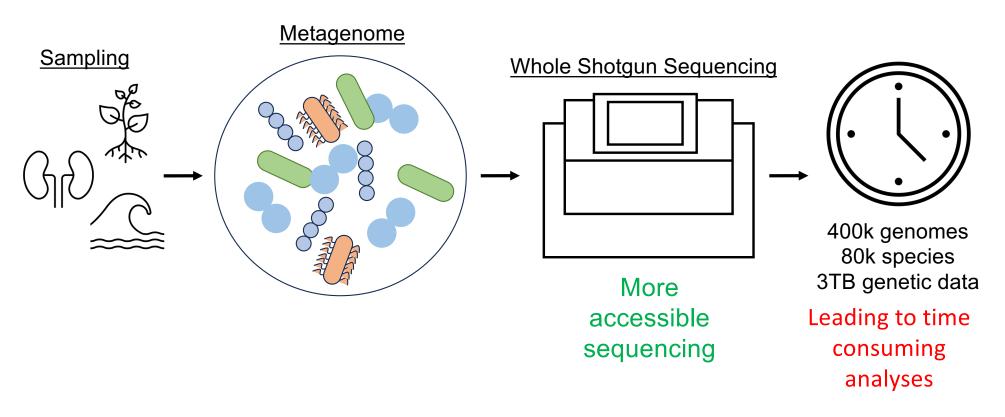


sourmash a tutorial on the FracMinHash sketch approach for metagenomic data

Judith Rodriguez^{1,2,3} and David Koslicki^{1,2,3,4}

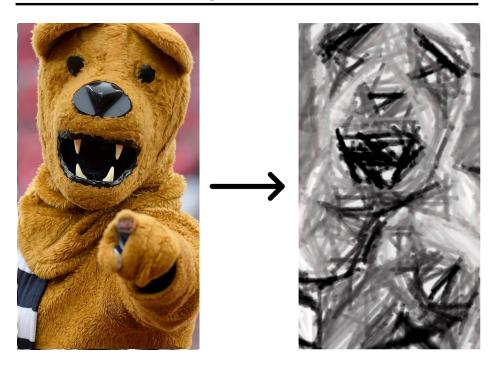
- 1. Bioinformatics and Genomics Program, Penn State
- 2. Life Sciences Huck Institute, Penn State
- 3. Department of Electrical Engineering and Computer Science, Penn State
- 4. Department of Biology, Penn State

Genomics and metagenomics research can be computationally overwhelming



sketching approaches can help with big data

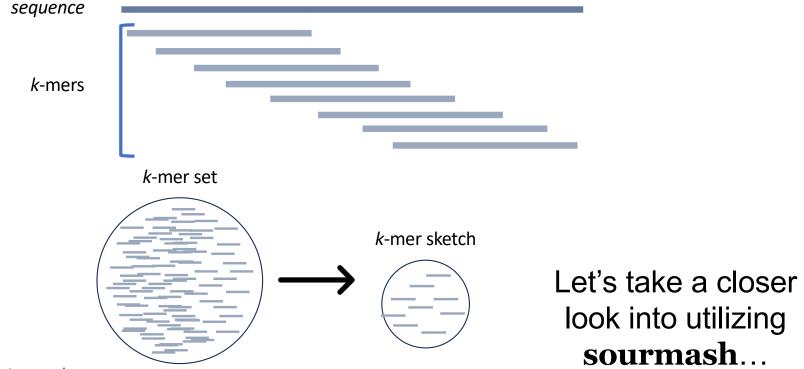
Do we recognize who this is?



Condense data to a simpler representation



sourmash facilitates sketching and analyses of large sequencing data



https://sourmash.readthedocs.io/

Brown, C. T., & Irber, L. (2016). sourmash: a library for MinHash sketching of DNA. Journal of open source software, 1(5), 27.

