

>>>py[cellerator] Cell Simulation Symbolic Processing in Python

Reference Notes

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Preface

The goal of `pycellerator` is to provide a software tool for biological simulation that

- Has all of the capabilities of the `Cellerator` arrow based reaction language[16];
- Is open source;
- Runs under Linux, Windows, and Mac OS X;
- Is written in a multi-platform language;
- May be used either as a stand-alone program, at the command-line, or as a library within a high-level computer language;
- Is written in a computer language that is freely available (at no cost to users) on all platforms;
- Does not depend on any external libraries except for freely available multi-platform libraries, so that it would not be necessary to maintain separated executable (or any executable) builds.

Python 2.7 was chosen as the programming platform because it is implemented on all three platforms, is freely available, has a large user support community, and has an extensive collect of external libraries that support it.

The last requirement is more complicated, because it rules out a lot of useful libraries that are written in C or Fortran and wrapped in Python, and hence required complicated installations (read: recompilations) for each Operating System. It is our contention that this software should not be operating system dependent and thus the final requirement was added.

`pycellerator` was developed using standard distributions provided by the Python community (<http://www.python.org>). It is fully compatible with standard distributions available commercially such as Anaconda and Enthought Canopy. It can also be used in ipython notebooks.

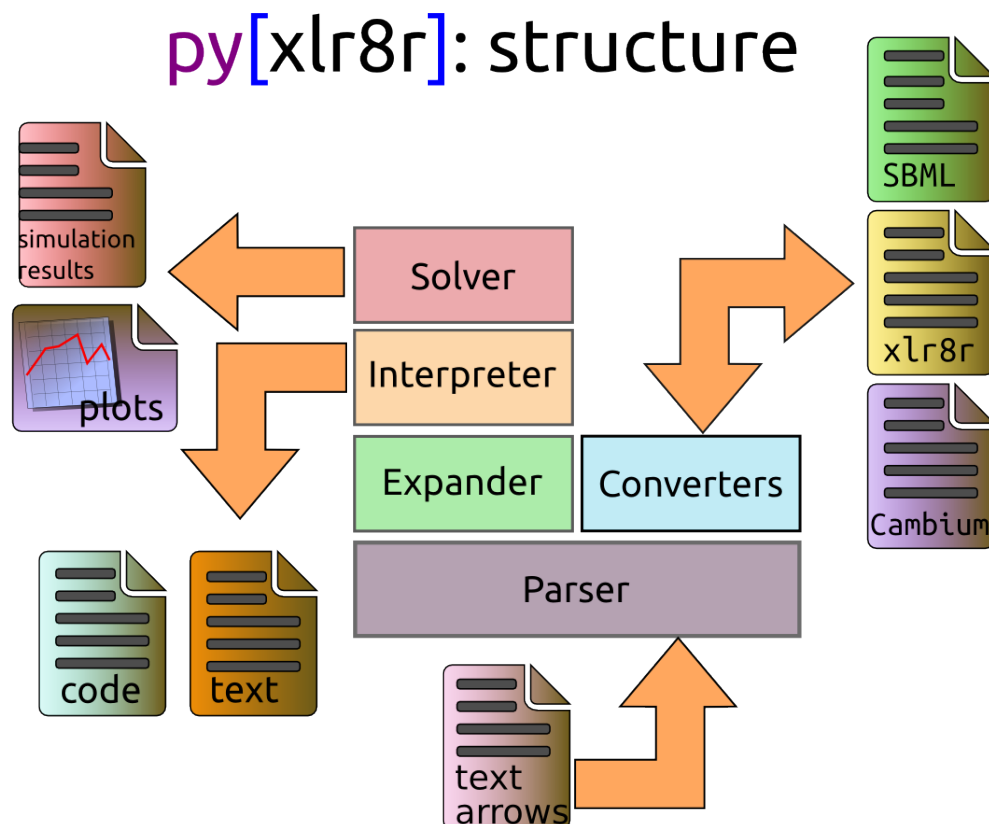
Chapter 1

Introduction

This chapter will provide an overview of pycellerator structure and functionality.

pycellerator has the following components, as illustrated in figure 1.1. The functional relationship between these components is illustrated in figure 1.2. In brief, pycellerator first reads a model file with the **parser** and converts it to an internal database. This database is then processed by the **expander** module converts the reactions in the database to their most basic form. Control is then passed to the **interpreter**, which converts the reactions to a second database of terms of terms in differential equations, and then combines all of these terms together to form a system of differential equations out of the model. Finally, the **solver** writes the system of differential equations to a python program and then runs the program, if required to perform a numerical simulation.

Figure 1.1: Schematic of pycellerator structure.



A Parser: The parser module reads lists of text-formatted reactions, typically from a text file that you will manually edit by hand, that describe a particular biochemical phenomenon. For example, a simplified version of the famous Belousov-Zhabotinsky reaction[1, 22] known as the Oregonator[5, 6] can be written in this language as:

```
[Br + BrO3 -> HBrO2 + HOBr, k1]
[Br + HBrO2 -> 2 HOBr, k2]
[BrO3 + HBrO2 -> 2 Ce + 2 HBrO2, k3]
[2 HBrO2-> BrO3 + HOBr, k4]
[Ce -> 0.5 Br, k5]
```

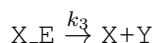
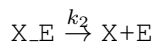
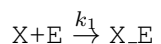
The parser module converts the reactions into an internal data structure that can be interrogated by the other modules with such questions as “What are the products of this reaction?”

The parser module is not normally invoked directly by the user.

Expander: The expander module converts a list of complex reactions into its most basic form. For example, the input reaction

```
[X => Y, mod[E], rates[k1,k2,k3]]
```

is a shorthand that represents the set of biochemical reactions



where X_E is the name of the complex formed when E is bound to X. The expander converts the input reaction to its three component reactions:

```
[X+E->X_E,k1]
[X_E->X+E,k2]
[X_E->Y+E,k3]
```

While it is possible for the user to call the expander directly to see what the component reactions look like, usually this will be done automatically by the interpreter or solver modules and the user will generally not be required to directly interact with the expander.

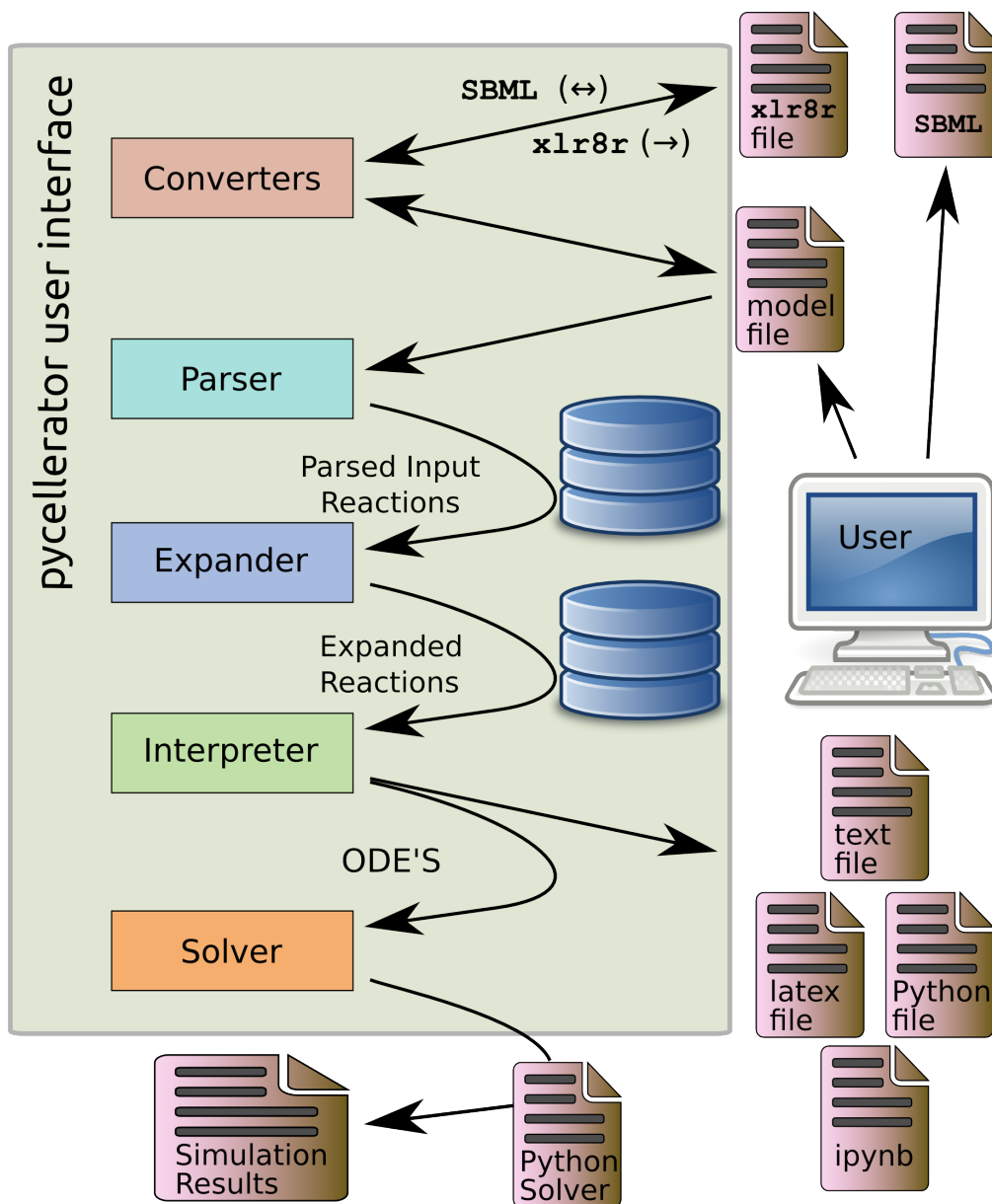
Interpreter: The interpreter can provide a list of differential equations, a simple python program that instantiates the system of differential equations, or a $\text{L}^{\text{A}}\text{T}_{\text{E}}\text{X}$ representation of the system of differential equations.

The interpreter can be invoked directly, if that is what is required, but if the user is only interested in a simulation, the interpreter need not be invoked manually.

Solver: The primary function of the solver module is to produce a python function that is compatibly with `scipy.tt.odeint`. If requested the solver module will also run a simulation utilizing the python function, plot selected (or all) simulation variables, and/or write the results to an output file in CSV or TSV format. Because this function is a completely stand-alone program – fully independent of all other `pycellerator` components – multiple instantiations can be easily parallelized, e.g., for parameter optimization or other chores.

Converters: The converters module is used to convert to and from other formats, particularly `Cellerator` (`x1r8r`) arrows. Every arrow implemented in `pycellerator` is equivalent to some arrow in `x1r8r` and vice-versa.

Figure 1.2: Functional overview of pycellerator operation. Users will interface with pycellerator in any of the following ways: via the pycellerator command line interface, e.g., in the mac terminal or or windows command prompt; in an ipython notebook; in the python shell; or by direct functions calls from other programs. Every function is available via any of these techniques. [The database icon is taken from the Wikimedia Commons and has a CC-BY-SA 3.0 license. The Computer icon was taken from Wikimedia commons and has a GPL 2.0 license. These licenses supercede the copyright license of this document. This picture may be copied and reused under the terms of the CC-BY-SA 3.0 and GPL 2.0 licenses.]



Chapter 2

Installation

2.1 Install a Basic Python System

To use `pycellerator`, python 2.7 and several python libraries must be installed on your computer. Python is free, open source, and available on all major operating systems. This section provides an overview of how to install a basic python system on Windows, Macintosh OS, and Linux operating systems.

2.1.1 Installing on Windows

There are two basic ways to install python: (1) install a “commercial” base distribution or (2) use a binary installer from <https://www.python.org/downloads/windows/>. There are several commercial distributions that provide free installers that will install the base python system for you. If you use a binary distribution, you will also have to install a number of other packages. The easiest commercial distributions to use are **Anaconda** and **Enthought Canopy**.

Anaconda Python

To install Anaconda Python, go to <http://continuum.io/downloads> and scroll down to select the appropriate installer for your operating system. This will download a file (e.g., `Anaconda-2.3.0-x86_64.exe`, or something similar).

Double click on the installer and follow the instructions to install python.

After you are done, locate the Anaconda Command Prompt from the windows search menu and type in the following. Type enter after line and wait until the command prompt (the name of the current folder) is shown.

```
conda update conda
conda update ipython ipython-notebook ipython-qtconsole
conda update numpy scipy sympy matplotlib
python -m pip install --upgrade pip
pip install pyparsing
pip install pulp
```

Enthought Canopy

As an alternative to Anaconda, Enthought Canopy is at <https://store.enthought.com/downloads/>. Pick your operating system from the button on the top of page and download the binary installer.

After downloading the installation package (e.g., `canopy-1.5.5-win-64.msi` or a similarly named file), double click on the installer to install python. It will be sufficient to answer all of the questions in the dialogs with the default values.

When the installer is finished, the canopy dashboard should open automatically. You must run the dashboard at least once to finish the installation. The dashboard will open a menu that says “Welcome to Canopy” at the top. If the dashboard does not open, there should be a Canopy icon on your desktop. Click this to open the Canopy dashboard. If this icon is not installed, open Canopy from the windows search menu. If it does not appear, the installation has not completed properly.

From the Canopy dashboard select `Edit > Preferences > General` and verify that Canopy is set as your default python environment. If it is not, click on the button that says “Set as Default”, then click on “OK.”

Then go to `tools > package manager > available packages` and click on “install all available packages.”

Then exit from Canopy by selecting `File > Exit`.

From the windows search menu, open Command Prompt and type in the following commands, one at a time. Type enter after line and wait until the command prompt (the name of the current folder) is shown.

```
python -m pip install --upgrade pip
pip install --upgrade numpy scipy sympy matplotlib
pip install pyparsing
pip install pulp
```

Using the Binary Installers

From <https://www.python.org/downloads/windows>, download the latest MSI installer for Python 2.7. At the time this was written, the file name was `python-2.7.9.amd64.msi`.

Locate the installer file, double click, and answer all the prompts. Make sure during the installation that `python.exe` and `pip` are checked off as visible to all.

Next, download a Microsoft Visual C compiler for Python from <http://www.microsoft.com/en-us/download/details.aspx?id=44266>. The installer file is called `VCforPython27.msi`. Locate the installer, double click, and follow the instructions. Restart your computer after the installation is complete.

Open a command prompt (Windows search, type `command prompt`) and enter the following:

```
python -m pip install --upgrade pip
pip install sympy pulp pyparsing setuptools
pip install numpy scipy matplotlib
pip install ipython[notebook]
```

2.1.2 Installing on Macintosh OS

If you have a Mac, then Python is already installed on your computer. The base system is pre-installed as part of the Mac operating system. The base system does not include the numerical libraries that are also required. You can either install these using `pip` or install one of the commercial systems like Anaconda or Enthought Canopy.

Follow the instructions for Windows if you want to install the commercial system. You will not need to install Visual C; instead, you may be prompted to download and install XCode tools from Apple during the installation process.

Upgrade Mac Python using pip

Locate the terminal application in the utilities folder and open it. Enter

```
sudo easy_install pip
```

When requested, enter your password (you must have administrator access on your Mac). If (when) you are prompted to install XCode from Apple, click yes, and follow the instructions on any dialog that follows.

When the XCode installation is completed (or if it was not suggested), open a new terminal session and type in the following. Hit the enter key after each line and wait for the prompt (the name of the current working directory) before typing in the next line.

```
pip install sympy pulp pyparsing  
pip install numpy scipy matplotlib  
pip install ipython[notebook]
```

If you have a virtual operating system like Parallels Desktop installed on your computer, make sure that Safari (or some other web browser such as Firefox or Chrome) is set as your default browser, and not Parallels. Otherwise ipython will try to open your virtual operating system every time it runs python.

2.1.3 Installing on Linux

You can either install a base python system from your package manager, download binaries from python.org, or build from source (also available at python.org). Pycellator requires python 2.7.X but is not compatible with python 3.x.

The standard python installation includes its own package manager called pip. If you install the base system from python.org this should automatically be installed for you. Otherwise, you should look to also install pip from your package manager. This allows you to bypass your package manager when updating python.

To upgrade to the latest version of pip,

```
python -m pip install --upgrade pip
```

To add any missing packages to python,

```
pip install packagename
```

To upgrade to the latest version of any package,

```
pip install --upgrade package
```

Pycellator needs the following packages which you can get from pip: pyparsing, pulp, sympy, numpy, scipy, and matplotlib.

```
pip install pyparsing  
pip install pulp  
pip install sympy  
pip install numpy  
pip install scipy  
pip install matplotlib
```

You can also put all the function names on a single line:

```
pip install pyparsing pulp sympy numpy scipy matplotlib
```

Binary installers and source code versions are also available for each of these packages.

Package	Project Home Page
numpy	http://numpy.org/
scipy	http://scipy.org/
matplotlib	http://matplotlib.sourceforge.net/
pyparsing	http://pyparsing.wikispaces.com/
sympy	http://code.google.com/p/sympy/
pulp	https://github.com/coin-or/pulp

2.2 Install libSBML

If you want to work with SBML files, you will also need to install a version of libSBML that is appropriate for your operating system. Make sure to install a version that is compatible with Python.

Follow the instructions at <http://sbml.org/Software/libSBML> to find the appropriate binary installer for your operating system.

You do not have to have libSBML installed if you do not plan on using SBML files. If libSBML is not installed, all non-SBML related functionality of pycellerator will be unaffected.

The following describes how to install libSBML in ubuntu if you are primarily interested in using libSBML for python and not for other languages. It was taken from <http://sourceforge.net/projects/sbml/files/libsbml/5.11.6-experimental/binaries/Linux/> on 5 Sept 2015, and you should check for the latest information.

1. Install the prerequisite software packages installed on your operating system: **python-dev**, **libxml2-dev**, **libz-dev**, and **libbz2-dev**.

```
sudo apt-get install python-dev libxml2-dev libz-dev libbz2-dev
```

2. From the terminal type:

```
sudo pip install python-libsbml
```

For windows, the latest (as of 5 Sept 2015) binary python installers are at <http://sourceforge.net/projects/sbml/files/libsbml/5.11.4/stable/Windows/64-bit/python/>. These installers will only install libSBML for python, and not for other languages.

The Mac installers are at <http://sourceforge.net/projects/sbml/files/libsbml/5.11.4/stable/Mac%20OS%20X/> and the instructions given for installation are similar to those for linux.

2.3 Install pycellerator

The procedure to install pycellerator follows.

1. Get the latest **release** from github. This can be found at <https://github.com/biomathman/pycellerator/releases>. Download the file **Install-pycellerator-v-X.zip**, where **X** is the

latest version number. This will be a compressed archive (zip file) of the files you need to install pycellerator.

2. Unzip the archive. Copy the folder **pycellerator** (e.g., drag and drop the entire folder) to wherever you want to keep your pycellerator files. For example, this may be a folder in your home directory. If your home folder is **johnsmith**, and you drag **pycellerator** onto your home folder, then your **pycellerator** folder will be **johnsmith/pycellerator**, i.e., the folder **pycellerator** inside your home folder.
3. Inside the **pycellerator** folder is detailed documentation in the file **pycellerator.pdf** (the file you are reading now). Chapter 2 gives detailed instructions on how to install Python (if you don't already have it installed); and what additional Python libraries are needed and how to get them.
4. A quickstart check of the command line utility, from inside the **pycellerator** folder, type the following into the terminal:

```
python pycellerator.py solve -in Gold1.model -plot
```

5. For a quickstart check of the ipython notebook, type the following into the terminal:

```
ipython notebook
```

Navigate to the notebook **demo.ipynb** and open it.

