

#### Canadian Bioinformatics Workshops

www.bioinformatics.ca

bioinformaticsdotca.github.io



#### Attribution-ShareAlike 4.0 International

Canonical URL: https://creativecommons.org/licenses/by-sa/4.0/

See the legal code

#### You are free to:

 $\label{eq:Share-copy} \textbf{Share} - \textbf{copy} \ \text{and} \ \textbf{redistribute} \ \textbf{the material in any medium} \ \textbf{or format for any} \\ \textbf{purpose, even commercially.}$ 

 $\label{eq:Adapt-prop} \mbox{\bf Adapt}-\mbox{remix}, \mbox{transform, and build upon the material for any purpose, even commercially.}$ 

The licensor cannot revoke these freedoms as long as you follow the license terms.

#### Under the following terms:

Attribution — You must give <u>appropriate credit</u>, provide a link to the license, and <u>indicate if changes were made</u>. You may do so in any reasonable manner, but not in any way that suggests the licensor endorses you or your use.

**ShareAlike** — If you remix, transform, or build upon the material, you must distribute your contributions under the <u>same license</u> as the original.

No additional restrictions — You may not apply legal terms or <u>technological</u> measures that legally restrict others from doing anything the license permits.

#### Notices:

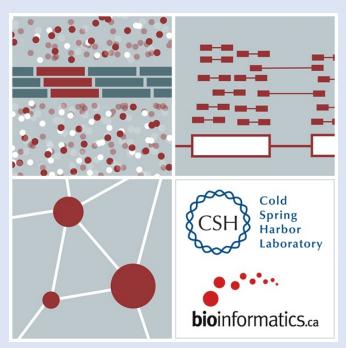
You do not have to comply with the license for elements of the material in the public domain or where your use is permitted by an applicable exception or limitation.

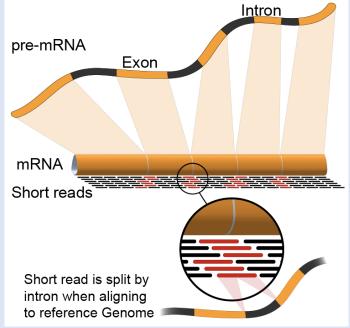
No warranties are given. The license may not give you all of the permissions necessary for your intended use. For example, other rights such as <u>publicity</u>, <u>privacy</u>, <u>or moral rights</u> may limit how you use the material.

## RNA-Seq Module 2: Alignment QC



Malachi Griffith, Obi Griffith, Isabel Risch, Vida Talebian RNA-seq Analysis 2024. June 17-19, 2024



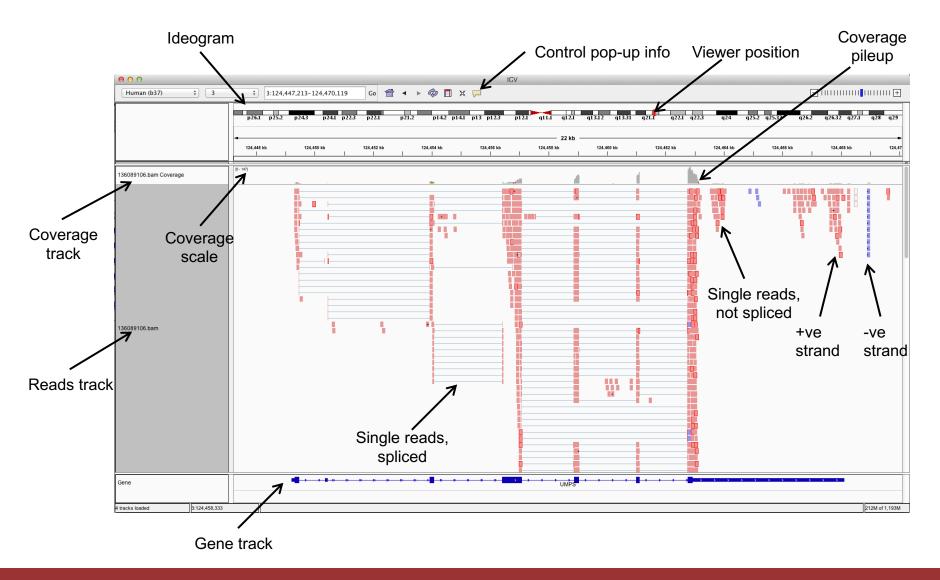




#### Learning objectives of module 3

- Visualization of RNA-seq alignments in IGV
- Alignment QC Assessment
- BAM read counting and determination of variant allele expression status

