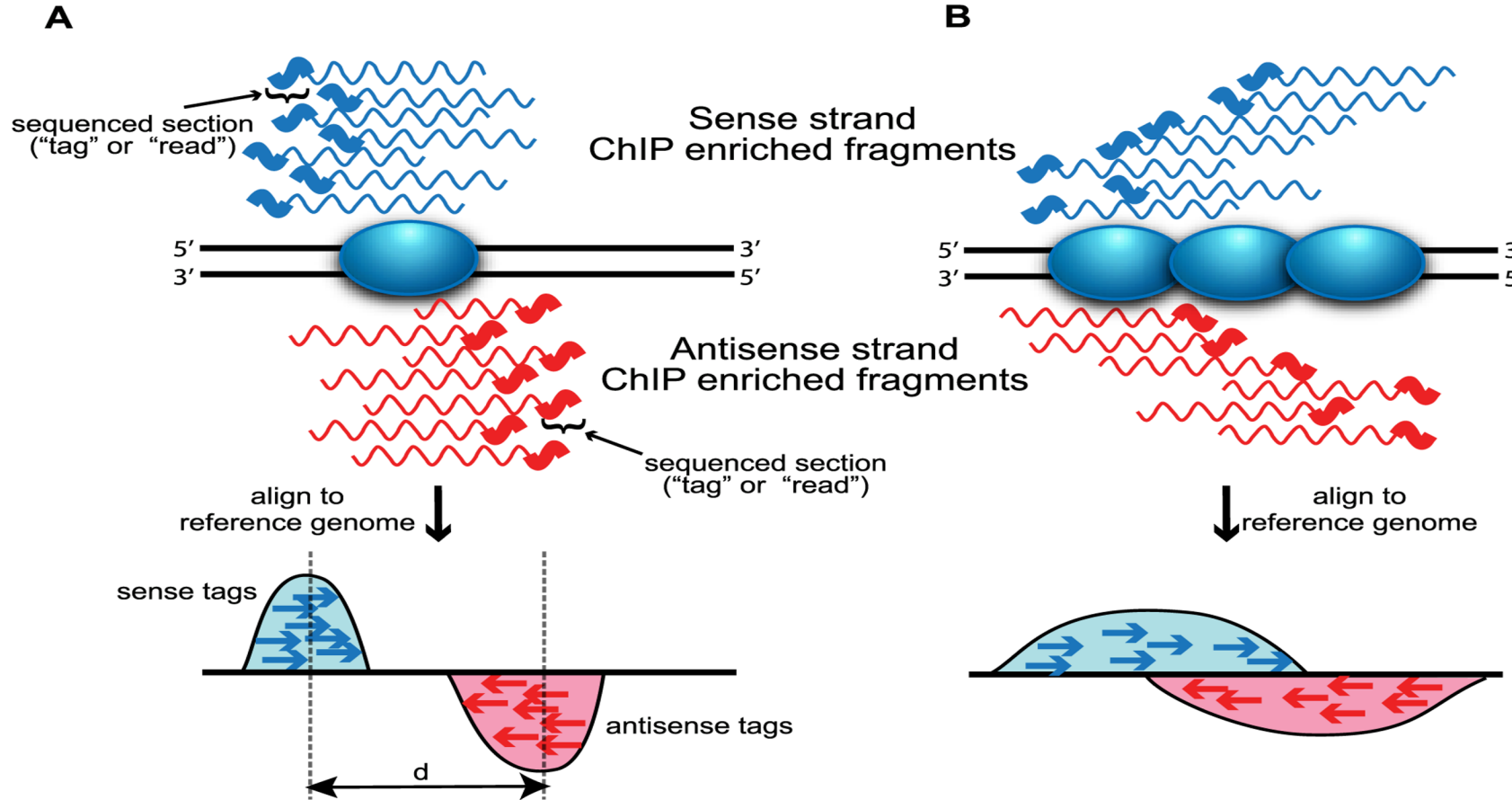
Visually

Computationally (Alignment)

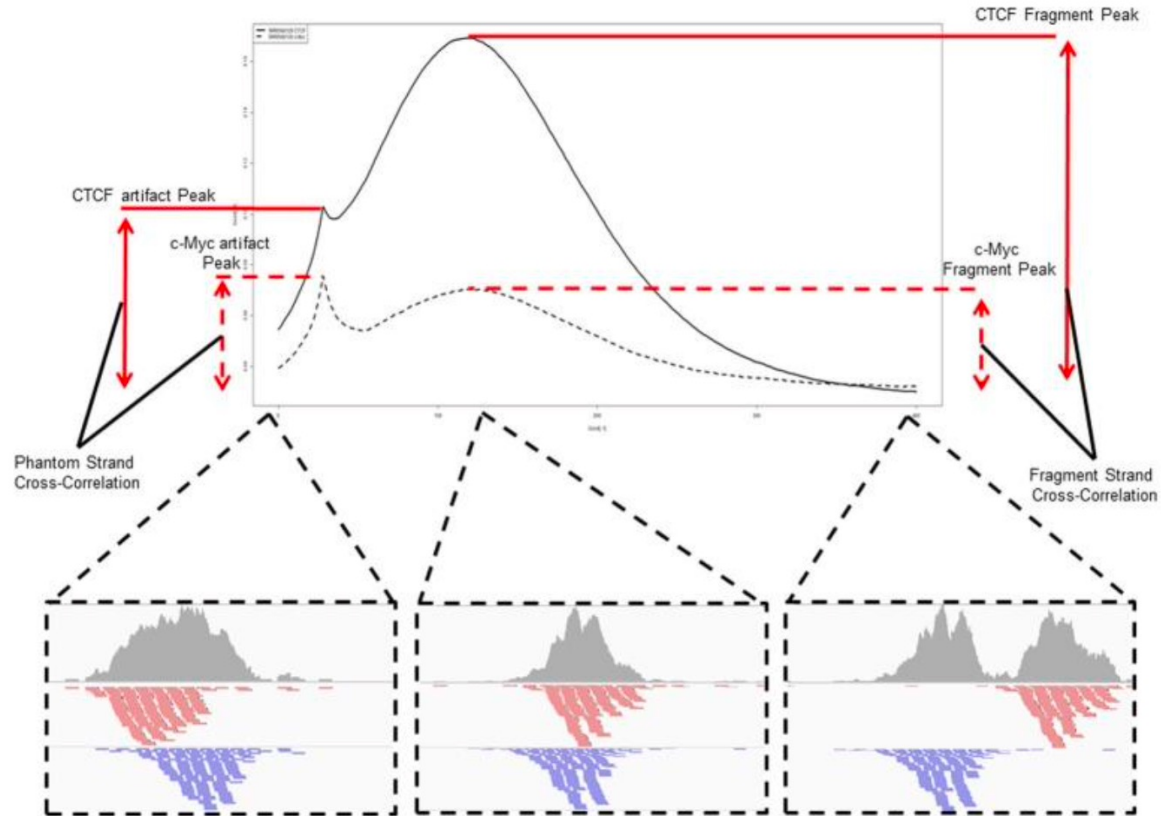
- Remove Blacklisted regions
- **Strand cross-correlation**
- PCR Bottleneck coefficient (PBC)