Chapter 3

pycellerator Arrow Reference

3.1 Introduction

Reactions in pycellerator are specified in textfiles using a arrow based language that can be typed using any standard ASCII or UTF keyboard. All of the characters needed to type a reaction exist on standard US keyboards. A summary of the arrow forms available in pycellerator is given in Table 3.1.

The canonical form for an arrow form is

$$[\text{LHS } \textit{arrow} \text{ RHS}, \text{ mod}[\textit{modifiers}], \text{ rates}[\textit{rate constants}]]$$

or

$$[\text{LHS } \textit{arrow} \text{ RHS}, \text{ rates}[\textit{rate constants}]]$$

All of the square brackets are required and:

LHS indicates a species, list of species, or sum of species that give the input to the reaction. Depending on the type of reaction, their concentrations or amounts may or may not change as a result of the reaction, but they will always affect the calculation of the output species.

RHS indicates a species, list of species, or sum of species that give the output of the reaction. Each species in RHS will normally change in concentration or amount as a result of the reaction.

arrow determines the type of reaction. It typically looks something like \rightarrow , \leftrightarrow , \Rightarrow , $\mid\rightarrow$, etc.

modifiers indicates a list of species that affect the output of the reaction; like input reaction, their concentrations may or may not be affected by the reactions. The definition of whether a species goes in the mod or *modifiers* list depends on the definition of the reaction in the following sections, and not on the biochemical process. This is a computational distinction only. In general, *modifiers* correspond to catalysts in enzymatic reactions. The normal distinction used is that for **catalytic** arrows (e.g., $\mid\rightarrow$) the amount of catalyst does not change, but in **enzymatic** arrows (e.g., \Rightarrow , $\Leftarrow\Rightarrow$) the amount of enzyme may, in fact, change.

Table 3.1

Name	Arrow	Typical ODE Term	Ref
Simple Mass Action Mass Action, Stoichiometry Mass Action, Reversible Simple Catalytic Catalytic, w/ Intermediate Complex Catalytic, Reversible, w/ Int. Complex Catalytic, Two Int. Species	1 [X -> Y, k]	$(s_{i,r} - s_{i,l})k \prod_{i \in \text{LHS}} X_i^{e_i}$	3.5.1
	2 [e1 X1 + e2 X2 + ... -> s1 Y1 + s2 Y2 + ..., k]		3.5.2
	3 [e1 X1 + ... <-> s1 Y1 + ..., rates[k1,k2]]	Expanded into a pair of type (1) or (2) reactions.	3.5.4
	4 [e1 A1 + e2 A2 + ... --> s1 B1 + s2 B2 + ..., mod[X], k]	Expanded into a single type (1) or (2) reaction.	3.5.3
	5 [A => B, mod[X], rates[k1, k2, k3, k4]]	Expanded into three or four type (1) reactions	3.5.5
	6 [A <=> B, mod[X,Y], rates[k1, k2, ..., k8]]	Expanded into two type (5) reactions.	3.5.6
	7 [A :=> B, mod[E], rates[k1,k2, ..., k6]]	Expanded into six type (1) reactions	3.5.7
Hill Functions	8 [A -> B, Hill[v,n,K,α,T]]	$\frac{vEk(\sum T_i A_i + \alpha)^n}{K^n + (\sum T_i A_i + \alpha)^n}$	3.7.1
	9 [A --> B, mod[E], Hill[v,n,K,α,T]]		
GRN (Genetic Regulatory Network Model)	10 [A -> B, GRN[v,β,n,h]]	$\frac{vE}{1 + \exp(-h - \sum \beta_i A_i^{n_i})}$	3.7.2
	11 [A --> B, mod[E], GRN[v,β,n,h]]		
S-System	12 [A -> B, SSystem[τ, k+, k-, c+, c-]]	$\frac{k_+ \prod A_i^{C_{i,+}} - k_- \prod A_i^{C_{i,-}}}{\tau}$	3.7.3
	13 [A --> B, mod[E], SSystem[τ, k+, k-, c+, c-]]		
MMH (Michaelis-Menten- Henri)	14 [A :->B, MMH[K,v]] or [A :->B, MMH[k1,k2,k3]]	$\frac{vAE}{K+A}$ or $\frac{k_3AE}{(k_2+k_3)/k_1+A}$	3.6.1
	15 [A :-->B, mod[X], MMH[K,v]] or [A :->B,mod[X], MMH[k1,k2,k3]]		
Rational Function	16 [[A1,A2,...],[X1,X2,...]]==>S,rational[a,d,m,n]	$\frac{a_0 + \sum a_i A_i^{n_i}}{d_0 + \sum d_i X_i^{m_i}}$	3.7.5
MWC (Monod-Wyman-Changeaux)	17 [S==>P, mod[E], MWC[k, n, c, L, K]]	$\frac{k\mathcal{E} (cLs(cs+1)^{n-1} + s(s+1)^{n-1})}{L(cs+1)^n + (s+1)^n}$	3.6.2
	18 [S==>P, mod[E, [[A1,A2,...],[I1,I2,...]], MWC[k, n, c, L, K]]	See reference for multiple Activator/Inhibitor equations.	
NHCA (Non-hierarchical coop. act.	19 A -> B, NCHA[v, Tp, Tm, n, m, k]	$\frac{vA^m}{kB^m + A^m}, A=1+T_pX^n, B=1+T_mX^n$	3.7.4

Table 3.1 (continued)

Name	Arrow	Typical ODE Term	Ref
USER	20 <code>[X1+X2+... ->Y, USER[v, T, n, h, f]]</code> f should be a Python lambda expression of a single variable	$vf(h - \sum T_i P_i^{X_i})$	3.7.6
using	21 <code>[s1*X1+s2*X2+...->q1*X1+..., using["expr"]]</code> expr should be Python infix expression depending on variables defined in the model.	$X'_i = q_i - s_i)X_i$	3.8

rates is a keyword that varies with the type of arrow; typical keywords include MWC (for Monod-Wyman-Changeaux); MMH (for Michaelis-Menten-Henri); Hill (for Hill functions); and so forth.

rate constants is a list of symbols or numbers that give the rate constants or other parameters used in the reaction equation. The definition of these parameters is different for each type of reaction, and is described in the following sections and subsections.

3.2 Substituting Equations for Rate Constants

In any of the arrow expressions that follow in the remainder of the chapter, any rate constant may be replaced by any valid Python expression in quotation marks. For example, the hypothetical reaction

```
[X -> Y, "v/(K+X)"]
```

is perfectly valid. Since the reaction itself is written in the form of a mass action equation, the expression is treated as a rate constant k and the ODEs are $[Y]' = [X]' = k[X]$, where k is to be replaced with the expression in quotes. It will thus produce the differential equation terms:

$$[Y]' = -[X]' = \frac{v[X]}{K + [X]}$$

This allows users to define virtually any rate law in a reaction.

See also User Defined Regulatory Arrows in section 3.7.6 and User Defined Stoichiometric Arrows in section 3.8. User defined stoichiometric arrows will not multiply the expression by the mass-action product (the product of the species on the left, each raised to their individual stoichiometries), as this method will, but will retain a balance of stoichiometry, with each reactant and product changing unless their stoichiometries are balanced. The user defined regulatory reactions, on the other hand, will not generate any ODE terms for the reactants on the left-hand side of the arrow.

3.3 Indexed Species

An array index may be attached to a species variable using parenthesis as a delimiter, as in the following example:

```
[K(3,0) <=> K(3,1), mod[RAFK, RAFph], rates[a1,d1,k1,0,a2,d2,k2,0]]
```

The index may also be applied to a modifier variable, and within the initial condition section of the model file, e.g.,

```
K[3,0] = 0.7
```

See the examples chapter (MAPK cascade with indexed stages) for an example.

Implementation Note: When instantiated in the `Reaction` class, the indices are expanded into the variable name, and separated with underscores e.g., `K[i, j]` becomes `K_i_j`; this variables are implemented in this expanded form in the solver and displayed this way on plots.

3.4 The Nil and EmptySet Species

The species `Nil` is used to represent the vacuum or the empty set. No ODE term is ever generated for the species `Nil`.

The variable `EmptySet` is predefined in Sympy and should not be used in models. If it is used in an SBML file it will be converted to `Nil`.

For example, the reaction

```
[Nil->A, k]
```

denotes creation and represents a special case of mass-action reaction, in which $A' = k$ (rather than $A' = k \times [\text{Nil}]$ as it would be in any other reaction).

Similarly, the reaction

```
[A->Nil, k]
```

represents destruction, and is treated normally for the species `A`, but no differential equation term is generated for `Nil`.

3.5 Mass Action Arrows

3.5.1 Simple Mass Action

pycellerator Arrow:

```
[X -> Y, k]}}
```

`X`, `Y` are identifiers representing chemical species; `k` is either an identifier or a number representing a rate constant.

Equivalent xlr8r Arrow:

$$\{X \rightarrow Y, k\}$$

Typical biochemical notation:

$$X \xrightarrow{k} Y$$

Interpreted differential equation terms:

$$\frac{dY}{dt} = -\frac{dX}{dt} = kX$$

3.5.2 Simple Mass Action, with stoichiometry

pycellerator Arrows:

```
[e1 X1 + e2 X2 + ... -> s1 Y1 + s2 Y2 +... , k]
[e1*X1 + e2*X2 + ... -> s1*Y1 + s2*Y2 +... , k]
```

Species: X1, X2, ..., Y1, Y2, ... are identifiers representing chemical species.

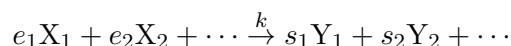
Stoichiometry: e1, e2, ..., s1, s2, ... are either identifiers or numbers representing stoichiometries before and after the reaction. The multiplication symbols (*) are optional.

If any stoichiometry is an identifier, there must be either an * or a blank space between it and the corresponding species identifier. If the stoichiometry is numerical the blank space is optional. For example 3X, 3*X, and 3 X, will all be treated as a species X with stoichiometry 3, and s H2O and s*H2O will both be interpreted as a species H2O with stoichiometry s, whereas sH2O will be interpreted as a single species sH2O with a stoichiometry of 1.

Equivalent x1r8r Arrow:

$$\{e_1 X_1 + e_2 X_2 + \dots \rightarrow s_1 Y_1 + s_2 Y_2 + \dots, k\}$$

Typical biochemical notation:



Interpreted differential equation terms:

$$\frac{dU_i}{dt} = (s_{i,r} - s_{i,l}) k X_1^{e_1} X_2^{e_2} \dots$$

where $s_{i,r}$ and $s_{i,l}$ are the stoichiometry of U_i on the right-hand-side and left-hand-side of the reaction, respectively, and U_i refers to any species either on either side of the equation (either an X_i or a Y_i). The product is taken over all the terms on the left-hand-side of the reaction, each term raised to its respective stoichiometry (this is also known as the law of mass action).

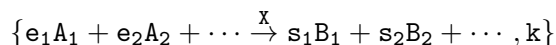
3.5.3 Simple Mass Action, Catalyzed

pycellerator Arrows:

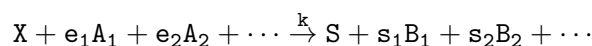
```
[e1 A1 + e2 A2 + ... --> s1 B1 + s2 B2 + ..., mod[X], k]
[e1*A1 + e2*A2 + ... --> s1*B1 + s2*B2 + ..., mod[X], k]
```

The stoichiometries are optional. Multiplication symbols (*) or spaces are optional if numerical stoichiometries are used, but are required if symbolic stoichiometries are used.

Equivalent x1r8r Arrow:



Typical biochemical notation:



Interpreted differential equation terms: The reaction is expanded into a single reaction of the form

```
[X + e1 A1 + e2 A2 + ... -> X + s1 B1 + s2 B2 + ..., mod[X], k]
```

as described in section 3.5.2.

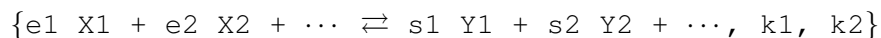
3.5.4 Reversible Mass Action

pycellerator Arrow:

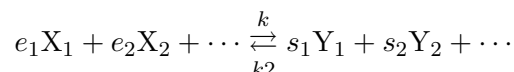
```
[e1 X1 + e2 X2 + ... <-> s1 Y1 + s2 Y2 + ... , rates[k1,k2]]
```

The stoichiometries are optional.

Equivalent xlr8r Arrow:



Typical biochemical notation:



Interpreted differential equation terms:

This arrow is reduced to the pair of simple mass action equations, as described in section 3.5.2:

```
[e1 X1 + e2 X2 + ... -> s1 Y1 + s2 Y2 + ... , k1]
[s1 Y1 + s2 Y2 + ... -> e1 X1 + e2 X2 + ... , k2]
```

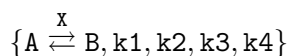
The contributions to the differential equations are then computed as described in section 3.5.2.

3.5.5 Catalytic Mass Action, Intermediate Substrate/Catalyst Complex Formation

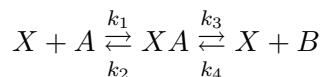
pycellerator Arrow:

```
[A => B, mod[X], rates[k1, k2, k3, k4]]
```

Equivalent xlr8r Arrow:



Typical biochemical notation:



Typically k_4 is omitted, in which case it is assumed to be zero. If fewer than four parameters are given they are zero-filled to the right.

This arrow is expanded into the following collection of simple mass action arrows as described in section 3.5.2:

```
[X + A -> A_X, k1]
[A_X -> X + A, k2]
[A_X -> X + B, k3]
[X + B -> B_X, k4]
```

The name of the intermediate complex is automatically generated by concatenating the names of the substrate and the catalyst with an underscore. The resulting contributions to the differential equations for $[A]$, $[B]$, $[X]$ and $[XA]$ in the above scheme are then:

$$\begin{aligned}
[A]' &= k_2[A_X] - k_1[A][X] \\
[B]' &= k_3[A_X] - k_4[B][X] \\
[X]' &= -k_1[A][X] + (k_2 + k_3)[A_X] - k_4[B][X] \\
[A_X]' &= k_1[A][X] - (k_2 + k_3)[A_X] + k_4[B][X]
\end{aligned}$$

3.5.6 Catalytic Mass Action, Intermediate Substrate/Catalyst Complex Formation, Reversible

pycellerator Arrow:

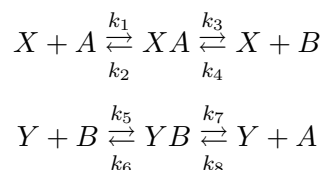
```
[A <=> B, mod[X, Y], rates[k1, k2, k3, k4, k5, k6, k7, k8]]
```

If fewer than 8 rate constants are given they are zero-filled to the right.

Equivalent xlr8r Arrow:

$$\{A \xrightleftharpoons[X]{X} B, k1, k2, k3, k4, k5, k6, k7, k8\}$$

Typical biochemical notation:



When computing the differential equation contribution, this arrow is expanded into the following pair of catalytic mass action reactions with intermediate complex formation, as described in section 3.5.5:

```
[A => B, mod[X], rates[k1, k2, k3, k4]]
[B => A, mod[Y], rates[k5, k6, k7, k8]]
```

These reactions are subsequently broken down in simple mass action reactions as described in section 3.5.2.

```
[A+X->A_X, k1] }
[A_X->A+X, k2] }
[A_X->B+X, k3] }
[B+X->A_X, k4] }
[B+Y->B_Y, k5] }
[B_Y->B+Y, k6] }
[B_Y->A+Y, k7] }
[A+Y->B_Y, k8] }
```

The contributions to the ODE are computed as described based on these reduced reactions, as described in 3.5.2:

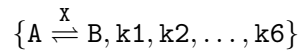
$$\begin{aligned}
[A]' &= k_2[A_X] + k_7[B_Y] - A * X * k1 - A * Y * k8 \\
[B]' &= k_3[A_X] + k_6[B_Y] - B * X * k4 - B * Y * k5 \\
[A_X]' &= -(k_2 + k_3)[A_X] + k_1[A][X] + k_4[B][X] \\
[B_Y]' &= -(k_6 + k_7)[B_Y] + k_8[A][Y] + k_5[B][Y] \\
[Y]' &= (k_6 + k_7)[B_Y] - k_8[A][Y] - k_5[B][Y] \\
[X]' &= (k_2 + k_3)[A_X] - k_8[A][X] - k_4[B][X]
\end{aligned}$$

3.5.7 Catalytic Mass Action with Substrate/Enzyme and Product/Enzyme Intermediate Complexes

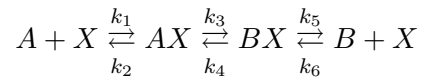
pycellerator Arrow:

```
[A :=> B, mod[X], rates[k1, k2, k3, k4, k5, k6]]
```

Equivalent xlr8r Arrow:



Typical biochemical notation:



Interpreted differential equation terms: This arrow is expanded into the following collection of simple mass action arrows as described in section 3.5.2:

```
[A+X->A_X, k1]
[A_X->A+X, k2]
[A_X->B_X, k3]
[B_X->A_X, k4]
[B_X->B+X, k5]
[B+X->B_X, k6]
```

These are then converted into terms in the differential equations as described above in 3.5.2:

$$\begin{aligned} [A]' &= k_2[A_X] - k_1[A][X] \\ [X]' &= k_2[A_X] + k_5[B_X] - k_1[A][X] - k_6[B][X] \\ [B]' &= k_5[B_X] - k_6[B][X] \\ [B_X]' &= k_3[A_X] - (k_4 + k_5)[B_X] + k_6[B][X] \\ [A_X]' &= k_4[B_X] - (k_2 + k_3)[A_X] + k_1[A][X] \end{aligned}$$

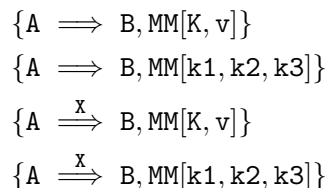
3.6 Equilibrium / Steady-State Models

3.6.1 Michaelis-Menten-Henri (MMH) Reactions

pycellerator Arrows:

```
[A :-> B, MMH[K, v]]
[A :-> B, MMH[k1, k2, k3]]
[A :-> B, mod[X], MMH[K, v]]
[A :-> B, mod[X], MMH[k1, k2, k3]]
```

Equivalent xlr8r Arrow:



Typical Biochemical Notation:

Non-standard.

Interpreted Differential Equation:

$$[B]' = -[A]' = \frac{v[A]}{K + [A]} \quad \text{for first form}$$

$$[B]' = -[A]' = \frac{k_3[A]}{(k_2 + k_3)/k_1 + [A]} \quad \text{for second form}$$

$$[B]' = -[A]' = \frac{v[A][X]}{K + [A]} \quad \text{for third form}$$

$$[B]' = -[A]' = \frac{k_3[A][X]}{(k_2 + k_3)/k_1 + [A]} \quad \text{for fourth form}$$

3.6.2 Monod-Wyman-Changeaux (MWC) Reactions

pycellerator Arrows:

1. Basic MWC:

```
[S==>P,mod[E],MWC[k,n,c,L,K]]
```

2. MWC with additional activators and inhibitors:

```
[[S1,S2,...]==>P,mod[E,[A1,A2,...],[I1,I2,...]],MWC[k,n,c,L},{K1}]
```

where **K1** = **[[KS1,KS2,...],[KA1,KA1,...],[KI1,KI2,...]]**.