## Visualization of RNA-seq alignments in IGV browser



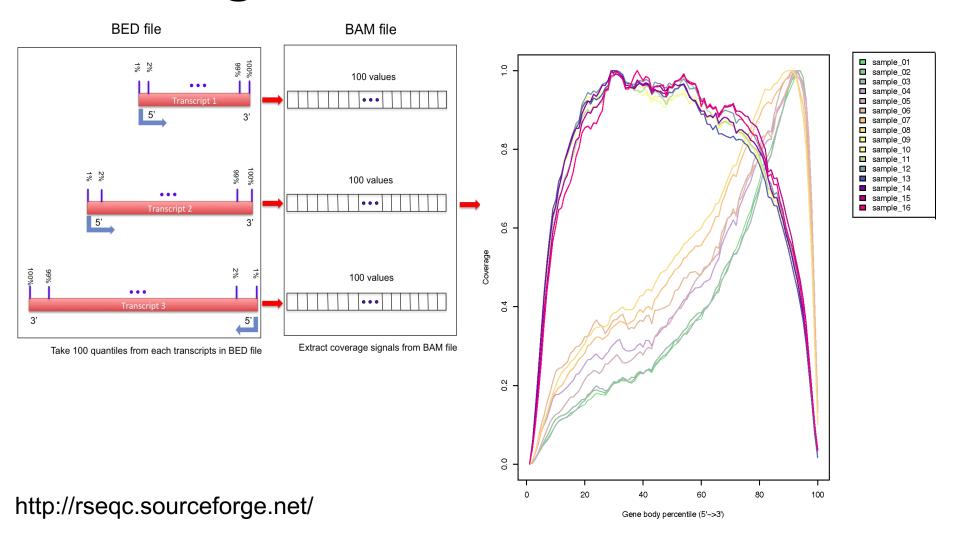
#### Alternative viewers to IGV

- Alternative viewers to IGV
  - http://www.biostars.org/p/12752/
  - http://www.biostars.org/p/71300/
- Artemis, BamView, Chipster, gbrowse2, GenoViewer, MagicViewer,
   Savant, Tablet, tview

#### Alignment QC Assessment

- 3' and 5' Bias
- Nucleotide Content
- Base/Read Quality
- PCR Artifact
- Sequencing Depth
- Base Distribution
- Insert Size Distribution

## Alignment QC: 3' & 5' Bias



## Alignment QC: Nucleotide Content

- Random primers are used to reverse transcribe RNA fragments into double-stranded complementary DNA (dscDNA)
- Causes certain patterns to be over represented at the beginning (5'end) of reads
- Deviation from expected A%=C%=G%=T%=25%

Journal List > Nucleic Acids Res > v.38(12); 2010 Jul > PMC2896536



Nucleic Acids Res. 2010 Jul; 38(12): e131.

Published online 2010 Apr 14. doi: 10.1093/nar/qkq224

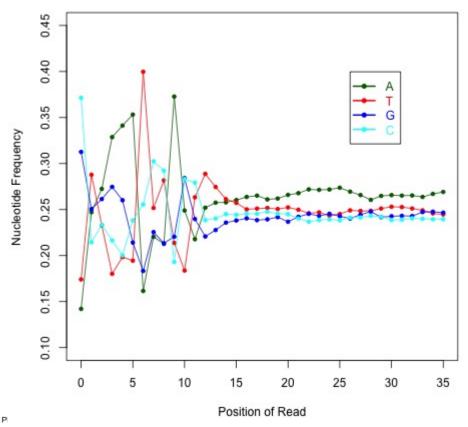
Biases in Illumina transcriptome sequencing caused by random hexamer priming

Kasper D. Hansen, 1,\* Steven E. Brenner, 2 and Sandrine Dudoit 1,3

Author information ▶ Article notes ▶ Copyright and License information ▶

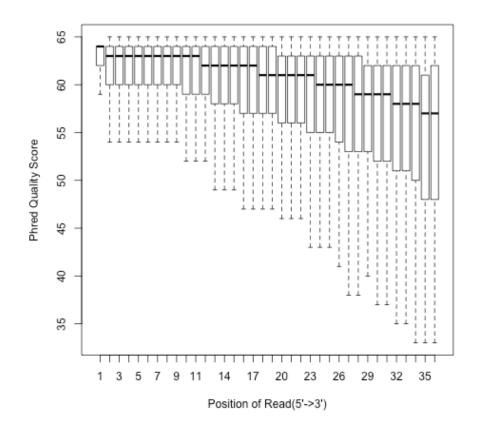
This article has been cited by other articles in PMC.

http://rseqc.sourceforge.net/



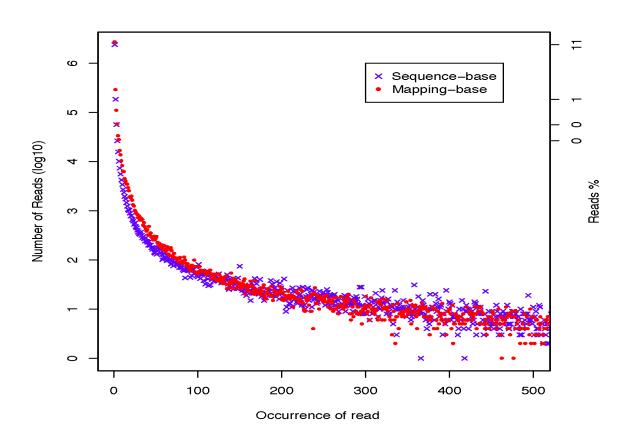
## **Alignment QC: Quality Distribution**

