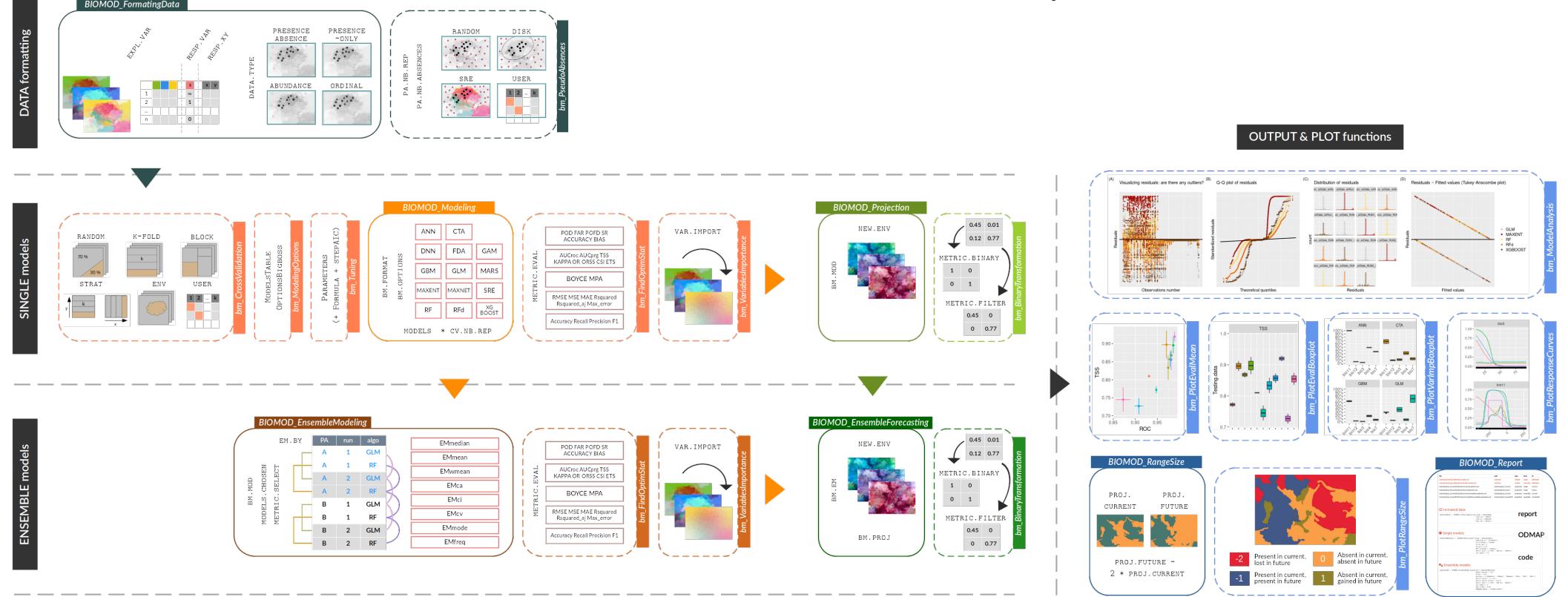


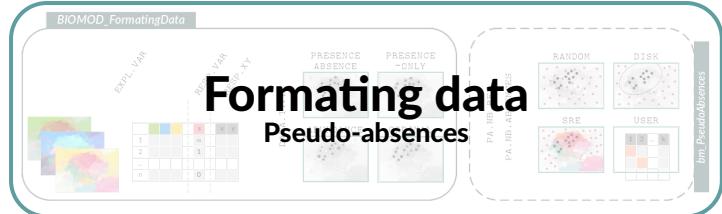
Species distribution modeling, calibration and evaluation, ensemble modeling



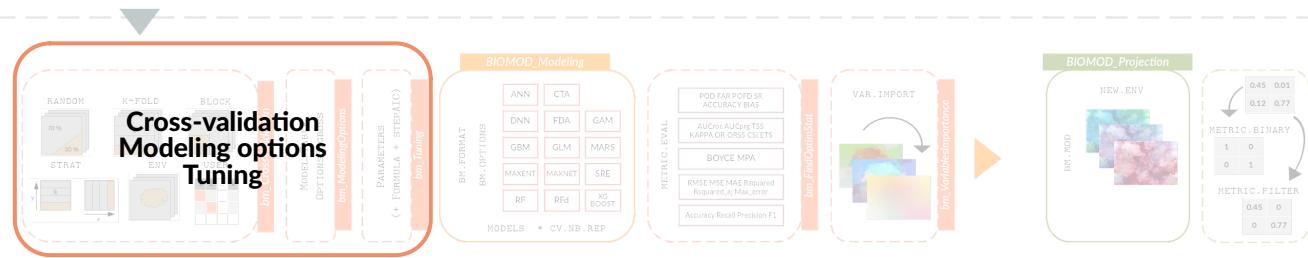
Species distribution modeling, calibration and evaluation, ensemble modeling



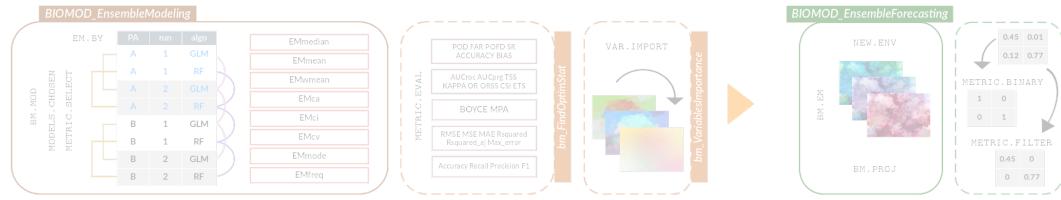
DATA formatting



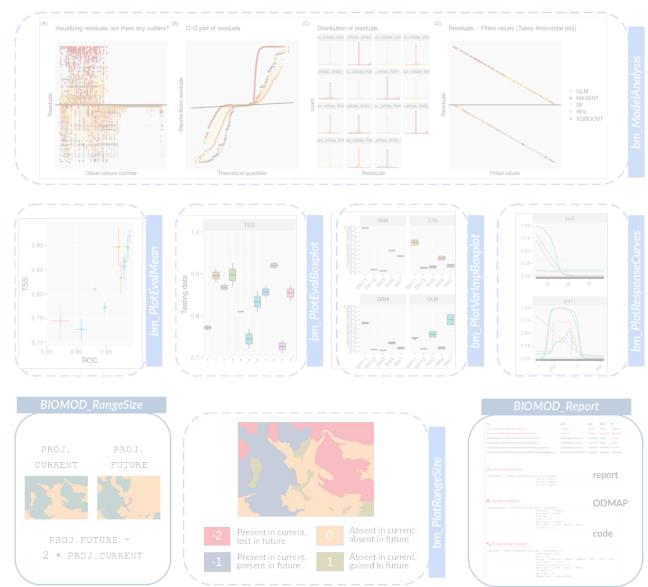
SINGLE models



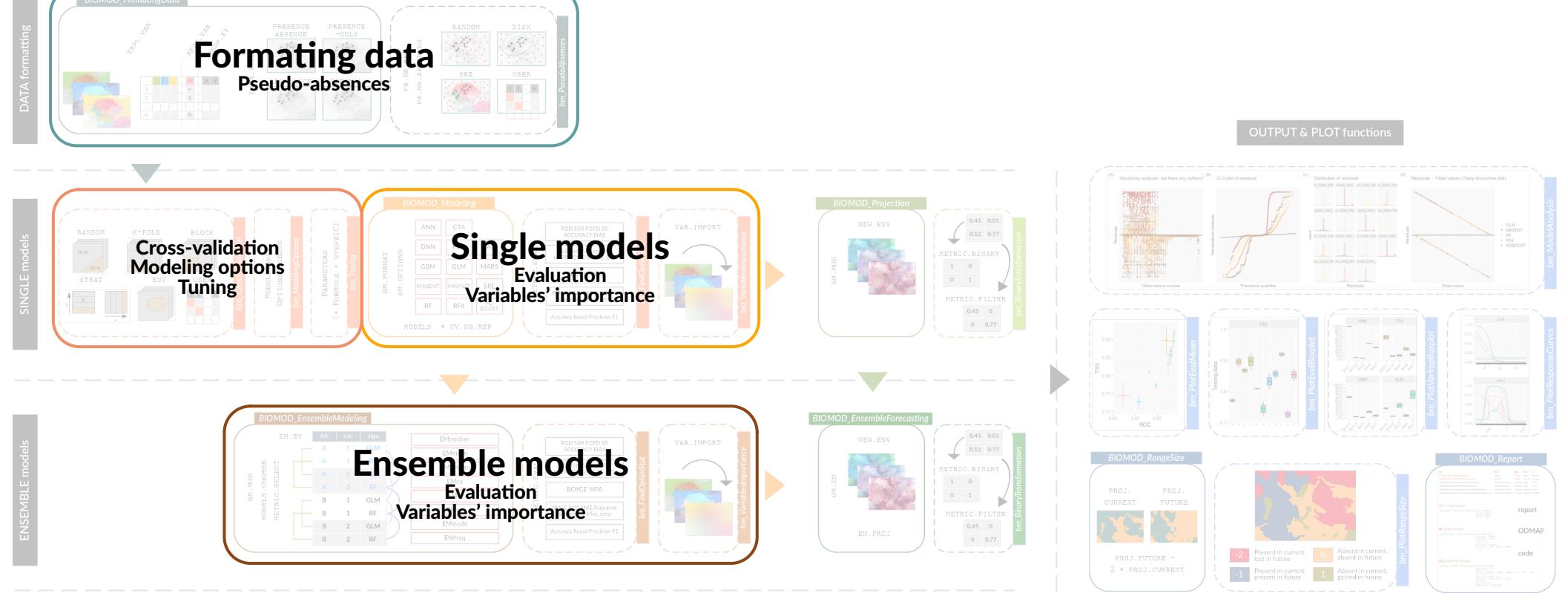
ENSEMBLE models



OUTPUT & PLOT functions



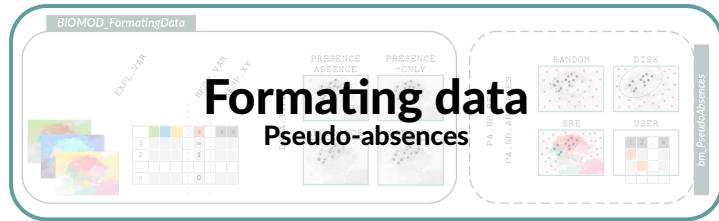
Species distribution modeling, calibration and evaluation, ensemble modeling



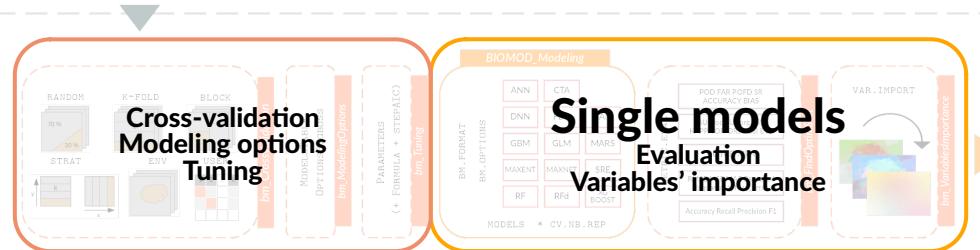
Species distribution modeling, calibration and evaluation, ensemble modeling



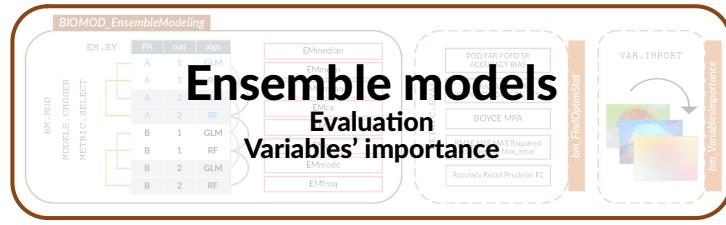
DATA formatting



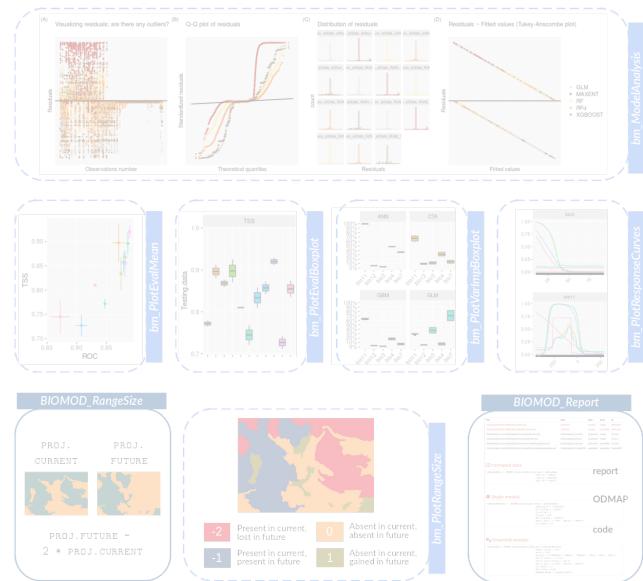
SINGLE models



ENSEMBLE models



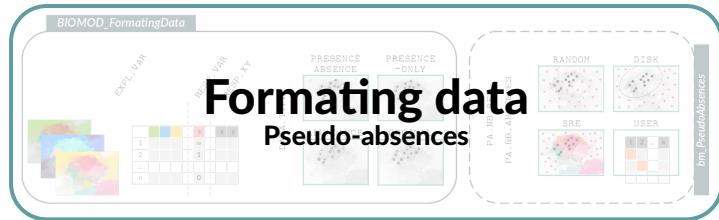
OUTPUT & PLOT functions



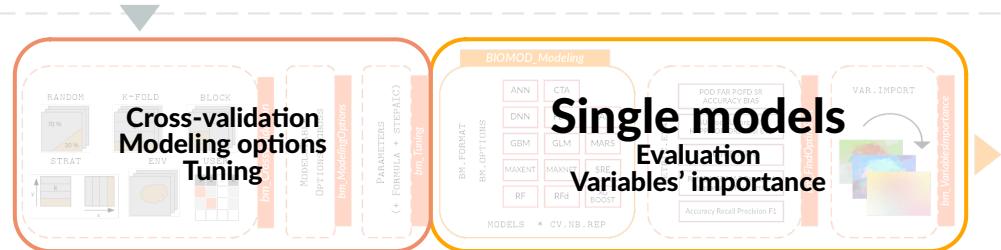
Species distribution modeling, calibration and evaluation, ensemble modeling