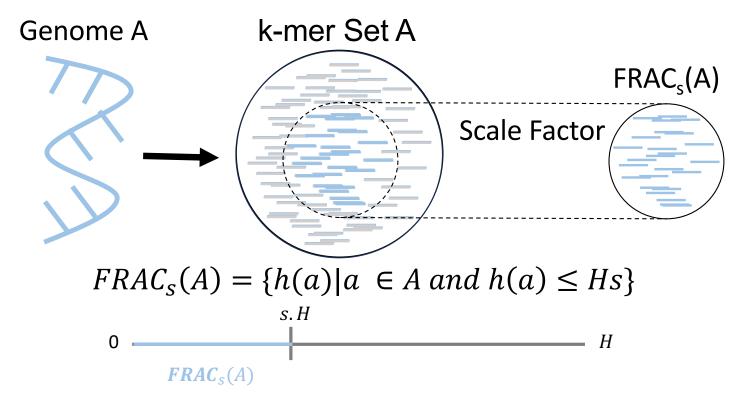
sourmash sketch

first step in sourmash analyses



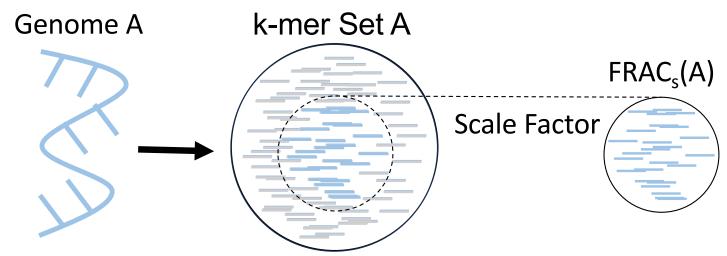


The FracMinHash sketch



Hera, M. R., Pierce-Ward, N. T., & Koslicki, D. (2023). Deriving confidence intervals for mutation rates across a wide range of evolutionary distances using FracMinHash. Genome research, 33(7), 1061-1068.