- Phred quality score is widely used to characterize the quality of base-calling
- Phred quality score = -10xlog(10)P, here
   P is probability that base-calling is wrong
- Phred score of 30 means there is 1/1000 chance that the base-calling is wrong
- The quality of the bases tend to drop at the end of the read, a pattern observed in sequencing by synthesis techniques



#### **Alignment QC: PCR Duplication**

- Duplicate reads are reads that have the same start/end positions and same exact sequence
- In DNA-seq, reads/start point is used as a metric to assess PCR duplication rate
- In DNA-seq, duplicate reads are collapsed using tools such as picard
- How is RNA-seq different from DNA-seq?

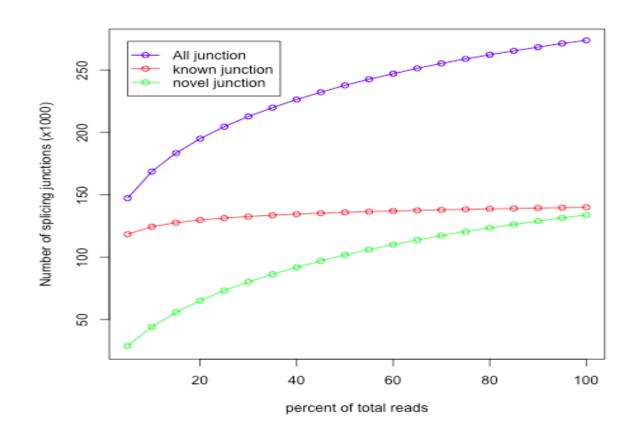


http://rseqc.sourceforge.net/

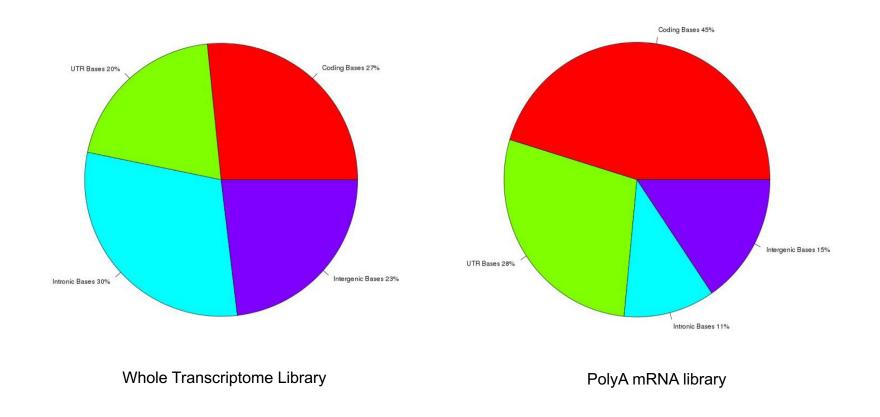
## Alignment QC: Sequencing Depth

#### • Have we sequenced deep enough?

- In DNA-seq, we can determine this by looking at the average coverage over the sequenced region. Is it above a certain threshold?
- In RNA-seq, this is a challenge due to the variability in gene abundance
- Use splice junctions detection rate as a way to identify desired sequencing depth
- Check for saturation by resampling 5%, 10%, 15%, ..., 95% of total alignments from aligned file, and then detect splice junctions from each subset and compare to reference gene model.
- This method ensures that you have sufficient coverage to perform alternative splicing analyses



## **Alignment QC: Base Distribution**



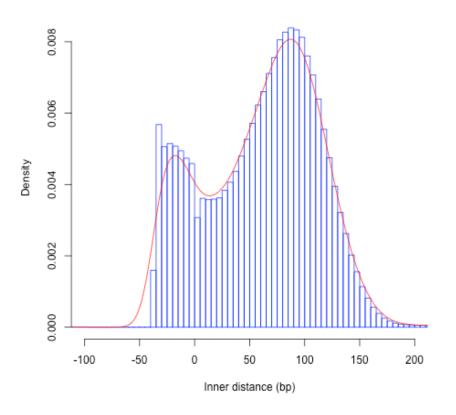
Your sequenced bases distribution will depend on the library preparation protocol selected

#### **Alignment QC: Insert Size**

http://thegenomefactory.blogspot.ca/2013/08/paired-end-read-confusion-library.html

## **Alignment QC: Insert Size**

Mean=60;SD=52



Consistent with library size selection?

## BAM read counting and variant allele expression status



- A variant C->T is observed in 12 of 25 reads covering this position. Variant allele frequency (VAF) 12/25 = 48%.
- Both alleles appear to be expressed equally (not always the case) -> heterozygous, no allele specific expression
- How can we determine variant read counts, depth of coverage, and VAF without manually viewing in IGV?

# We are on a Coffee Break & Networking Session

#### Workshop Sponsors:









