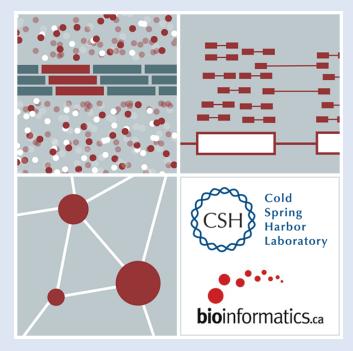
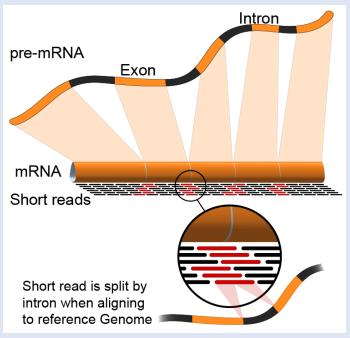


# Introduction to single cell RNA sequencing and analysis

Kelsy Cotto, Malachi Griffith, Obi Griffith, Evelyn Schmidt, Kartik Singhal, Zach Skidmore CRI Bioinformatics Workshop. May 17-22, 2025







## Acknowledgements

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- Jennifer Foltz, Washington University, Genomics in Medicine
- Trevor Pugh, PMH, OICR, CBW
- Allegra Petti, MGH/Harvard
- Brian Haas, Broad Institute







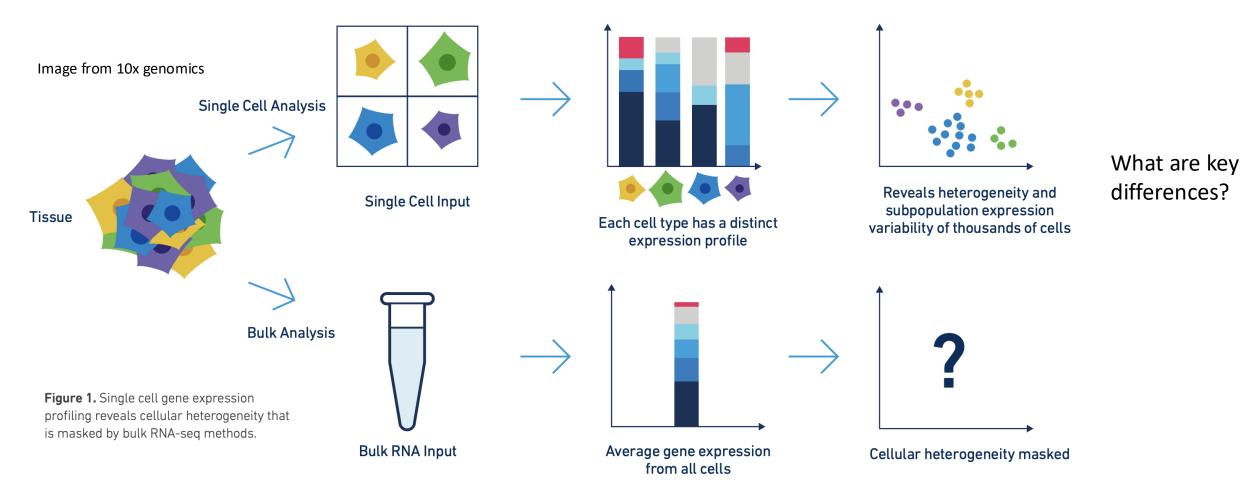




# Single-cell omics - A new paradigm in biology and medicine?

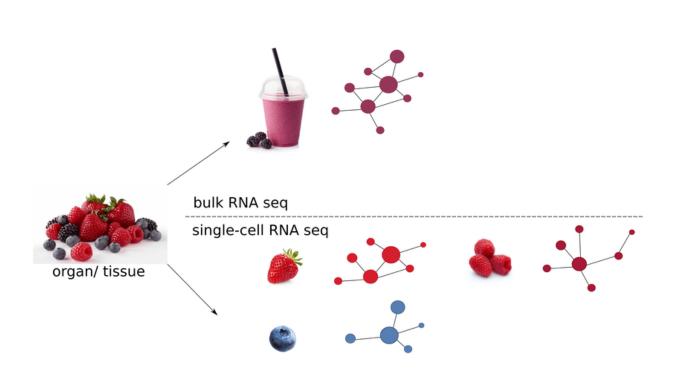
- Redefine "cell type"
  - Catalog known cell types in healthy and diseased tissues
  - Define/discover new cell types
- Redefine relationships between cells
- Interpret genetic variation at single-cell resolution
- And much more...

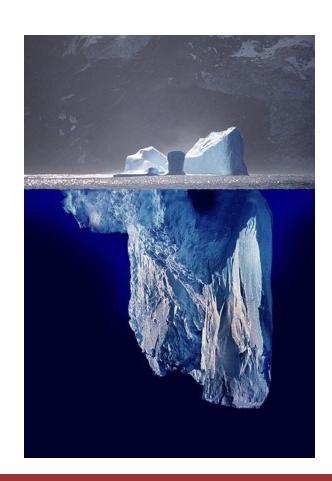
# Discussion of bulk vs single cell RNA-seq



Factors to compare: Cost, complexity of library prep, complexity of analysis, qualitative and quantitative differences in richness of information obtained.