3. MWC with Competitive inhibition.

```
[[S1,S2,...]==>P], mod[E,[A1,A2,...],[I1,I2,...],X ], MWC[k,n,c,L},{K2 }]
```

where **x** has the form

```
[[[CS11, CS12,...],      # S1 Competitive Inhibitors
  [CS21, CS22,...],...] # S2 Competitive Inhibitors
 [[CA11, CA11,...],      # A1 Competitive Inhibitors
  [CA21, CA22,...],...] # A2 Competitive Inhibitors
```

and **K2**= **[[KS1, KS2,...], [KA1, KA2, ...], [KI1, KI2, ...], [KCS11, CS12, ...], [KCS21, KCS22, ...], [KCA11, KCA12, ...], [KCA21, KCA22, ...]]**.

Equivalent xlr8r Arrow:

$$\{S \xrightarrow{\text{Enz}} P, \text{MWC}[k, n, c, L, K]\} \quad (\text{for (1)})$$

$$\left\{ \frac{S \xrightarrow{\text{Enz}} P}{\{\{A1, A2, \dots\}, \{I1, I2, \dots\}\}}, \text{MWC}[k, n, c, L, K, \dots] \right\} \quad (\text{for (2)})$$

$$\left\{ \frac{S \xrightarrow{\text{Enz}} P}{\{\{A1, A2, \dots\}, \{I1, I2, \dots\}, \{CS1, \dots\}, \dots, \{CA1, \dots\}, \dots\}}, \text{MWC}[k, n, c, L, K, \dots] \right\} \quad (\text{for (3)})$$

Typical Biochemical Notation:

Not standardized.

Interpreted Differential Equation:

The ODE terms are based on the theory of [12]. Let $c = [S]/K$. Then

$$[P]' = -[S]' = [E] \frac{s(1+s)^{n-1} + Lsc(1+sc)^{n-1}}{(1+s)^n + L(1+sc)^{n-1}} \quad \text{for form (1)}$$

The terms for forms (2) and (3) are described in [13]. For the first arrow, let

$$s_j = \frac{[S]_j}{K_{Sj}}, a_j = \frac{[A]_j}{K_{Aj}}, i_j = \frac{[I]_j}{K_{Ij}}$$

Then

$$[P]' = -[S]' = [E] \frac{\prod(1+a_j)^n \prod s_j \prod(1+s_j)^{n-1} + L \prod(1+i_j)^n \prod(cs_j) \prod(1+cs_j)^{n-1}}{\prod(1+a_j)^n \prod(1+s_j)^n + L \prod(1+i_j)^n \prod(1+cs_j)^{n-1}} \quad \text{for form (2)}$$

For form (3) we can also normalize the competitive inhibitors as

$$\overline{s_j} = c \sum_k \frac{[CS]_{jk}}{K_{CSjk}}, \quad \overline{a_j} = c \sum_k \frac{[CA]_{jk}}{K_{CAjk}}$$

and then

$$\begin{aligned} [P]' &= -[S]' \\ &= [E] \frac{\prod(1+a_j\overline{a_j})^n \prod s_j \prod(1+s_j+\overline{s_j})^{n-1} + L \prod(1+i_j)^n \prod(cs_j) \prod(1+cs_j+\overline{s_j})^{n-1}}{\prod(1+a_j+\overline{a_j})^n \prod(1+s_j)^n + L \prod(1+i_j)^n \prod(1+cs_j+\overline{s_j})^{n-1}} \quad \text{for form(3)} \end{aligned}$$

3.6.3 Non-regulatory Hill Functions

These are described below in section 3.7.1.

3.7 Regulatory Arrows

3.7.1 Hill Functions

pycellerator Arrows:

There are three Hill function forms.

<code>[A -> B, Hill[v, n, K, a, T]]</code>	<code># Hill (1)</code>
<code>[[P,Q,...] -> R, Hill[v, n, K, a, [TP,TQ,...]]]</code>	<code># Hill (2)</code>
<code>[[X,Y,Z,...] --> U, mod[E], Hill[v, n, K, a, [TX,TY, TZ,...]]]</code>	<code># Hill (3)</code>

Note that the third form (Hill(3)) is not actually a regulatory arrow, as each of the species on the left is also affected by the arrow.

Equivalent xlr8r Arrows:

$$\begin{aligned} \{A \mapsto B, \text{Hill}[v, n, K, a, T]\} & \quad \text{for Hill (1)} \\ \{\{P, Q, \dots\} \mapsto R, \text{Hill}[v, n, K, a, \{TP, TQ, \dots\}]\} & \quad \text{for Hill (2)} \\ \{\{X, Y, Z, \dots\} \xrightarrow{E} U, \text{Hill}[v, n, K, a, \{TX, TY, TZ, \dots\}]\} & \quad \text{for Hill (3)} \end{aligned}$$

Typical Biochemical Notation:

Not standardized.

Interpreted Differential Equation:

For (Hill (1)) and (Hill (2)) only the amount of the product changes, and not the amount of any of the reactants on the left-hand-side of the reaction:

$$[B]' = \frac{v(a + T[A])^n}{K^n + (a + T[A])^n} \quad \text{for Hill (1)} \quad (3.1)$$

$$[R]' = \frac{v(a + T_P[P] + T_Q[Q] + \dots)^n}{K^n + (a + T_P[P] + T_Q[Q] + \dots)^n} \quad \text{for Hill (2)} \quad (3.2)$$

$$[U]' = \frac{v[E](a + T_X[X] + T_Y[Y] + \dots)^n}{K^n + (a + T_X[X] + T_Y[Y] + \dots)^n} \quad \text{for Hill (3)} \quad (3.3)$$

For the third form (Hill (3)), the products on the left all change also,

$$[X]' = [Y]' = \dots = -\frac{v[E](a + T_X[X] + T_Y[Y] + \dots)^n}{K^n + (a + T_X[X] + T_Y[Y] + \dots)^n} \quad \text{for Hill (3) only} \quad (3.4)$$

Difference from xlr8r: In xlr8r the combination of arrows

$$\begin{aligned} & \{ \{A1 \mapsto B, \text{Hill}[v, n, K, a, T1] \}, \\ & \{A2 \mapsto B, \text{Hill}[v, n, K, a, T2] \}, \\ & \{A3 \mapsto B, \text{Hill}[v, n, K, a, T3] \}, \dots \} \end{aligned} \quad (3.5)$$

will be given ODE terms

$$[B]' = \frac{v(a + T_1[A1] + T_2[A2] + T_3[A3] + \dots)^n}{K^n + (a + T_1[A1] + T_2[A2] + T_3[A3] + \dots)^n} \quad (3.6)$$

whereas in pycellerator it will be interpreted as:

$$[B]' = \frac{v(a + T_1[A1])^n}{K^n + (a + T_1[A1])^n} + \frac{v(a + T_2[A2])^n}{K^n + (a + T_2[A2])^n} + \frac{v(a + T_3[A3])^n}{K^n + (a + T_3[A3])^n} \quad (3.7)$$

To get an interpretation of the form (3.6) one must use the arrow form Hill (2). Unfortunately, it is not possible to represent equations of the form given by (3.7) in xlr8r, even though such combinations are frequently used in the modeling literature, unless the user explicitly coded the equation as part of the rate constant. This is the motivation for the change in pycellerator.

3.7.2 GRN Arrows

pycellerator Arrows:

There are three forms of GRN arrows in pycellerator.

<code>[A -> B, GRN[v, T, n, h]]</code>	<code># GRN (1)</code>
<code>[[P,Q,...] -> R, GRN[v, [TP,TQ,...], n, h]]</code>	<code># GRN (2)</code>
<code>[[X,Y,...] --> U, mod[E], GRN[v, [TX,TY,...], n, h]]</code>	<code># GRN (3)</code>

Equivalent xlr8r Arrow:

$$\begin{aligned} \{A \mapsto B, \text{GRN}[v, T, n, h]\} & \quad \text{for GRN (1)} \\ \{\{P, Q, \dots\} \mapsto R, \text{GRN}[v, \{TP, TQ, \dots\}, n, h]\} & \quad \text{for GRN (2)} \\ \{\{X, Y, \dots\} \xrightarrow{E} U, \text{GRN}[v, \{TX, TY, \dots\}, n, h]\} & \quad \text{for GRN (3)} \end{aligned}$$

Note that the third form of this arrow, GRN (3) is not actually implemented in xlr8r, only in pycellerator.

Typical Biochemical Notation:

Not standardized.

Interpreted Differential Equation:

$$[B]' = \frac{v}{1 + \exp(-h - T[A]^n)} \quad \text{for GRN (1)} \quad (3.8)$$

$$[R]' = \frac{v}{1 + \exp(-h - T_P[P]^n - T_Q[Q]^n + \dots)} \quad \text{for GRN (2)} \quad (3.9)$$

$$[U]' = \frac{v}{1 + \exp(-h - T_X[X]^n - T_Y[Y]^n + \dots)} \quad \text{for GRN(3)} \quad (3.10)$$

Difference from xlr8r: In xlr8r the combination of arrows

$$\begin{aligned} & \{\{A1 \mapsto B, \text{GRN}[v, T1, n, h]\}, \\ & \{A2 \mapsto B, \text{GRN}[v, T2, n, h]\}, \\ & \{A3 \mapsto B, \text{GRN}[v, T2, n, h]\}, \dots \} \end{aligned} \quad (3.11)$$

will be given ODE terms

$$[B]' = \frac{v}{1 + \exp(-h - T_1[A1]^n - T_2[A2]^n - \dots)} \quad (3.12)$$

whereas in pycellerator it will be given the ODE

$$[B]' = \frac{v}{1 + \exp(-h - T_1[A1]^n)} + \frac{v}{1 + \exp(-h - T_2[A2]^n)} + \dots$$

The version given by equation (3.12) can still be obtained by using the arrow form $[[A1, A2, \dots] | \text{-->} B]$.

3.7.3 S-Systems

pycellerator Arrows:

There are two forms of S-System arrows in pycellerator:

<code>[S -> P, SSystem[tau, a, b, g, h]]</code>	<code># S-System (1)</code>
<code>[[S1, S2, ...] -> P, SSystem[tau, a, b, [g1, g2, ...], [h1, h2, ...]]</code>	<code># S-System (2)</code>

Equivalent xlr8r Arrow:

$$\begin{aligned} \{S \mapsto P, \text{SSystem}[\tau, a, b, g, h]\} & \quad \text{for S-System (1)} \\ \{S1, S2, \dots \mapsto P, \text{SSystem}[\tau, a, b, \{g1, g2, \dots\}, \{h1, h2, \dots\}]\} & \quad \text{for S-System (2)} \end{aligned}$$

Typical Biochemical Notation:

Not standardized.

Interpreted Differential Equation:

$$[P]' = \frac{1}{\tau} (a[S]^g - b[S]^h) \quad \text{for S-System(1)} \quad (3.13)$$

$$[P]' = \frac{1}{\tau} (a \prod_i [S]_i^{g_i} - b \prod_i [S]_i^{h_i}) \quad \text{for S-System (2)} \quad (3.14)$$

S-System reactions are numerically unstable when any $[S]_i \rightarrow 0$ if the corresponding $g_i < 0$, because this would lead to a division by zero condition. The numerical “trick” that is used to fix this is to set concentrations to a small number ϵ rather than 0 in this case. The default value for ϵ is 10^{-37} but may be reset in the solver module with the keyword `-epsilon value`.

This reaction is described in detail in [14, 15].

3.7.4 NHCA

pycellerator Arrows:

There are two pycellerator Arrows for non-hierarchical cooperative activation.

<code>[A -> B, mod[X], NHCA[v, TP, TM, n, m, k]]</code>	<code># NHCA1</code>
<code>[[A1, A2, ...] -> B, mod[X], NHCA[v, [TP1, ...], [TM1, ...], [n1, ...], m, k]]</code>	<code># NHCA2</code>

Equivalent xlr8r Arrow:

$\{A \xrightarrow{X} B, \text{NHCA}[v, \{TP, TM\}, n, m, k]\}$ for NHCA1

$\{\{A1, A2, \dots\} \xrightarrow{X} B, \text{NHCA}[v, \{\{TP1, TP2, \dots\}, \{TM1, TM2, \dots\}\}, \{n1, n2, \dots\}, m, k]\}$ for NHCA2

Note that the second form, (NHCA2), is not actually implemented in xlr8r, only in pycellerator.

Typical Biochemical Notation:

Not standardized.

Interpreted Differential Equation:

This reaction is described in more detail in [17].

$$[B]' = v[E] \frac{(1 + T_P [A]^n)^m}{k(1 + T_M [A]^n)^m + (1 + T_P [A]^n)^m} \quad \text{for NHCA1}$$

$$[B]' = v[E] \frac{\prod_i (1 + T_{Pi} [A]_i^{n_i})^m}{k \prod_i (1 + T_{Mi} [A]_i^{n_i})^m + \prod_i (1 + T_{Pi} [A]_i^{n_i})^m} \quad \text{for NHCA2}$$

3.7.5 Rational Functions

pycellerator Arrows:

<code>[[[X1, X2, ...], [Y1, Y2, ...]] ==> Z,</code>	
<code>rational[[a0, a1, a2, ...],</code>	<code># coefficients of numerator</code>
<code>[d0, d1, d2, ...],</code>	<code># coefficients of denominator</code>
<code>[m0, m1, m2, ...],</code>	<code># exponents, numerator</code>
<code>[n0, n1, n2, ...]]]</code>	<code># exponents, denominator</code>

As with `xlr8r`, any of the input species may represent a product, for example,

```
[[[O*S,O*S*N], [O,O*S,O*S*N,O*G]] ==>N,
  rational[[e0,e1,e2],[1,f0,f1,f2,f3],[],[[]]]]
```

The asterisks must be specified between the species if the product-form is used.

Equivalent `xlr8r` Arrow:

$$\{\{\{X1, X2, \dots\}, \{Y1, Y2, \dots\}\}\} \Rightarrow Z, \text{ rational}\{\{a0, a1, a2, \dots\}, \\ \{d0, d1, d2, \dots\}, \\ \{m1, m2, \dots\}, \\ \{n1, n2, \dots\}\}$$

Typical Biochemical Notation:

Not standardized.

Interpreted Differential Equation: For the input reaction

```
[[[X1,X2,X3,...], [Y1,Y2,...]]==>Z,
  rational[[a0,a1,a2,a3,...], [d0,d1,d2,d3,...],
           [m0,m1,m2,m3,...], [n0,n1,n2,n3,...]]]
```

the ODE term produced is:

$$[Z]' = \frac{a_0^{m_0} + a_1[X1]^{m_1} + a_2[X2]^{m_2} + a_3[X3]^{m_3} + \dots}{d_0^{n_0} + d_1[Y1]^{n_1} + d_2[Y2]^{n_2} + d_3[Y3]^{n_3} + \dots}$$

Difference From `xlr8r`: In `pycellerator` the first value in each list of exponents is applied to the leading constants, rather than to the first parameter, so that `pycellerator` expects the same number of exponents as coefficients. In `xlr8r` there is no way to specify an exponent on the first coefficient, and `xlr8r` expects one fewer exponents than coefficients. Thus `xlr8r` would ignore the extra exponents and interpret the above reaction as follows:

$$[Z]' = \frac{a_0 + a_1[X1]^{m_0} + a_2[X2]^{m_1} + a_3[X3]^{m_2} + \dots}{d_0 + d_1[Y1]^{n_0} + d_2[Y2]^{n_1} + d_3[Y3]^{n_2} + \dots}$$

3.7.6 User Defined Regulatory Arrows

`pycellerator` Arrows:

There are several different forms. In the first form, there is a single reactant forming a single product.

```
[A -> B, USER[v, T, n, h, f]] # USER (1)
```

In the second form, multiple reactants interact to produce a single product vectorally.

```
[[P1,P2,...]|-> Q, USER[v, T, n, h, f]] # USER (2)
```

In the third form, a modifier can be specified.

```
[[X1,X2,...]|--> Y, mod[E], USER[v, T, n, h, f]] # USER (3)
```

In USER(2) and USER (3) the expressions for **T** and **n** may be either single symbols, numbers, or lists of length up to the length of the number of reactants on the left-hand side of the arrow. If there are fewer values, the list will be extended with the final value provided (the rightmost value).

In each case, the function f must be a valid Python lambda expression of a single variable, enclosed in quotation marks. For example,

```
[[P,Q] |-> R, USER[v, T, n, h, "lambda x: 1/(1+exp(-x))"]]]
```

represents a USER equivalent version of the standard GRN reaction.

Equivalent xlr8r Arrow:

$$\begin{aligned} \{A \mapsto B, \text{USER}[v, T, n, h, f]\} & \quad \text{for USER (1)} \\ \{\{P_1, P_2, \dots\} \mapsto Q, \text{USER}[v, T, n, h, f]\} & \quad \text{for USER (2)} \end{aligned}$$

The equivalent of example in xlr8r would be

$$\{\{P, Q\} \mapsto R, \text{USER}[v, T, n, h, 1/(1+\text{Exp}[\#])\&]\}$$

In xlr8r there is no equivalent reaction for USER (3); it is new in pycellerator.

Typical Biochemical Notation:

Not standardized.

Interpreted Differential Equation: For the input reactions USER (1) through USER (3),

$$\begin{aligned} [B]' &= v f(h - T[A]^n) & \text{for USER (1)} \\ [Q]' &= v f(h - T_1[P_1]^{n_1} - T_2[P_2]^{n_2} - \dots) & \text{for USER (2)} \\ [Y]' &= v [E] f(h - T_1[X_1]^{n_1} - T_2[X_2]^{n_2} - \dots) & \text{for USER (3)} \end{aligned}$$

Difference From xlr8r:

In pycellerator the head of the rate list must be the word USER; in xlr8r it may be any symbol not otherwise assigned.

In pycellerator the function f must be a Python lambda expression. In xlr8r, while a Mathematica lambda expression (otherwise known as a pure function, such as $1/(1+\text{Exp}[\#])\&$) is permitted but it is not required, and the name of any previously defined function may be substituted.

3.8 User Defined Stoichiometric Arrows

pycellerator Arrows:

```
[s1*A1+s2*A2+... -> q1*A1+q2*A2+..., using["expression"]]
```

where the s_1, s_2, \dots and q_1, q_2, \dots are (possibly zero) stoichiometries and "expression" is a valid mathematical expression expressed in syntactically correct python. It must be enclosed in quotation marks. The asterisks between the stoichiometries and the species in the reaction are optional.