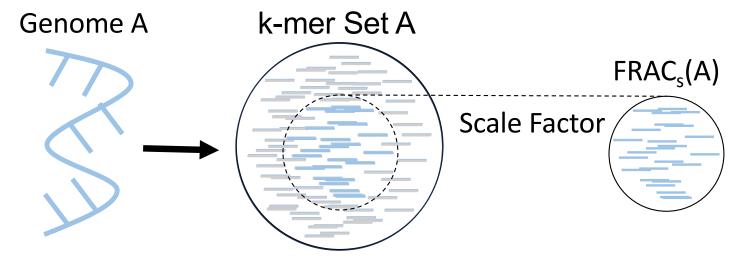




sourmash sketch <dna, protein, translate> <FASTA/Q> -p k=<ksize>,scaled=<int>

sourmash command

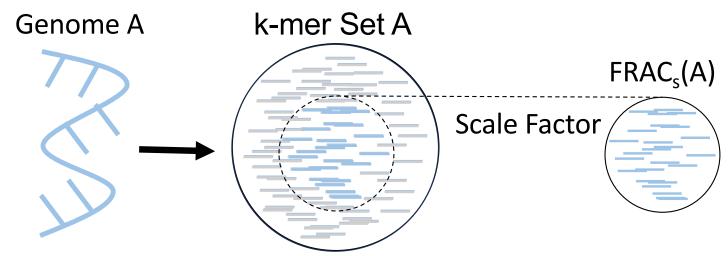




sourmash sketch <dna, protein, translate> <FASTA/Q> -p k=<ksize>,scaled=<int>

identify the type of input sequences

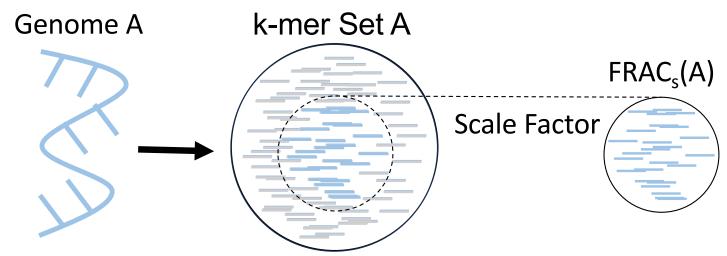




sourmash sketch <dna, protein, translate> <FASTA/Q> -p k=<ksize>,scaled=<int>

Identify the filename with sequences

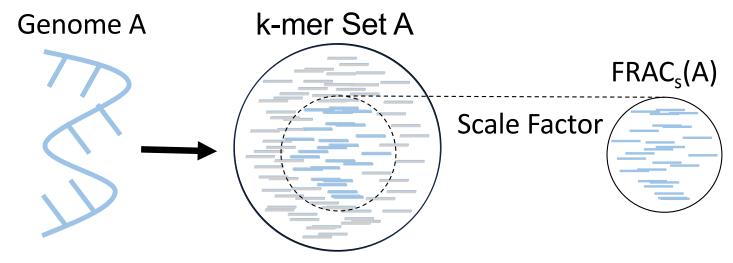




sourmash sketch <dna, protein, translate> <FASTA/Q> -p k=<ksize>,scaled=<int>

Flag required to tune parameters

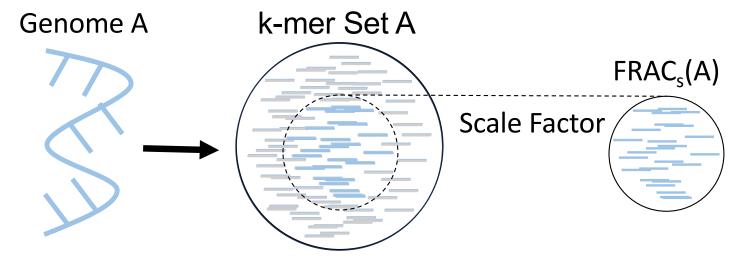




sourmash sketch <dna, protein, translate> <FASTA/Q> -p k=<ksize>,scaled=<int>

Identify the filename with sequences

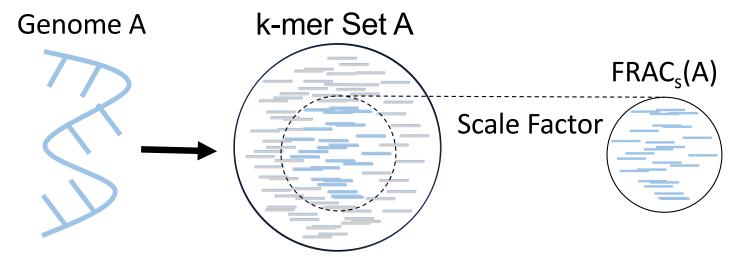




sourmash sketch <dna, protein, translate> <FASTA/Q> -p k=<ksize>,scaled=<int>

scales down sketch





sourmash sketch <dna, protein, translate> <FASTA/Q> -p k=<ksize>,scaled=<int>

Example: scaled=10 will keep 1 of 10 k-mers

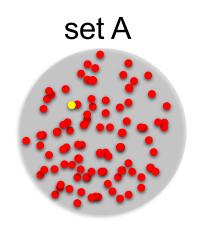
sourmash compare

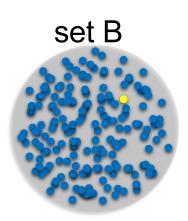
perform pairwise similarity estimations between metagenomic samples





Estimating similarity indexes: jaccard



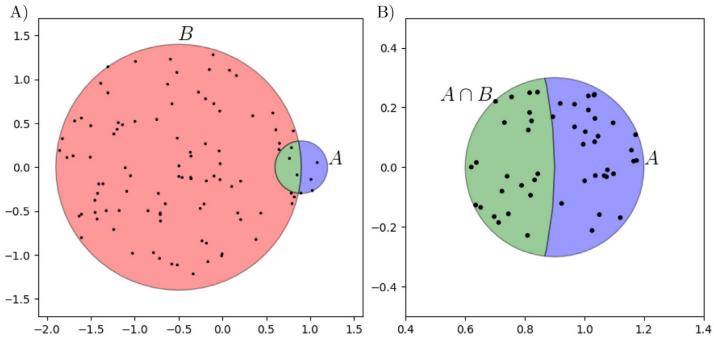


$$J(A,B) = \frac{|A \cap B|}{|A \cup B|} \approx \frac{|S(A \cup B) \cap S(A) \cap S(B)|}{|S(A \cup B)|}$$

Indexes close to 1 are interpreted as more similar



However, the **jaccard** estimate becomes worse with differing set sizes



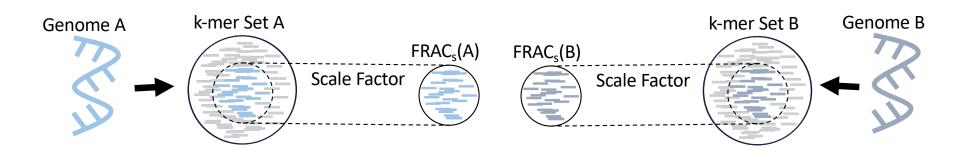
...further, bloom filters are limited to smaller datasets