## Fruit salad or tip of the iceberg?

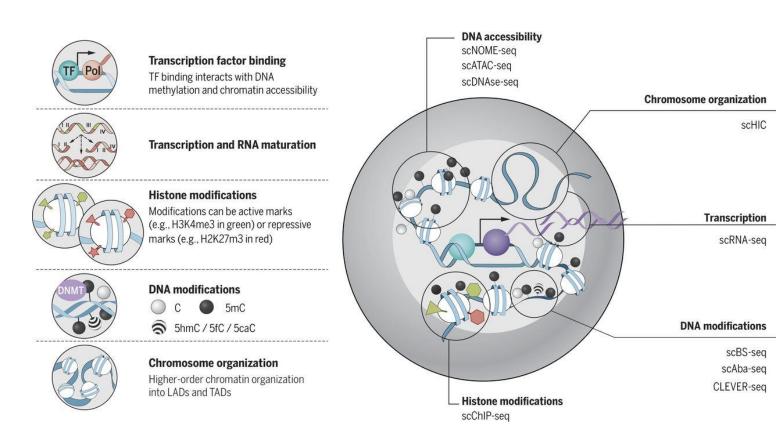




scRNA

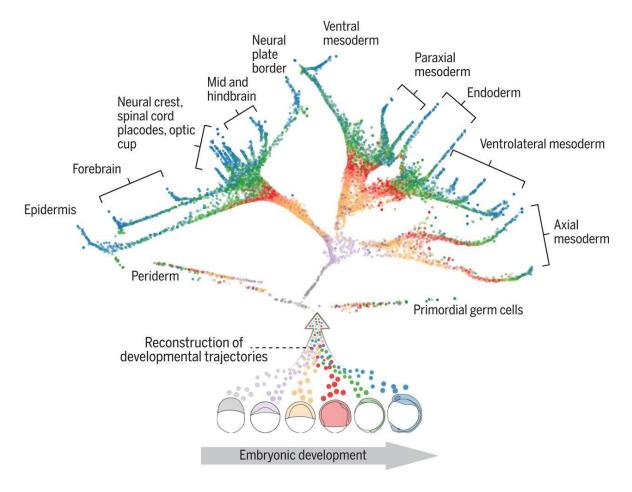
## Multimodal single-cell measurements

- scRNA/snRNA
- CITE-seq
- TCR sequencing
- scATAC-seq
- scMethyl-seq
- TARGET-seq, G&T-seq (scDNA & scRNA)
- scCRISPRi/Perturb-seq
- Patch-seq
- Spatial transcriptomics