Strand dependent bimodality



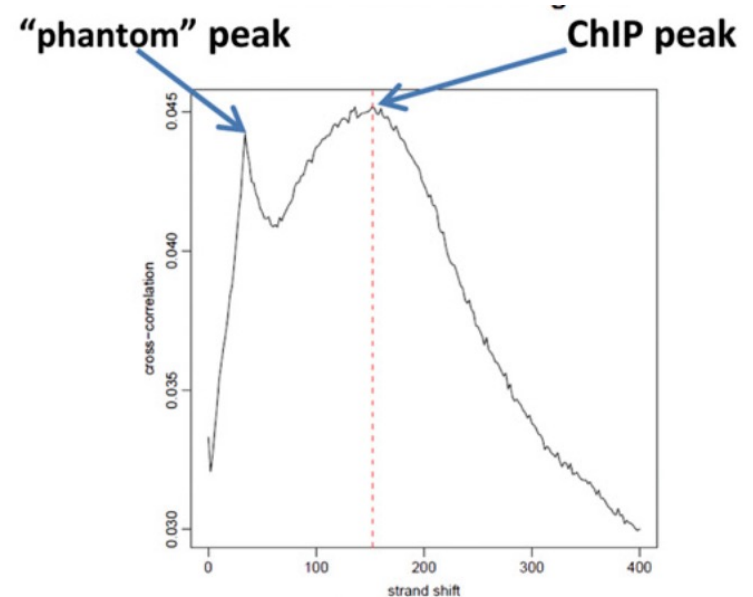
Strand cross-correlation



Strand cross-correlation plot

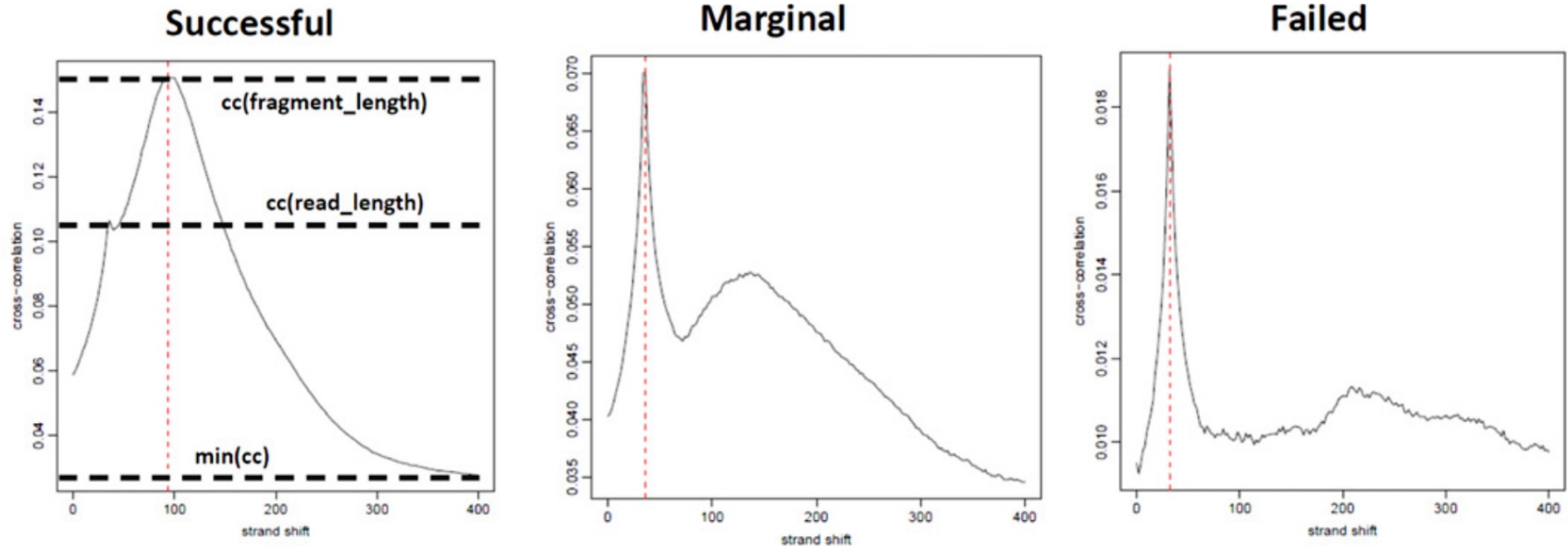
The cross-correlation plot typically produces two peaks:

- ChIP peak - fragment length
- Phantom peak - read length



Strand cross-correlation plot

G

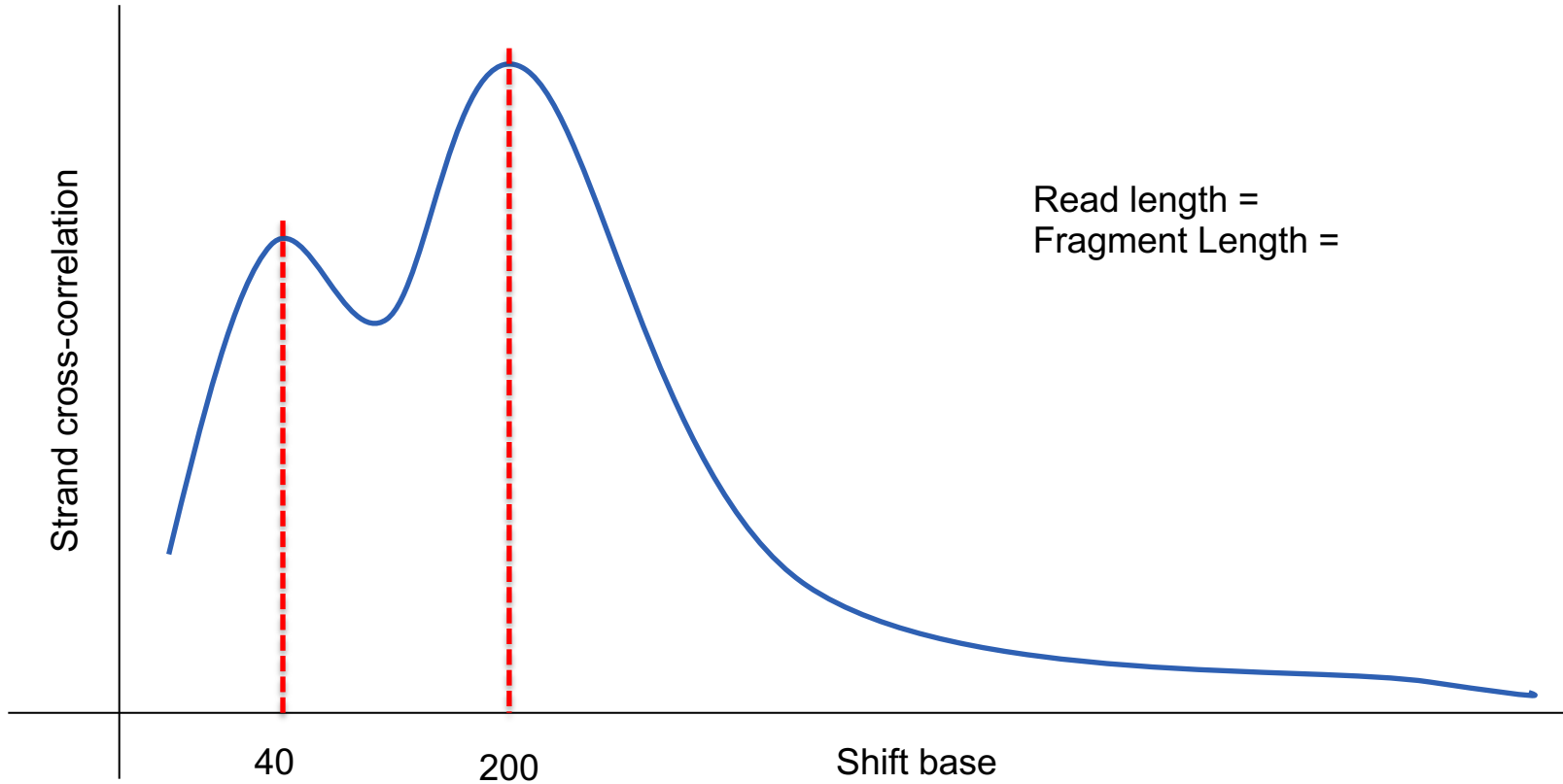


$$NSC = \frac{cc(fragment\ length)}{min(cc)}$$

$$RSC = \frac{cc(fragment\ length) - min(cc)}{cc(read\ length) - min(cc)}$$

Very successful ChIP experiments generally have $NSC > 1.05$ and $RSC > 0.8$

Strand cross-correlation plot



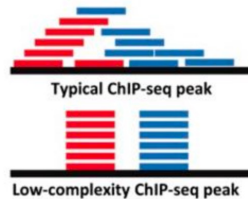
Quality control – ChIP-seq data

Visually

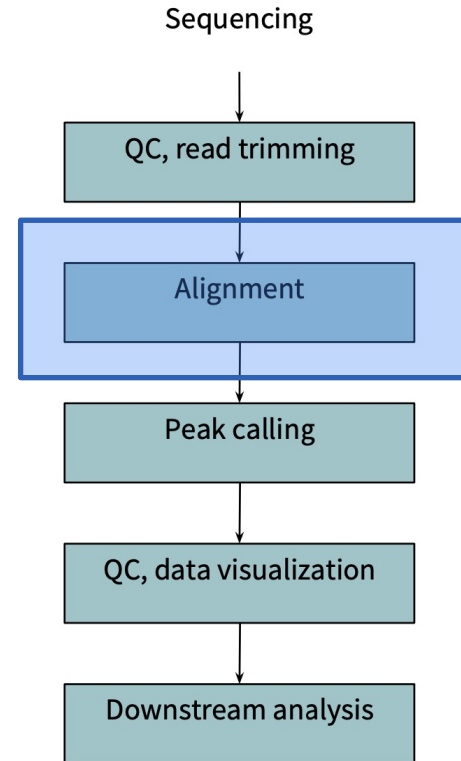
Computationally (Alignment)

- Remove Blacklisted regions
- Strand cross-correlation
- PCR Bottleneck coefficient (PBC)

$$PBC = \frac{N_1}{N_2}$$



N1: # genomic positions with one read aligned (**higher**)
N2: # genomic positions with one or more reads

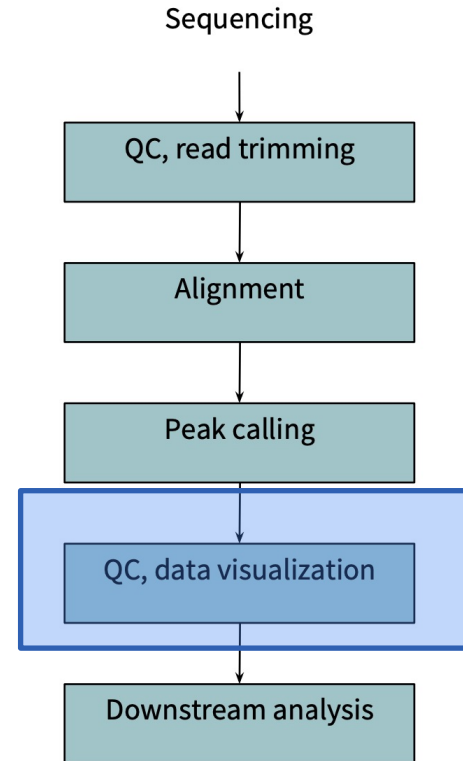


Quality control – ChIP-seq data

Visually

Computationally (peak)

- Num of peaks with good FDR(<0.05)

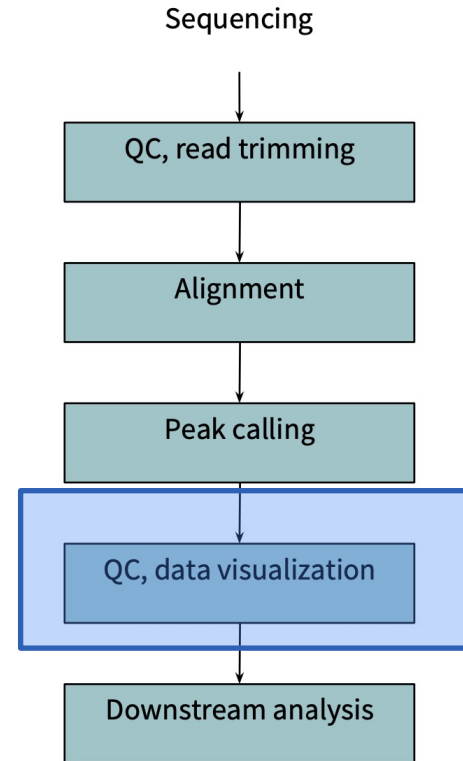


Quality control – ChIP-seq data

Visually

Computationally (peak)