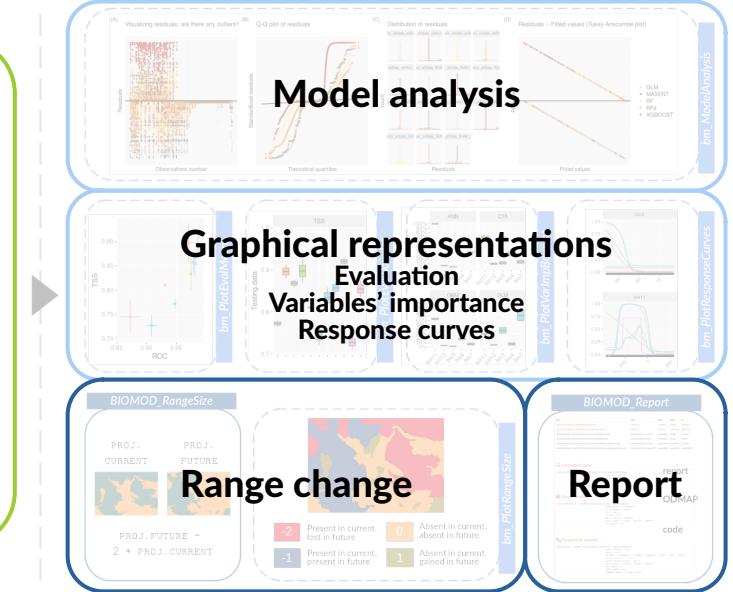
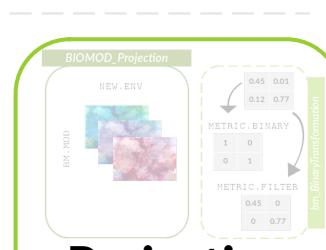
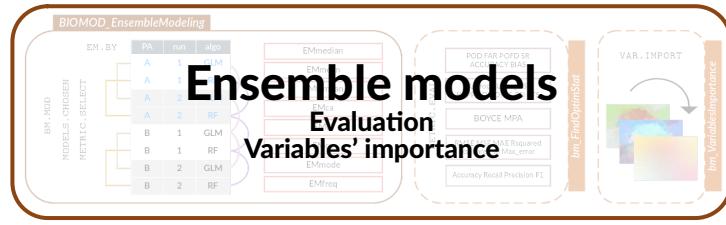
DATA formatting



SINGLE models

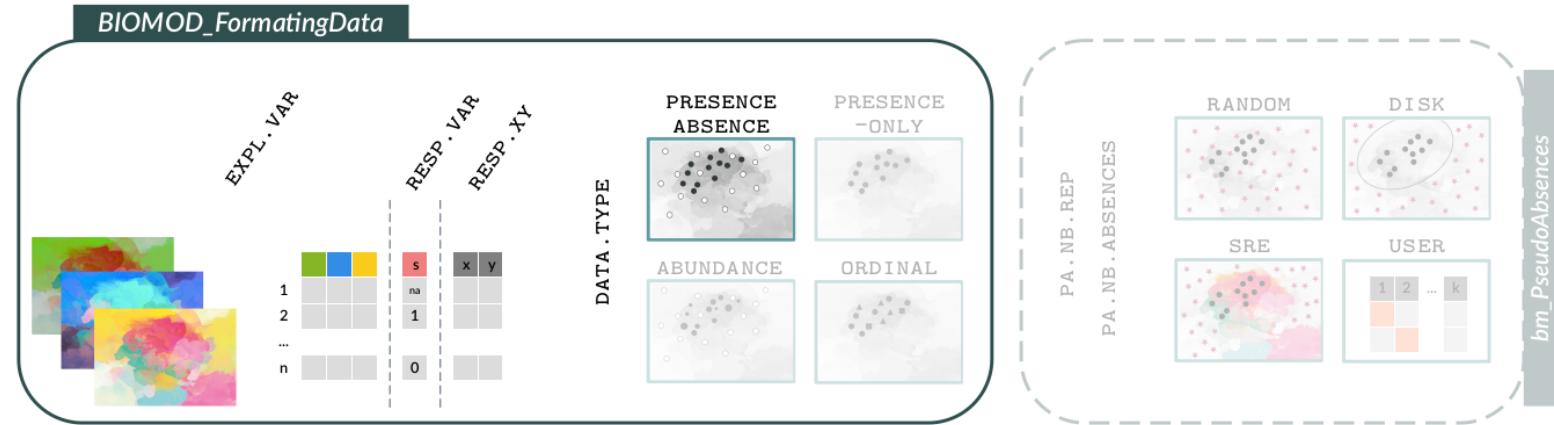


ENSEMBLE models



1. Formating data

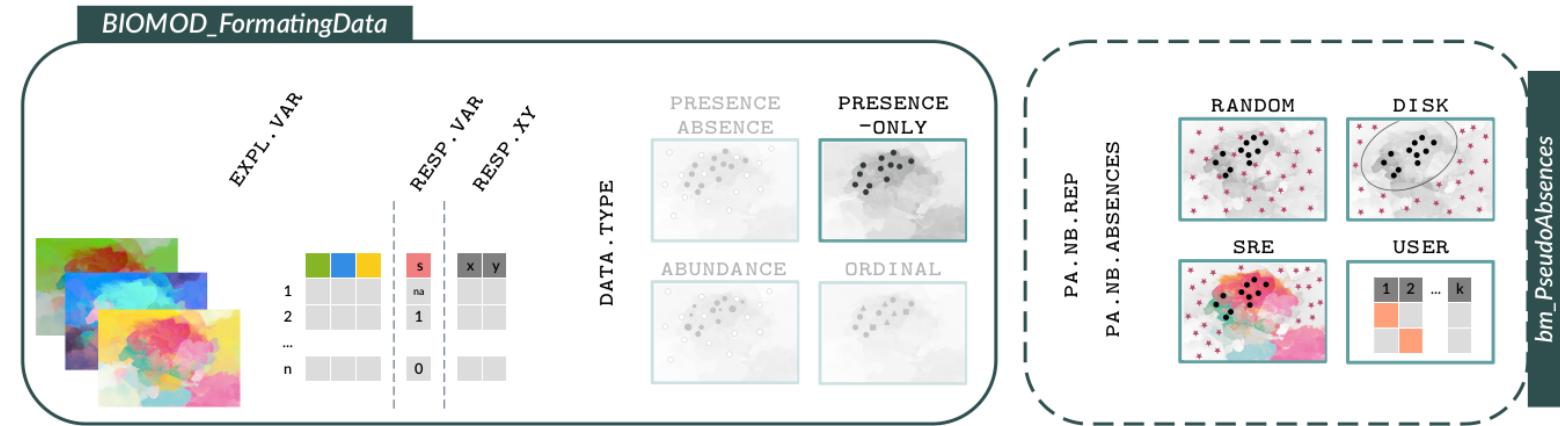
» presences-absences



Species occ
Environment

1. Formating data

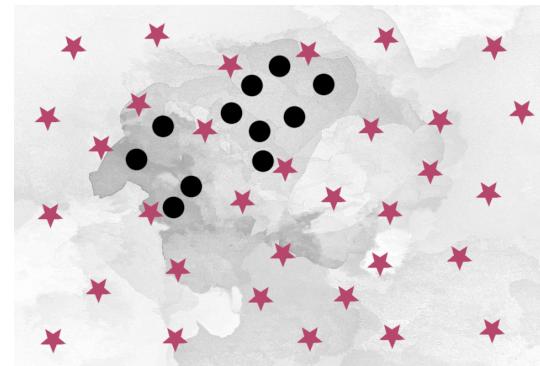
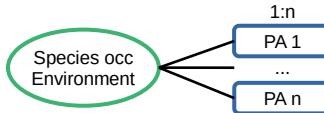
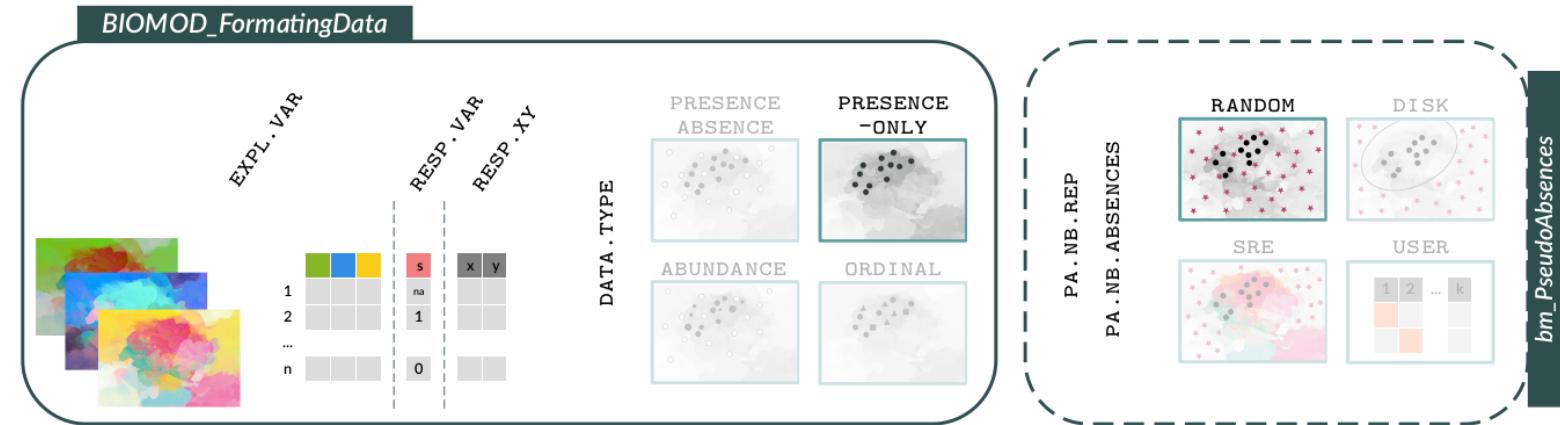
- » presence-only data
- » avoid to mix with real absences



Species occ
Environment

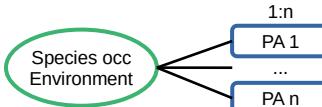
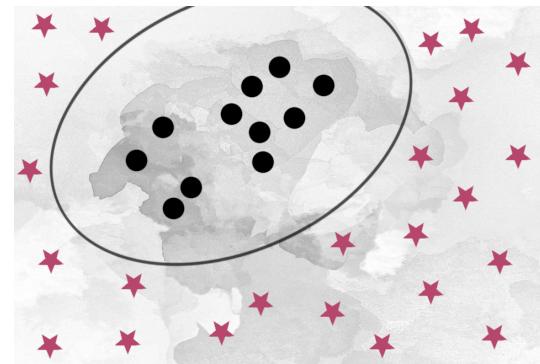
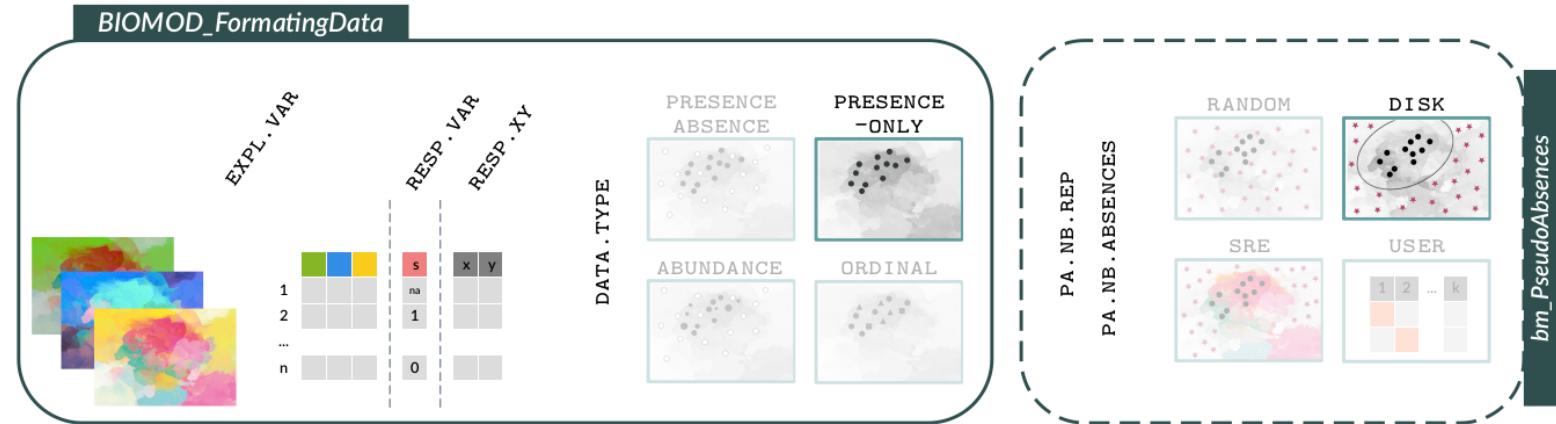
1. Formating data

- » presence-only data
- » avoid to mix with real absences
- » **random** : sampling potentially biased / non-exhaustive



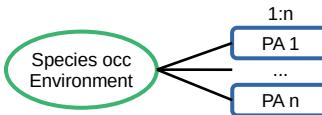
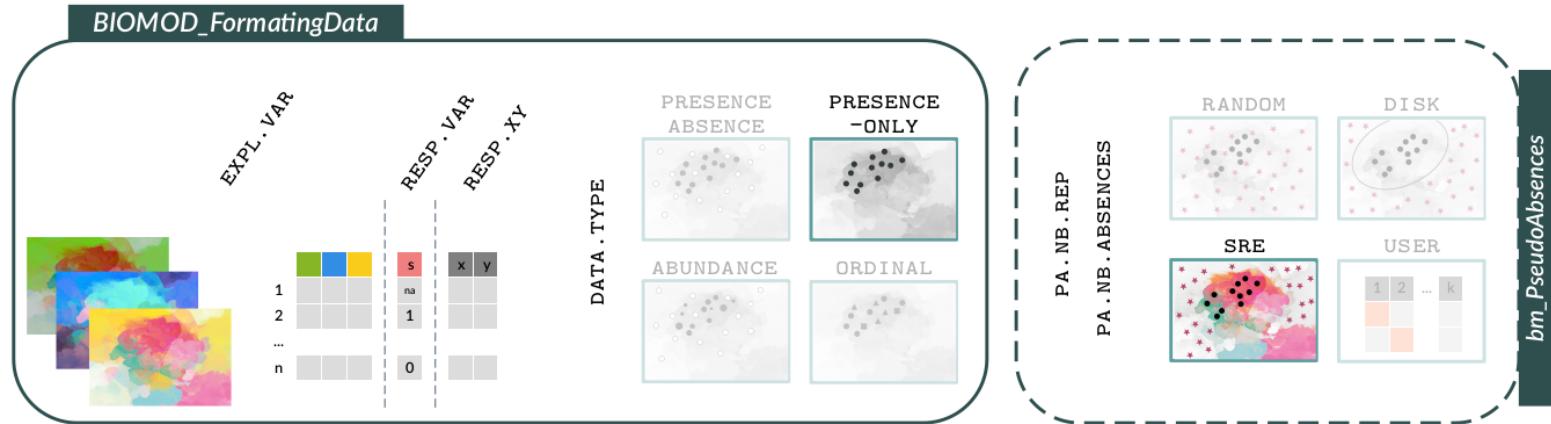
1. Formating data

- » **presence-only** data
- » avoid to mix with real absences
- » **random** : sampling potentially biased / non-exhaustive
- » **disk** : geographic niche well sampled



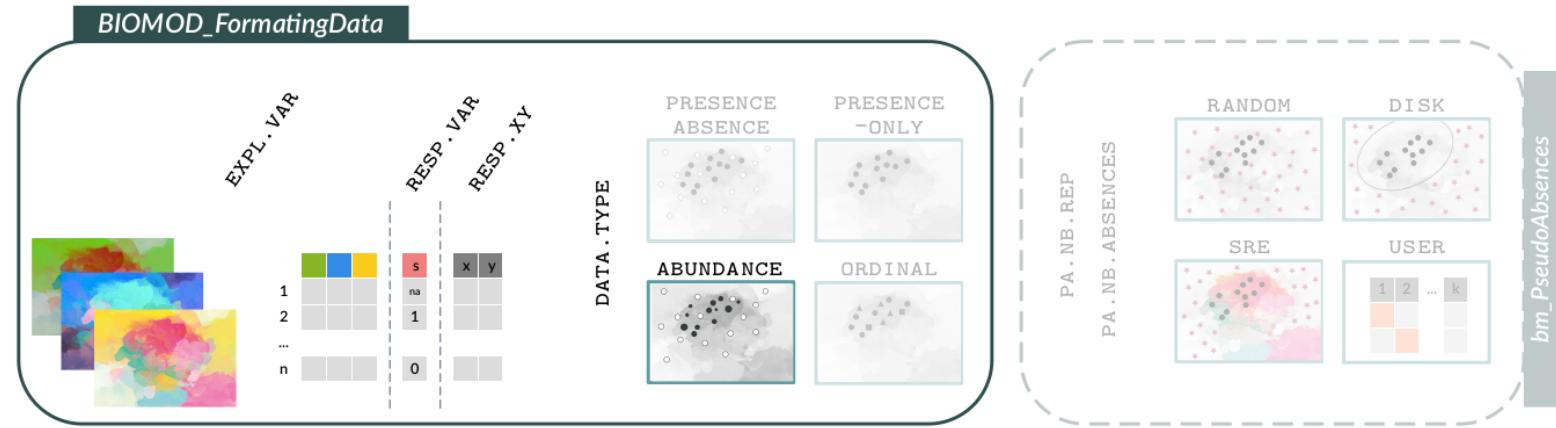
1. Formating data

- » presence-only data
- » avoid to mix with real absences
- » **random** : sampling potentially biased / non-exhaustive
- » **disk** : geographic niche well sampled
- » **SRE** : environmental niche well sampled



1. Formating data

» abundance data



Species occ
Environment

1. Formating data

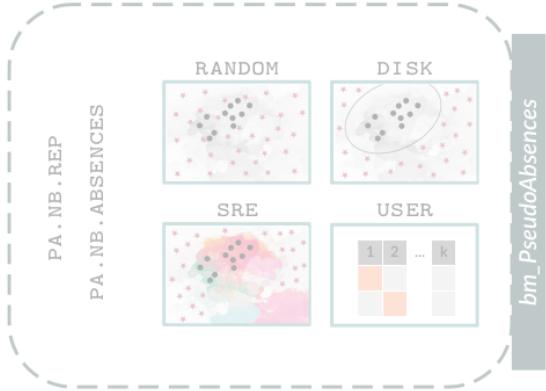
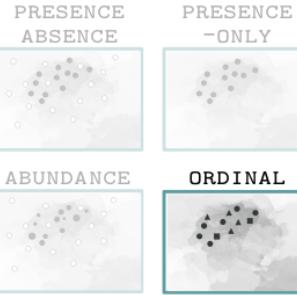
BIOMOD_FormatingData

EXPL · VAR
RESP · VAR
RESP · X_T



1			
2			
..			
n			

DATA · TYPE



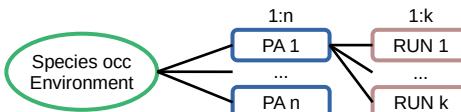
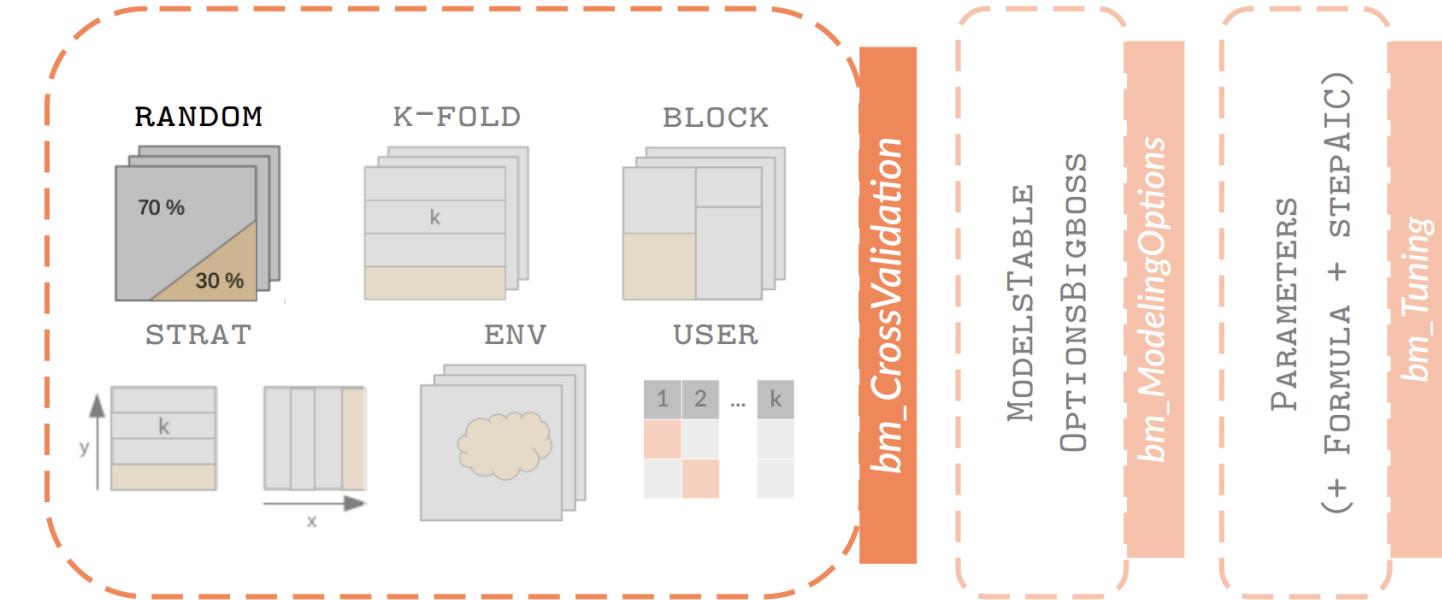
» **ordinal** data



Species occ
Environment

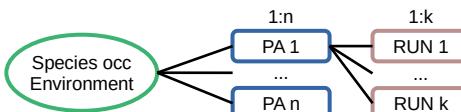
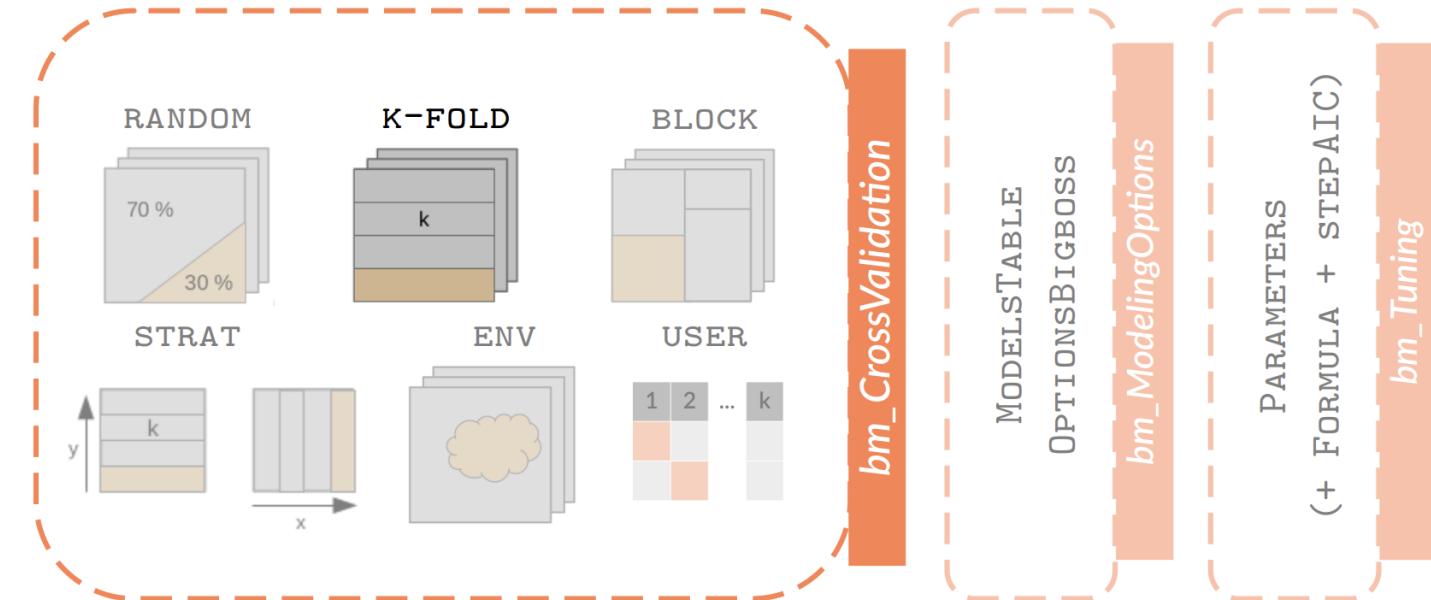
1. Formating data

- » simple calibration / validation split at the modeling step, and repeated nb.rep times



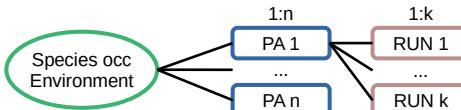
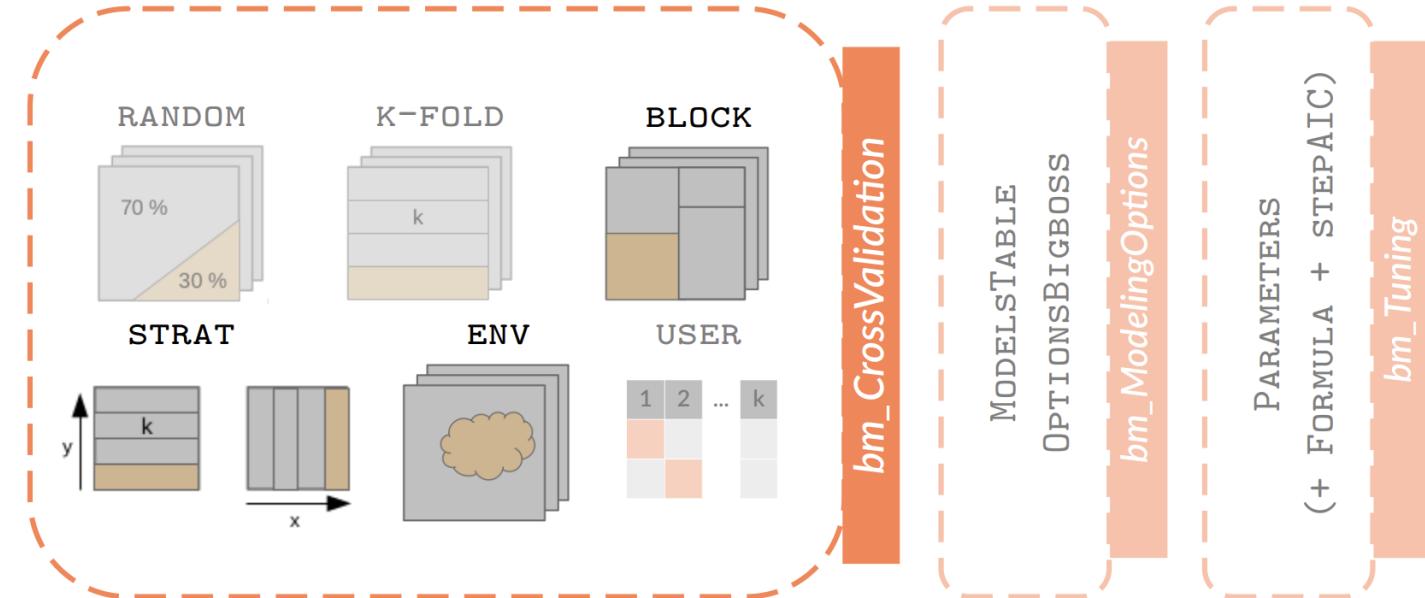
1. Formating data

- » simple calibration / validation split at the modeling step, and repeated nb.rep times
- » **k-fold** : partition data into k sub-dataset, and repeated nb.rep times



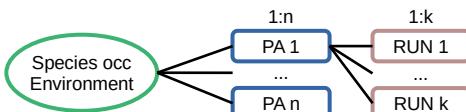
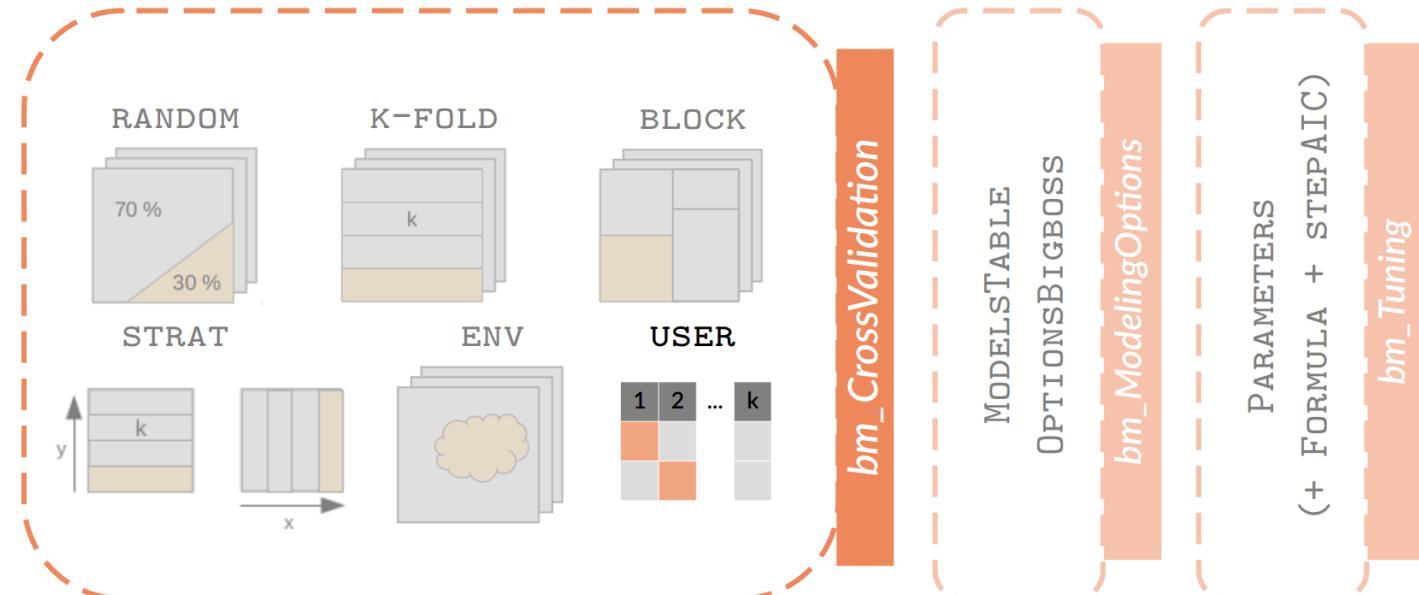
1. Formating data

- » simple calibration / validation split at the modeling step, and repeated nb.rep times
- » **k-fold** : partition data into k sub-dataset, and repeated nb.rep times
- » **stratified** : partition data into k sub-dataset (x, y, both, block, env)



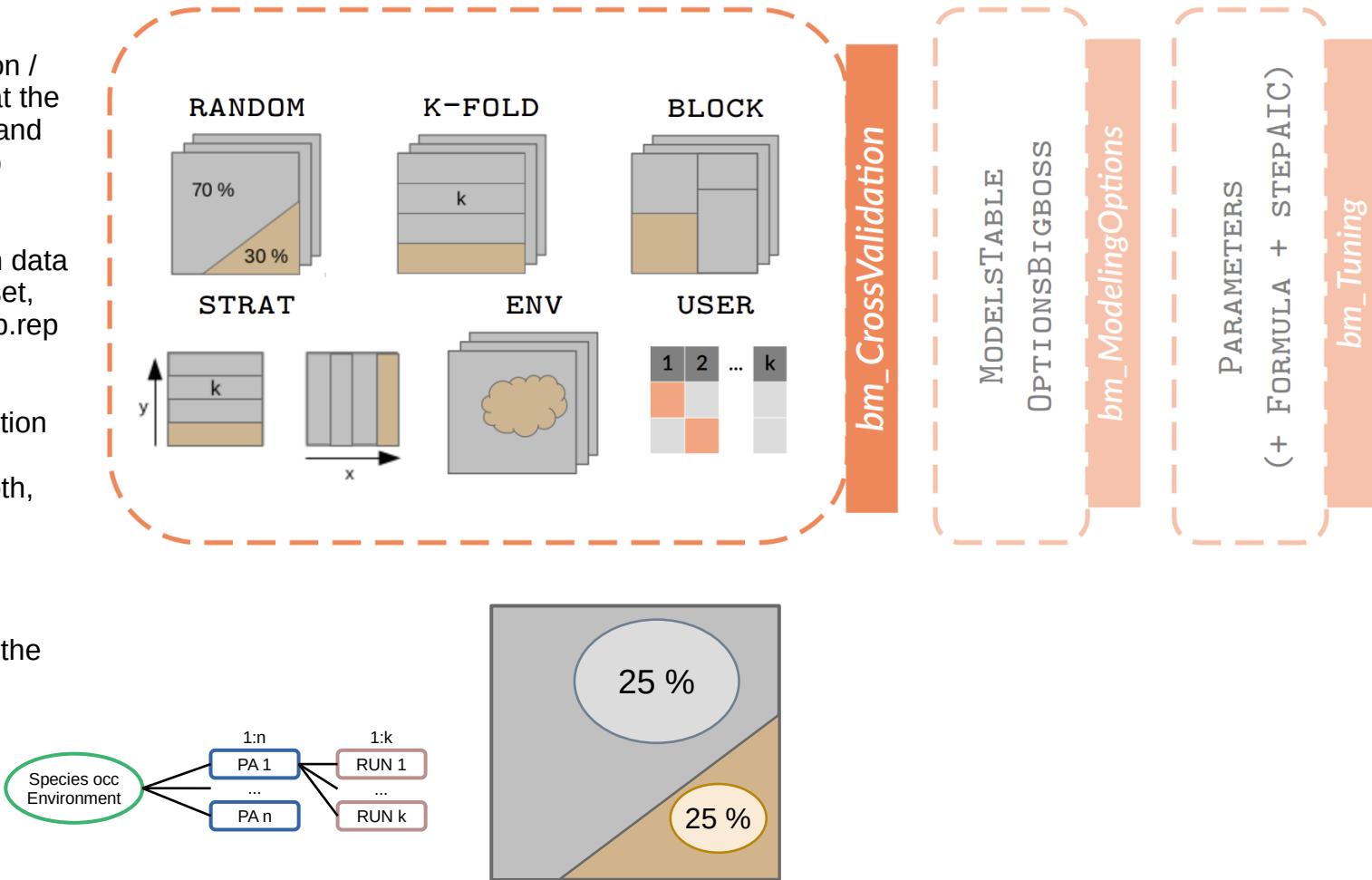
1. Formating data

- » simple calibration / validation split at the modeling step, and repeated nb.rep times
- » **k-fold** : partition data into k sub-dataset, and repeated nb.rep times
- » **stratified** : partition data into k sub-dataset (x, y, both, block, env)
- » **user defined**



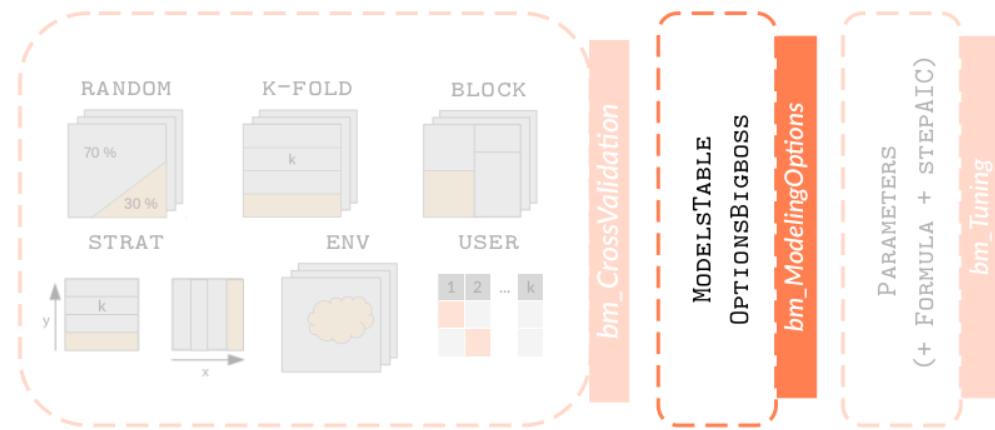
1. Formating data

- » simple calibration / validation split at the modeling step, and repeated nb.rep times
- » **k-fold** : partition data into k sub-dataset, and repeated nb.rep times
- » **stratified** : partition data into k sub-dataset (x, y, both, block, env)
- » **user defined**
- » **balance** : keep the prevalence of presences (or absences) in sub-dataset

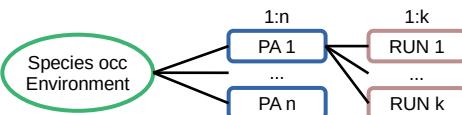


1. Formating data

- » 12 types of models,
16 single models,
11 that can use non-binary data
- » 1 coded in biomod2,
1 external software,
13 other R packages



	model	type	package	func	train
1	ANN	binary	nnet	nnet	avNNet
2	CTA	binary	rpart	rpart	rpart
3	DNN	binary	cito	dnn	tune
4	FDA	binary	mda	fda	fda
5	GAM	binary	gam	gam	gamLoess
6	GAM	binary	mgcv	bam	bam
7	GAM	binary	mgcv	gam	gam
8	GBM	binary	gbm	gbm	gbm
9	GLM	binary	stats	glm	glm
10	MARS	binary	earth	earth	earth
11	MAXENT	binary	MAXENT	MAXENT	ENMEvaluate
12	MAXNET	binary	maxnet	maxnet	maxnet
13	RF	binary	randomForest	randomForest	rf
14	Rfd	binary	randomForest	randomForest	rf
15	SRE	binary	biomod2	bm_SRE	bm_SRE
16	XGBOOST	binary	xgboost	xgboost	xgbTree
17	CTA	nonbinary	rpart	rpart	rpart
18	DNN	nonbinary	cito	dnn	tune
19	FDA	nonbinary	mda	fda	fda
20	GAM	nonbinary	gam	gam	gamLoess
21	GAM	nonbinary	mgcv	bam	bam
22	GAM	nonbinary	mgcv	gam	gam
23	GBM	nonbinary	gbm	gbm	gbm
24	GLM	nonbinary	stats	glm	glm
25	MARS	nonbinary	earth	earth	earth
26	RF	nonbinary	randomForest	randomForest	rf
27	XGBOOST	nonbinary	xgboost	xgboost	xgbTree



1. Formating data

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16 single models,
11 that can use non-binary data
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13 other R packages
- » **default** : extracted from functions

nnet package:nnet

[Fit Neural Networks](#)

Description:

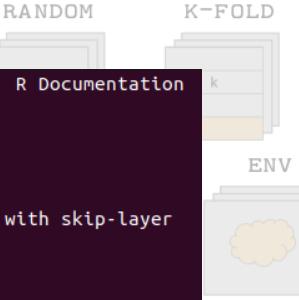
Fit single-hidden-layer neural network, possibly with skip-layer connections.

Usage:

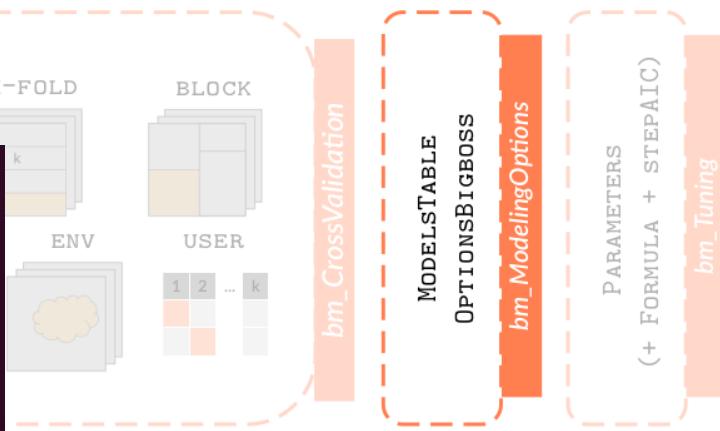
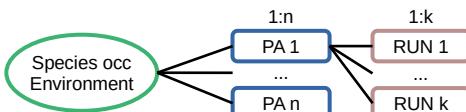
```
nnet(x, ...)

## S3 method for class 'formula'
nnet(formula, data, weights, ...,
     subset, na.action, contrasts = NULL)

## Default S3 method:
nnet(x, y, weights, size, Wts, mask,
     linout = FALSE, entropy = FALSE, softmax = FALSE,
     censored = FALSE, skip = FALSE, rang = 0.7, decay = 0,
     maxit = 100, Hess = FALSE, trace = TRUE, MaxNWts = 1000,
     abstol = 1.0e-4, reltol = 1.0e-8, ...)
```



	model	type	package	func	train
1	ANN	binary	nnet	nnet	avNNet
2	CTA	binary	rpart	rpart	rpart
3	DNN	binary	cito	dnn	tune
4	FDA	binary	mda	fda	fda
5	GAM	binary	gam	gam	gamLoess
6	GAM	binary	mgcv	bam	bam
7	GAM	binary	mgcv	gam	gam
8	GBM	binary	gbm	gbm	gbm
9	GLM	binary	stats	glm	glm
10	MARS	binary	earth	earth	earth
11	MAXENT	binary	MAXENT	MAXENT	ENMEvaluate
12	MAXNET	binary	maxnet	maxnet	maxnet
13	RF	binary	randomForest	randomForest	rf
14	Rfd	binary	randomForest	randomForest	rf
15	SRE	binary	biomod2	bm_SRE	bm_SRE
16	XGBOOST	binary	xgboost	xgboost	xgbTree
17	CTA	nonbinary	rpart	rpart	rpart
18	DNN	nonbinary	cito	dnn	tune
19	FDA	nonbinary	mda	fda	fda
20	GAM	nonbinary	gam	gam	gamLoess
21	GAM	nonbinary	mgcv	bam	bam
22	GAM	nonbinary	mgcv	gam	gam
23	GBM	nonbinary	gbm	gbm	gbm
24	GLM	nonbinary	stats	glm	glm
25	MARS	nonbinary	earth	earth	earth
26	RF	nonbinary	randomForest	randomForest	rf
27	XGBOOST	nonbinary	xgboost	xgboost	xgbTree



1. Formating data

- » 12 types of models,
16 single models,
11 that can use non-binary data
- » 1 coded in biomod2,
1 external software,
13 other R packages
- » **default** : extracted from functions
- » **bigboss** : redefined by biomod2 team

nnet package:nnet

[Fit Neural Networks](#)

Description:

Fit single-hidden-layer neural network, possibly with skip-layer connections.