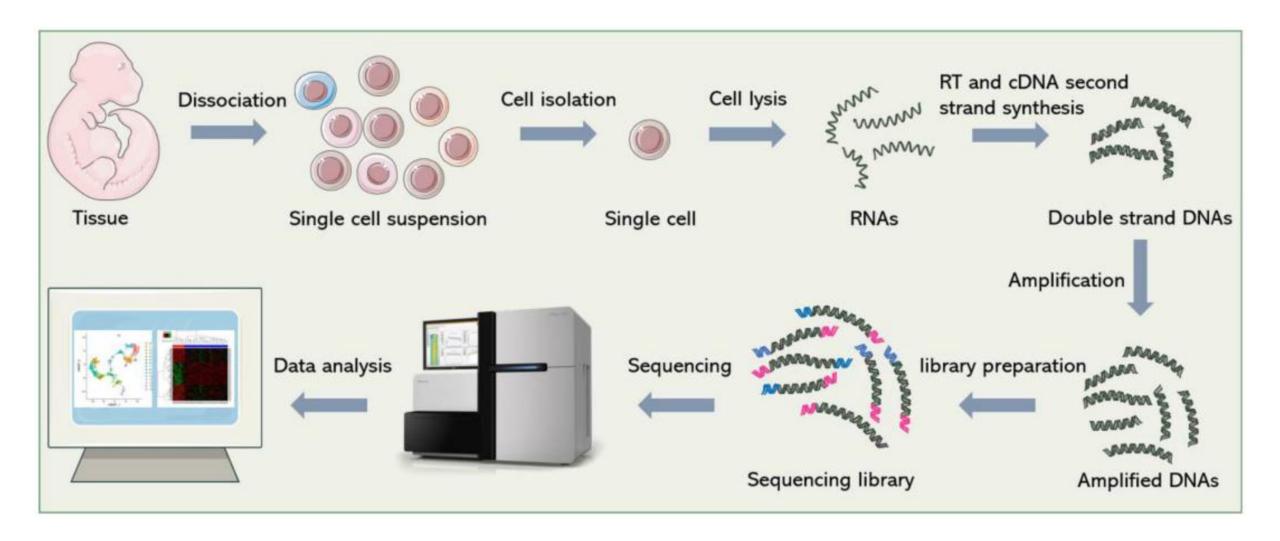
Kelsey et al. Science 2017

## Cell type vs cell lineage vs cell state

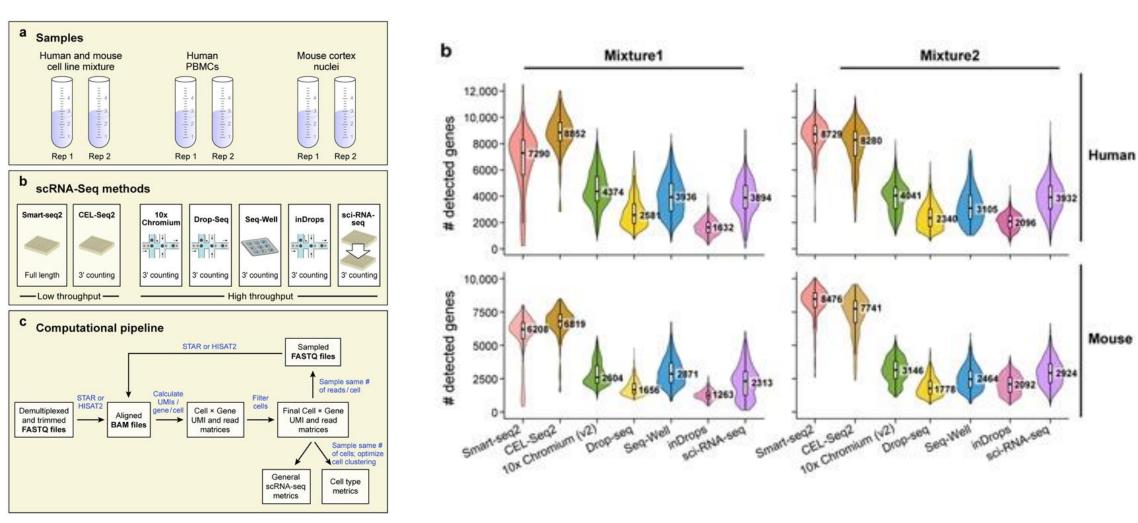


https://www.science.org/doi/10.1126/science.aar3131

# Basic overview of single cell RNA analysis



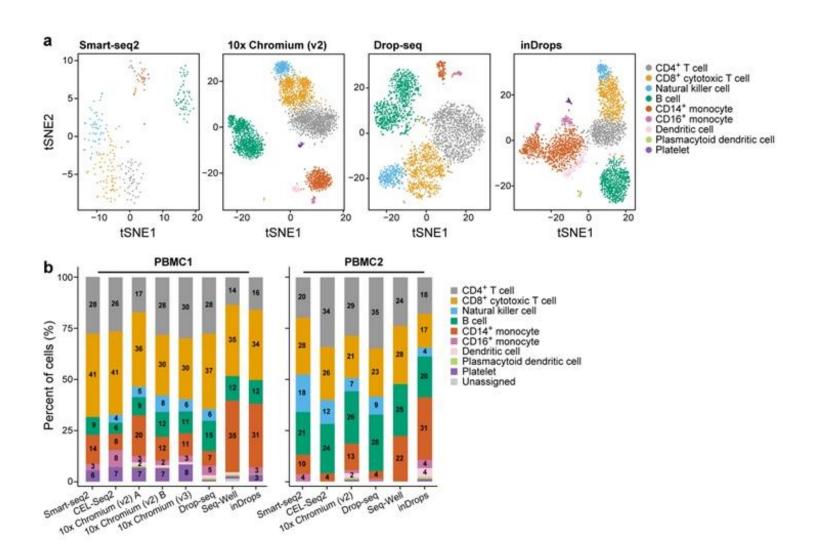
#### Multiple platforms have emerged ... Each with trade-offs...