- Num of peaks with good FDR(<0.05)
- **Fold Change**

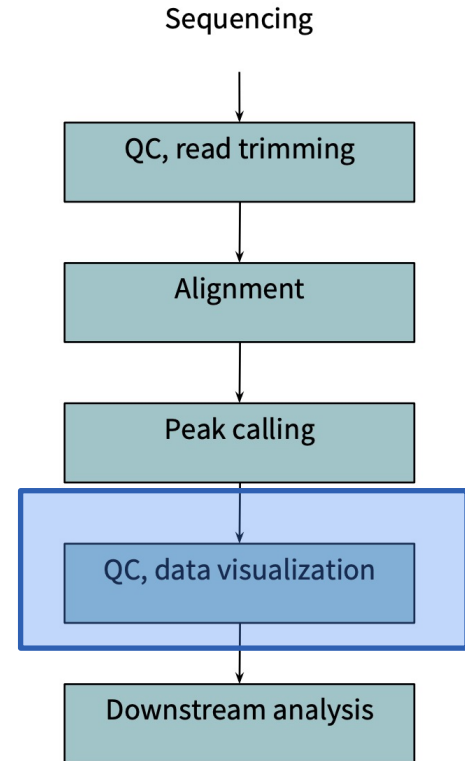
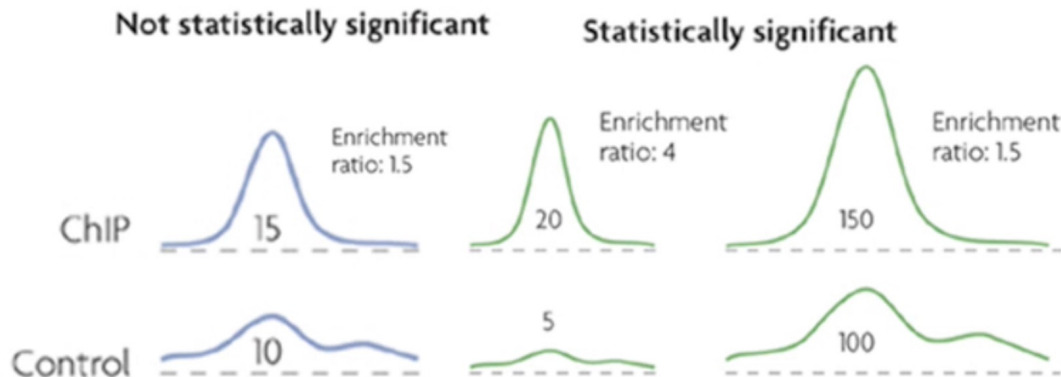


Quality control – ChIP-seq data

Visually

Computationally (peak)

- Num of peaks with good FDR(<0.05)
- **Fold Change (enrichment ratio)**



Quality control – ChIP-seq data

Visually

Computationally (peak)

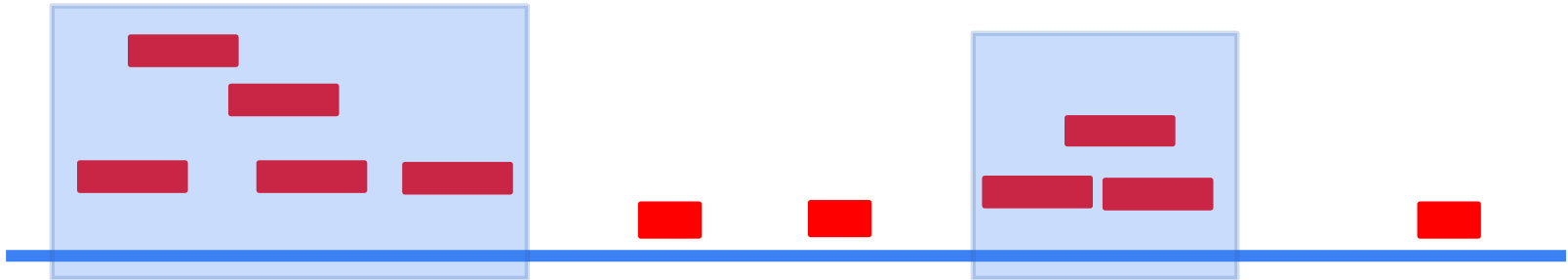
- Num of peaks with good FDR (0.05)
- Fold Change
- **Fraction of Reads in Peaks (FRiP>5%)**

Quality control – ChIP-seq data

Visually

Computationally (peak)

- Num of peaks with good FDR (0.05)
- Fold Change
- Fraction of Reads in Peaks (FRiP>5%)

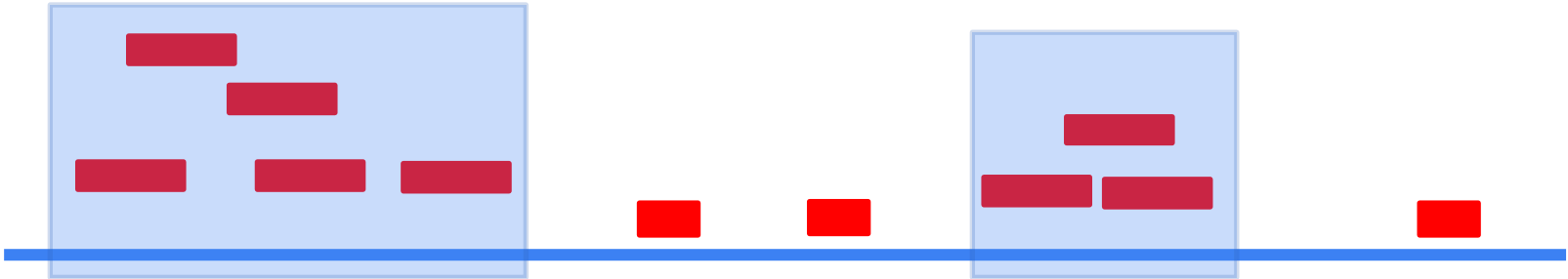


Quality control – ChIP-seq data

Visually

Computationally (peak)

- Num of peaks with good FDR (0.05)
- Fold Change
- Fraction of Reads in Peaks (FRiP>5%)
$$\text{FRiP} = \frac{\text{reads} \in \text{peaks}}{\text{total reads}}$$