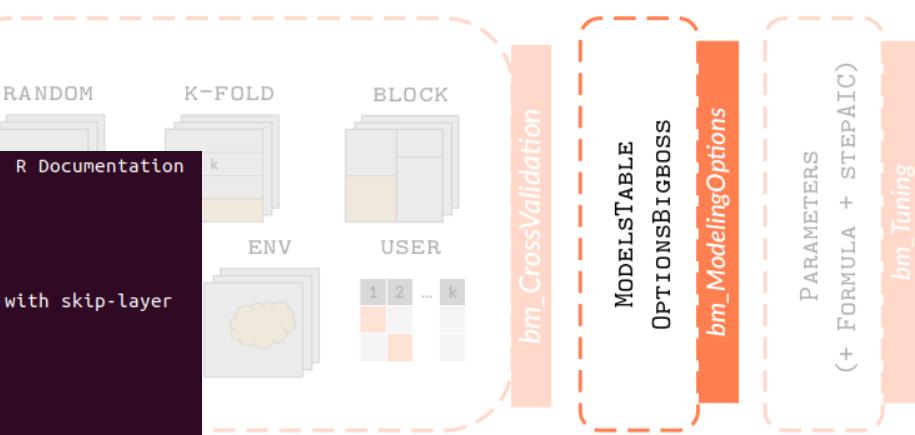
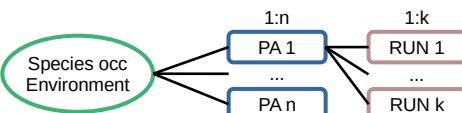
Usage:

```
nnet(x, ...)

## S3 method for class 'formula'
nnet(formula, data, weights, ...,
     subset, na.action, contrasts = NULL)

## Default S3 method:
nnet(x, y, weights, size, Wts, mask,
     linout = FALSE, entropy = FALSE, softmax = FALSE,
     censored = FALSE, skip = FALSE, rang = 0.7, decay = 0,
     maxit = 100, Hess = FALSE, trace = TRUE, MaxNWts = 1000,
     abstol = 1.0e-4, reltol = 1.0e-8, ...)
```

```
> ANN options (datatype: binary , package: nnet , function: nnet ) :
( dataset_allData_allRun )
  - size = 5 (default: 2 )
  - decay = 5 (default: NULL )
  - trace = FALSE (default: NULL )
  - rang = 0.1 (default: NULL )
  - maxit = 200 (default: NULL )
```



	model	type	package	func	train
1	ANN	binary	nnet	nnet	avNNet
2	CTA	binary	rpart	rpart	rpart
3	DNN	binary	cito	dnn	tune
4	FDA	binary	mda	fda	fda
5	GAM	binary	gam	gam	gamLoess
6	GAM	binary	mgcv	bam	bam
7	GAM	binary	mgcv	gam	gam
8	GBM	binary	gbm	gbm	gbm
9	GLM	binary	stats	glm	glm
10	MARS	binary	earth	earth	earth
11	MAXENT	binary	MAXENT	MAXENT	ENMEvaluate
12	MAXNET	binary	maxnet	maxnet	maxnet
13	RF	binary	randomForest	randomForest	rf
14	Rfd	binary	randomForest	randomForest	rf
15	SRE	binary	biomod2	bm_SRE	bm_SRE
16	XGBOOST	binary	xgboost	xgboost	xgbTree
17	CTA	nonbinary	rpart	rpart	rpart
18	DNN	nonbinary	cito	dnn	tune
19	FDA	nonbinary	mda	fda	fda
20	GAM	nonbinary	gam	gam	gamLoess
21	GAM	nonbinary	mgcv	bam	bam
22	GAM	nonbinary	mgcv	gam	gam
23	GBM	nonbinary	gbm	gbm	gbm
24	GLM	nonbinary	stats	glm	glm
25	MARS	nonbinary	earth	earth	earth
26	RF	nonbinary	randomForest	randomForest	rf
27	XGBOOST	nonbinary	xgboost	xgboost	xgbTree

1. Formating data

- » 12 types of models, 16 single models, 11 that can use non-binary data
- » 1 coded in biomod2, 1 external software, 13 other R packages
- » **default** : extracted from functions
- » **bigboss** : redefined by biomod2 team
- » **user-defined**

nnet package:nnet

[Fit Neural Networks](#)

Description:

Fit single-hidden-layer neural network, possibly with skip-layer connections.

Usage:

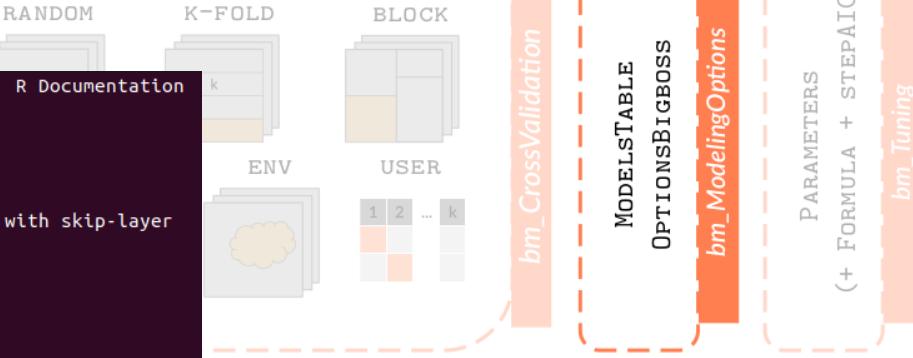
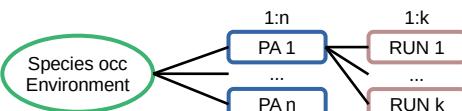
```
nnet(x, ...)

## S3 method for class 'formula'
nnet(formula, data, weights, ...,
     subset, na.action, contrasts = NULL)

## Default S3 method:
nnet(x, y, weights, size, Wts, mask,
     linout = FALSE, entropy = FALSE, softmax = FALSE,
     censored = FALSE, skip = FALSE, rang = 0.7, decay = 0,
     maxit = 100, Hess = FALSE, trace = TRUE, MaxNWts = 1000,
     abstol = 1.0e-4, reltol = 1.0e-8, ...)
```

```
> ANN options (datatype: binary , package: nnet , function: nnet ) :
( dataset_allData_allRun )
  - size = 5 (default: 2 )
  - decay = 5 (default: NULL )
  - trace = FALSE (default: NULL )
  - rang = 0.1 (default: NULL )
  - maxit = 200 (default: NULL )
```

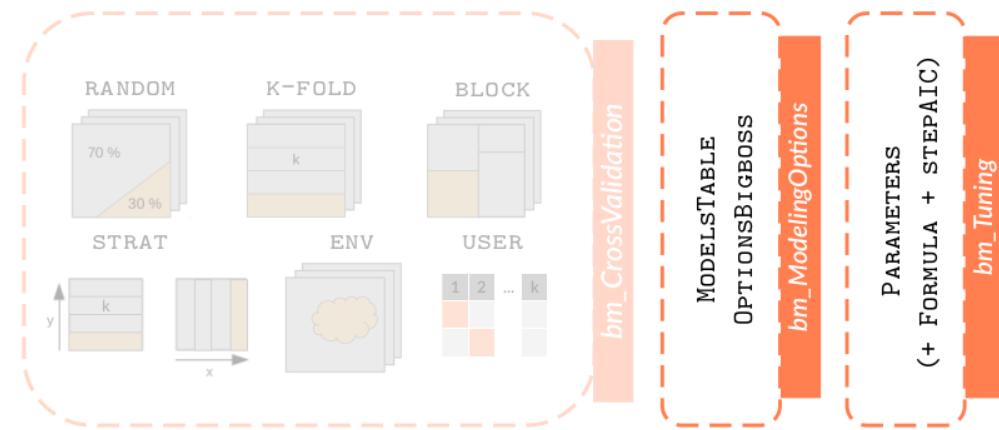
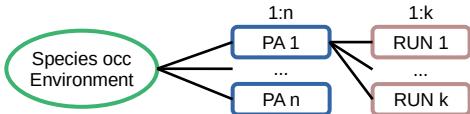
```
user.ANN = list(' allData_allRun' =
  list(size = 5,
       decay = 0.5,
       trace = FALSE,
       rang = 0.1,
       maxit = 500))
```



	model	type	package	func	train
1	ANN	binary	nnet	rpart	rpart
2	CTA	binary	rpart	rpart	rpart
3	DNN	binary	cito	dnn	tune
4	FDA	binary	mda	fda	fda
5	GAM	binary	gam	gam	gamLoess
6	GAM	binary	mgcv	bam	bam
7	GAM	binary	mgcv	gam	gam
8	GBM	binary	gbm	gbm	gbm
9	GLM	binary	stats	glm	glm
10	MARS	binary	earth	earth	earth
11	MAXENT	binary	MAXENT	MAXENT	ENMEvaluate
12	MAXNET	binary	maxnet	maxnet	maxnet
13	RF	binary	randomForest	randomForest	rf
14	Rfd	binary	randomForest	randomForest	rf
15	SRE	binary	biomod2	bm_SRE	bm_SRE
16	XGBOOST	binary	xgboost	xgboost	xgbTree
17	CTA	nonbinary	rpart	rpart	rpart
18	DNN	nonbinary	cito	dnn	tune
19	FDA	nonbinary	mda	fda	fda
20	GAM	nonbinary	gam	gam	gamLoess
21	GAM	nonbinary	mgcv	bam	bam
22	GAM	nonbinary	mgcv	gam	gam
23	GBM	nonbinary	gbm	gbm	gbm
24	GLM	nonbinary	stats	glm	glm
25	MARS	nonbinary	earth	earth	earth
26	RF	nonbinary	randomForest	randomForest	rf
27	XGBOOST	nonbinary	xgboost	xgboost	xgbTree

1. Formating data

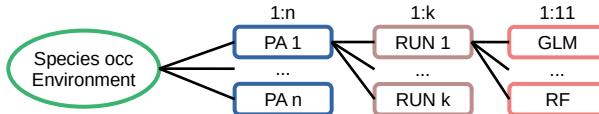
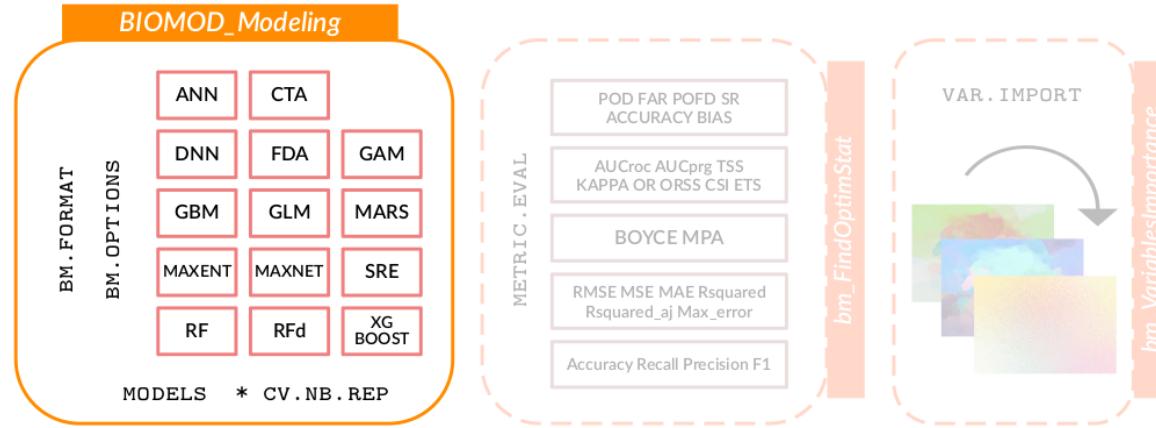
- » 12 types of models,
16 single models,
11 that can use non-binary data
- » 1 coded in biomod2,
1 external software,
13 other R packages
- » **default** : extracted from functions
- » **bigboss** : redefined by biomod2 team
- » **user-defined**
- » **tuned** : with *train* function from *caret* package



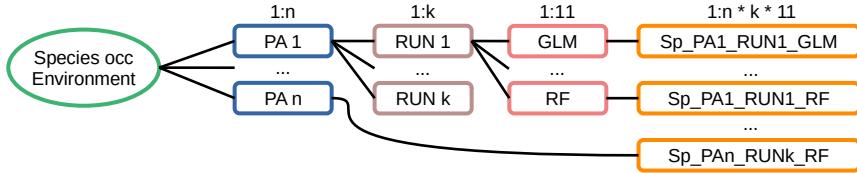
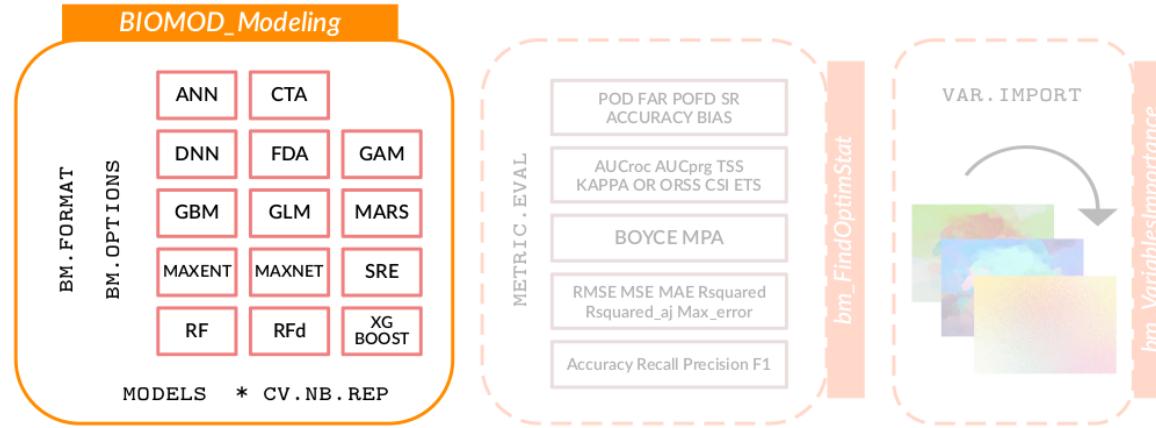
- » test a bunch of parameters, and try to keep the « best » according to some evaluation metrics (TSS or ROC)

	model	type	package	func	package	func
1	ANN	binary	nnet	rpart	rpart	avNNet
2	CTA	binary	rpart	dnn	tune	rpart
3	DNN	binary	cito	fda	fda	tune
4	FDA	binary	mda	gam	gam	fda
5	GAM	binary	gam	mgcv	bam	gamLoess
6	GAM	binary	mgcv	gbm	gam	bam
7	GAM	binary	gbm	gbm	gbm	gbm
8	GBM	binary	stats	earth	glm	glm
9	GLM	binary	earth	earth	earth	earth
10	MARS	binary	MAXENT	MAXENT	ENMevaluate	earth
11	MAXENT	binary	maxnet	maxnet	maxnet	maxnet
12	MAXNET	binary	randomForest	randomForest	rf	rf
13	RF	binary	randomForest	randomForest	randomForest	rf
14	Rfd	binary	randomForest	randomForest	randomForest	rf
15	SRE	binary	biomod2	bm_SRE	bm_SRE	xgbTree
16	XGBOOST	binary	xgboost	xgboost	xgboost	rpart
17	CTA	nonbinary	rpart	dnn	tune	tune
18	DNN	nonbinary	cito	fda	fda	fda
19	FDA	nonbinary	mda	gam	gam	gamLoess
20	GAM	nonbinary	gam	mgcv	bam	bam
21	GAM	nonbinary	mgcv	gbm	gbm	gbm
22	GAM	nonbinary	gbm	gbm	gbm	gbm
23	GBM	nonbinary	stats	glm	glm	glm
24	GLM	nonbinary	glm	earth	earth	earth
25	MARS	nonbinary	earth	earth	earth	rf
26	RF	nonbinary	randomForest	randomForest	randomForest	xgbTree
27	XGBOOST	nonbinary	xgboost	xgboost	xgboost	xgbTree