PMC7289686

Plate-based vs Droplet-based

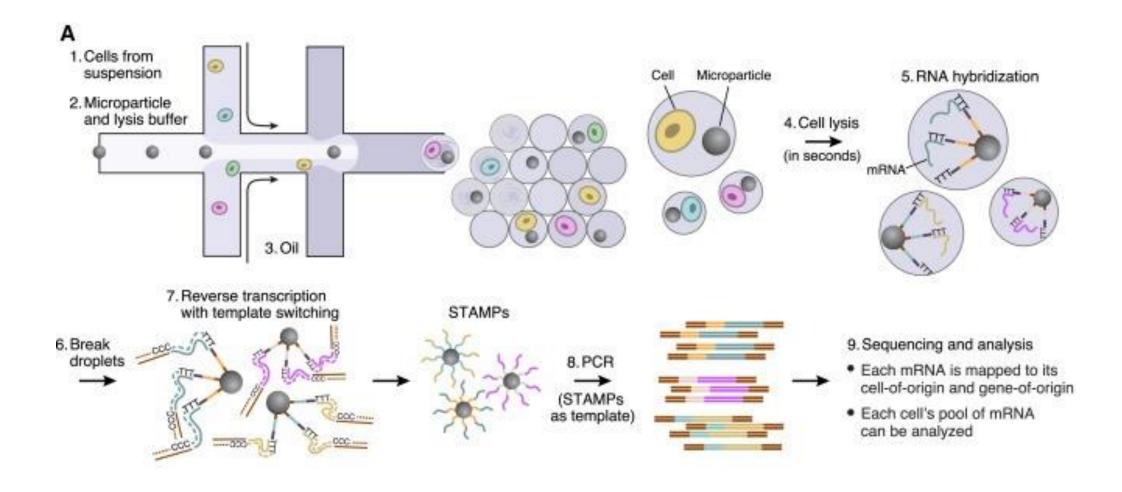
### Multiple platforms have emerged ... Each with trade-offs...



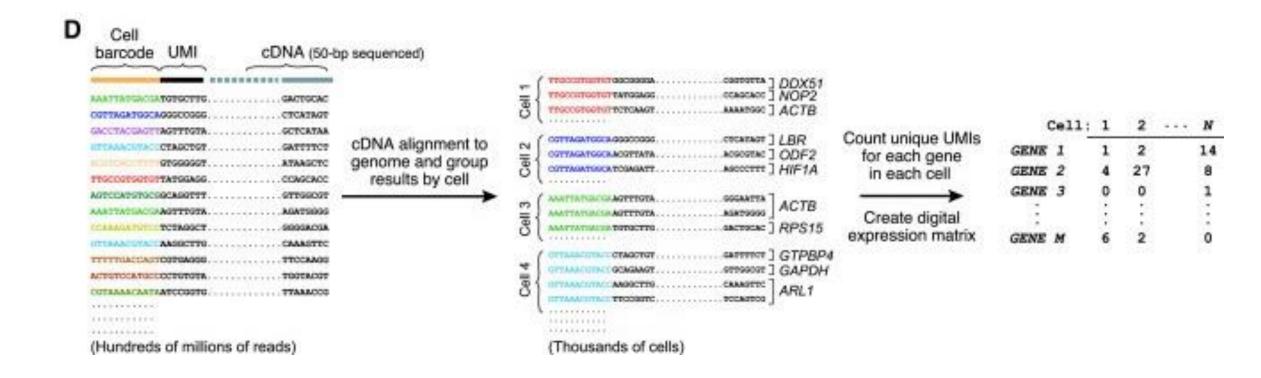
#### Additional considerations

- Size of cells
- Expected % of cell population of interest
- Depth of detection needed for experimental question
- Accessibility