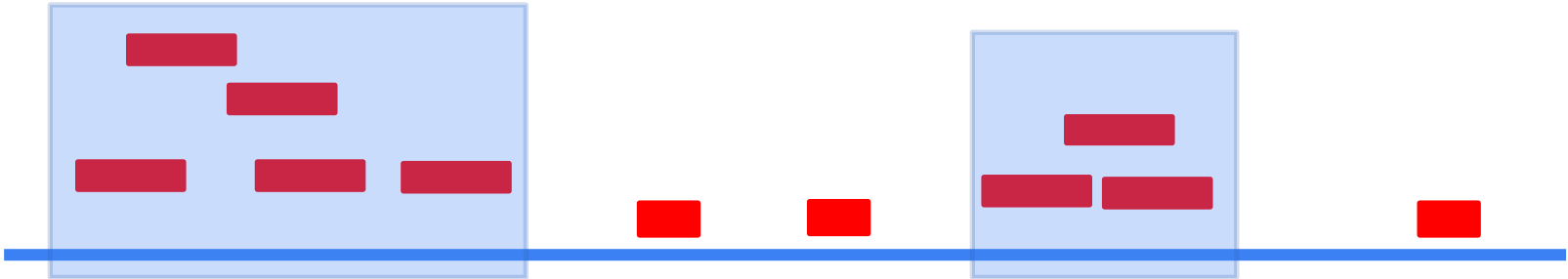


Quality control – ChIP-seq data

Visually

Computationally (peak)

- Num of peaks with good FDR (0.05)
- Fold Change
- Fraction of Reads in Peaks (FRiP > 5%)
$$\text{FRiP} = \frac{\text{reads} \in \text{peaks}}{\text{total reads}}$$



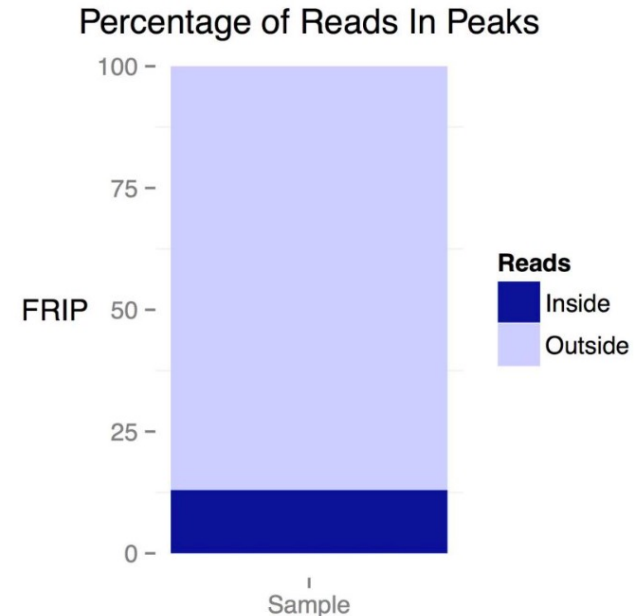
N.B. FRiP is sensitive to the specifics of peak calling method, antibody & target factor pair, so FRiP < 1% does not automatically mean failure

Quality control – ChIP-seq data

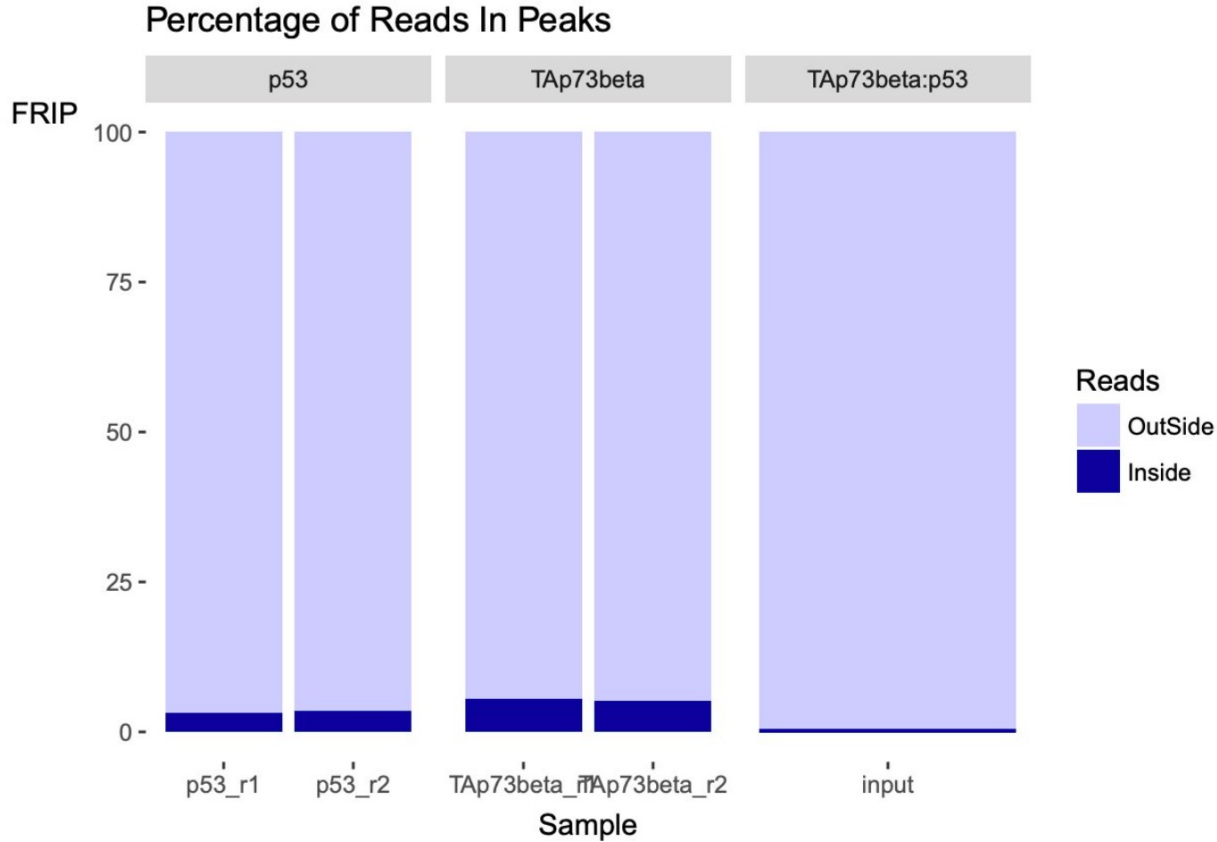
Visually

Computationally (peak)

- Num of peaks with good FDR (0.05)
- Fold Change
- **Fraction of Reads in Peaks (FRiP > 5%)**



What do you see here?