2.a Single models

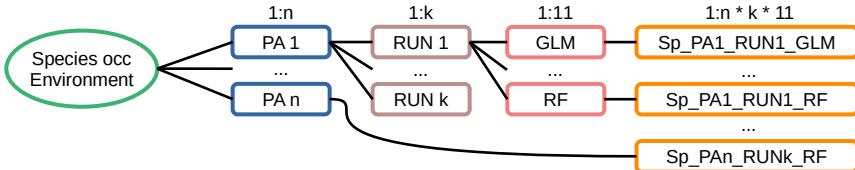
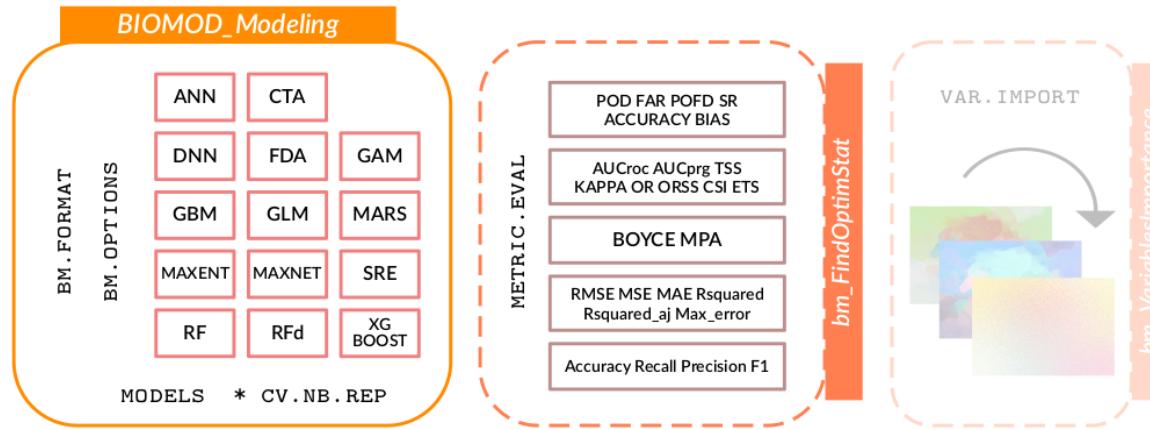


2.a Single models



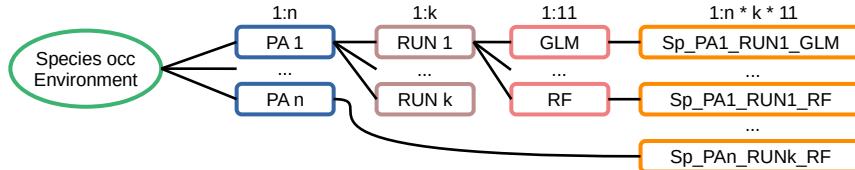
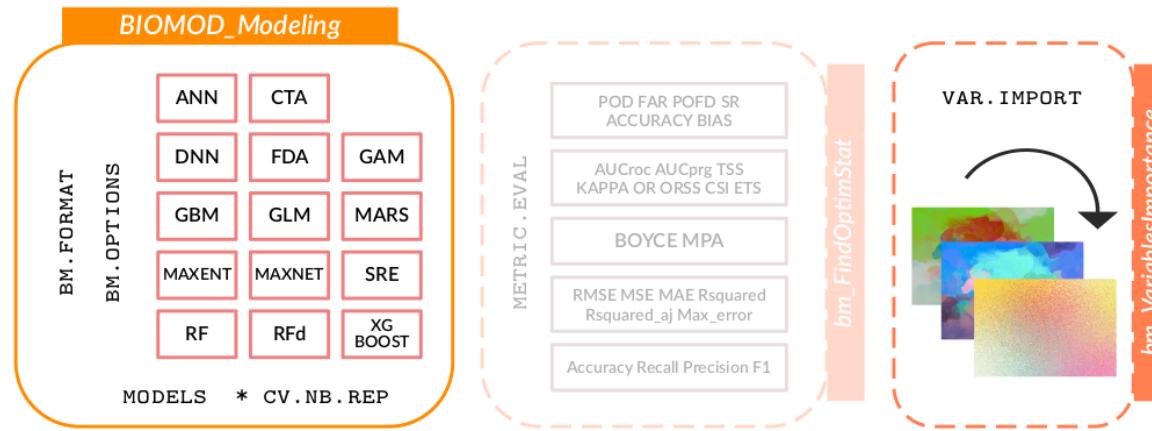
2.a Single models

- » except ROC, all evaluation metrics obtained from contingency table (containing *TP*, *FP*, *TN*, *FN*)
- » require a **binary transformation** :
 - range of thresholds tested
 - keep threshold optimising the evaluation metric

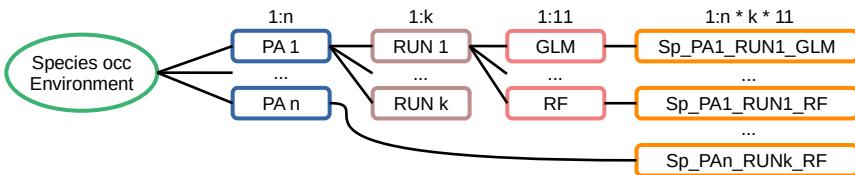
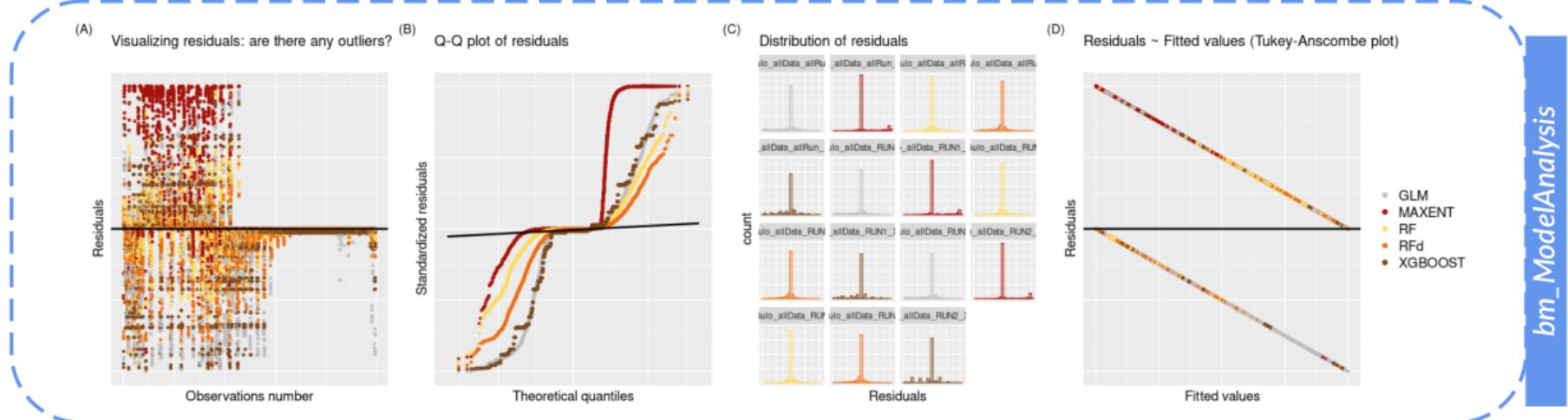


2.a Single models

- » comparison of importance of variables between models
- » **Pearson correlation** between :
 - normal prediction
 - prediction with 1 variable randomised



3.a Exploring single models

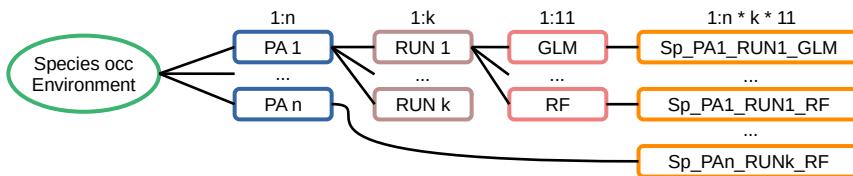
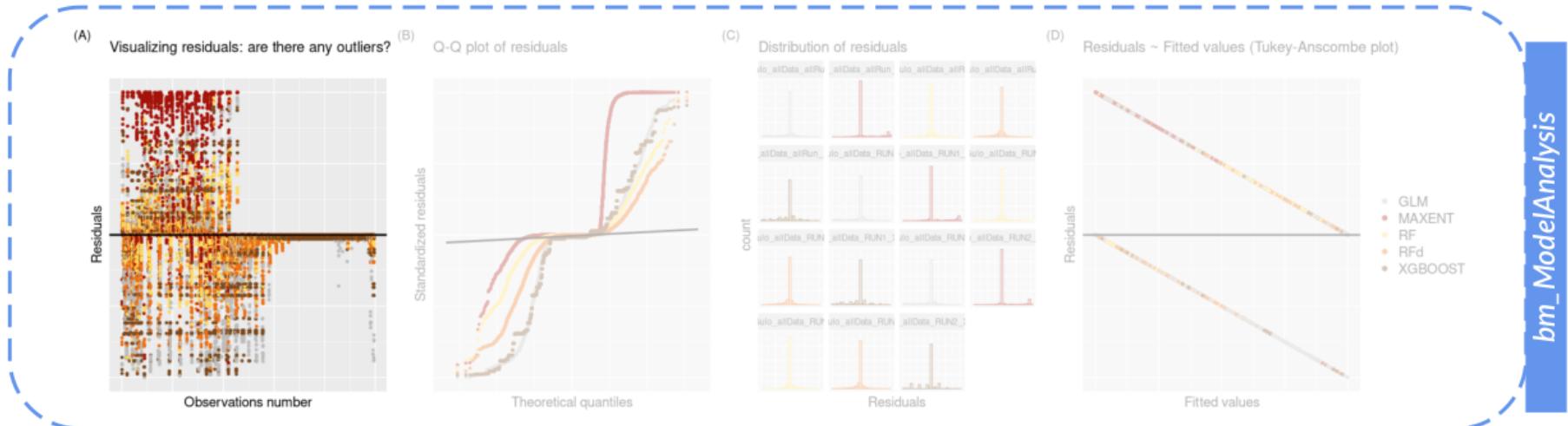


3.a Exploring single models

» $\text{residuals} \sim f(\text{observations number})$

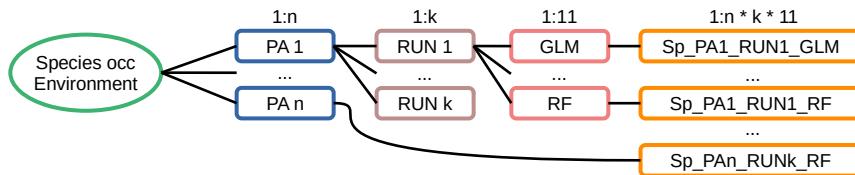
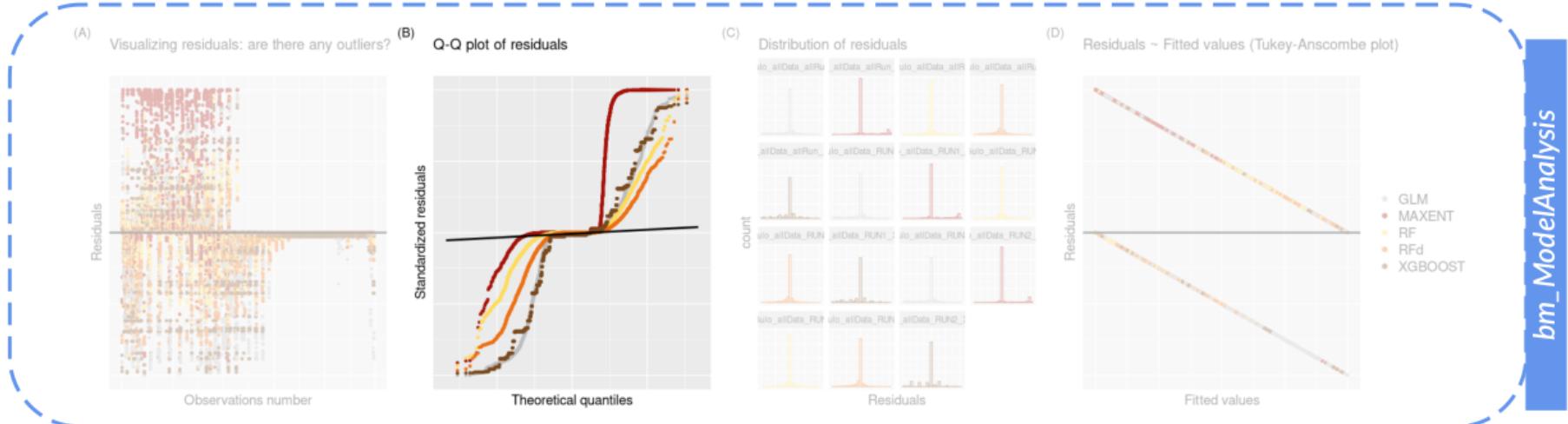
» visualize the potential outliers

- x-axis only helps to find the outlier number



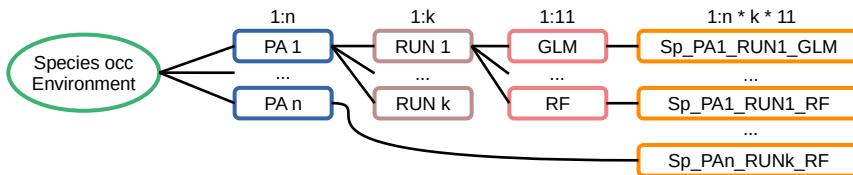
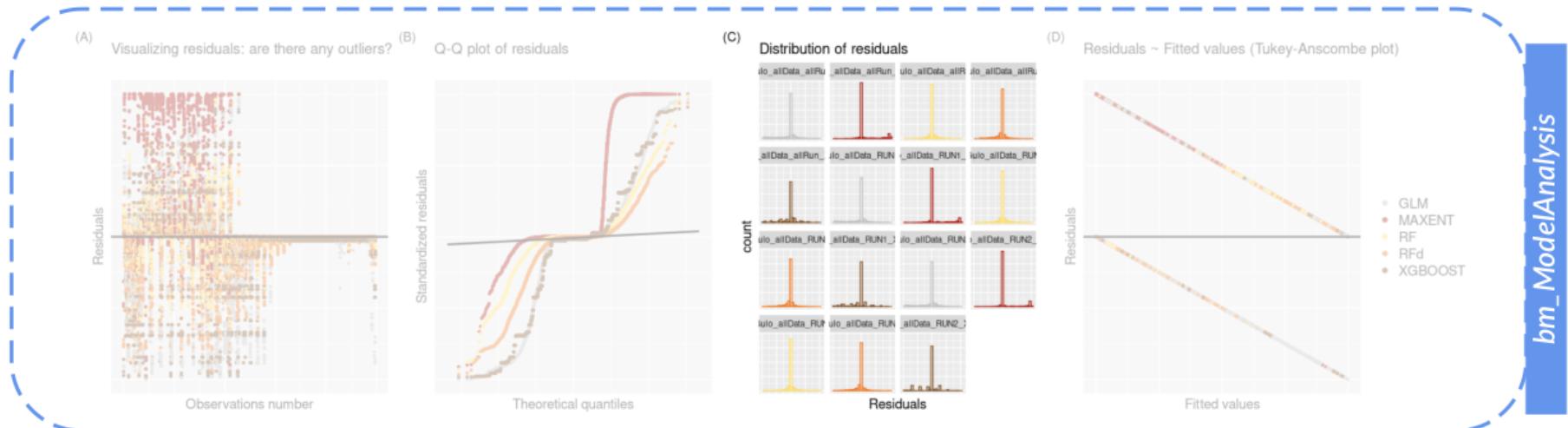
3.a Exploring single models