Interpreted Differential Equation:

$$\frac{d[X]}{dt} = (q_i - s_i) \times (\text{expression}) \quad (3.15)$$

For example:

```
[B + 3S -> 4P + 5S, using["A*B**3"]]
```

returns the collection of ODE terms:

$$\begin{aligned} [P]' &= 4[A][B]^3 \\ [S]' &= 2[A][B]^3 \\ [B]' &= -[A][B]^3 \end{aligned}$$

Compare with User Defined Regulatory Arrows (Section 3.7.6) and Equations as Rate Constants (Section 3.2), which are similar but produce slightly different results.

3.9 Cascades

Any mass action, catalytic mass action, MMH, Hill Function, GRN, S-System, or NHCA reaction can be written in a cascade as a single reaction. A **cascade** is defined as sequence of repeated reactions with the same arrow and the same rate constants. For example, the reactions

```
[A => B, mod[E], rates[k1,k2,k3]]
[B => C, mod[F], rates[k1,k2,k3]]
[C => D, mod[G], rates[k1,k2,k3]]
```

can be written as a single reaction cascade:

```
[A => B => C => D, mod[E, F, G], rates[k1,k2,k3]]
```

Reactions without modifies can also be written as cascades:

```
[P :-> Q :-> R, MMH[KD, v]]
```

which represents the pair of reactions

```
[P :-> Q, MMH[KD, v]]
[Q :-> R, MMH[KD, v]]
```

Note that if different rate constants are required at different states in the cascade, the reactions must be written separately, and not as part of a cascade.

Different types of arrows cannot be combined together, thus a cascade cannot be written as $A \rightarrow B \mid \rightarrow C$ but must be written as two separate reactions.

An example of a model with a cascade is given by the MAP-Kinase model with oscillations in section [5.1](#).

3.10 Flux Arrows

Reactions that represent fluxes are a fundamentally different type of entity than reactions used in kinetic models, as described in the previous sections. This is because Flux reactions do not (necessarily) have a rate law (or ODE) associated with them, although they normally have a total rate, given by product of a velocity and a stoichiometry.

Normally a model will be composed **either** entirely of kinetic arrows **or** entirely of flux arrows. In the present implementation of `pycellerator` one is not allowed to combine the two in a model. The Mathematica implementation of `xlr8r` does not support Flux reactions at all (at the current time).

The format of a flux arrow is

$$[\text{lhs} \rightarrow \text{rhs}, \text{Flux}[\text{low} < \text{id} < \text{up}, \text{obj}, \text{flux}]] \quad (3.16)$$

where *lhs* and *rhs* are stoichiometric expressions such as $3A + B$ or X , and the other terms are defined in the following table:

pycellerator	COBRA SBML Variable	Description
<i>low</i>	LOWER_BOUND	numerical lower bound or <code>-inf</code> if unbounded
<i>up</i>	UPPER_BOUND	numerical upper bound or <code>inf</code> if unbounded
<i>id</i>	SBML reaction Id	variable name used to refer to reaction flux
<i>obj</i>	OBJECTIVE_COEFFICIENT	Used to determine objective function component of \mathbf{f} vector.
<i>flux</i>	FLUX_VALUE	Value to assign to flux. Not required to perform FBA optimization. Sometimes used for SBML L2.4 kinetic law.

An example is

```
[ES -> E + S, Flux[0 < v < 1, 1, 0]]
```

Note that the “less-than” sign must be used, though the constraint interval is typically closed, and not open. In the case of equality, a constraint of the form

$$1 < v < 1 \tag{3.17}$$

would be used to indicate a constraint meaning $v = 1$.

Flux optimization will solve the linear programming problem

$$\text{maximize } \mathbf{v}^T \mathbf{f} \tag{3.18}$$

$$\text{subject to } \mathbf{N} \mathbf{v} = \mathbf{0} \tag{3.19}$$

$$\text{and } low_1 < v_1 < up_1 \tag{3.20}$$

$$\text{and } low_2 < v_2 < up_2 \tag{3.21}$$

$$\vdots$$

where \mathbf{v} is the vector of fluxes $(v_1, v_2, \dots)^T$, \mathbf{N} is the stoichiometry matrix, and \mathbf{f} is the vector of objective coefficients.

Chapter 4

pycellerator Function Reference

Conventions used in this chapter:

For the command line syntax:

`typewriter font` is used to represent required input exactly as it should be typed

italicized Roman font is used to represent input that should be replaced with a value, such as the name of an input file.

Items enclosed in square brackets, e.g., [`like this`] are optional. This remark only applies to expressions typed on the command line (e.g., the terminal in linux or MacOS, or at the command prompt in Windows). It **does not apply** to reactions.

The \$ is used to indicate the command prompt in several examples.

Reaction Syntax:

Examples of reactions, such as those that should be written in input files, are written in `typewriter font like this`. Note that reaction syntax requires the use of square brackets, and the contents of square brackets are not optional in this case.

4.1 Command Line Interface

The generic command line syntax is

```
$ python pycellerator.py keyword [options]
```

where **keyword** is one of **EXPAND**, **INTERPRET**, **CONVERT**, **PARSE**, **SOLVE**, **SBML**, **FLUX** and **VERSION**, and [options] is an optional sequence of command line options that is difference for each keyword. The case (upper or lower) of the tt keyword does not matter. The syntax and options for each keyword are discussed in the following sections.

4.2 parser

The parser converts from text-based arrow notation into an internal python class (data structure) called a `Reaction`.

Command Line Syntax:

```
$ python pycellerator parse -in \textit{filename} [-dump] [-trace]}
```

```
$ python parser.py -in \textit{filename} [-dump] [-trace]}
```

The `in filename` specifies the input file name, which contains a list of text reactions. An example is given by the following:

```
[X <=> XP,    mod[Z, ZPP], rates[a, d, k, 0, a, d, k]] # first reaction
[XP <=> XPP,   mod[Z, ZPP], rates[a, d, k, 0, a, d, k]] # second reaction
[Y <=> YP,     mod[X, XPP], rates[a, d, k, 0, a, d, k]]
[YP <=> YPP,   mod[X, XPP], rates[a, d, k, 0, a, d, k]]
[Z <=> ZP,     mod[Y, YPP], rates[a, d, k, 0, a, d, k]]
[ZP <=> ZPP,   mod[Y, YPP], rates[a, d, k, 0, a, d, k]]
```

One reaction may be placed per line; additional white space is ignored. The `#` character indicates a comment; anything written to the right of this character will be ignored.

If the `-trace` option is used, the text reaction will be written to the screen after each reaction is successfully read and parsed.

If the `-dump` option is used, the `Dump()` method will be applied (and written to the screen) for each reaction after it is processed, whether the reaction is successfully parsed or not.

Command Line in Shell:

```
>>> import pycellerator
>>> pycellerator.run("parse -in filename [-dump] [-trace]")
```

API (functions):

`parser(*keywords)`

input: (keyword arguments only)
return value: list of `Reactionss`

Optional keyword arguments:

`inputfile=""` Name of input file containing reactions.
`trace=False` Turns on tracing, same as command line `-trace`

`ParseArrow(r)`

input: `r`: String reaction in pycellerator format.
return value: Reaction object representing the input text reaction. Operationally, there is no difference between invoking `ParseArrow(r)` and `Reaction(r)`; the difference is that one invokes the class directly and the other does it via a functional interface.

API (class):`Reaction(r)`**input:** `r`: String reaction in pycellerator format.**Methods:**

`r.Input()` value of the original text reaction
`r.Parsed()` output of the `pyparsing` module
`r.Number()` number of sequential calls to parser
`r.Length()` length of the `pyparsing` object
`r.ArrowType()` Cellerator arrow type as a string
`r.Arrow()` Actual arrow as a string
`r.LHS()` List of reactants
`r.RHS()` List of products
`r.LH_STOIC()` Stoichiometry of reactants (mass action only)
`r.RH_STOIC()` Stoichiometry of products (mass action only)
`r.MHS()` List of modifiers
`r.Rates()` List of rate constants or parameters in reaction
`r.Dump()` Produce a formatted dump of the `Reaction` class object

Comparison of `Arrow()` and `ArrowType()`. See previous chapter for descriptions of corresponding reactions.

<code>r.Arrow()</code>	<code>r.ArrowType()</code>
"->"	"Mass Action"
"-->"	"Mass Action"
"<->"	"Mass Action"
"=>"	"Mass Action"
"<=>"	"Mass Action"
":->"	"MMH"
":-->"	"MMH"
":=>"	"Mass Action"
" ->"	"GRN"
" -->"	"GRN"
"==>"	"Multiuse-Arrow"

Examples.1. Input with `-trace`

```

$more catalytic.dat
[X => Y, mod[E], rates[k1,k2,k3]]
[Y => X, mod[F], rates[k4,k5,k6]]
$python pycellerator.py -in catalytic.dat -trace
py[cellerator]: parser (2015-03-02 17:58:18)
input: [X => Y, mod[E], rates[k1,k2,k3]]
input: [Y => X, mod[F], rates[k4,k5,k6]]
[<__main__.Reaction object at 0x7f0442369a50>,
 <__main__.Reaction object at 0xe47dd0>]

```

2. Input with -dump

```

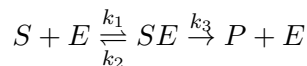
$cat sto.dat
[ 3A + 4B -> P + Q + R,  k]
$ python pyx.py -in sto.dat -dump
py[xlr8r]: parser (2012-03-02 18:08:54)
===== Reaction Number 1 =====
Input:      [ 3A + 4B -> P + Q + R,  k]
Parsed:      [(['3', 'A', '4', 'B'], {}), '->',
              (['P', 'Q', 'R'], {}), (['k'], {})]
Arrow:      ->
ArrowType:   Mass Action
LHS:         ['A', 'B']
LH_STOIC:    ['3', '4']
RHS:         ['P', 'Q', 'R']
RH_STOIC:    ['1', '1', '1']
modifiers:   []
rates:       ['k']
[<__main__.Reaction object at 0x7f6b1c3d2c90>]

```

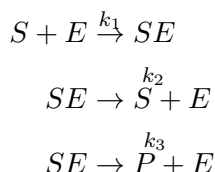
4.3 expander

Normally there is no need for the user to call the expander directly.

The expander module expands complex py[xlr8r] reactions into their constituent parts; for example, the catalytic reaction



is broken down into its constituent reactions



The input to the expander is a list of reactions in text form, or the name of a file containing a list of such reactions, one line per file.

The output of the expander is a list of the expanded reactions, either as Reaction objects, or a list of text reactions, or both.

Command Line Syntax:

```
$ python expander.py -in inputfile [-dump] [-out outputfile]
```

```
$ python pycellerator.py expand -in inputfile [-dump] [-out outputfile]
```

The *inputfile* may be either a reaction file or a model file. If it is a model file it must have file extension `.model`.

If the `-dump` option is specified the text output of the expander will be spewed to the screen as a list of text output reactions (compatible in format to input text reactions).

If the `-out` option is specified the text output of the expander will be written to the requested file. If no file name is given a default file name will be generated (e.g., `expanded-reactions.out`).

Existing files will not be over-written; if the requested file already exists a new file name will be generated that is based on the requested file name but has a unique string attached to the end of it.

Command Line in Shell:

```
>>> import pycellerator
>>> pycellerator.run ("expand -in inputfile [-dump] [-out ] outputfile]")
```

API:

```
expand(r)
  input:      single Reaction or list of Reactions
  return value: list of expanded Reactions (see parser)
```

Optional keyword arguments:

none

```
expandReactions(filename)
  input:      name of input file
  return value: list of Reactions (see parser)
```

Optional keyword arguments:

`dump = False` if True, print text reactions to screen
`text = False` if True, return list of text reactions instead of Reactions.

Example Input File: Here is an example of a typical input file, followed by a full example of the dump command using th an shorter input file.

```
[X <=> XP,    mod[Z, ZPP], rates[a, d, k, 0, a, d, k]]
[XP <=> XPP,   mod[Z, ZPP], rates[a, d, k, 0, a, d, k]]
[Y <=> YP,     mod[X, XPP], rates[a, d, k, 0, a, d, k]]
[YP <=> YPP,   mod[X, XPP], rates[a, d, k, 0, a, d, k]]
[Z <=> ZP,     mod[Y, YPP], rates[a, d, k, 0, a, d, k]]
[ZP <=> ZPP,   mod[Y, YPP], rates[a, d, k, 0, a, d, k]]
```

Full Example In the following example, we will dump a model that is generated from a single line file.

```
$ more test.dat
[X :=> Y,    mod[Z], rates[k1,k2,k3,k4,k5]]
$
$ python expander.py -in test.dat -dump -out test.out
py[cellerator]: expander (2015-09-01 10:39:34)
[X+Z->X_Z,k1]
[X_Z->X+Z,k2]
[X_Z->Z_Y,k3]
[Z_Y->X_Z,k4]
```

```
[Z_Y->Y+Z,k5]
[Y+Z->Z_Y,0]
Output written to: test.out
$
$ more test.out
[X+Z->X_Z,k1]
[X_Z->X+Z,k2]
[X_Z->Z_Y,k3]
[Z_Y->X_Z,k4]
[Z_Y->Y+Z,k5]
[Y+Z->Z_Y,0]
$
```

4.4 interpreter

The interpreter converts reactions from arrow format into differential equations. The functionality of the interpreter module is analogous to that of the Cellerator `interpret` command.

Command Line Syntax:

```
$ python interpreter.py -in filename -out filename]
[ -dump ]
[ -format FORMAT]
[ -frozen variable variable ...]
```

```
$ python pycellerator.py interpret -in filename -out filename]
[ -dump ]
[ -format FORMAT]
[ -frozen variable variable ...]
```

The input *filename* may be either a reaction file or a model file. If it is a model file it must have a file extension of **.model**. Here is an example reaction file:

```
[Br + BrO3 -> HBrO2 + HOBr, k1]
[Br + HBrO2 -> 2*HOBr, k2]
[BrO3 + HBrO2 -> 2*Ce + 2*HBrO2, k3]
[2*HBrO2-> BrO3 + HOBr, k4]
[Ce -> 0.5*Br, k5]
```

If the **-dump** option is specified the **Reaction** object database will be dumped to the screen.

If the **-out** option is specified the text output of the interpreter will be written to the requested file. If no file name is given a default file name will be generated (e.g., **interpreted -out reactions.out**).

If the **-out** option is not used, the result will be written to the screen.

Existing files will not be over-written; if the requested file already exists a new file name will be generated that is based on the requested file name but has a unique string attached to the end of it.

Output formats are **ODE** (default), **CODE**, **PYTHON**, **LATEX**, **DICT**, **JACOBIAN**, **STEADYSTATE** and are not case-sensitive.

-format ODE will return a text-formatted table of differential equations. Here is an example of **ODE** format:

```
BrO3' = -Br*BrO3*k1 - BrO3*HBrO2*k3 + k4*HBrO2**2.0
HBrO2' = Br*BrO3*k1 + 2.0*Br*HBrO2*k2 + k4*HBrO2**2.0
Br' = 0.5*Ce*k5 - Br*BrO3*k1 - Br*HBrO2*k2
HBrO2' = Br*BrO3*k1 + BrO3*HBrO2*k3 - Br*HBrO2*k2 - 2.0*k4*HBrO2**2.0
Ce' = -Ce*k5 + 2.0*BrO3*HBrO2*k3
```

-format CODE or **-format=PYTHON** will return a python function that will evaluate the differential equations. Here is an example of the output of the **CODE** or **PYTHON** format:

```
def f(y,t):
    y[0] = BrO3
    y[0] = HBrO2
    y[0] = Br
    y[0] = HBrO2
    y[0] = Ce
    yp[0] = -Br*BrO3*k1 - BrO3*HBrO2*k3 + k4*HBrO2**2.0
    yp[1] = Br*BrO3*k1 + 2.0*Br*HBrO2*k2 + k4*HBrO2**2.0
    yp[2] = 0.5*Ce*k5 - Br*BrO3*k1 - Br*HBrO2*k2
    yp[3] = Br*BrO3*k1 + BrO3*HBrO2*k3 - Br*HBrO2*k2 - 2.0*k4*HBrO2**2.0
    yp[4] = -Ce*k5 + 2.0*BrO3*HBrO2*k3
    return (yp)
```

Note that this code is not directly useable because the values of parameters are not specified. Also, this is not the same code that is returned by the solver module, as that code has the rate constants fully instantiated, unless the *inputfile* following the **-in** option is a model file with file extension **.model** that has all of the rate constants fully defined in the **\$RATES** block.

-format LATEX will return a L^AT_EX encoding of the equations within a minimal wrapper that makes it compatible with both **latex** and **pdflatex**. This option uses the **sympy latex** function and may not produce the expected output. For example, the above file will produce

```
\documentclass[12pt,letterpaper]{article}
\usepackage[latin1]{inputenc}
\usepackage{amsmath, amsfonts, amssymb}
\author{py[xlr8r]}
\date{\today}
\title{Automatically Generated Equations}
\begin{document}
\maketitle
\begin{align*}
BrO3' &= Br BrO_{3} k_{1} - BrO_{3} HBrO_{2} k_{3} + \\
&\quad HBrO_{2}^{2.0} k_{4} \\
HBrO2' &= r BrO_{3} k_{1} + 2.0 Br HBrO_{2} k_{2} + \\
&\quad HBrO_{2}^{2.0} k_{4} \\
Br' &= Br BrO_{3} k_{1} - Br HBrO_{2} k_{2} + 0.5 Ce k_{5} \\
HBrO2' &= r BrO_{3} k_{1} - Br HBrO_{2} k_{2} + \\
&\quad BrO_{3} HBrO_{2} k_{3} - 2.0 HBrO_{2}^{2.0} k_{4} \\
Ce' &= .0 BrO_{3} HBrO_{2} k_{3} - Ce k_{5} \\
\end{align*}
\end{document}
```

-format DICT will return a python dictionary of the form `"variable":ode, "variable":ode,...` where each `ode` is the right-hand side of the symbolic differential equation for the corresponding variable. Here is an example of a dictionary:

```
{'BrO3': -Br*BrO3*k1 - BrO3*HBrO2*k3 + k4*HBrO2**2.0,
 'HBrO2': Br*BrO3*k1 + 2.0*Br*HBrO2*k2 + k4*HBrO2**2.0,
 'Br': 0.5*Ce*k5 - Br*BrO3*k1 - Br*HBrO2*k2,
 'HBrO2': Br*BrO3*k1 + BrO3*HBrO2*k3 - Br*HBrO2*k2 - 2.0*k4*HBrO2**2.0,
 'Ce': -Ce*k5 + 2.0*BrO3*HBrO2*k3}
```

-format JACOBIAN will return the Jacobian matrix of the system:

```
$ python pyx.py interpret -in oregonator.dat -format Jacobian
[-Br*k1 - HBrO2*k3, 0, -BrO3*k1, -BrO3*k3 + 2*HBrO2*k4, 0]
[ Br*k1, 0, BrO3*k1 + 2*HBrO2*k2, 2*Br*k2 + 2*HBrO2*k4, 0]
[ -Br*k1, 0, -BrO3*k1 - HBrO2*k2, -Br*k2, 0.5*k5]
[ Br*k1 + HBrO2*k3, 0, BrO3*k1 - HBrO2*k2, BrO3*k3 - Br*k2 - 4*HBrO2*k4, 0]
[ 2*HBrO2*k3, 0, 0, 2*BrO3*k3, -k5]
```

-format STEADYSTATE will return the steady state of the system. Consider, for example, the following simple model `simple.model`:

```
$Reactions
[2*A -> B, k]
[B -> Nil, k1]
[B -> A, k2]
[Nil -> A, k3]
$IC
A=1
B=1
$Rates
k=1
k1=1
k2=.1
k3 = 1
$
```

This model has a single realizable (physical) steady state and a second non-realizable steady state (with negative concentrations).

```
$ python pycellerator.py interpret -in simple.model -format steadystate
There are 2 steady states
(A, B) = (-0.723746864455746, 0.476190476190476) has eigenvalues
[2.85841999579395, -1.06343253797097]
(A, B) = (0.723746864455746, 0.476190476190476) has eigenvalues
[-1.02268894521036, -2.97229851261262]
$
```

The **-symbolic** option may be used in conjunction with **-format STEADYSTATE** to return a tuple (**vars**, **ss**, **evs**), where **vars** is a tuple of the system variables; **ss** is a list of the steady states, each as a tuple; and **evs** is a list of the eigenvalues of the Jacobian at that steady state. This format is probably more meaningful form within the Python Shell:

```
>>> import pycellerator.interpreter as interp
>>> s,ss,evs=interp.interpret("simple.model",
    format="steadystate",symbolic=True)
>>> s
[A, B]
>>> ss
[(-0.723746864455746, 0.476190476190476),
 (0.723746864455746, 0.476190476190476)]
>>> evs
[[2.85841999579395, -1.06343253797097],
 [-1.02268894521036, -2.97229851261262]]
>>>
```

If the **-frozen** option is used it should be followed by one or more unquoted variable names in the model. These variables will be treated as frozen variables, i.e., the right hand sides of their differential equations will be set to zero. Note that these may also be additional variables that are not otherwise specified in the model equations, in which case additional equations will be added to the file.

Command Line in Shell:

```
>>> import pycellerator
>>> pycellerator.run ("interpret -in inputfile [ -out filename]
    [ -dump ] [ -format FORMAT][ -frozen variable variable ..]
    [ -symbolic] ")
```

API:

makeODETerms(r, s, *keywords)

input: r: list of reactions as Reaction class objects
s: dictionary of sympy symbols as {"var":symbol,"var":symbol,...}
return value: dictionary of ode right-hand-sides by text name of variable, as in {"var":ode, "var":ode,...} - the var's in the dictionary are the same as those in s.

Optional keyword arguments:

frozen = [] list of variables, given by the text equivalents, whose differential equations are held fixed, as in frozen=["var1","var2",...]. The entries in the return dictionary will be .."var1":0,"var2":0,.. for these variables.

interpret(filename, *keywords)

input: name of input file
return value: text table of differential equations, as per keywords

Optional keyword arguments:

format = "ODE" same as command line ODE format
 other options are "DICT", "CODE", "PYTHON",
 "JACOBIAN", "STEADYSTATE"
symbolic = False set to True to get a symbolic Steady state tuple
dumpparser="False" same as command line Dump
ofile="" name of output file

Example: This example uses the input file illustrated above.

```
$ python interpreter.py -in oregonator.dat -frozen BrO3 -format code
py[cellerator]: interpreter (2015-09-02 11:53:09)
def f(y,t):
    y[0] = Ce
    y[0] = HOBr
    y[0] = HBrO2
    y[0] = BrO3
    y[0] = Br
    yp[0] = -Ce*k5 + 2.0*BrO3*HBrO2*k3
    yp[1] = Br*BrO3*k1 + 2.0*Br*HBrO2*k2 + k4*HBrO2**2.0
    yp[2] = Br*BrO3*k1 + BrO3*HBrO2*k3 - Br*HBrO2*k2 - 2.0*k4*HBrO2**2.0
    yp[3] = 0
    yp[4] = 0.5*Ce*k5 - Br*BrO3*k1 - Br*HBrO2*k2
    return (yp)
$
```

4.5 solver

The solver will write a stand-alone python script to run a simulation on a model, and then will execute that script.

Command Line Syntax:

```
$ python pycellerator.py solve -in filename
    [-run stepsize duration]
    [-plot variable variable variable ...]]
    [-plotcolumns n]
    [-sameplot]
    [-out filename] ]
    [-format FORMAT] ]
    [-pyfile filename]]
    [-norun]
```

or

```
$ python solver.py -in filename
    [-run stepsize duration]
    [-plot variable variable variable ...]
    [-plotcolumns n]
    [-sameplot]
    [-out filename] ]
    [-format FORMAT] ]
    [-pyfile filename]]
    [-norun]
```

-in filename specifies the name of the model file.

-run stepsize duration specifies the step size (for plotting and output files) and duration of the run. The default values are 100 and 1.

-plot variable variable variable - if **-plot** is used alone, without any variable names, all variables are plotted on a grid. If any variables are listed, then only the requested variables are plotted.

-plotcolumns n defines the number of columns to use in the grid of plots (default is 3). Only used in conjunction with the **-plot** option.

-sameplot - the variables will all be plotted on a single graph rather than a grid. This options supersedes **-plotcolumns**.

-out filename - optional output file to write the solution to at the requested step size (as given in **-run** option).

-format FORMAT - format of output file; values of **FORMAT** are **CSV** (comma-separated values); **TSV** (tab-separated values); and **TABLE** (space-separated values). The default format is **CSV**.

-norun - just generate the code, don't run it.

-mxstep n - the default number of steps in **ODEINT** is 500. This is increased to 500000 by **pycellerator**.

-pyfile filename gives the name of the python code to be generated, otherwise a default file name will be used. Existing files will not be overwritten, so if the file already exists, and approximation to the requested file name will be used instead.

Command Line in Shell:

```
>>>import pycellerator
>>> pycellerator.run("interpret -in inputfile
    [-run stepsize duration]
    [-plot [ variable variable variable ...]
    [-plotcolumns n]
    [-sameplot]
    [-out filename] ]
    [-format FORMAT] ]
    [-pyfile filename] ]
    [-norun]")
```

4.6 converter

The converter module will convert **pycellerator** text reactions to Cellerator (xlr8r) *Mathematica* reaction notebooks.

The reactions will be specified in their *Mathematica* FullForm notation, so that they can be written as ASCII text files. This notation can be read directly or copied and pasted directly into an xlr8r notebook and is fully compatible with the xlr8r `interpret` and `run` commands.

Note that \LaTeX output is provided by the interpreter module, and SBML input and output is provided by the SBML module.

Command Line Syntax:

```
$ python pycellerator.py convert -in filename [-out filename] [-dump]
```

The **-in filename** specifies the input file name, which is a list of text pycellerator reactions, identical in format to the reactions sent to the parser, such as:

```
[X <=> XP,    mod[Z, ZPP], rates[a, d, k, 0, a, d, k]]
[XP <=> XPP,   mod[Z, ZPP], rates[a, d, k, 0, a, d, k]]
[Y <=> YP,     mod[X, XPP], rates[a, d, k, 0, a, d, k]]
[YP <=> YPP,   mod[X, XPP], rates[a, d, k, 0, a, d, k]]
[Z <=> ZP,     mod[Y, YPP], rates[a, d, k, 0, a, d, k]]
[ZP <=> ZPP,   mod[Y, YPP], rates[a, d, k, 0, a, d, k]]
```

The input **filename** may also be a model file (with file extension **.model**). In this case the output will include not only xlr8r reactions, but lists of initial conditions and rate constants in formats compatible with the xlr8r `run` function.

The **-out filename** specifies the output file name. which is typically a *Mathematica* notebook. If it is not specified a unique file name will be generated. The contents of this file will typically look something like this:

```
model = {
  List[Underoverscript[RightArrowLeftArrow[X, XP], ZPP, Z],
    a, d, k, 0, a, d, k, 0],
  List[Underoverscript[RightArrowLeftArrow[XP, XPP], ZPP, Z],
    a, d, k, 0, a, d, k, 0],
  List[Underoverscript[RightArrowLeftArrow[Y, YP], XPP, X],
    a, d, k, 0, a, d, k, 0],
  List[Underoverscript[RightArrowLeftArrow[YP, YPP], XPP, X],
    a, d, k, 0, a, d, k, 0],
  List[Underoverscript[RightArrowLeftArrow[Z, ZP], YPP, Y],
    a, d, k, 0, a, d, k, 0],
  List[Underoverscript[RightArrowLeftArrow[ZP, ZPP], YPP, Y],
    a, d, k, 0, a, d, k, 0]}
```

The **-dump** option indicates that the intermediate Reaction database should be dumped to the screen.

Command Line in Shell:

```
>>> import pycellerator
>>> pycellerator.run("convert -in filename [-out filename] [-dump]")
```

API:**convert (*keywords)**

input: (by keyword only)
return value: full path name of output file

Optional keyword arguments:

inputfile="" Name of file containing pycellerator reactions to be converted
outputfile="" Requested name of outputfile. If the file already exists
the name will be modified to generate a unique file name.
If not specified a unique default name will be generated.

4.7 SBML Support

4.7.1 Reading SBML Files

When an SBML model is read, it is converted to a `py[cellerator]` model file. A model file that is generated from an SBML model will typically look somewhat different from a hand-generated model because the reactions and other structures will not normally correspond to the typical biological entities that we think of as reactions.

Rather than running models directly from SBML, the **SBML read** command converts an SBML model directly into a pycellerator model file. This arrow file can be used like any other pycellerator model file. SBML entities that are not implemented in pycellerator but are referenced in the SBML file cannot be converted to pycellerator.

Syntax

```
python pycellerator.py sbml read -in filename [-model filename]
```

If **-model filename** is specified, the output will be written to the specified file. Otherwise the output will be written to the file **tmp.model**. Existing files will not be overwritten, and an approximation to the requested name will be used if the file already exists.

Caution: Unexpected errors could occur if variables (e.g., **species**) in the SBML model are the same as predefined functions in python, particularly **sympy** functions. For example, model 9 (Huang, 1996, “Ultra-sensitivity in the MAPK Cascade”) in the Biomodels database (<https://www.ebi.ac.uk/biomodels-main/BIOMD0000000009>) contains the variable **E1** which is also the name of an elliptic integral function in **sympy**. Attempting to convert this model directly would produce the somewhat obscure looking error

```
TypeError: unsupported operand type(s) for *: 'Symbol' and 'function'
```

To fix this, the user is fore-warned to change the names of all symbols, such as **E1**, to non-conflicting symbols.

In the following example, Biocompare 9 species **E1** was converted globally (in a text editor) to a new species **Species_E1** prior to use. The revised SBML file was saved in a new file **BM9.xml**. To convert this file to a pycellator model file **BM9.model**, use

```
python pycellator.py SBML READ -in BM9.xml -model BM9.model
```

The following model file was generated from the biomodels file

```
$ASSIGNMENTS
K_PP_norm=(KPase_PP_K + PP_K)/(K + KPase_PP_K + KPase_P_K +
PP_K + PP_KK_K + PP_KK_P_K + P_K)
rel_K_PP_max=1.11105062057732*K_PP_norm
KK_PP_norm=(KKPase_PP_KK + PP_KK + PP_KK_K + PP_KK_P_K)/(KK + KKPase_PP_KK +
KKPase_P_KK + PP_KK + PP_KK_K + PP_KK_P_K + P_KK + P_KKK_KK + P_KKK_P_KK)
KKK_P_norm=(P_KKK + P_KKK_KK + P_KKK_P_KK)/(KKK + P_KKK + P_KKK_KK + P_KKK_P_KK)
r1a=compartment*(1000.0*KKK*Species_E1 - 150.0*Species_E1_KKK)
r1b=150.0*Species_E1_KKK*compartment
r2a=compartment*(1000.0*E2*P_KKK - 150.0*E2_P_KKK)
r2b=150.0*E2_P_KKK*compartment
r3a=compartment*(1000.0*KK*P_KKK - 150.0*P_KKK_KK)
r3b=150.0*P_KKK_KK*compartment
r4a=compartment*(1000.0*KKPase*P_KK - 150.0*KKPase_P_KK)
r4b=150.0*KKPase_P_KK*compartment
r5a=compartment*(1000.0*P_KK*P_KKK - 150.0*P_KKK_P_KK)
r5b=150.0*P_KKK_P_KK*compartment
r6a=compartment*(1000.0*KKPase*PP_KK - 150.0*KKPase_PP_KK)
r6b=150.0*KKPase_PP_KK*compartment
r7a=compartment*(1000.0*K*PP_KK - 150.0*PP_KK_K)
r7b=150.0*PP_KK_K*compartment
r8a=compartment*(1000.0*KPase*P_K - 150.0*KPase_P_K)
r8b=150.0*KPase_P_K*compartment
r9a=compartment*(1000.0*PP_KK*P_K - 150.0*PP_KK_P_K)
r9b=150.0*PP_KK_P_K*compartment
r10a=compartment*(1000.0*KPase*PP_K - 150.0*KPase_PP_K)
r10b=150.0*KPase_PP_K*compartment
$REACTIONS
[Species_E1 -> Nil, using["r1a/compartment"]]
[Nil -> Species_E1_KKK, using["r1a/compartment"]]
[KKK -> Nil, using["r1a/compartment"]]
[Nil -> Species_E1, using["r1b/compartment"]]
[Species_E1_KKK -> Nil, using["r1b/compartment"]]
[Nil -> P_KKK, using["r1b/compartment"]]
[Nil -> E2_P_KKK, using["r2a/compartment"]]
[P_KKK -> Nil, using["r2a/compartment"]]
[E2 -> Nil, using["r2a/compartment"]]
[E2_P_KKK -> Nil, using["r2b/compartment"]]
[Nil -> KKK, using["r2b/compartment"]]
[Nil -> E2, using["r2b/compartment"]]
[KK -> Nil, using["r3a/compartment"]]
[Nil -> P_KKK_KK, using["r3a/compartment"]]
[P_KKK -> Nil, using["r3a/compartment"]]
[P_KKK_KK -> Nil, using["r3b/compartment"]]
[Nil -> P_KK, using["r3b/compartment"]]
[Nil -> P_KKK, using["r3b/compartment"]]
[Nil -> KKPase_P_KK, using["r4a/compartment"]]
[P_KK -> Nil, using["r4a/compartment"]]
[KKPase -> Nil, using["r4a/compartment"]]
[KKPase_P_KK -> Nil, using["r4b/compartment"]]
```



```

[Nil -> KK, using["r4b/compartment"]]
[Nil -> KKPase, using["r4b/compartment"]]
[Nil -> P_KKK_P_KK, using["r5a/compartment"]]
[P_KK -> Nil, using["r5a/compartment"]]
[P_KKK -> Nil, using["r5a/compartment"]]
[P_KKK_P_KK -> Nil, using["r5b/compartment"]]
[Nil -> PP_KK, using["r5b/compartment"]]
[Nil -> P_KKK, using["r5b/compartment"]]
[Nil -> KKPase_PP_KK, using["r6a/compartment"]]
[PP_KK -> Nil, using["r6a/compartment"]]
[KKPase -> Nil, using["r6a/compartment"]]
[KKPase_PP_KK -> Nil, using["r6b/compartment"]]
[Nil -> P_KK, using["r6b/compartment"]]
[Nil -> KKPase, using["r6b/compartment"]]
[Nil -> PP_KK_K, using["r7a/compartment"]]
[K -> Nil, using["r7a/compartment"]]
[PP_KK -> Nil, using["r7a/compartment"]]
[PP_KK_K -> Nil, using["r7b/compartment"]]
[Nil -> PP_KK, using["r7b/compartment"]]
[Nil -> P_K, using["r7b/compartment"]]
[Nil -> KPase_P_K, using["r8a/compartment"]]
[P_K -> Nil, using["r8a/compartment"]]
[KPase -> Nil, using["r8a/compartment"]]
[KPase_P_K -> Nil, using["r8b/compartment"]]
[Nil -> K, using["r8b/compartment"]]
[Nil -> KPase, using["r8b/compartment"]]
[Nil -> PP_KK_P_K, using["r9a/compartment"]]
[PP_KK -> Nil, using["r9a/compartment"]]
[P_K -> Nil, using["r9a/compartment"]]
[Nil -> PP_K, using["r9b/compartment"]]
[PP_KK_P_K -> Nil, using["r9b/compartment"]]
[Nil -> PP_KK, using["r9b/compartment"]]
[PP_K -> Nil, using["r10a/compartment"]]
[Nil -> KPase_PP_K, using["r10a/compartment"]]
[KPase -> Nil, using["r10a/compartment"]]
[Nil -> KPase, using["r10b/compartment"]]
[KPase_PP_K -> Nil, using["r10b/compartment"]]
[Nil -> P_K, using["r10b/compartment"]]

$RATES
K_PP_norm_max=0.900049
compartment=4e-12

$IC
Species_E1=3e-05
E2=0.0003
KKK=0.003
P_KKK=0.0
KK=1.2
P_KK=0.0
PP_KK=0.0
K=1.2
P_K=0.0
PP_K=0.0
KPase=0.12
KKPase=0.0003
Species_E1_KKK=0.0
E2_P_KKK=0.0
P_KKK_KK=0.0

```

```
P_KKK_P_KK=0.0
PP_KK_K=0.0
PP_KK_P_K=0.0
KKPase_PP_KK=0.0
KKPase_P_KK=0.0
KPase_PP_K=0.0
KPase_P_K=0.0
K_PP_norm=0.0
KK_PP_norm=0.0
KKK_P_norm=0.0
rel_K_PP_max=0.0
```

This file can support simulations as with any other model file, e.g.

```
python pycellerator.py solve -in BM9.model -plot
```

Supported Features

The following SBML features are not supported by `pycellerator` and will be ignored in SBML models: **unitDefinitions**, **compartmentTypes**, **speciesTypes**, **initialAssignments**, **algebraicRules**, **constraints**, and **events**. Support for these features may be added at a later date. Since some of the structures need to be translated, and the way things are evaluated may not correspond to the expected rules of SBML evaluation, as described in the following paragraphs.

1. All variables that have either their **boundaryCondition** or **constant** flag set to **True** are converted to **Frozen** variables in the model. This means that they may be set by **using** reactions or by **assignment** statements. Either of these could potentially conflict with the intent of the SBML model. For example, if the **constant** field is set to **True**, SBML says that such variables should never change, but if either a rate or assignment rule is present in the SBML model it will be used in the `pycellerator` model (technically this would be an invalid SBML model, so should not lead to problems). However, if both values are set to **False** and the variable is set in both a rule and a reaction, then there could be a combination of assignments and ODES for that variable, in conflict with the intent of the SBML (again this would be invalid SBML, so the situation should not arise).
2. If a **parameter** is a **variable** in an **assignmentRule** and has a **value** in a **parameter** statement, then the **value** in the **parameter** statement will take precedence and the **assignmentRule** will be ignored.
3. All **rateRules** are converted to "using" reactions of the form **Nil -> Variable** and the **variable** is added to the list of **frozen** variables, so that it cannot be changed by any other reactions.
4. All reaction **kineticLaws** are assigned to assignment statements with the same variable as the reaction **id**.
5. Each **reaction** is converted to a collection of **using** creation and annihilation reactions, with one reaction for each species in the reaction. The reaction will always have the form of **X -> Nil** or **Nil -> X**, depending on the net stoichiometry of the species in the reaction. Thus, for example, if the SBML file has a reaction

```
A + 3B -> 2A + B + C
```

with a kinetic law **K**, the following `pycellerator` reactions will be generated:

```
[Nil -> A, using["K/VA"]]
[B -> Nil, using["2*K/VB"]]
[Nil -> C, using["K/VC"]]
```

where **VA**, **VB**, and **VC** are the volumes of the compartments that species **A**, **B** and **C** reside in, respectively. All of the stoichiometries inside the **using** expression are expressed as positive quantities because the sign will be calculated automatically by the interpreter module, because the reaction `[X -> Nil, using["K"]]` will produce the same ODE term as `[Nil -> X, using["-K"]]`, namely, $[X]' = -K$.