Adapted from Dora Bihary's slides

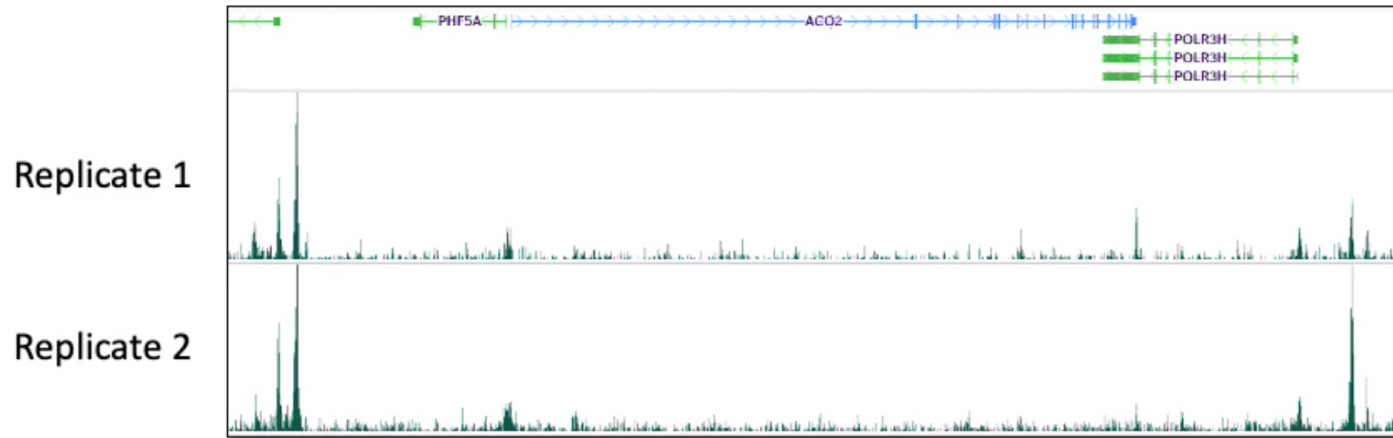
Quality control – ChIP-seq data

Visually

Computationally (peak)

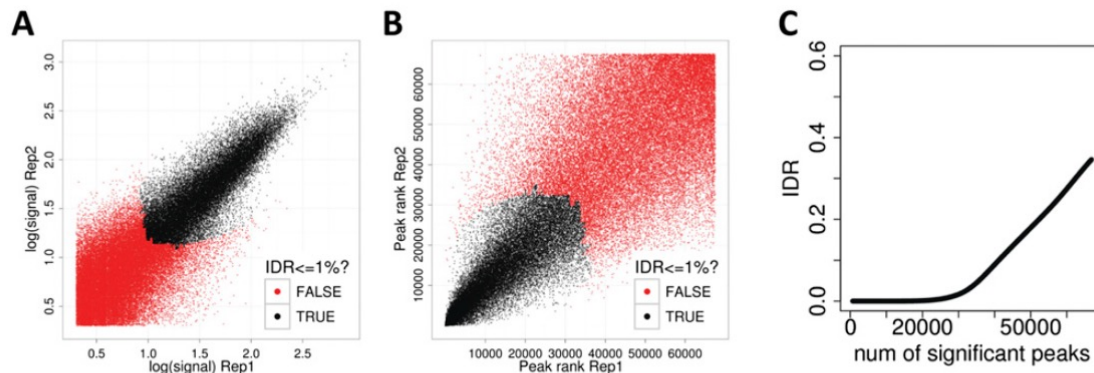
- Num of peaks with good FDR (0.05)
- Fold Change
- Fraction of Reads in Peaks (FRiP>5%)
- Irreproducible discovery rate (IDR – replicates)

Irreproducible discovery rate (IDR)

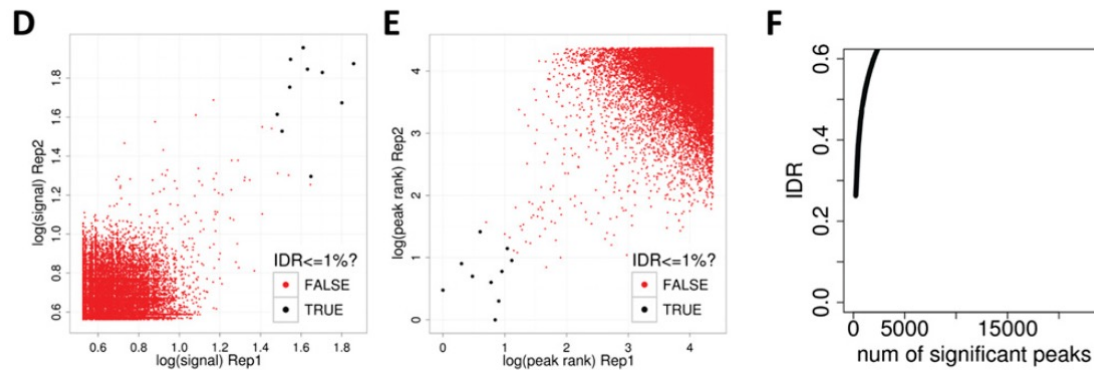


IDR

RAD21 Replicates (high reproducibility)



SPT20 Replicates (low reproducibility)



Quality control – ChIP-seq data

Visually

Computationally (peak)

- Num of peaks with good FDR (0.05)
- Fold Change
- Fraction of Reads in Peaks (FRiP>5%)
- Irreproducible discovery rate (IDR – replicates)
- **Standardised standard deviation (SSD)**

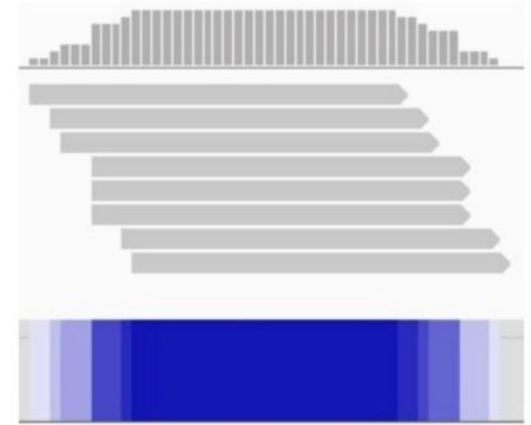
Dispersion of coverage

The depth of coverage:

- The number of fragments at a specific genomic region

Expectation:

The depth to have large diversity in an enriched ChIP dataset !



Depth	Base Pairs
1	3
2	4
3	3
5	3
6	4
7	3
8	26

Measure the dispersion of coverage

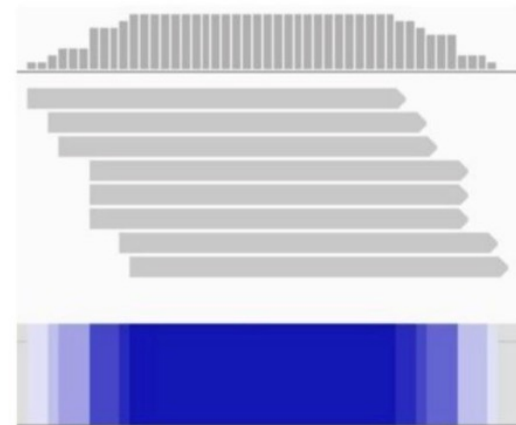
- Based on whole genome pile-up signal

$$SSD = \frac{SD}{\sqrt{n}}$$

An enriched sample: significant pile-up

SSD (higher the better)

- High for samples with enriched regions
- Low for controls with uniform coverage