- » standardized residuals $\sim f(\text{theoretical quantiles})$
- » Q-Q plot to assess if residuals follow a normal distribution



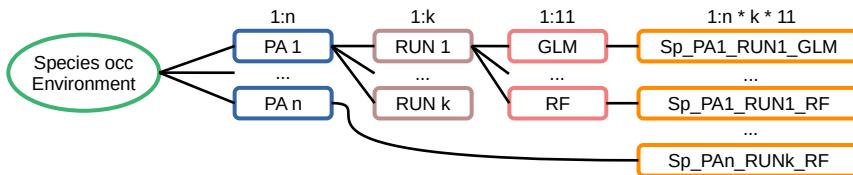
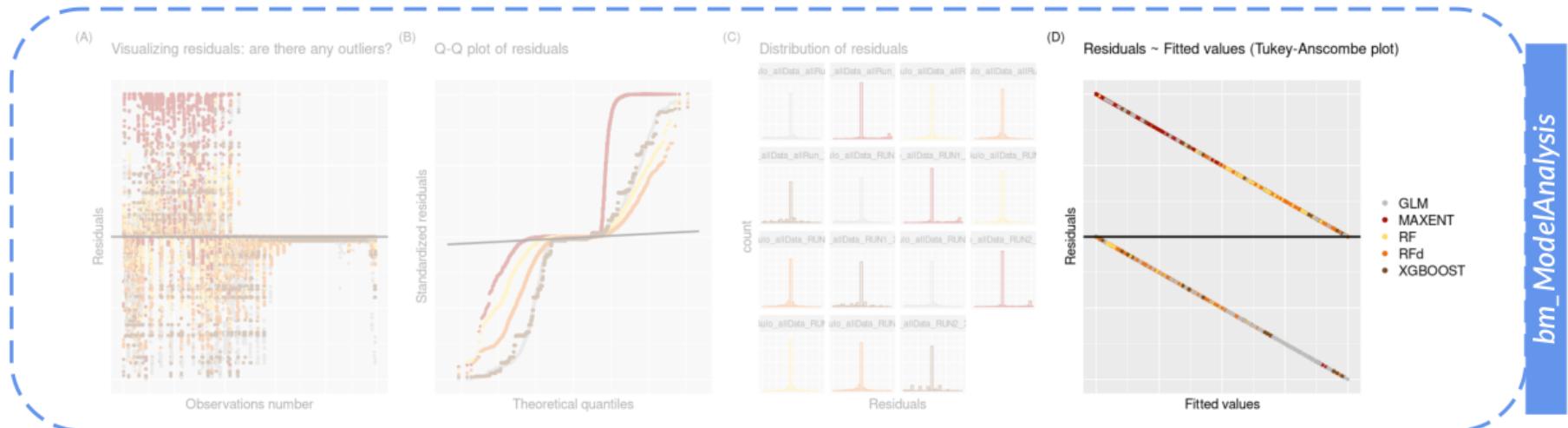
3.a Exploring single models

- » residuals distribution
- » histogram to assess if residuals follow a normal distribution

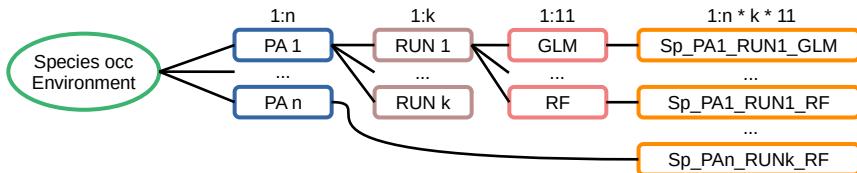
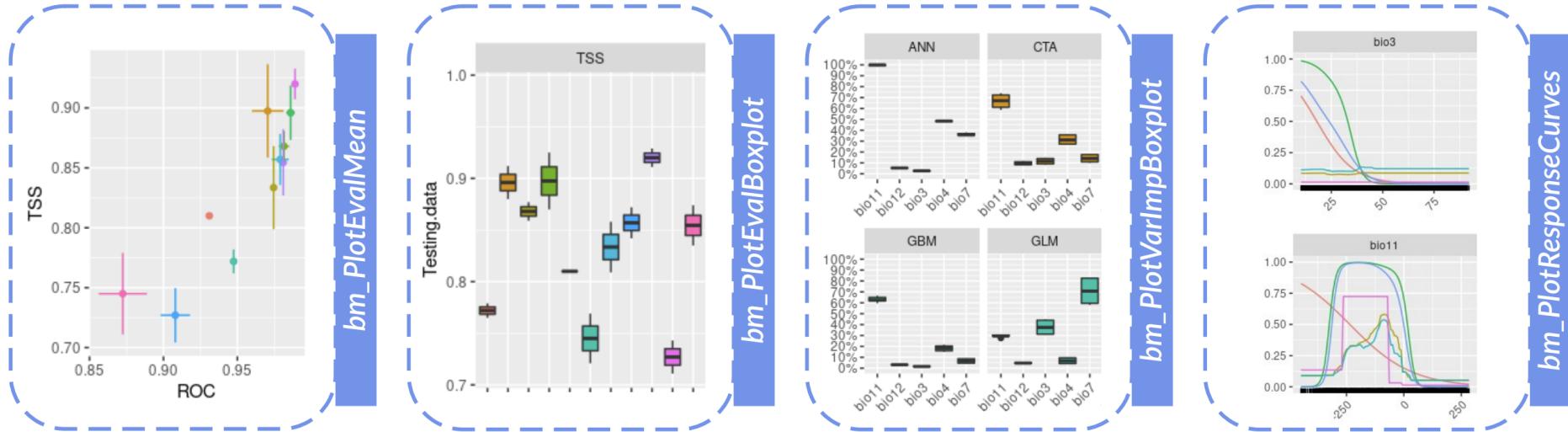


3.a Exploring single models

- » $\text{residuals} \sim f(\text{fitted values})$
- » Tukey-Anscombe plot to detect heteroscedasticity
 - meaning residuals do not have a constant variance

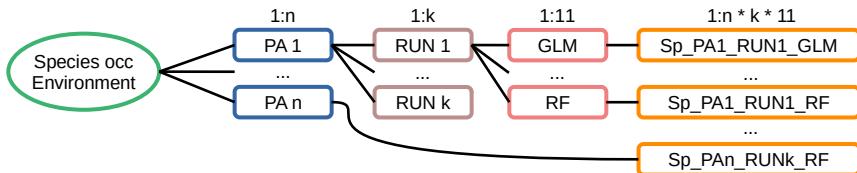
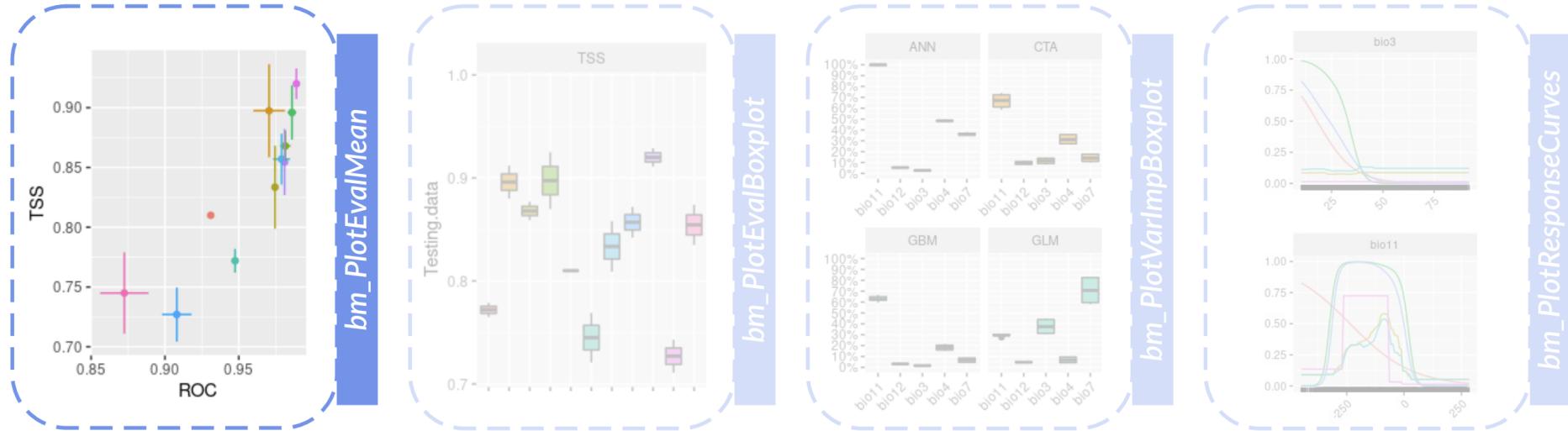


3.a Exploring single models



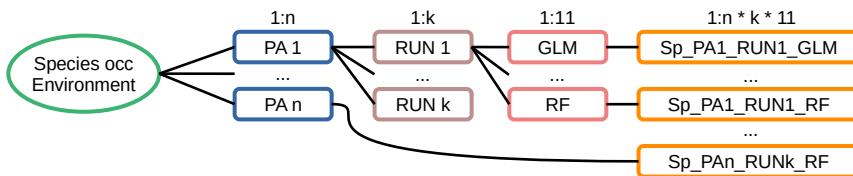
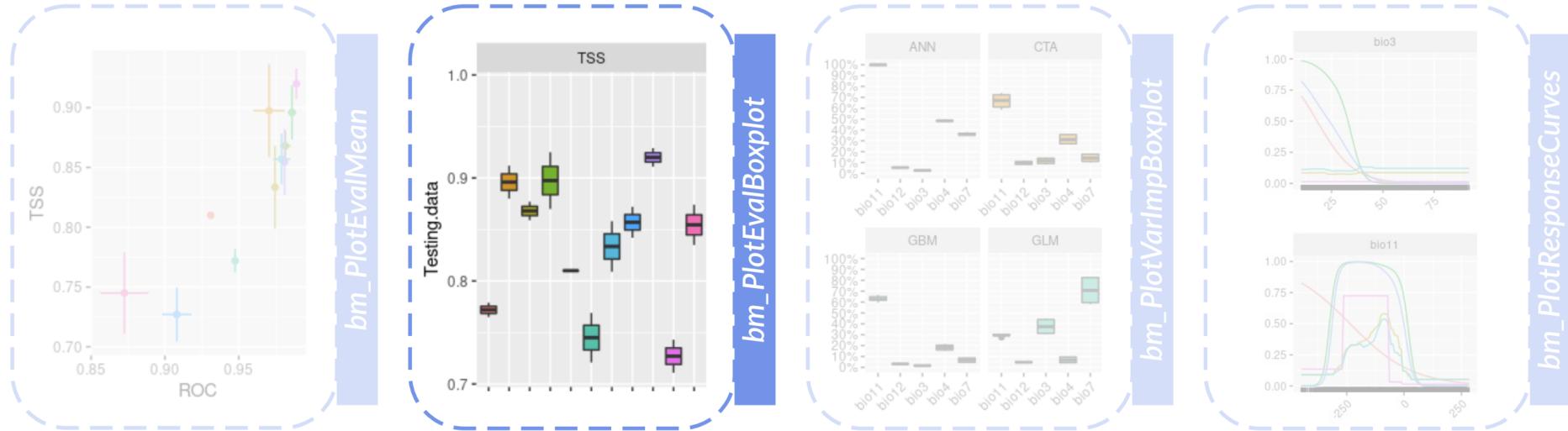
3.a Exploring single models

- » « evaluation space »
- » visualize the metrics consistency between models



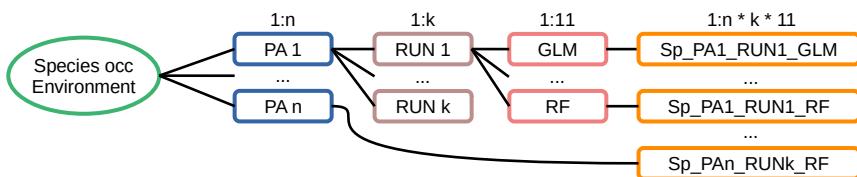
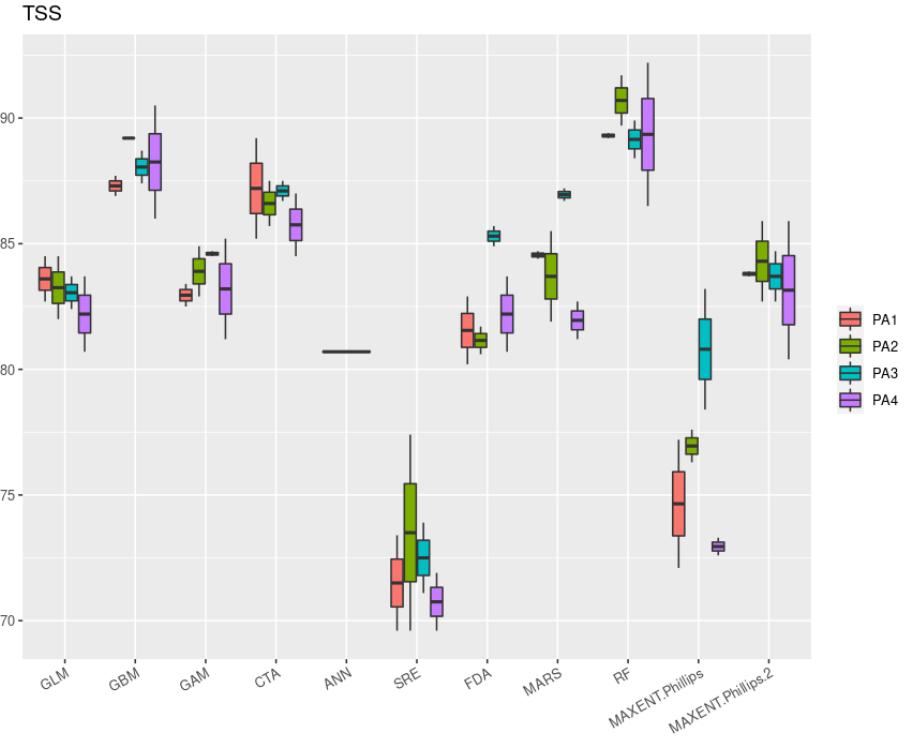
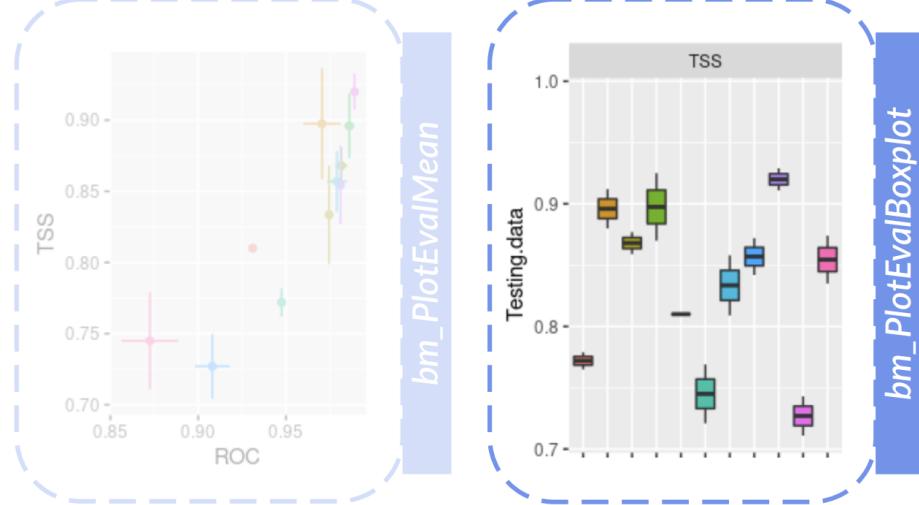
3.a Exploring single models