Koslicki and Zabeti 2019



Estimating similarity indexes: containment

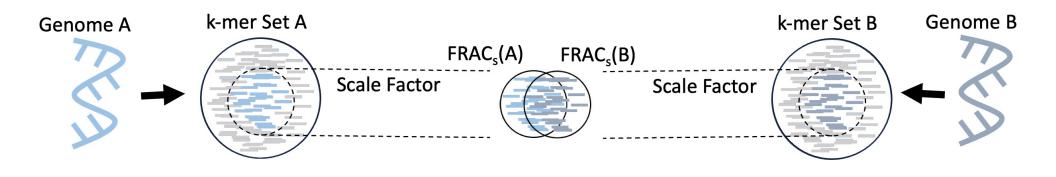
...addresses issues using in jaccard



$$C_{frac}(A,B) \coloneqq \frac{|FRAC_s(A) \cap FRAC_s(B)|}{|FRAC_s(A)|(1-(1-s)^{|A|})} \mathbb{1}_{|FRAC_s(A)|>0}$$



sourmash compare Estimate the containment between two genomes

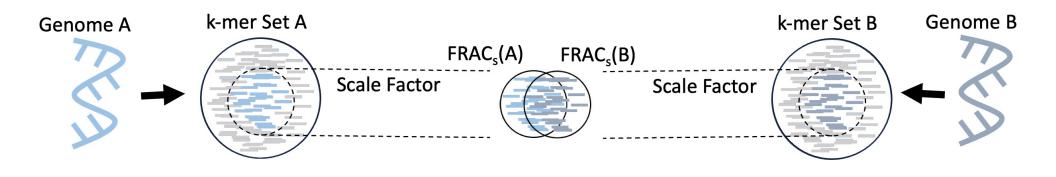


sourmash compare <ref signature> <query signature> --containment --ksize <ksize>
sourmash command



sourmash compare

Estimate the containment between two genomes



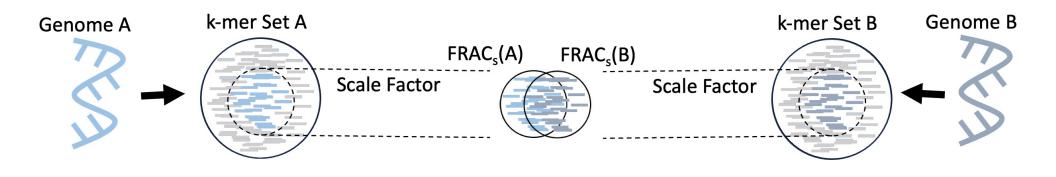
sourmash compare <query signature> <ref signature> --containment --ksize <ksize>