Depth	Base Pairs
1	3
2	4
3	3
5	3
6	4
7	3
8	26

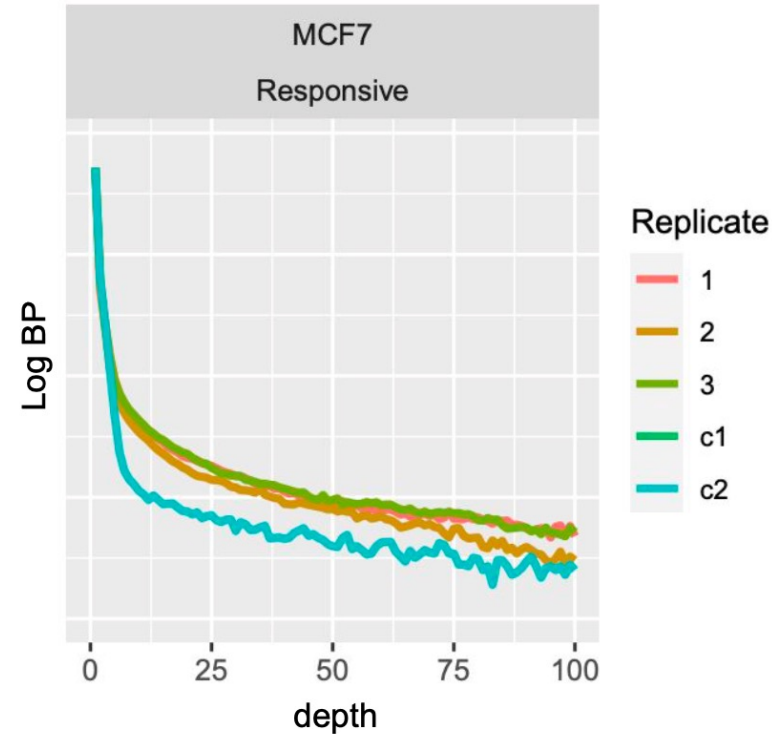
Visualisation

Coverage histogram

X: depth – read pileup height at a base pair position

Y: logBP – the number of positions that have this pileup height in log scale

- Good enrichment (1,2,3)
- Input (c1, c2)



Documentation from bioconductor ChIPQC
(<https://bioconductor.riken.jp/packages/3.4/bioc/html/ChIPQC.html>)
Carroll and Stark

Database

- <http://cistrome.org/db>

Batch	Species	Biological Source	Factor	Publication	Quality Control Library Complexity: PCR bottleneck coefficient (PBC)
<input type="checkbox"/>	Homo sapiens	HeLa; Epithelium; Cervix	BTAF1	Johannes F, et al. Bioinformatics 2010	
<input type="checkbox"/>	Homo sapiens	HeLa; Epithelium; Cervix	GAPDH	Johannes F, et al. Bioinformatics 2010	
<input type="checkbox"/>	Homo sapiens	K562; Erythroblast; Bone Marrow	EGR1	Tang C, et al. Electrophoresis 2010	
<input type="checkbox"/>	Homo sapiens	LS174T; Epithelium; Colon	TCF4	Mokry M, et al. PLoS ONE 2010	
<input type="checkbox"/>	Homo sapiens	LS174T; Epithelium; Colon	TCF4	Mokry M, et al. PLoS ONE 2010	
<input type="checkbox"/>	Homo sapiens	LS174T; Epithelium; Colon	TCF4	Mokry M, et al. PLoS ONE 2010	
<input type="checkbox"/>	Homo sapiens	LS174T; Epithelium; Colon	TCF4	Mokry M, et al. PLoS ONE 2010	
<input type="checkbox"/>	Homo sapiens	LS174T; Epithelium; Colon	TCF4	Mokry M, et al. PLoS ONE 2010	
<input type="checkbox"/>	Homo sapiens	BJ; Fibroblast; Skin	TERF1	Simonet T, et al. Cell Res. 2011	
<input type="checkbox"/>	Homo sapiens	BJ; Fibroblast; Skin	TERF2	Simonet T, et al. Cell Res. 2011	
<input type="checkbox"/>	Homo sapiens	22RV1; Epithelium; Prostate	AR	Yu J, et al. Cancer Cell 2010	
<input type="checkbox"/>	Homo sapiens	HEK293T; Epithelium; Embryonic Kidney	PHF8	Fortschegger K, et al. Mol. Cell. Biol. 2010	
<input type="checkbox"/>	Homo sapiens	aTconv; T Lymphocyte; Blood	H3K4me1	Tian Y, et al. PLoS ONE 2011	
<input type="checkbox"/>	Homo sapiens	aTconv; T Lymphocyte; Blood	H3K4me3	Tian Y, et al. PLoS ONE 2011	
<input type="checkbox"/>	Homo sapiens	BG01; Embryonic Stem Cell; Embryo	H3K27me3	Guenther MG, et al. Cell Stem Cell 2010	