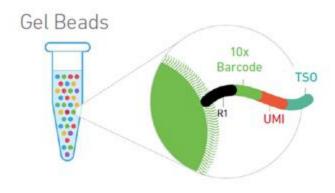
## Intro to 10X approach

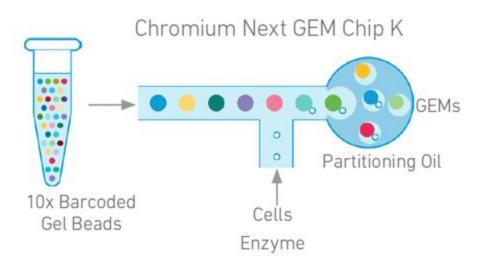


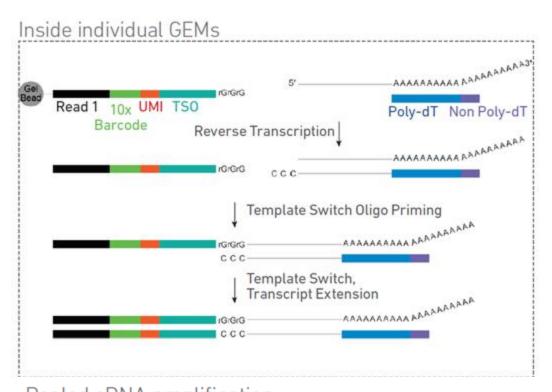
## Intro to 10X approach

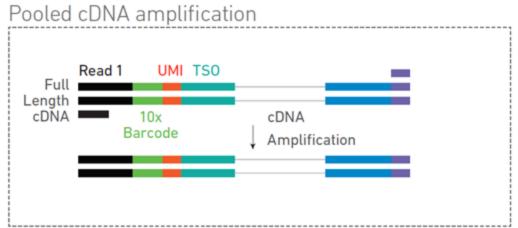


#### 10X Approach (5')



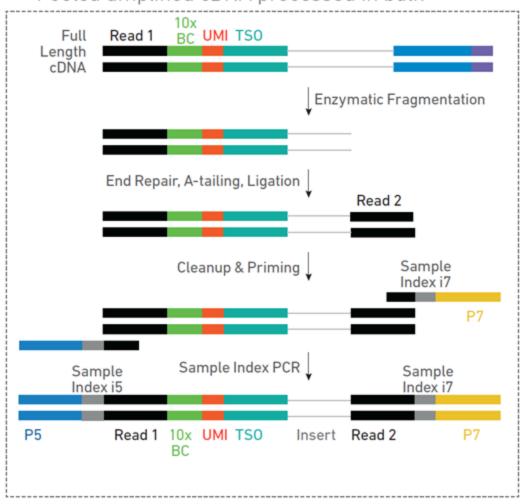


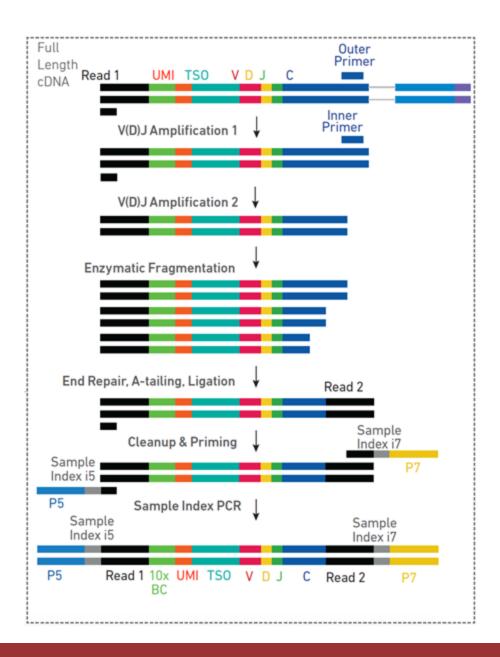




#### **10X Approach (5' v2)**

#### Pooled amplified cDNA processed in bulk



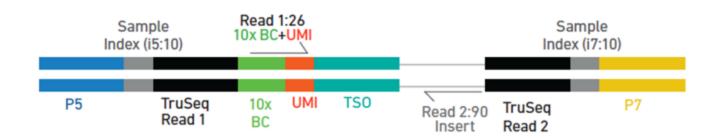


### 10X Single Cell 5' GEX and V(D)J dual index libraries



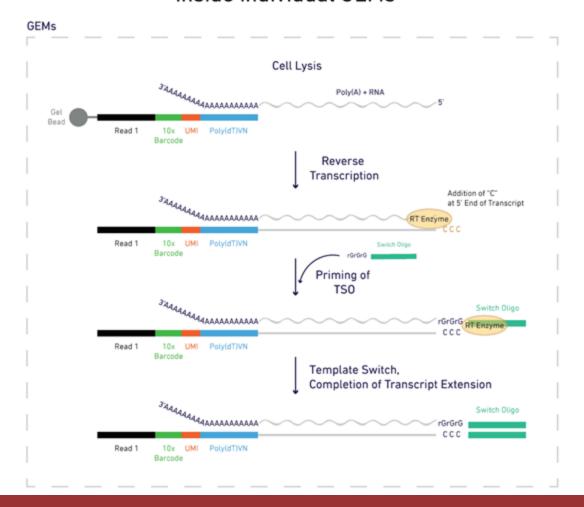


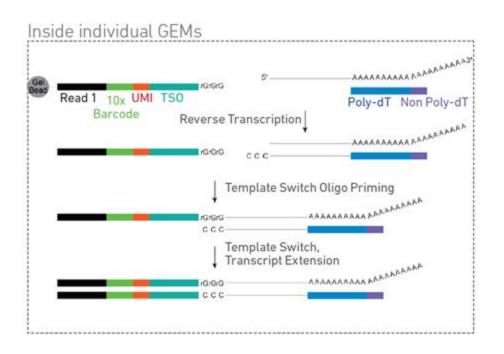
#### Chromium Single Cell 5' Gene Expression Dual Index Library



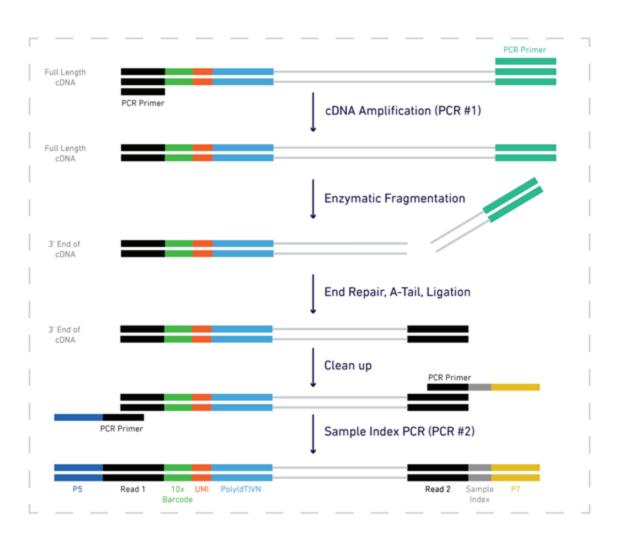
# 3' vs 5' approach - polyT and TSO switch places