sample of interest



sourmash compare

Estimate the containment between two genomes

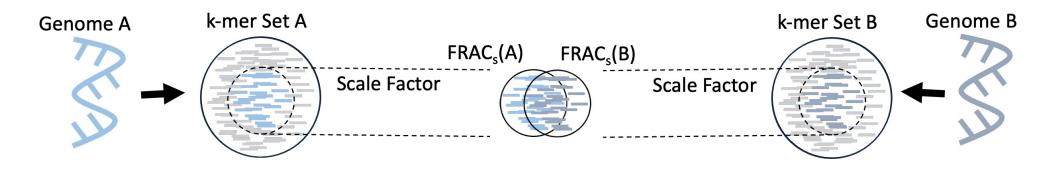


sourmash compare <query signature> <ref signature> --containment --ksize <ksize>

compared to another sample



sourmash compare Estimate the containment between two genomes

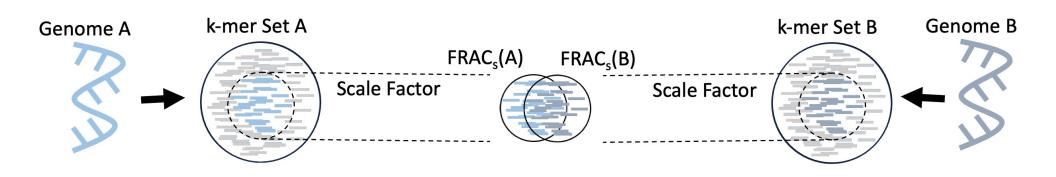


sourmash compare <query signature> <ref signature> --containment --ksize <ksize>

indicate the similarity index



sourmash compare Estimate the containment between two genomes



sourmash compare <query signature> <ref signature> --containment --ksize <ksize>

report highly similar sequences data between samples





First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o MANIFEST.csv

command for further options to information from signatures



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o MANIFEST.csv

option to create MANIFEST



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o MANIFEST.csv

input signature query of interest



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o MANIFEST.csv

output name flag



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o sample_001.manifest.csv

SOURMASH-MANIFEST-VERSION: 1.0 internal_location,md5,md5short,ksize,moltype,num,scaled,n_hashes,with_abundance,name,filename sample_001.fna.singleton.sig,d3513280b35b2a918a7181875c0683c8,d3513280,31,DNA,0,500,22,0,genome_A,sample_001.fna sample_001.fna.singleton.sig,2d96ee330b6b295a06b70fdbfb75af34,2d96ee33,31,DNA,0,500,21,0,genome_B,sample_001.fna sample_001.fna.singleton.sig,364c20a1ca43d3ae3e9ae7d9e9a6c837,364c20a1,31,DNA,0,500,19,0,genome_C,sample_001.fna

Column Name	Description
internal_location	Name of signature file
md5	Name for sketch
ksize	Size used to produce k-mers
moltype	Type of sequence: DNA, Protein
scaled	Parameter used for sourmash sketch to reduce k-mer set
n_hashes	Total k-mers in final sketch
name	Sequence name in FASTA/Q
filename	Input FASTA/Q name



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o sample_001.manifest.csv

SOURMASH-MANIFEST-VERSION: 1.0 internal_location,md5,md5short,ksize,moltype,num,scaled,n_hashes,with_abundance,name,filename sample_001.fna.singleton.sig,d3513280b35b2a918a7181875c0683c8,d3513280,31,DNA,0,500,22,0,genome_A,sample_001.fna sample_001.fna.singleton.sig,2d96ee330b6b295a06b70fdbfb75af34,2d96ee33,31,DNA,0,500,21,0,genome_B,sample_001.fna sample_001.fna.singleton.sig,364c20a1ca43d3ae3e9ae7d9e9a6c837,364c20a1,31,DNA,0,500,19,0,genome_C,sample_001.fna

sourmash search <signature query> <signature ref> --md5 d3513280b35b2a918a7181875c0683c8 --containment

command to searching signatures



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o sample_001.manifest.csv

SOURMASH-MANIFEST-VERSION: 1.0 internal_location,md5,md5short,ksize,moltype,num,scaled,n_hashes,with_abundance,name,filename sample_001.fna.singleton.sig,d3513280b35b2a918a7181875c0683c8,d3513280,31,DNA,0,500,22,0,genome_A,sample_001.fna sample_001.fna.singleton.sig,2d96ee330b6b295a06b70fdbfb75af34,2d96ee33,31,DNA,0,500,21,0,genome_B,sample_001.fna sample_001.fna.singleton.sig,364c20a1ca43d3ae3e9ae7d9e9a6c837,364c20a1,31,DNA,0,500,19,0,genome_C,sample_001.fna

sourmash search <signature query> <signature ref> --md5 d3513280b35b2a918a7181875c0683c8 --containment

signature file where md5 of interest is found



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o sample_001.manifest.csv

SOURMASH-MANIFEST-VERSION: 1.0 internal_location,md5,md5short,ksize,moltype,num,scaled,n_hashes,with_abundance,name,filename sample_001.fna.singleton.sig,d3513280b35b2a918a7181875c0683c8,d3513280,31,DNA,0,500,22,0,genome_A,sample_001.fna sample_001.fna.singleton.sig,2d96ee330b6b295a06b70fdbfb75af34,2d96ee33,31,DNA,0,500,21,0,genome_B,sample_001.fna sample_001.fna.singleton.sig,364c20a1ca43d3ae3e9ae7d9e9a6c837,364c20a1,31,DNA,0,500,19,0,genome_C,sample_001.fna

sourmash search <signature query> <signature ref> --md5 d3513280b35b2a918a7181875c0683c8 --containment

signature file that will be searched against



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o sample_001.manifest.csv

SOURMASH-MANIFEST-VERSION: 1.0 internal_location,md5,md5short,ksize,moltype,num,scaled,n_hashes,with_abundance,name,filename sample_001.fna.singleton.sig,d3513280b35b2a918a7181875c0683c8,d3513280,31,DNA,0,500,22,0,genome_A,sample_001.fna sample_001.fna.singleton.sig,2d96ee330b6b295a06b70fdbfb75af34,2d96ee33,31,DNA,0,500,21,0,genome_B,sample_001.fna sample_001.fna.singleton.sig,364c20a1ca43d3ae3e9ae7d9e9a6c837,364c20a1,31,DNA,0,500,19,0,genome_C,sample_001.fna

