Handling Categorical Predictors: plyr, ANOVA, and more

Group Properties: Kelp

- Kelp sampled at multiple sites annually
- At each transect, holdfast diameter and # of fronds counted

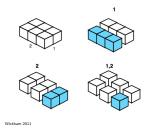


How can we get quick summaries by site?, year, or both?

```
YEAR MONTH
                    DATE SITE TRANSECT QUAD SIDE FRONDS
    2000
             9 2000-09-28 BULL
    2000
             9 2000-09-28 BULL
                                        20
                                                    11
    2000
          9 2000-09-28 BULL
                                    2 20
# 10 2000
          9 2000-09-28 BULL
                                    2 20
                                                    34
# 16 2000
             9 2000-09-28 BULL
                                    3 20
                                                    27
# 17 2000
            9 2000-09-28 BULL
                                                    38
    HLD_DIAM
# 2
          7
          65
# 9
          55
# 10
          55
# 16
          65
# 17
```

For loops for Summarization by Site

The Split, Apply, Combine Strategy



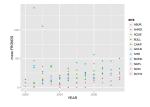
ddply from Hadley Wickham's plyr library

```
kelpMeans
     SITE mean.FRONDS
     ABUR
                29.26
     AHND
                17.63
     AQUE
                21.04
     BULL
                27.30
     CARP
                13.11
     GOLB
                42.16
     IVEE
                25.81
     MOHK
                20.04
     NAPL
                13.16
# 10 SCDI
                 0.00
# 11 SCTW
                14.73
```

ddply from Hadley Wickham's plyr library

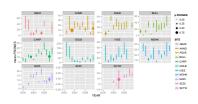
Multiple Groups & ddply

Multiple Groups & ddply



Complex Functions & ddply

Complex Functions & ddply



Exercise: Correlation!

- Evaluate the correlation between fronds and holdfasts by site and year
- ▶ Plot it
- Extra: include the SE of the correlation visually



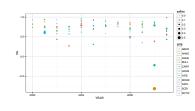
Exercise: Correlation!

```
kelpCor <- ddply(kelp, .(YEAR, SITE), function(adf){
    #first get the correlation
    cors <- cor(adf$FROND, adf$HLD_DIAM)

    #use this to calculate it's SE
    seCor <- sqrt(1-cors"2) / (nrow(adf)-2))

#return both
    return(c(rho = cors, seRho = seCor))
})</pre>
```

Exercise: Correlation!



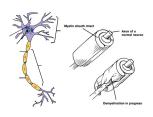
Many plyr Functions

Input Output	Array	Data frame	List	Discarded
Array	aaply	adply	alply	a_ply
Data frame	daply	ddply	dlply	d_ply
List	lanly	ldply	llnly	1 ply

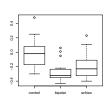
Also r*ply to replicate an action and return an object. Great for simulation.

See also colwise and each for everyday use!

Categorical Predictors: Gene Expression and Mental Disorders



Categorical Predictors

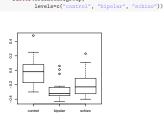


How do we determine the importance of categorical predictors?

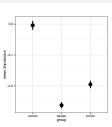
Categorical Predictors Ubiquitous

- ► Treatments in an Experiment
- Spatial groups plots, Sites, States, etc.
 Individual sampling units
- ► Temporal groups years, seasons, months

Aside: Reordering Factors brainGene\$group <- factor(brainGene\$group,

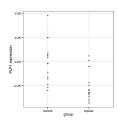




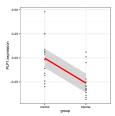


What is the variance between groups v. within groups?

But How is the Model Fit?

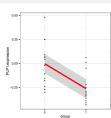


But How is the Model Fit?



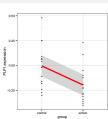
Underlying linear model with ${\sf control} = {\sf intercept},$ dummy variable for bipolar

But How is the Model Fit?



Underlying linear model with control = intercept, dummy variable for bipolar

But How is the Model Fit?