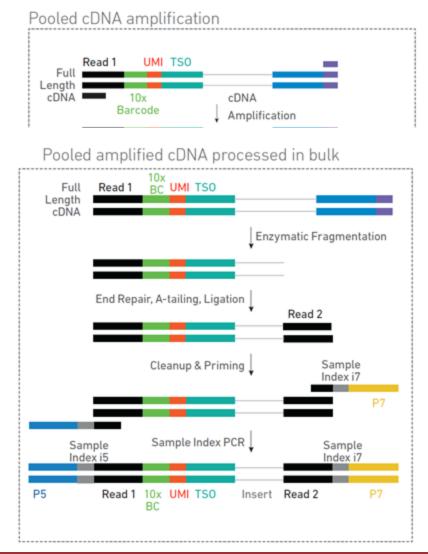
#### Inside individual GEMs



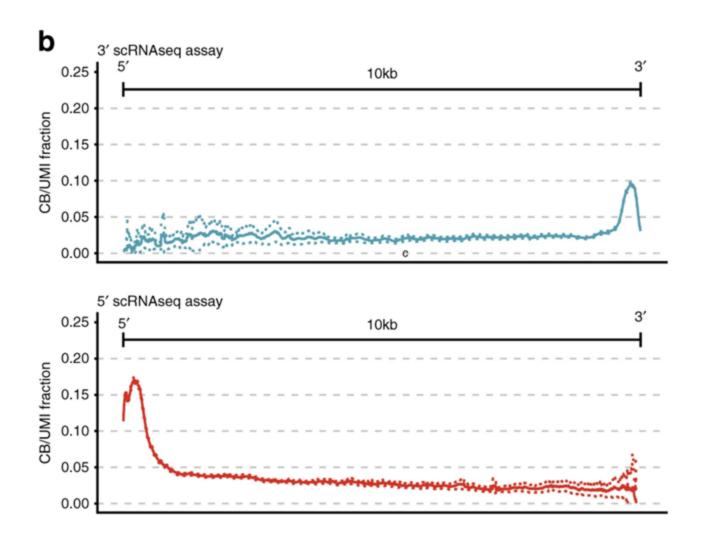


# 3' vs 5' approach - Fragmentation and PCR steps result in different ends of transcripts being kept





### 3' vs 5' approach results in different end bias



PMC6694122

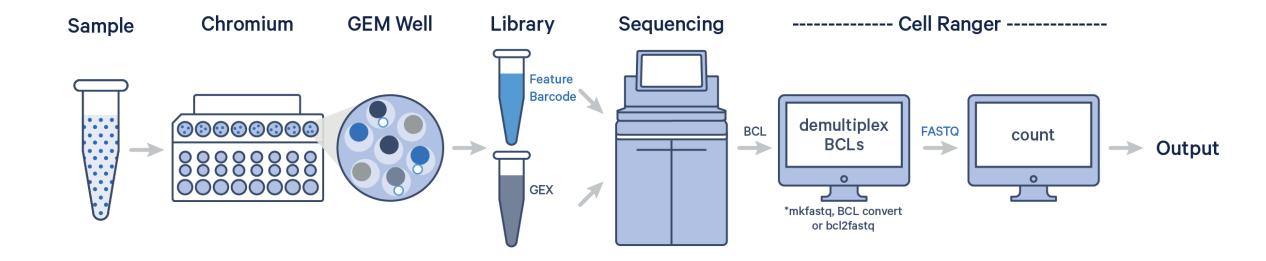
Fig1b. Petti et al. 2019. Nat Comm

### 3' vs 5' approach

- Both require polyA transcripts
- Both involve end bias (choose 3' or 5')
  - 5' coverage more evenly distributed?
- Only 5' approach is compatible with V(D)J enrichment
  - Because business end of V(D)J transcript is at 5' followed by large C region at 3' end
  - In order to amplify TCR from 3' 10X library you would need very large fragments and would have to design primers off the variable V genes instead of constant C gene

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### Overview of 10x genomics pipeline