sourmash search <signature query> <signature ref> --md5 d3513280b35b2a918a7181875c0683c8 --containment

md5 identifier of interest indicated



First, produce a MANIFEST.csv to search using an md5 identifier as the query

sourmash sig manifest <signature file> -o sample_001.manifest.csv

SOURMASH-MANIFEST-VERSION: 1.0 internal_location,md5,md5short,ksize,moltype,num,scaled,n_hashes,with_abundance,name,filename sample_001.fna.singleton.sig,d3513280b35b2a918a7181875c0683c8,d3513280,31,DNA,0,500,22,0,genome_A,sample_001.fna sample_001.fna.singleton.sig,2d96ee330b6b295a06b70fdbfb75af34,2d96ee33,31,DNA,0,500,21,0,genome_B,sample_001.fna sample_001.fna.singleton.sig,364c20a1ca43d3ae3e9ae7d9e9a6c837,364c20a1,31,DNA,0,500,19,0,genome_C,sample_001.fna

sourmash search <signature query> <signature ref> --md5 d3513280b35b2a918a7181875c0683c8 --containment

indicate the similarity index



First, produce a MANIFEST.csv to search using an md5 identifier as the query

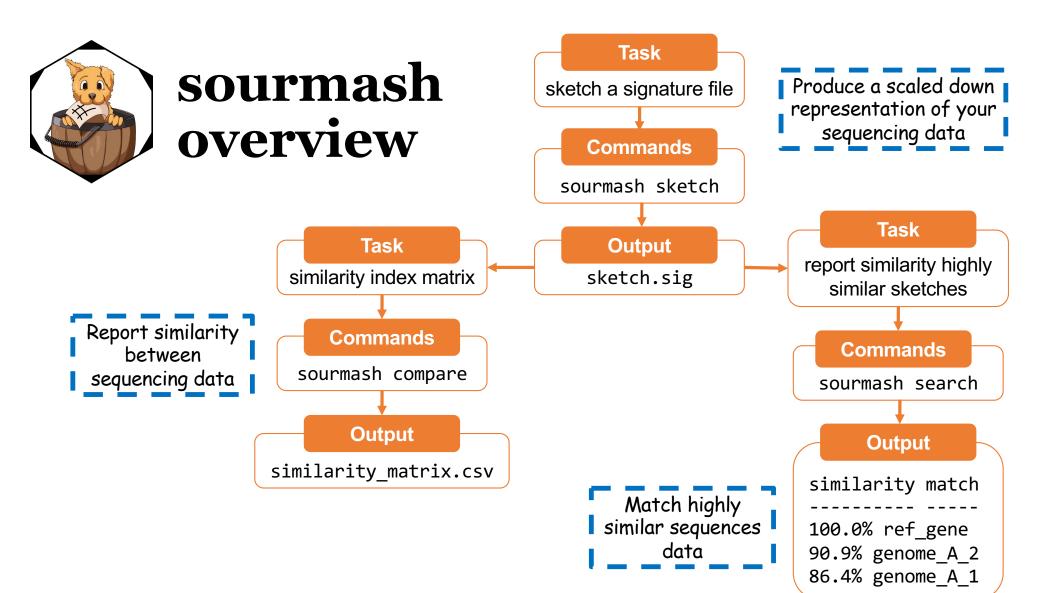
sourmash sig manifest <signature file> -o sample_001.manifest.csv

SOURMASH-MANIFEST-VERSION: 1.0 internal_location,md5,md5short,ksize,moltype,num,scaled,n_hashes,with_abundance,name,filename sample_001.fna.singleton.sig,d3513280b35b2a918a7181875c0683c8,d3513280,31,DNA,0,500,22,0,genome_A,sample_001.fna sample_001.fna.singleton.sig,2d96ee330b6b295a06b70fdbfb75af34,2d96ee33,31,DNA,0,500,21,0,genome_B,sample_001.fna sample_001.fna.singleton.sig,364c20a1ca43d3ae3e9ae7d9e9a6c837,364c20a1,31,DNA,0,500,19,0,genome_C,sample_001.fna

sourmash search <signature query> <signature ref> --md5 d3513280b35b2a918a7181875c0683c8 --containment

sourmash overview





sourmash tutorial

https://github.com/KoslickiLab/sourmash_tutorial