- At each integration step, all **assignmentRules** (with their parameter values instantiated) are evaluated before the differential equations are updated. Thus a differential equation may refer to any variable defined by an **assignmentRule**
- All functions defined in the **listOfFunctions** are instantiated globally so that any rule or reaction may refer to them.

4.7.2 Writing SBML Files

Not all features of SBML are supported. Only reactions, initial conditions, and parameters are written to the SBML file. Functions and assignment rules are not written to SBML in the present version.

Syntax

```
python pycellerator.py SBML write -in file [-out filename]
```

SBML files are very lengthy so only a very simple example will be given here. Consider the model file **catalytic.model**

```
$Reactions
[X => Y, mod[E], rates[k1,k2,k3]]
[Y -> Nil, k]
$IC
X = 1.0
Y = 0.0
E = 1.0
$Rates
k1 = 1.0
k2 = 1.0
k3 = 1.0
k = .001
```

To convert this to SBML, the syntax is

```
python pycellerator.py SBML WRITE -in catalytic.model
```

Here is the output.

```
<?xml version="1.0" encoding="UTF-8"?>
<sbml xmlns="http://www.sbml.org/sbml/level3/version1/core" level="3"
      version="1">
  <model substanceUnits="item" timeUnits="dimensionless" extentUnits="item">
    <listOfCompartments>
      <compartment id="compartment" spatialDimensions="3" size="1"
```

```

        units="dimensionless" constant="true"/>
</listOfCompartments>
<listOfSpecies>
  <species id="Y" compartment="compartment" initialAmount="0"
    hasOnlySubstanceUnits="true" boundaryCondition="false"
    constant="false"/>
  <species id="X" compartment="compartment" initialAmount="1"
    hasOnlySubstanceUnits="true" boundaryCondition="false"
    constant="false"/>
  <species id="E" compartment="compartment" initialAmount="1"
    hasOnlySubstanceUnits="true" boundaryCondition="false"
    constant="false"/>
  <species id="Nil" compartment="compartment" initialAmount="0"
    hasOnlySubstanceUnits="true" boundaryCondition="false"
    constant="false"/>
</listOfSpecies>
<listOfParameters>
  <parameter id="k3" value="1" units="dimensionless" constant="true"/>
  <parameter id="k2" value="1" units="dimensionless" constant="true"/>
  <parameter id="k1" value="1" units="dimensionless" constant="true"/>
  <parameter id="k" value="0.001" units="dimensionless"
    constant="true"/>
</listOfParameters>
<listOfReactions>
  <reaction id="r1" reversible="false" fast="false">
    <listOfReactants>
      <speciesReference species="X" stoichiometry="1" constant="false"/>
      <speciesReference species="E" stoichiometry="1" constant="false"/>
    </listOfReactants>
    <listOfProducts>
      <speciesReference species="X_E" stoichiometry="1" constant="false"/>
    </listOfProducts>
    <kineticLaw>
      <math xmlns="http://www.w3.org/1998/Math/MathML">
        <apply>
          <times/>
          <ci> E </ci>
          <ci> X </ci>
          <ci> k1 </ci>
        </apply>
      </math>
    </kineticLaw>
  </reaction>
  <reaction id="r2" reversible="false" fast="false">
    <listOfReactants>
      <speciesReference species="X_E" stoichiometry="1" constant="false"/>
    </listOfReactants>
    <listOfProducts>
      <speciesReference species="X" stoichiometry="1" constant="false"/>
      <speciesReference species="E" stoichiometry="1" constant="false"/>
    </listOfProducts>
    <kineticLaw>
      <math xmlns="http://www.w3.org/1998/Math/MathML">
        <apply>
          <times/>
          <ci> X_E </ci>
          <ci> k2 </ci>

```

```

        </apply>
      </math>
    </kineticLaw>
  </reaction>
  <reaction id="r3" reversible="false" fast="false">
    <listOfReactants>
      <speciesReference species="X_E" stoichiometry="1" constant="false"/>
    </listOfReactants>
    <listOfProducts>
      <speciesReference species="Y" stoichiometry="1" constant="false"/>
      <speciesReference species="E" stoichiometry="1" constant="false"/>
    </listOfProducts>
    <kineticLaw>
      <math xmlns="http://www.w3.org/1998/Math/MathML">
        <apply>
          <times/>
          <ci> X_E </ci>
          <ci> k3 </ci>
        </apply>
      </math>
    </kineticLaw>
  </reaction>
  <reaction id="r4" reversible="false" fast="false">
    <listOfReactants>
      <speciesReference species="Y" stoichiometry="1" constant="false"/>
    </listOfReactants>
    <listOfProducts>
      <speciesReference species="Nil" stoichiometry="1" constant="false"/>
    </listOfProducts>
    <kineticLaw>
      <math xmlns="http://www.w3.org/1998/Math/MathML">
        <apply>
          <times/>
          <ci> Y </ci>
          <ci> k </ci>
        </apply>
      </math>
    </kineticLaw>
  </reaction>
</listOfReactions>
</model>
</sbml>

```


Chapter 5

Examples

This chapter present some models that are illustrative of pycellerator. The following matrix maps some of the features that are used against the various models.

Model	Mass Action ($->$)	Mass Action ($-->$)	Mass Action ($<->$)	Mass Action ($=>$)	Mass Action ($<=>$)	Mass Action ($:=>$)	MMH ($:>$, $:<$)	GRN ($ >$, $ <$)	Hill ($ >$, $ <$)	NHCA ($ >$, $ <$)	SSystem ($ >$)	rational ($==>$)	MWC ($==>$)	Equations / USER	Cascades
<i>E coli</i> Growth											✓				
Lineage Determination	✓											✓		✓	
MAPK Oscillations		✓		✓											✓
Mitotic Oscillator	✓								✓					✓	
NF- κ B	✓		✓						✓						
Oregonator	✓														
Repressilator	✓		✓						✓			✓		✓	
Ring Oscillator (MA)					✓										
Ring Oscillator (MMH)							✓								
Ring Oscillator (GRN)	✓							✓							
Ring Oscillator (NHCA)	✓									✓					

5.1 MAP-Kinase with Oscillations

The following model (unpublished) demonstrates MAPK oscillations and illustrates the use of both catalyzed reactions and enzymatic cascades.

```

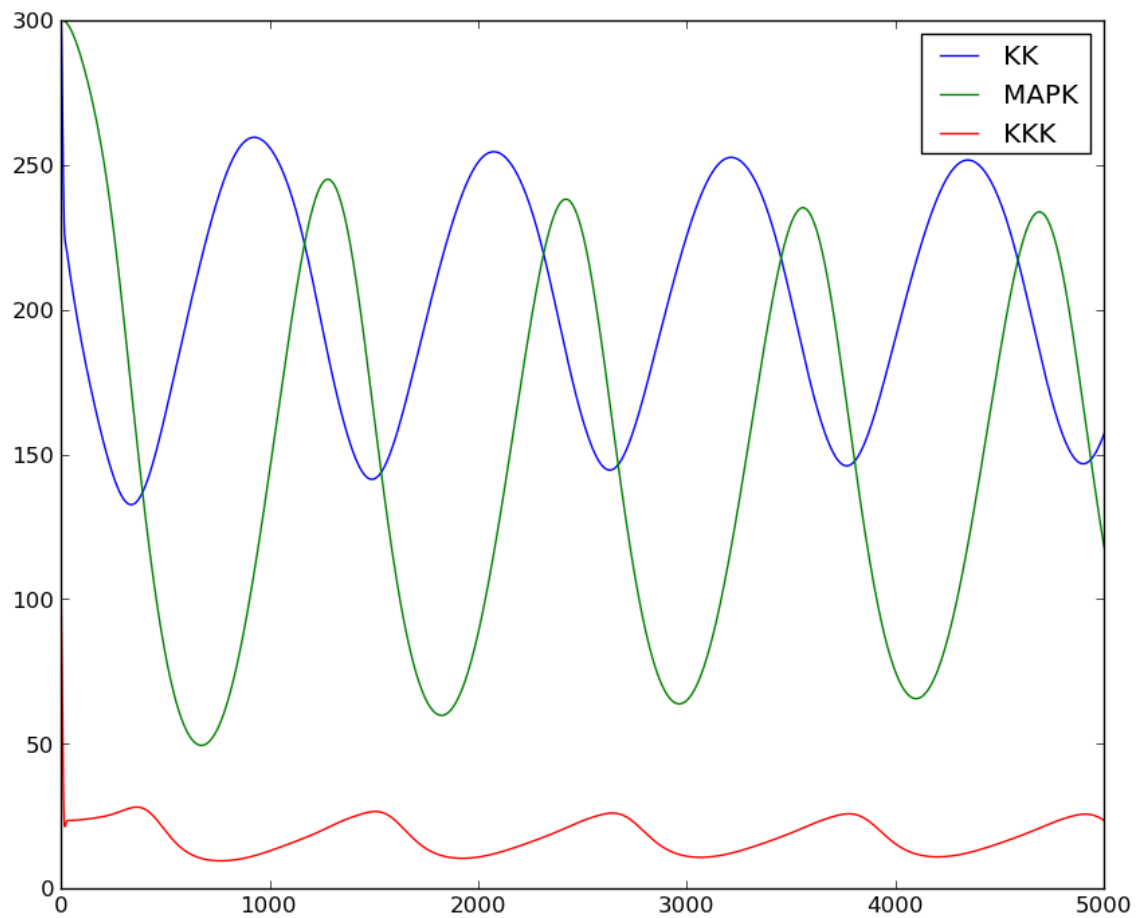
$Reactions
#
# kinases
#
[Nil<=>S, rates[a0, d0]]
[KKK => KKKp, mod[S], rates[a1,d1,k1]]
[KK => Kp => KKpp, mod[KKKp, KKKp], rates[a3,d3,k3]]
[MAPK => Kp => Kpp, mod[KKpp], rates[a3,d3,k3]]
#
# competitive inhibition
#
[KKK_S + Kpp <=> KKK_S_Kpp, rates[a7, d7]]
#
# phosphatases
#
[KKKp => KKK, mod[KKKph], rates[a4,d4,k4]]
[KKpp => Kp => KK, mod[KKph], rates[a5,d5,k5]]
[Kpp => Kp => MAPK, mod[Kph], rates[a6,d6,k6]]
$IC
  KKK = 100
  KKKp = 0
  KK = 300
  Kp = 0
  KKpp = 0
  MAPK = 300
  Kpp = 0
  S = 1
  Kph = 1
  KKph = 1
  KKKph = 10
$Rates
  a0 = 1
  d0 = 1
  a1 = 1
  d1 = 7.5
  k1 = 2.5
  a3 = 1
  d3 = 10
  k3 = 0.025
  a4 = 1
  d4 = 1
  k4 = 1
  a5 = 1
  d5 = 1
  k5 = 1
  a6 = 1
  d6 = 1
  k6 = 1
  a7 = 1
  d7 = 1
$

```



```
$ python pycelerator.py solve -in MAPK.model -plot KK KKK MAPK  
-sameplot -run 5000 2
```

/home/mathman/src/pyxlr8r/pyxlr8r-1204049/code/MAPK.model (2012-04-02 11:21:13)



5.2 Repressilator

This model demonstrates the use of Hill and rational arrows. An alternative version that uses USER reactions is given at the end of this section. A simulation model for the repressilator [4] is given by¹

```
$reactions
[PX -> Nil, Beta]
[PY -> Nil, Beta]
[PZ -> Nil, Beta]
[X -> X + PX, Beta]
[Y -> Y + PY, Beta]
[Z -> Z + PZ, Beta]
[PZ |-> X, Hill[alpha1, n, K, 0, 1]]
[PX |-> Y, Hill[alpha1, n, K, 0, 1]]
[PY |-> Z, Hill[alpha1, n, K, 0, 1]]
[X <-> Nil, rates[k1, alpha0]]
[Y <-> Nil, rates[k1, alpha0]]
[Z <-> Nil, rates[k1, alpha0]]
[[[Nil], [PY]] ==> Z, rational[[alpha], [K, 1], [1], [n, n]]]
[[[Nil], [PZ]] ==> X, rational[[alpha], [K, 1], [1], [n, n]]]
[[[Nil], [PX]] ==> Y, rational[[alpha], [K, 1], [1], [n, n]]]
$rates
alpha = 250
alpha0 = 0
alpha1 = 0
Beta = 5
n = 2.1
k1 = 1
K = 1
$ic
PX = 5
PZ = 15
```

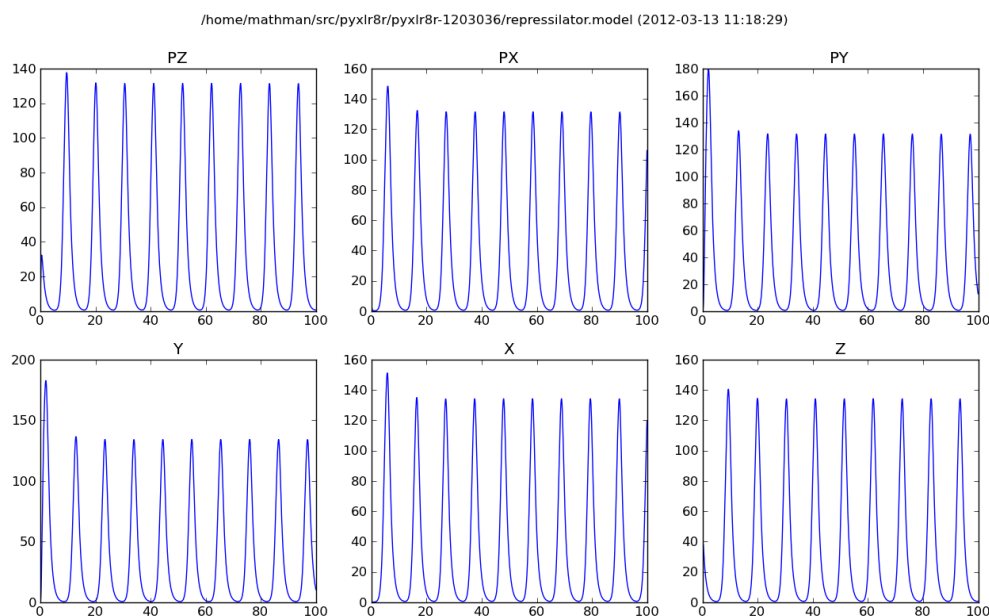
If we call this file `repressilator.model` then the command²

```
$ python pycellerator solve -in repressilator.model -plot -run 100 .1
```

will produce a plot such as this:

¹The terminating dollar sign at the end of the models is optional.

²The leading dollar sign in the command refers to the bash prompt and should not be manually entered.



as well as a CSV file that looks something like this:

```
0.0,15.0,5.0,0.0,0.0,0.0,0.0
0.1,14.2278260331,3.05384634588,0.247394702031,1.34912572721,0.0990958803047,23.562567514
0.2,21.504406326,1.90401628677,1.28225870666,4.57557848574,0.149799701424,37.9011356108
0.3,28.2774224855,1.21721312786,3.86093341284,11.2385014195,0.16314039654,38.2545583466
0.4,31.5798946122,0.803272174073,8.97682635583,22.2830086972,0.166187145597,35.2157392832
0.5,32.317811614,0.552727585611,17.2339097555,36.8646688505,0.166663798479,31.9877430887
...
99.8 0.648217893072,98.8724564077,14.3446430729,11.4817962739,113.312240676,0.649415887468
99.9,0.655884845799,105.886314242,12.9821534961,10.3905866114,119.451386878,0.685929274901
100.0,0.678921934741,112.385765424,11.7488271934,9.4030395272,124.765357623,0.74177680618
```

To see the differential equations produced by this model one could isolate the reactions in a file such as

```
[PX -> Nil, Beta]
[PY -> Nil, Beta]
[PZ -> Nil, Beta]
[X -> X + PX, Beta]
[Y -> Y + PY, Beta]
[Z -> Z + PZ, Beta]
[PZ |-> X, Hill[alpha1, n, K, 0, 1]]
[PX |-> Y, Hill[alpha1, n, K, 0, 1]]
[PY |-> Z, Hill[alpha1, n, K, 0, 1]]
[X <-> Nil, rates[k1, alpha0]]
[Y <-> Nil, rates[k1, alpha0]]
[Z <-> Nil, rates[k1, alpha0]]
[[[Nil], [PY]] ==> Z, rational[[alpha], [K, 1], [1], [n, n]]]
[[[Nil], [PZ]] ==> X, rational[[alpha], [K, 1], [1], [n, n]]]
[[[Nil], [PX]] ==> Y, rational[[alpha], [K, 1], [1], [n, n]]]
```

If we call this file **repressilator.dat** then the command

```
python interpreter.py -in repressilator.dat
```

will produce the output

```

PZ' = Beta*Z - Beta*PZ
PX' = Beta*X - Beta*PX
PY' = Beta*Y - Beta*PY
Y' = alpha0 - Y*k1 + alpha/(K**n + PX**n) + alpha1*PX**n/(K**n + PX**n)
X' = alpha0 - X*k1 + alpha/(K**n + PZ**n) + alpha1*PZ**n/(K**n + PZ**n)
Z' = alpha0 - Z*k1 + alpha/(K**n + PY**n) + alpha1*PY**n/(K**n + PY**n)

```

Within the ipython interface, the simpler command `cellerator.printODES()` would be sufficient to do this after loading model.