### How deeply do you need to sequence?

General Rule: Achieve 90% saturation

#### Official Recommendations (reads/cell):

3' - V3: 20K

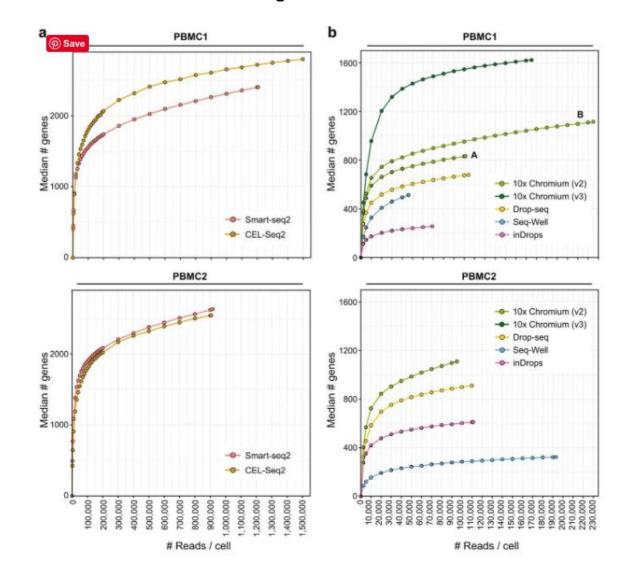
3' - V2: 50K

5' - 20K

5' with variant discovery - 200K

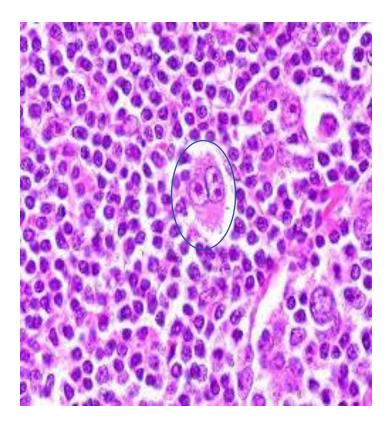
5' V(D)J - 5K

Fixed RNA: 10K



# Example application: Increased capacity and new technologies are allowing study of previously inaccessible tumor types

• Hodgkins Lymphoma - has remained a challenge because HRS cells account for  $^{\sim}1-5\%$  of cells in the tumor tissue.



- Isolate HRS cells and then apply genomic techniques
  - Flow sorting very challenging cell type
  - Laser capture microdissection extremely low inputs for sequencing
- Handful of cell lines have been profiled
- Almost no genome-wide sequencing data exists

Todd Fehniger



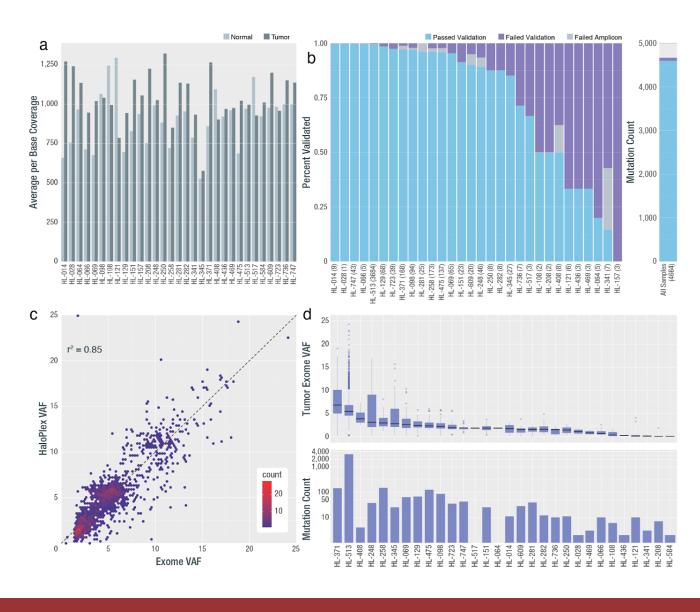
Felicia Gomez



# Brute force strategy - Ultra Deep Exome sequencing of 31 Hodgkins tumor/normal pairs

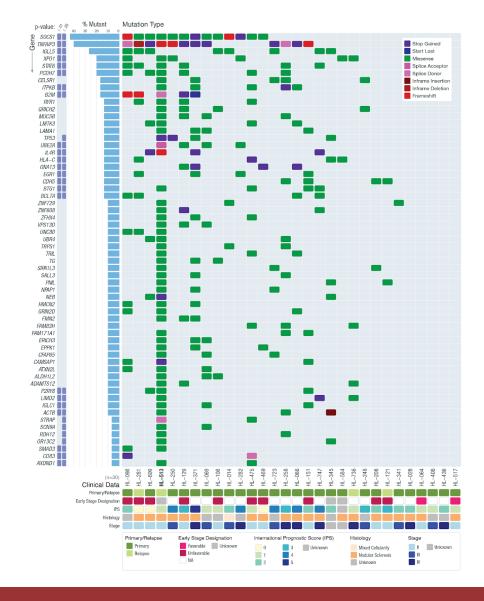
- IDT Exome capture reagent
- Three KAPA libraries were constructed/sample
- Libraries were sequenced across eight lanes of an Illumina HiSeq
- Somatic Variant Calling
  - SNV were called using 5 variant callers
  - Indels were called using 4 variant callers
- Target depth ~1000x
- Validation with targeted Haloplex technology