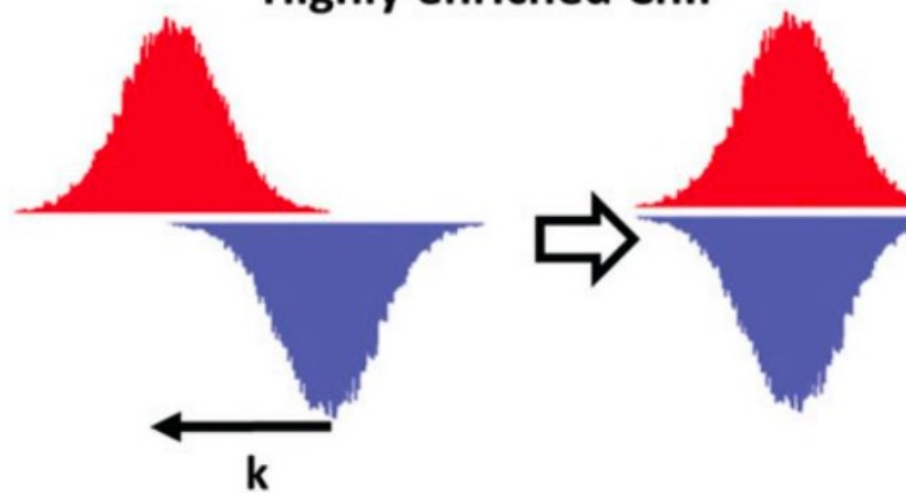
Lunch break before practical

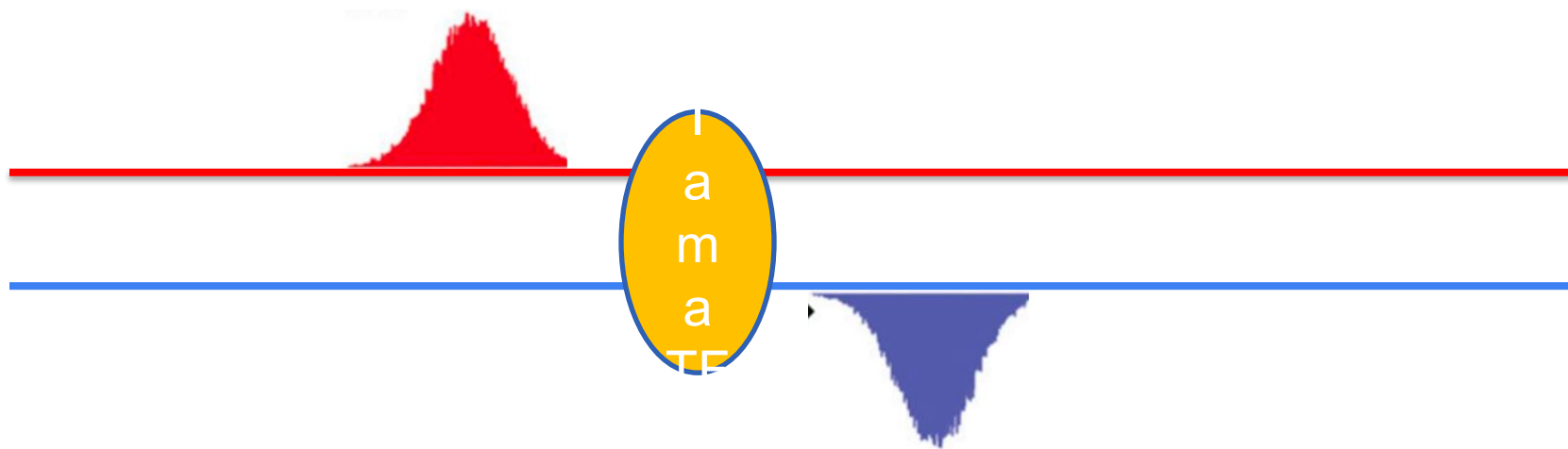
Back at 13.30

Supplementary

- Strand cross-correlation

Highly enriched ChIP

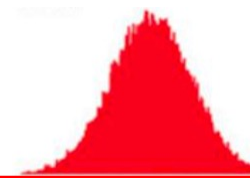






[1,2,3,5,7,8,7,5,3,2,1]





[1,2,3,5,7,8,7,5,3,2,1]

[1,2,3,5,7,8,7,5,3,2,1]





Red block 1

Red block 2

Red block 3

Red block 4

Red block 5



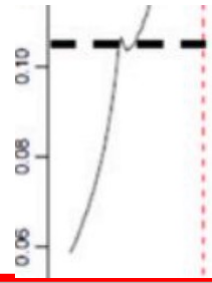
Blue block 1

Blue block 2

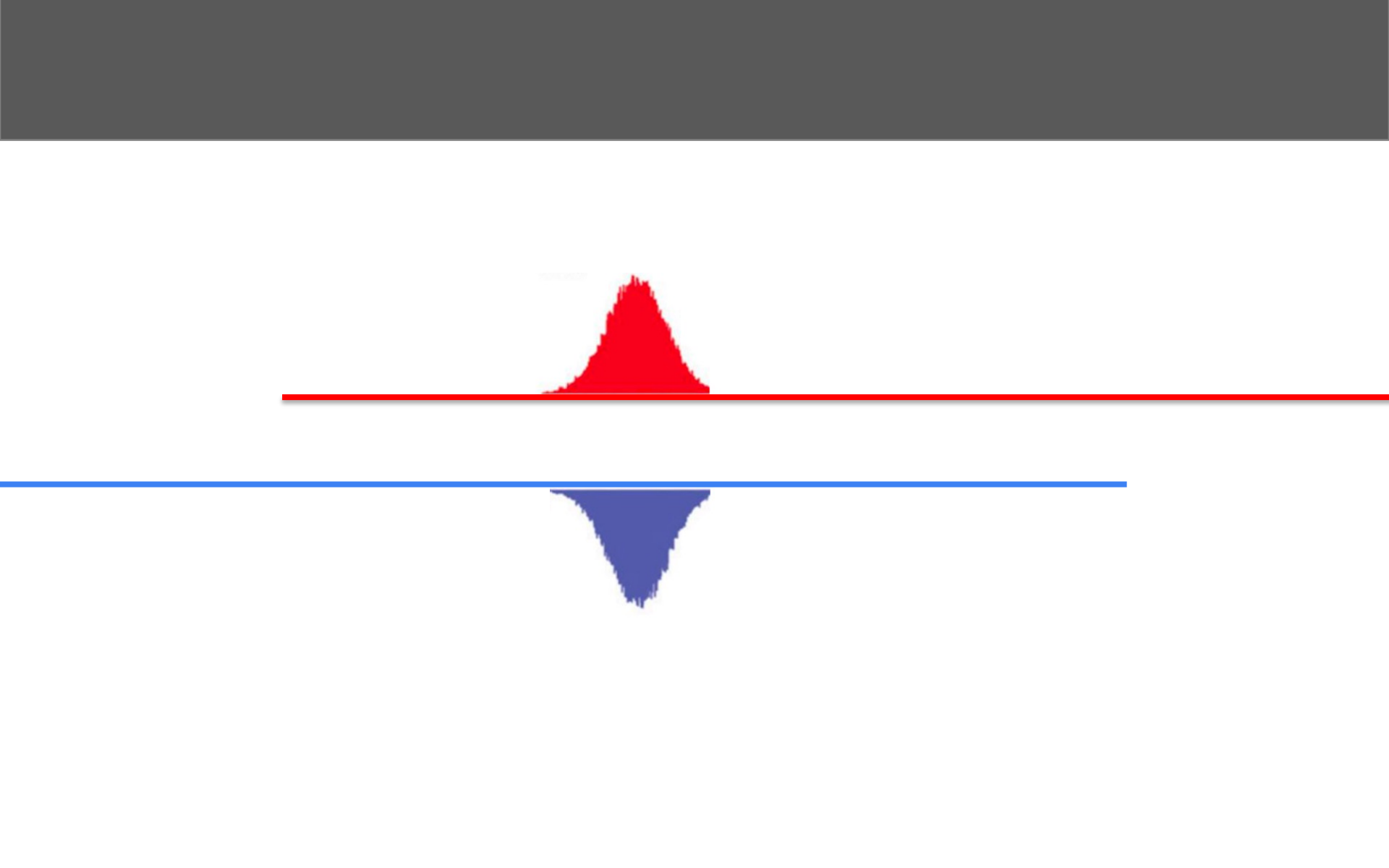
Blue block 3

Blue block 4

Blue block 5







- <https://www.youtube.com/watch?v=XWcWn8dt4c8>