The repressilator can also be implemented using **rational** and **USER** reactions:

```

[[[Nil], [PY]] ==> Z, rational[[alpha], [K, 1], [1], [n, n]]]
[[[Nil], [PZ]] ==> X, rational[[alpha], [K, 1], [1], [n, n]]]
[[[Nil], [PX]] ==> Y, rational[[alpha], [K, 1], [1], [n, n]]]
[PY |-> Z, USER[alpha, -1, n, 0, "lambda x: 1/(K**n+x)"]]
[PZ |-> X, USER[alpha, -1, n, 0, "lambda x: 1/(K**n+x)"]]
[PX |-> Y, USER[alpha, -1, n, 0, "lambda x: 1/(K**n+x)"]]

```

5.3 Lineage Determination

This model demonstrates the use of the rational with product notation. The model is based on [2].

```

$REACTIONS
[[[A,O*S,O*S*N],[A,O,O*S,O*S*N,C*O,GC]]==>O, rational[[a0,a1,a2,a3],
[1,b0,b1,b2,b3,b4,b5],[],[[]]]
[O->Nil,gamma1]
#
[[[O*S,O*S*N],[O,O*S,O*S*N]] ==>S, rational[[c0,c1,c2],
[1,d0,d1,d2],[],[[]]]
[S->Nil,gamma2]
#
[[[O*S,O*S*N],[O,O*S,O*S*N,O*G]] ==>N, rational[[e0,e1,e2],
[1,f0,f1,f2,f3],[],[[]]]
[N->Nil,gamma3]
#
[[[C],[C,C*O]]==>C, rational[[g0,g1],[1,h0,h1],[],[[]]]
[C->Nil,gamma4]
#
[[[C,G],[C,G]]==>GC, rational[[i0,i1,i2],[1,j0,j1],[],[[]]]
[GC->Nil,gamma5]
#
[[[O,G],[O,G,N]]==>G, rational[[p0,p1,p2],[1,q0,q1,q2],[],[[]]]
[G->Nil,gammag]
#
[A->A,1]
$IC
O=5
N=5
S=5
G=0
GC=0
C=0
A=1
$Rates
a0 = 0.001
a1 = 1
a2 = 0.005
a3 = 0.025
b0 = 1
b1 = 0.001
b2 = 0.005
b3 = 0.025
b4 = 10
b5 = 10
gamma1 = 0.1
c0 = 0.001
c1 = 0.005
c2 = 0.025
d0 = 0.001
d1 = 0.005
d2 = 0.025
gamma2 = 0.1
e0 = 0.001
e1 = 0.1
e2 = 0.1

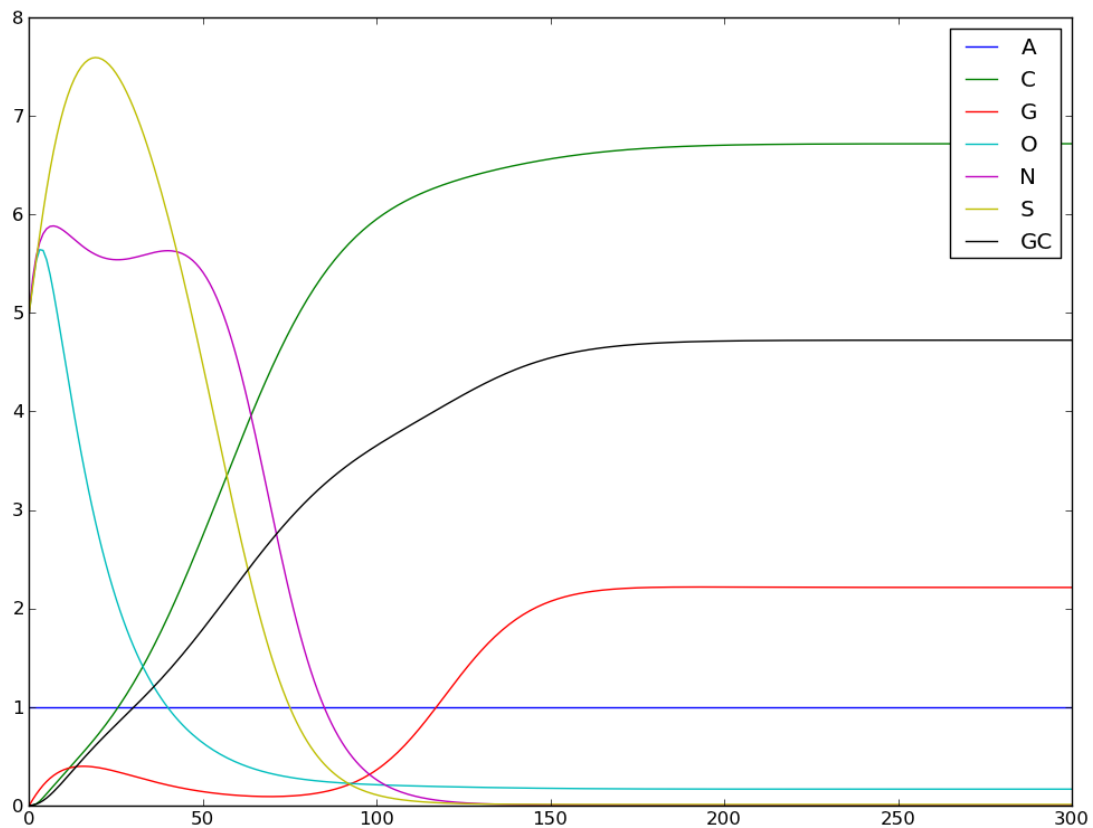
```

```

f0 = 0.001
f1 = 0.1
f2 = 0.1
f3 = 10
gamma3 = 0.1
g0 = 0.001
g1 = 2
h0 = 2
h1 = 5
gamma4 = 0.1
gamma5 = 0.1
i0 = 0.001
i1 = 0.1
i2 = 0.1
j0 = 0.1
j1 = 0.1
p0 = 0.1
p1 = 1.
p2 = 0.00025
q0 = 1.
q1 = 0.00025
q2 = 15
gammag = 0.1
$

```

/home/mathman/src/pyxlr8r/pyxlr8r-1203047/code/CP.model (2012-03-31 19:15:31)



5.4 NF- κ B Signaling Model

A pycellator model of the the NF- κ B signaling model of [8] is given by:

```

$Reactions
[IkBa + NFkB <=> IkBa_NFkB, rates[a1, d1]]
[IKK_IkBa + NFkB <=> IKK_IkBa_NFkB, rates[a2, d2]]
[NFkB <=> NFkBn, rates[tr1, tr2]]
[IkBa_NFkB -> NFkB, deg1]
[IkBan + NFkBn <=> IkBan_NFkBn, rates[a1, d1]]
[IkBa + IKK <=> IKK_IkBa, rates[a3, d3]]
[IkBat -> Nil, deg3]
[IkBa <=> IkBan, rates[tr3, tr4]]
[IKK_IkBa_NFkB -> IKK + NFkB, k1]
[IKK_IkBa -> IKK, k2]
[IkBat |-> IkBa, Hill[v1, n1, K1]]
[IkBa_NFkB <=> IkBan_NFkBn, rates[tr5, tr6]]
[IkBa -> Nil, deg2]
[NFkBn |-> IkBat, Hill[v2, n2, K2]]
[Nil -> IkBat, trb]
[IkBa_NFkB + IKK <=> IKK_IkBa_NFkB, rates[a4, d4]]
[IKK -> Nil, adapt]
$IC
NFkB = 0.003053
IkBa = 0.372
IkBa_NFkB = 0.09826
NFkBn = .00047638
IKBan = 0.11937
IKBan_NFkBn = 0.001985
IKK = 0.1
IkBat = 0.008454217
IKK_IkBa_NFkB = 0
IKK_IkBa = 0
$Rates
v1 = 2.448
K1 = 10
n1 = 1
v2 = 1.02713
K2 = 1
n2 = 2
a1 = 30
a2= 30
a3= 1.35
a4= 11.1
tr1= 5.4
tr2= 0.0048
tr3= 0.018
tr4= 0.012
tr5= 0
tr6= 0.82944
d1= 0.03
d2= 0.03
d3= 0.0075
d4= 0.105
deg1= 0.00135
deg2= 0.00675
deg3= 0.0168

```

```

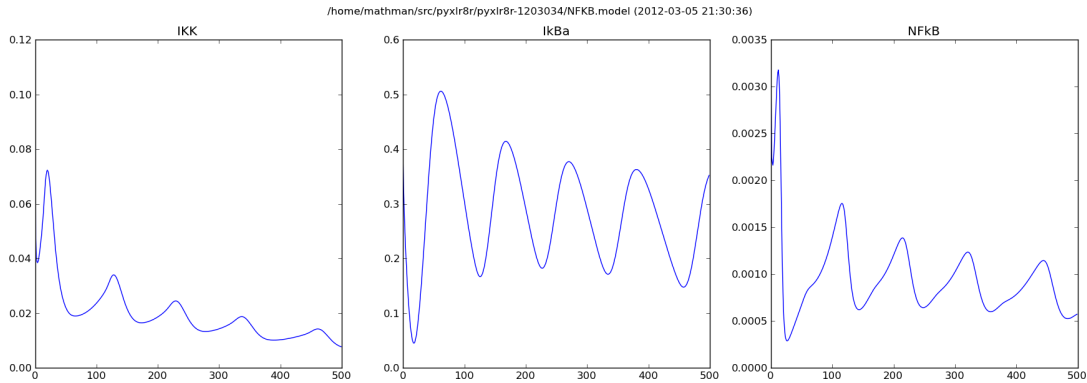
k1= 1.221
k2= 0.2442
trb=0.0000921375
adapt= 0.0072
$

```

Running a simulation with this file as `NFKB.model` using the command

```
python pycellerator solve -in NFKB.model -plot IKK IkBa NFkB -run 500 .1
```

will produce the following plot:



The differential equations can be obtained from

```
python pycellerator.py interpret -in NFKB.model
```

which produces the following output,

```

IkBa_NFkB' = IKK_IkBa_NFkB*d4 + IkBan_NFkBn*tr6 - IkBa_NFkB*d1 -
  IkBa_NFkB*deg1 - IkBa_NFkB*tr5 + IkBa*NfkB*a1 - IKK_IkBa_NFkB*a4
IkBan_NFkBn' = IkBa_NFkB*tr5 - IkBan_NFkBn*d1 - IkBan_NFkBn*tr6 +
  IkBan*NfkBn*a1
IKK_IkBa' = IKK_IkBa_NFkB*d2 - IKK_IkBa*d3 - IKK_IkBa*k2 +
  IKK_IkBa*a3 - IKK_IkBa*NfkB*a2
NfkBn' = IkBan_NFkBn*d1 + NfkB*tr1 - NfkBn*tr2 - IkBan*NfkBn*a1
IKK_IkBa_NFkB' = -IKK_IkBa_NFkB*d2 - IKK_IkBa_NFkB*d4 -
  IKK_IkBa_NFkB*k1 + IKK_IkBa_NFkB*a4 +
  IKK_IkBa*NfkB*a2
NfkB' = IKK_IkBa_NFkB*d2 + IKK_IkBa_NFkB*k1 + IkBa_NFkB*d1 +
  IkBa_NFkB*deg1 + NfkBn*tr2 - NfkB*tr1 -
  IKK_IkBa*NfkB*a2 - IkBa*NfkB*a1
IkBan' = IkBa*tr3 + IkBan_NFkBn*d1 - IkBan*tr4 - IkBan*NfkBn*a1
IkBa' = IKK_IkBa*d3 + IkBa_NFkB*d1 + IkBan*tr4 - IkBa*deg2 -
  IkBa*tr3 - IKK_IkBa*a3 - IkBa*NfkB*a1 +
  v1*IkBt**n1/(IkBat**n1 + K1**n1)
IkBat' = trb - IkBat*deg3 + v2*NfkBn**n2/(K2**n2 + NfkBn**n2)
IKK' = IKK_IkBa*d3 + IKK_IkBa*k2 + IKK_IkBa_NFkB*d4 +
  IKK_IkBa_NFkB*k1 - IKK*adapt - IKK_IkBa*a3 -
  IKK_IkBa_NFkB*a4

```

If the code option is requested, e.g.,

```
python pycellerator.py interpret -in NFKB.dat -format CODE -out nfkb.py
```


The file `nfkb.py` will look something like this:³

```
def f(y,t):
    y[0] = IkBa_NFkB
    y[1] = IkBan_NFkBn
    y[2] = IKK_IkBa
    y[3] = NFkBn
    y[4] = IKK_IkBa_NFkB
    y[5] = NFkB
    y[6] = IkBan
    y[7] = IkBa
    y[8] = IkBat
    y[9] = IKK
    yp[0] = IKK_IkBa_NFkB*d4 + IkBan_NFkBn*tr6 - IkBa_NFkB*d1 -
            IkBa_NFkB*deg1 - IkBa_NFkB*tr5 + IkBa*NfKB*a1 -
            IKK*IcBa_NFkB*a4
    yp[1] = IkBa_NFkB*tr5 - IkBan_NFkBn*d1 - IkBan_NFkBn*tr6 +
            IkBan*NfKBn*a1
    yp[2] = IKK_IkBa_NFkB*d2 - IKK_IkBa*d3 - IKK_IkBa*k2 +
            IKK*IcBa*a3
            - IKK_IcBa*NfKB*a2
    yp[3] = IkBan_NFkBn*d1 + NFkB*tr1 - NFkBn*tr2 -
            IkBan*NfKBn*a1
    yp[4] = -IKK_IcBa_NFkB*d2 - IKK_IcBa_NFkB*d4 - IKK_IcBa_NFkB*k1 +
            IKK*IcBa_NFkB*a4 + IKK_IcBa*NfKB*a2
    yp[5] = IKK_IcBa_NFkB*d2 + IKK_IcBa_NFkB*k1 + IkBa_NFkB*d1 +
            IkBa_NFkB*deg1 + NFkBn*tr2
            - NFkB*tr1 - IKK_IcBa*NfKB*a2 - IkBa*NfKB*a1
    yp[6] = IkBa*tr3 + IkBan_NFkBn*d1 - IkBan*tr4 - IkBan*NfKBn*a1
    yp[7] = IKK_IcBa*d3 + IkBa_NFkB*d1 + IkBan*tr4 - IkBa*deg2 -
            IkBa*tr3 - IKK*IcBa*a3
            - IkBa*NfKB*a1 + v1*IcBat**n1/(IcBat**n1 + K1**n1)
    yp[8] = trb - IcBat*deg3 + v2*NfKBn**n2/(K2**n2 + NFkBn**n2)
    yp[9] = IKK_IcBa*d3 + IKK_IcBa*k2 + IKK_IcBa_NFkB*d4 +
            IKK_IcBa_NFkB*k1 - IKK*adapt
            - IKK*IcBa*a3 - IKK_IcBa_NFkB*a4
    return (yp)
```

³While each line of code terminates with a newline character, individual lines of code are not line-wrapped and may be very line. The wrapping shown here was edited for illustration in this document, so in fact, it is likely that most lines of code will exceed the standard 80 character page width. This is why the usual python line wrap characters are not shown in the example.

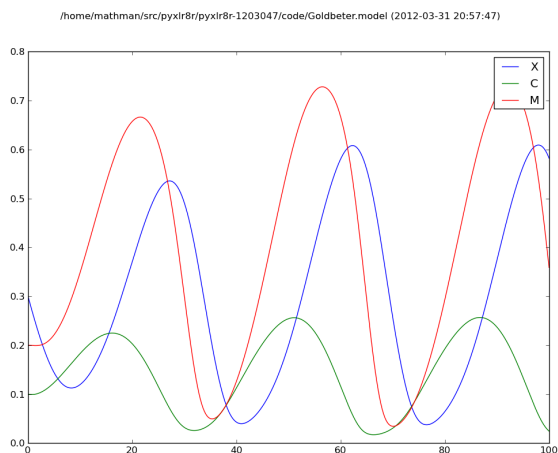
5.5 Minimal Mitotic Model

This is a standard minimal model of a mitotic oscillator^[7]. It illustrates the use non-catalytic Hill function arrows as well as the use of equations as part of the rate constants. The model is given by:

```
# Goldbeter, A. A minimal cascade model for the mitotic
# oscillator involving cyclin and cdc2 kinase. Proc. Natl.
# Acad. Sci. USA 88:9107-1101 (1991).
$REACTIONS
[C <=> Nil, rates[kd, vi]]
[C |--> Nil, mod[X], Hill[vd, 1, Kd, 0,1 ]]
[M |--> Nil, mod[Nil], Hill[v2, 1, K2, 0, 1]]
[X |--> Nil, mod[Nil], Hill[v4, 1, K4, 0, 1]]
[Nil -> X, " vm3 * M * (1-X)/(K3+1-X)"]
[C |-> M, Hill[" vm1*(1-M)/(K1+1-M)", 1, Kc, 0, 1]]
$IC
C = 0.1
M = 0.2
X = 0.3
$RATES
vd = 0.1
vi = 0.023
v2 = 0.167
v4 = 0.1
vm1 = 0.5
vm3 = 0.2
kd = 0.00333
K1 = 0.1
K2 = 0.1
K3 = 0.1
K4 = 0.1
Kc = 0.3
Kd = 0.02
$
```

The syntax for running the model is

```
$ python pycellerator.py solve -in Goldbeter.model -plot -run 100 .1
-sameplot
```



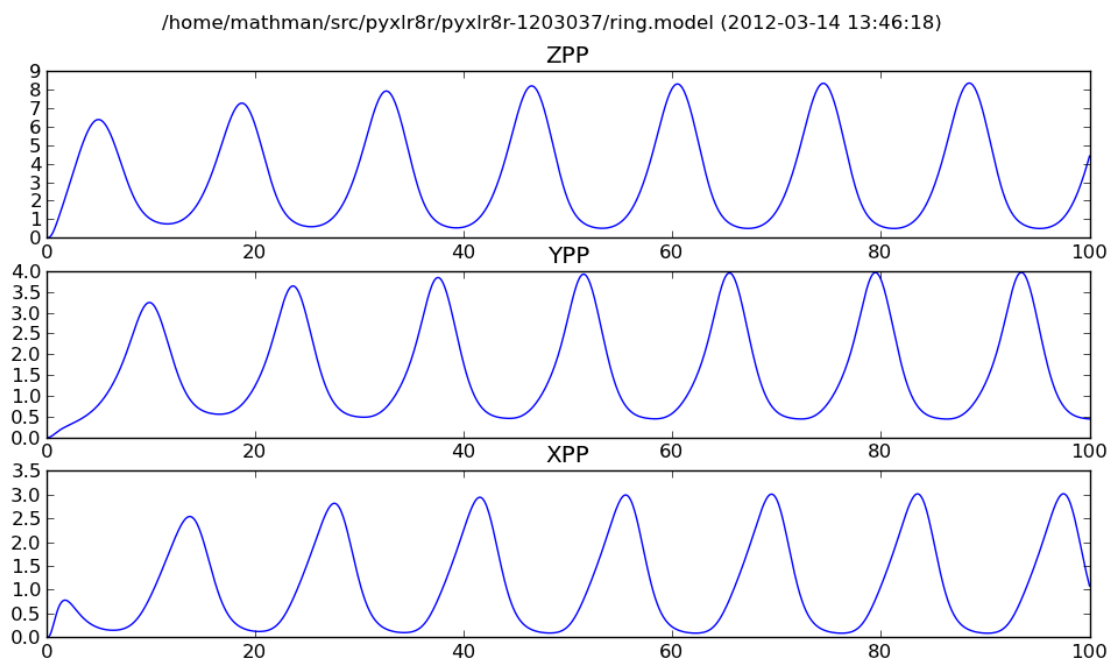
5.6 Ring Oscillator Using Mass Action Equations

The following model describes a ring oscillator using only mass-action equations:

```
$Reactions
[X <=> XP,    mod[Z, ZPP], rates[a, d, k, 0, a, d, k]]
[XP <=> XPP,   mod[Z, ZPP], rates[a, d, k, 0, a, d, k]]
[Y <=> YP,     mod[X, XPP], rates[a, d, k, 0, a, d, k]]
[YP <=> YPP,   mod[X, XPP], rates[a, d, k, 0, a, d, k]]
[Z <=> ZP,     mod[Y, YPP], rates[a, d, k, 0, a, d, k]]
[ZP <=> ZPP,   mod[Y, YPP], rates[a, d, k, 0, a, d, k]]
$IC
X = 10.0
Y = 15.0
Z = 20.0
$Rates
a = 1.0
d = 1.0
k = 1.0
$
```