### High coverage and concordance achieved



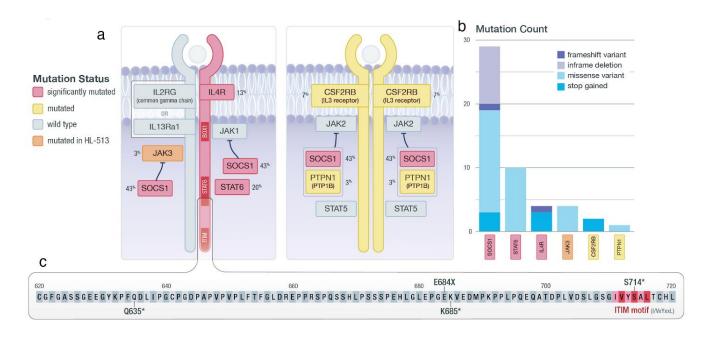
- ~1000X coverage achieved
- High VAF concordance between WES and haloplex validation
- Overall variant validation rate >90%
- Mean mutation burden = 33 protein-coding variants
  - One hypermutator 3,160 variants

# Represents one of the few comprehensive surveys of HL - mutation landscape reveals known and novel genes/pathways/hotspots

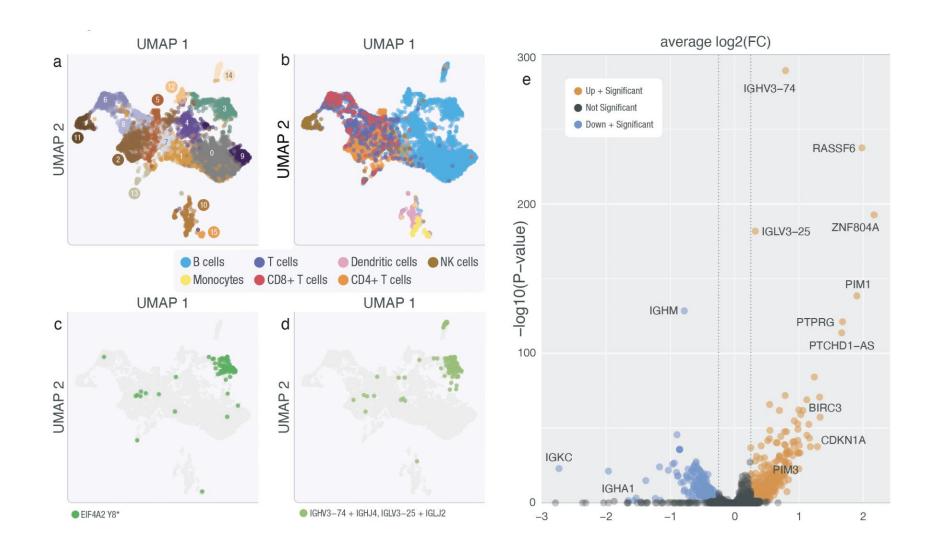


- ~50 genes have mutations in 3 or more samples
- JAK/STAT SOCS1 & STAT6
- NFKB TNFAIP3 & XPO1
- SWI/SNF BCL7A & SMAD3
- *PCDH7, IGLL5, CDH5, ...*

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#### Single nucleus sequencing identifies mutant-positive HRS cells







RESEARCH ARTICLE

https://doi.org/10.1158/2767-9764.CRC-23-0140

**OPEN ACCESS** 

#### Ultra-Deep Sequencing Reveals the Mutational Landscape of Classical Hodgkin Lymphoma

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Felicia Gomez<sup>1,2,3</sup>, Bryan Fisk<sup>1,2</sup>, Joshua F. McMichael<sup>2</sup>, Matthew Mosior<sup>1,2</sup>, Jennifer A. Foltz<sup>1</sup>, Zachary L. Skidmore<sup>1,2</sup>, Eric J. Duncavage<sup>4</sup>, Christopher A. Miller<sup>1,2</sup>, Haley Abel<sup>1,2</sup>, Yi-Shan Li<sup>4</sup>, David A. Russler-Germain<sup>1</sup>, Kilannin Krysiak<sup>1,2,3,4</sup>, Marcus P. Watkins<sup>1</sup>, Cody A. Ramirez<sup>1,2</sup>, Alina Schmidt<sup>1,2</sup>, Fernanda Martins Rodrigues<sup>1,2</sup>, Lee Trani<sup>2</sup>, Ajay Khanna<sup>1</sup>, Julia A. Wagner<sup>1</sup>, Robert S. Fulton<sup>2</sup>, Catrina C. Fronick<sup>2</sup>, Michelle D. O'Laughlin<sup>2</sup>, Timothy Schappe<sup>1</sup>, Amanda F. Cashen<sup>1</sup>, Neha Mehta-Shah<sup>1</sup>, Brad S. Kahl<sup>1</sup>, Jason Walker<sup>2</sup>, Nancy L. Bartlett<sup>1</sup>, Malachi Griffith<sup>1,2,3,5</sup>, Todd A. Fehniger<sup>1,3</sup>, and Obi L. Griffith<sup>1,2,3,5</sup>

# Introduction to scRNA dataset for hands-on exercises

https://rnabio.org/module-08-scrna/0008/01/02/scRNA\_Data/

Includes brief demo of CellRanger QC Reports and Loupe Browser