Underlying linear model with control = intercept, dummy variable for schizo

Different Ways to Write a Categorical Model	This is a Linear Model		
$y_{ij} = \bar{y} + (\bar{y}_i - \bar{y}) + (y_{ij} - \bar{y}_i)$ $y_{ij} = \mu + \alpha_i + \epsilon_{ij}, \qquad \epsilon_{ij} \sim N(0, \sigma^2)$ $y_j = \beta_0 + \sum \beta_i x_i + \epsilon_j, \qquad x_i = 0, 1$ $x_i \text{ inidicates presence/abscence of a category}$ Traditional ANOVA special case where all x_i are orthogonal Often one category set to β_0 for ease of fitting	bg.sub.lm <- lm(PLP1.expression ~ group, data=brainGene)		
Hypothesis Testing with a Categorical Model: ANOVA	Assumptions of Ordinary Least Squares Regression		
$H_0=\mu_1=\mu 2=\mu 3=\dots$ OR $eta_0=\mu, \qquad eta_i=0$	 Independence of data points Normality within groups Homoscedasticity (homogeneity of variance) 		

F-Test to Compare

 $SS_{Total} = SS_{Between} + SS_{Within}$

$$SS_{Between} = \sum_i \sum_j (ar{Y}_i - ar{Y})^2$$
, df=k-1
 $SS_{Within} = \sum_i \sum_j (Y_{ij} - ar{Y}_i)^2$, df=n-k

To compare them, we need to correct for different DF. This is the Mean Square.

$$MS = SS/DF$$
, e.g, $MS_W = \frac{SS_W}{n-k}$

ANOVA

anova(bg.sub.lm)

- # Analysis of Variance Table
- # Response: PLP1.expression
- # Df Sum Sq Mean Sq F value Pr(>F) # group 2 0.54 0.2701 7.82 0.0013
- # group 2 0.54 0.2701 7.82 0.0013 # Residuals 42 1.45 0.0345

F-Test to Compare

 $F=\frac{MS_R}{MS_W} \ \ {\rm with\ DF=k\text{-}1,n\text{-}k}$ (note similarities to SS_R and SS_E notation of regression)

Inspecting Assumptions par(mfrow=c(2,3))

Levene's Test of Homogeneity of Variance

```
library(car)

# Loading required package: MASS
# Loading required package: nnet

leveneTest(PLP1.expression ~ group, data=brainGene)

# Levene's Test for Homogeneity of Variance (center = median)

# Df F value Pr(>F)

# group 2 1.01 0.37

# 42

Levene's test robust to departures from normality
```

What do I do if I Violate Assumptions?

- ► Nonparametric Kruskal-Wallace (uses ranks)
- ► Transform?
- GLM with ANODEV

Kruskal Wallace Test

```
kruskal.test(PLP1.expression = group, data=brainGene)

# Kruskal-Wallis rank sum test

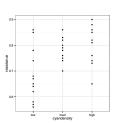
# data: PLP1.expression by group
# Kruskal-Wallis chi-squared = 13.2, df = 2, p-value =
# 0.001361
```

Exercise: Daphnia Resistance

- ► Plot the mean and SE of the data by group
- Evaluate whether the data is appropriate for ANOVA
- Fit an ANOVA and check diagnostics
- Evaluate results & compare to Kruskal-Wallace and a glm with a Gamma distribution

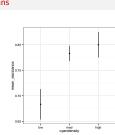


Daphnia Data



Daphnia Means

Daphnia Means



How about HOV?

```
leveneTest(resistance ~ cyandensity, data=daphnia)

# Levene's Test for Homogeneity of Variance (center = median)

# Df F value Pr(>F)

# group 2 2 0.15
```

ANOVA shows an Effect

anova(daphniaLM)

Analysis of Variance Table

```
# Response: resistance
            Df Sum Sq Mean Sq F value Pr(>F)
# cyandensity 2 0.0892 0.0446
                                 6.69 0.0041
# Residuals 29 0.1933 0.0067
```

daphniaLM <- lm(resistance ~ cyandensity, data=daphnia)

KW shows an Effect

```
Kruskal-Wallis rank sum test
# data: resistance by cyandensity
# Kruskal-Wallis chi-squared = 8.2, df = 2, p-value =
# 0.01658
```

Bad GLM Does Not

```
# Analysis of Deviance Table
# Model: Gamma, link: identity
# Response: resistance
# Terms added sequentially (first to last)
             Df Deviance Resid. Df Resid. Dev
# NIIII
                                 31
                                        0.529
# cvandensity 2
                   0.162
                                         0.367
```

Diagnostics Also Good