Syntax:

```
python pycellerator.py solver -in ringma.model -plot -run 100 .1
-plotcolumns 1
```



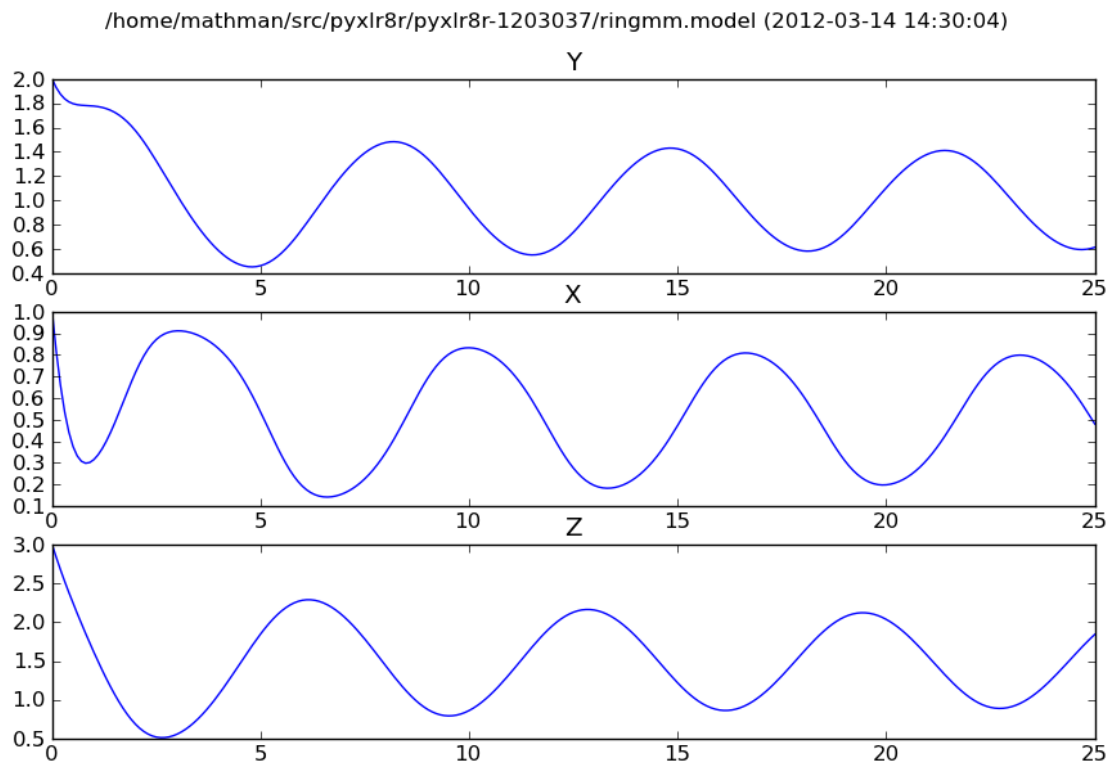
5.7 Ring Oscillator Using Michaelis-Menten-Henri Reactions

```

$Reactions
[X :--> XP, mod[Z], MMH[K,v]]
[XP:--> X, mod[XP],MMH[K,v]]
[Y :--> YP, mod[X], MMH[K,v]]
[YP:--> Y, mod[XP],MMH[K,v]]
[Z :--> ZP, mod[Y], MMH[K,v]]
[ZP:--> Z, mod[YP],MMH[K,v]]
$IC
X = 1.0
Y = 2.0
Z = 3.0
$Rates
v=1.0
K=0.5
$

```

```
python pycellerator solve -in ringmm.model -plot -run 25 .1 -plotcolumns 1
```



5.8 Ring Oscillator Using GRN Reactions

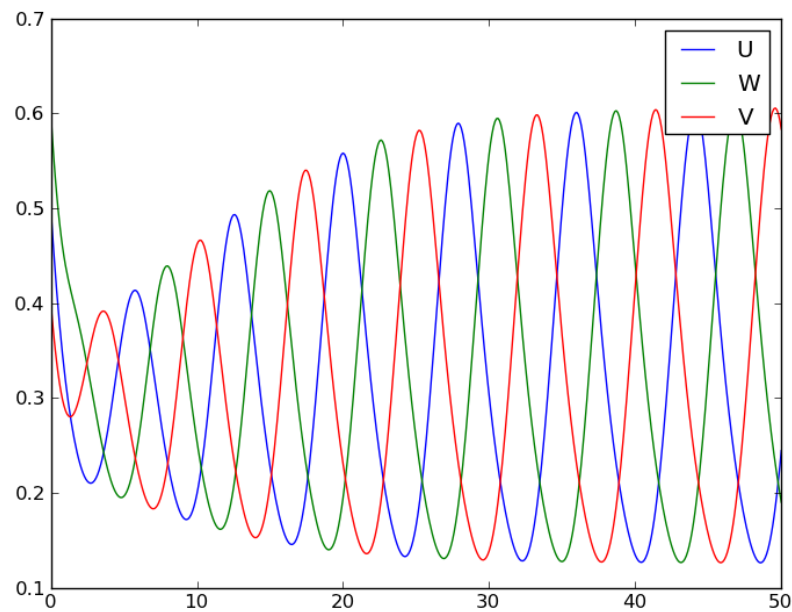
```

$Reactions
[U |-> V, GRN[v, b, n, h]]
[V |-> W, GRN[v, b, n, h]]
[W |-> U, GRN[v, b, n, h]]
[V -> Nil, k]
[U -> Nil, k]
[W -> Nil, k]
$IC
U = .5
V = .4
W = 0.6
$Rates
v = 1
n = 2
h = 1
k = 0.5
b = -10
$

```

```
python pycelerator solve -in ringgrn.model -plot -run 50 .1 -sameplot
```

/home/mathman/src/pyxl8r/pyxl8r-1203037/ringgrn.model (2012-03-14 22:47:47)



5.9 Ring Oscillator Using NHCA Reactions

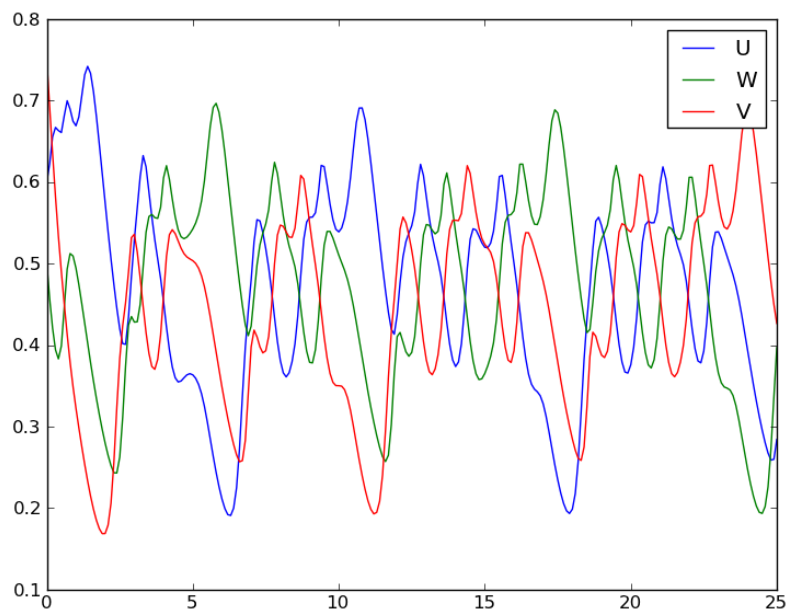
```

$Reactions
[U |-> V, NHCA[v, TP, TM, n, m, K] ]
[V |-> W, NHCA[v, TP, TM, n, m, K] ]
[W |-> U, NHCA[v, TP, TM, n, m, K] ]
[V -> Nil, k]
[U -> Nil, k]
[W -> Nil, k]
$IC
U = .6
V = .75
W = .5
$Rates
v = 1
n = 2
m = 2
K = 10
k = 1
TP=.2
TM=-5
$

```

```
python pycellerator.py solve -in ringnhca.model -plot -run 25 .1 -sameplot
```

/home/mathman/src/pyxl8r/pyxl8r-1203038/ringnhca.model (2012-03-15 11:13:14)

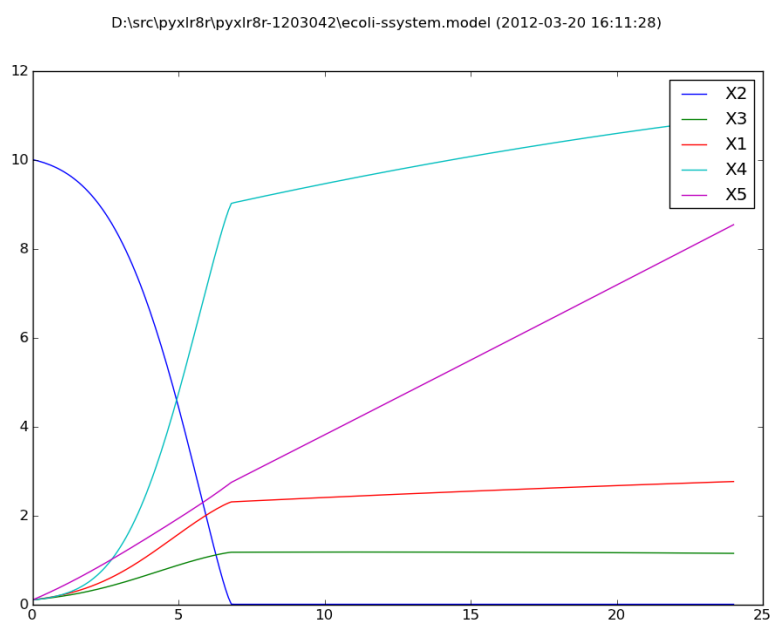


5.10 Recombinant *E coli* growth model

This model uses an S-System approach that is given by [11].

```
$reactions
[[X1,X2] |-> X1,    SSystem[1, a1, b1, [g11,g12], [h11,h12]]]
[[X1,X2] |-> X2,    SSystem[1, a2, b2, [0, 0], [h21, h22]]]
[[X1,X2,X3] |-> X3, SSystem[1, a3, b3, [g31,g32,0], [h31,h32,h33]]]
[[X1,X2,X4] |-> X4, SSystem[1, a4, b4, [g41,g42,0], [h41,h42,h44]]]
[[X1,X2,X5] |-> X5, SSystem[1, a5, b5, [g51,g52,0], [h51,h52,h55]]]
$ic
X1 = 0.1
X2 = 10
X3 = 0.1
X4 = 0.1
X5 = 0.1
$rates
a1 = 0.4973
a2 = 0.0817
a3 = 0.2858
a4 = 3.7124
a5 = 0.4562
b1 = 0.1648
b2 = 1.2484
b3 = 0.1285
b4 = 2.5318
b5 = 0.0335
g11 = 0.9099
g31 = 0.7366
g41 = 1.7076
g51 = 0.2292
g12 = 0.1301
g32 = 0.1311
g42 = 0.1252
g52 = 0.0277
h11 = 1.7514
h21 = 0.9325
h31 = 1.3535
h41 = 1.9875
h51 = 1.1978
h12 = 0.1292
h22 = 0.1927
h32 = 0.1175
h42 = 0.1210
h52 = 0.4462
h33 = -0.0110
h44 = -0.0100
h55 = -0.0426
$
```

```
python pycellerator.py solve -in ecoli.model -plot -run 24 .1 -sameplot
```



5.11 MAPK Cascade with Indexed Stages

```

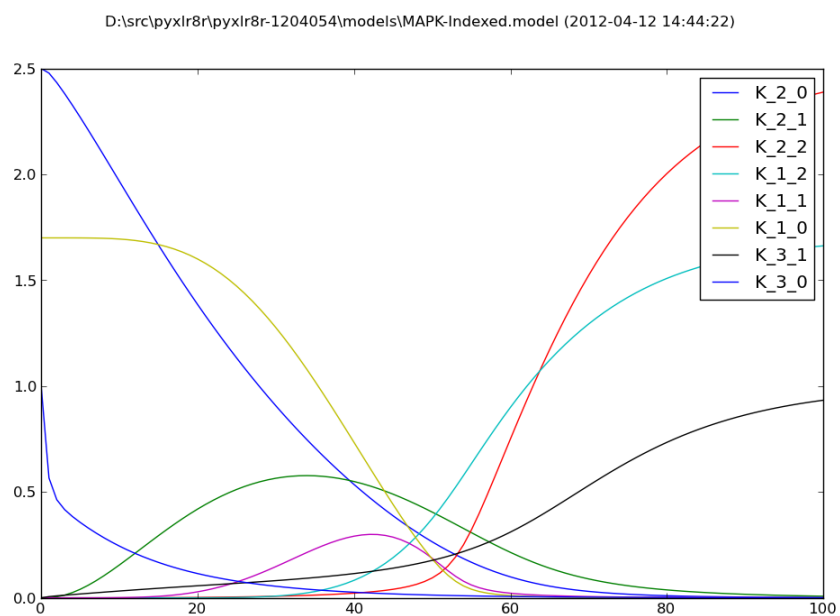
$Reactions
[K(3,0) <=> K(3,1), mod[RAFK, RAFph],
  rates[a1,d1,k1,0,a2,d2,k2,0]]
[K(2,0) <=> K(2,1) <=> K(2,2), mod[ K(3,1), MEKph ],
  rates[ a3, d3, k3, 0, a4, d4, k4, 0, a5, d5, k5, 0, a6, d6, k6, 0]]
[K(1,0) <=> K(1,1) <=> K(1,2), mod[ K(2,2), MAPKph ],
  rates[ a7, d7, k7, 0, a8, d8, k8, 0, a9, d9, k9, 0, a10, d10, k10, 0]]
$IC
K(3,0) = 1
K(2,0) = 2.5
K(1,0) = 1.7
RAFK = 1
$Rates
a1=1.
a2=0.5
a3=3.3
a4=10.
a5=3.3
a6=10.
a7=20.
a8=5.
a9=20.
a10=5.
d1=.4
d2=.5
d3=.42
d4=.8
d5=.4
d6=.8
d7=.6
d8=.4
d9=.6
d10=.4
k1=.1
k2=.1
k3=.1
k4=.1
k5=.1
k6=.1
k7=.1
k8=.1
k9=.1
k10=.1
$

```

```

python pycellerator.py solve -in MAPK-Indexed.model -solve -plot K(1,0)
  K(1,1) K(1,2) K(2,0) K(2,1) K(2,2) K(3,0) K(3,1) -sameplot

```



Appendix A

File Formats

A.1 Reaction Files

Reaction files are lists of cellerator reactions, with at most one reaction per line. A single reaction may be spread over multiple lines if the last character on the line is the backslash (\) character. Reaction syntax is as described in chapter 3. Blank lines are ignored, and the cross-hatch character (#) indicates the beginning of a comment.¹ Comments are terminated by an end-of-line. An example is given by the following:

```
# Oregonator Data File
# Reference: Field and Noyes, J Chem Phys 60: 1877 (1974)
#
[Br + BrO3 -> HBrO2 + HOBr, k1]
[Br + HBrO2 -> 2*HOBr, k2]
[BrO3 + HBrO2 -> 2*Ce + 2*HBrO2, k3]
[2*HBrO2-> BrO3 + HOBr, k4]
[Ce -> 0.5*Br, k5]
```

A.2 Model Files

A model file contains reactions, initial conditions, rate constants, function definitions (optional) and frozen variables (optional). Each of theses lists (except for the reactions) is optional. Each lists is indicated by an indicator **\$Assignments**, **\$Functions**, **\$Reactions**, **\$IC**, **\$Rates** or **\$Frozen** as illustrated in the following example. The order of the different sections does not matter, i.e., **Rates** may precede **IC**, etc. Comments and blank lines are ignored

The **\$Assignments** section should be followed by a list of assignment statements of the form

```
variable = expression
```

where **variable** is any model variable and **expression** is any valid python expression, such as

```
A = B + C - 4D
C = pow(B, 4) +A
```

The **\$Functions** section contains a list of functions that may be reference elsewhere in the model. Functions are defined with the syntax

¹The crosshatch was chosen as the comment delimiter because it will be familiar to Python users.

```
fname(var1,var2,...) = expression
```

where **fname** is the name of the function; **var1,var2,...** are the arguments of the function, and **expression** is any valid python infix expression. For example,

```
f(m, x) = m * (1-x) / (K3+1-x)
g(m) = (1-m) / (K1+1-m)
```

In the first function definition, the function has two arguments, **m** and **x**, which must be passed to it; the function must be then referenced as, say **f(P,Q)** in a kinetic law, which would represent the expression $P \cdot (1-Q) / (K3+1-Q)$. The reference to **K3** is a reference to a global parameter in the model. The second function has only one variable; a reference **g(U)** in a kinetic law would represent the expression $(1-U) / (K1+1-U)$ where **K1** is also a global parameter of the model.

An example of the **\$Functions** keyword is illustrated by this alternative version of the Goldbeter mitotic model (compare to the one given in section 5.5):

```
$REACTIONS
[C <=> Nil, rates[kd, vi]]
[C |--> Nil, mod[X], Hill[vd, 1, Kd, 0,1 ]]
[M |--> Nil, mod[Nil], Hill[v2, 1, K2, 0, 1]]
[X |--> Nil, mod[Nil], Hill[v4, 1, K4, 0, 1]]
[Nil -> X, "vm3*f(M,X)"]
[C |-> M, Hill["vm1*g(M)", 1, Kc, 0, 1]]
$IC
C = 0.1
M = 0.2
X = 0.3
$FUNCTIONS
f(m,x) = m * (1-x) / (K3+1-x)
g(m) = (1-m) / (K1+1-m)
$RATES
vd = 0.1
vi = 0.023
v2 = 0.167
v4 = 0.1
vm1 = 0.5
vm3 = 0.2
kd = 0.00333
K1 = 0.1
K2 = 0.1
K3 = 0.1
K4 = 0.1
Kc = 0.3
Kd = 0.02
```

The **\$Reactions** section contains a list of Reactions as described in Chapter 3, e.g.,

```
[Br + BrO3 -> HBrO2 + HOBr, k1]
```

The **\$Frozen** section contains a list of frozen variables, one variable per line. Frozen variables do not normally change their values in a reaction; however, they are allowed to change their values in (a) an assignment rule; or (b) a **using** reaction (which corresponds to an SBML rate rule).

An example of frozen variables is given in the Oregonator mode file.

```
# Oregonator Data File
# Reference: Field and Noyes, J Chem Phys 60: 1877 (1974)
#
$Reactions
[Br + BrO3 -> HBrO2 + HOBr, k1]
[Br + HBrO2 -> 2*HOBr, k2]
[BrO3 + HBrO2 -> 2*Ce + 2*HBrO2, k3]
[2*HBrO2-> BrO3 + HOBr, k4] # but BrO3 is frozen
[Ce -> 0.5*Br, k5]
$IC
HBrO2 = .001
Br = .003
Ce = .05
BrO3 = .1
HOBr = 0
$Rates
k1 = 1.3
k2 = 2.0E6
k3 = 34
k4 = 3000.0
k5 = 0.02
$Frozen
BrO3
```

The \$IC section contains initial conditions, one per line, in the form **variable = value**

Appendix B

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Version 3, 29 June 2007

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