- » more classical view
- » visualize the metrics consistency between models



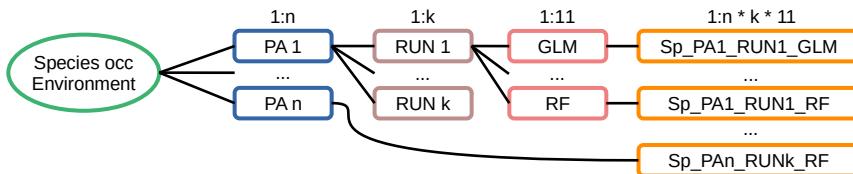
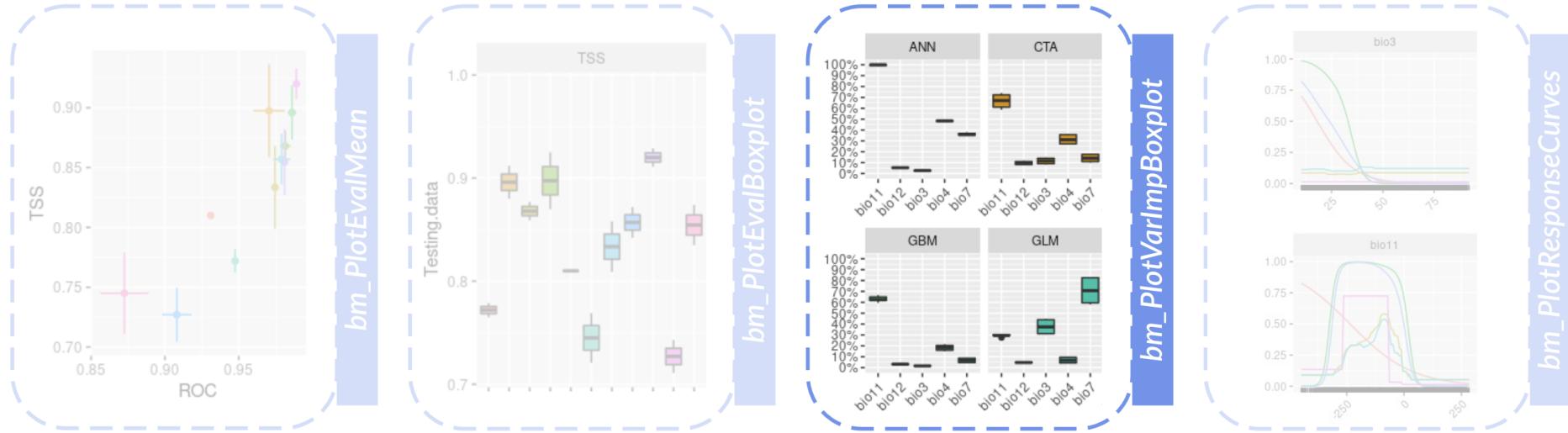
3.a Exploring single models

- » more classical view
- » visualize the metrics consistency between models
 - explore the different levels of subsets



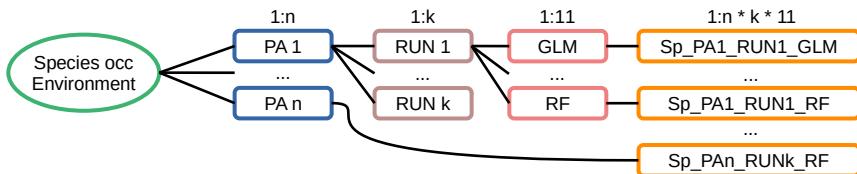
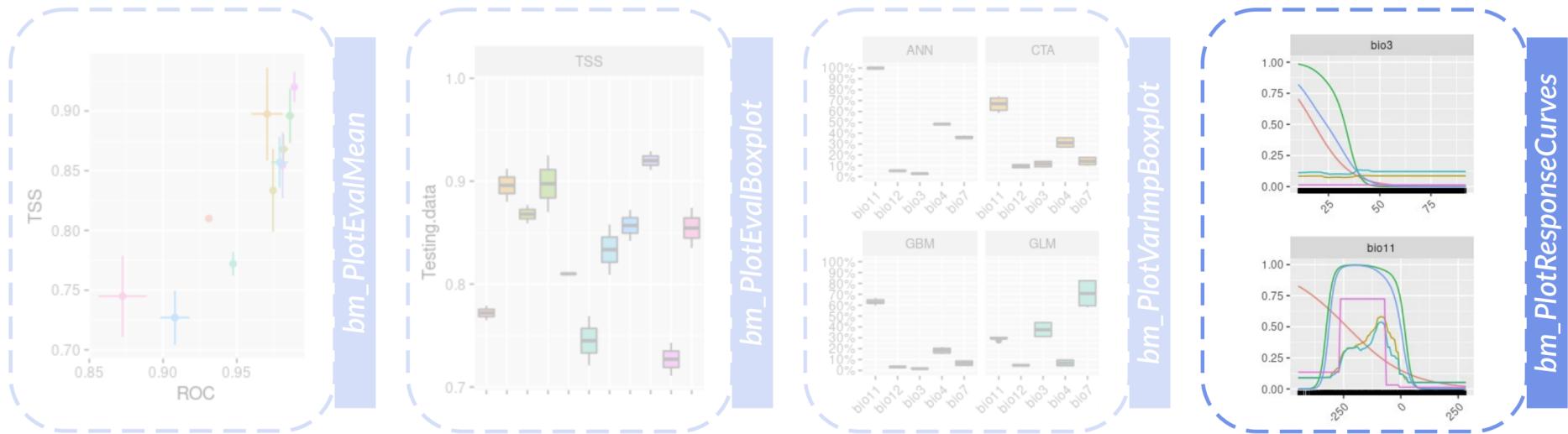
3.a Exploring single models

- » compare importance of variables between models
- » visualize the consistency between models
(and different types of models)

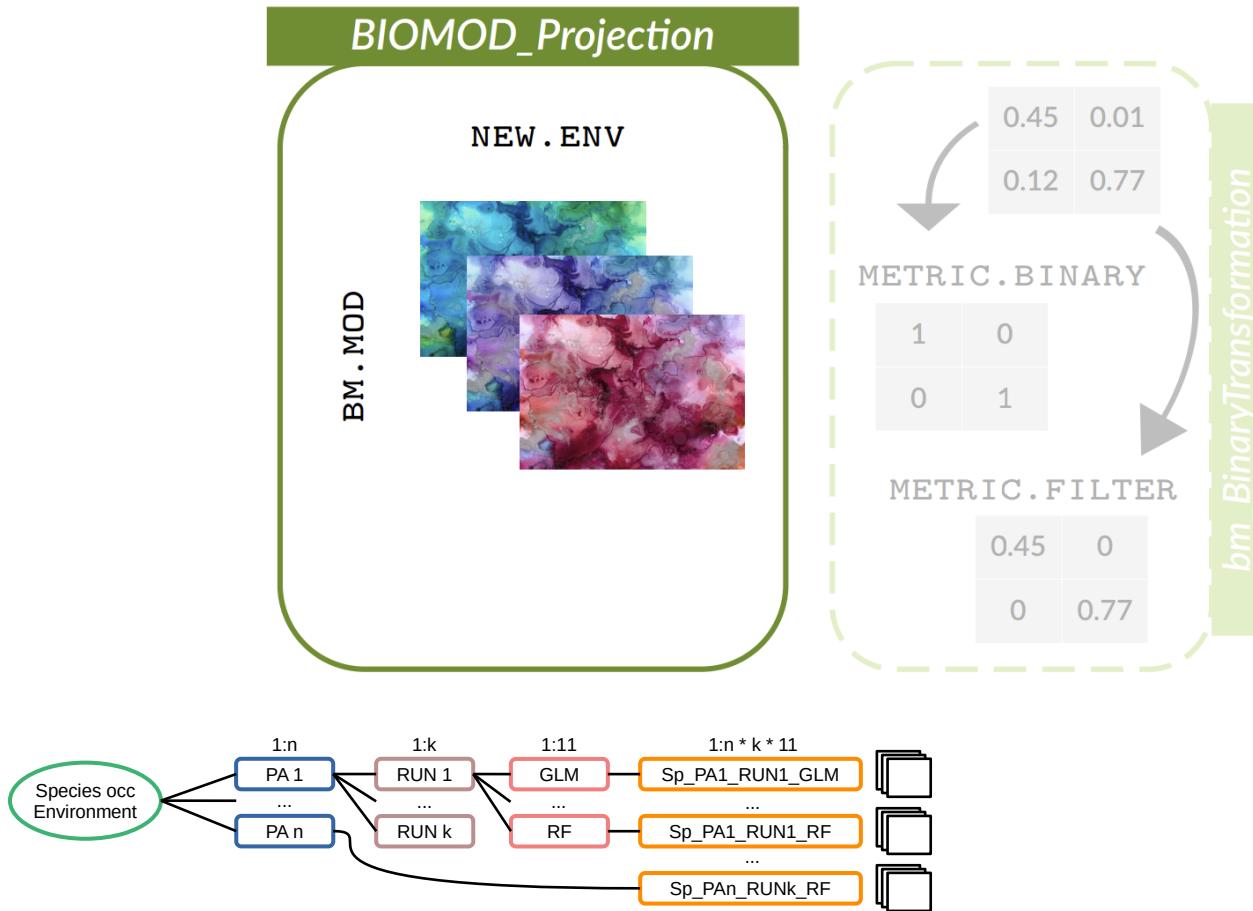


3.a Exploring single models

- » better understand the effect of each variable along its gradient onto the probability of presence
- » visualize the consistency between models (*and different types of models*)

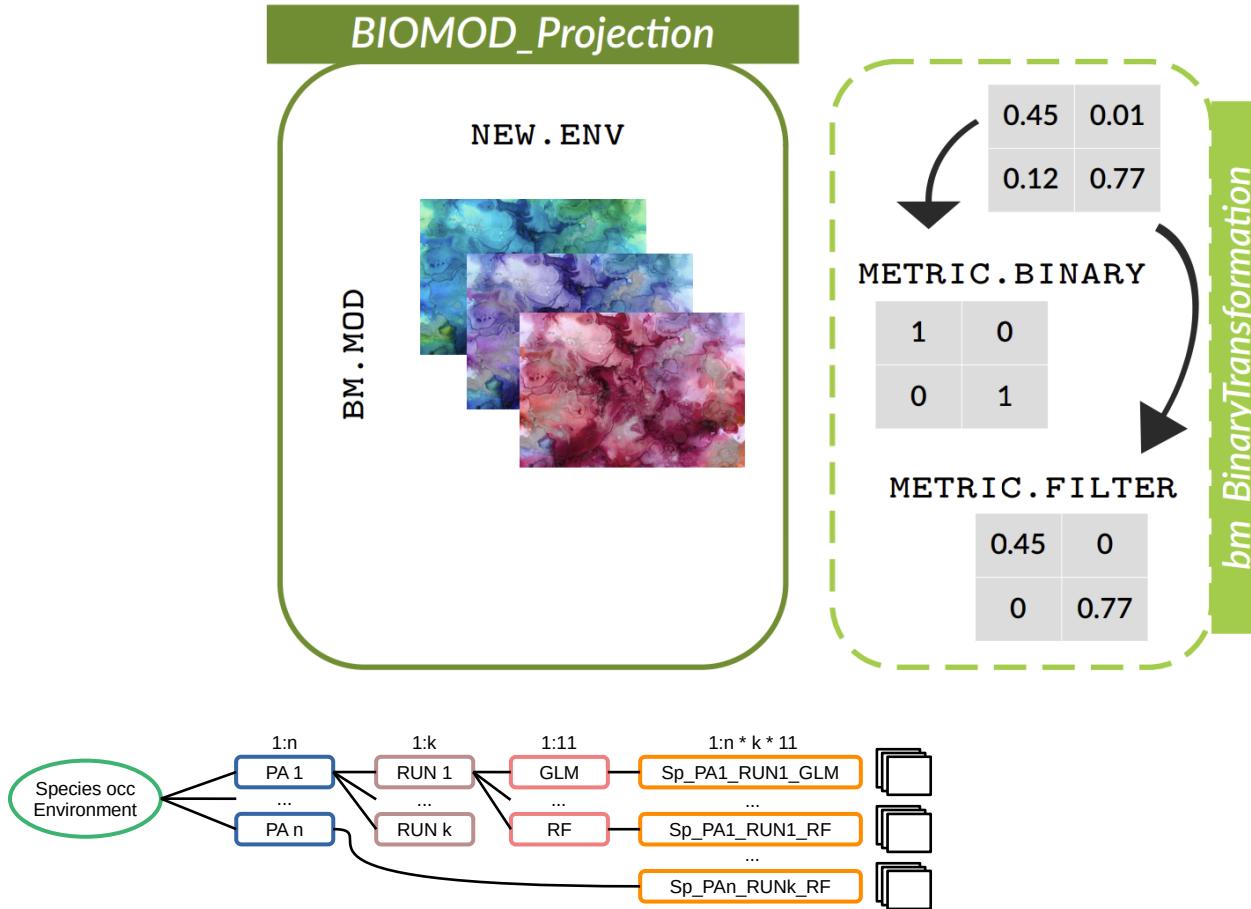


4.a Projecting single models

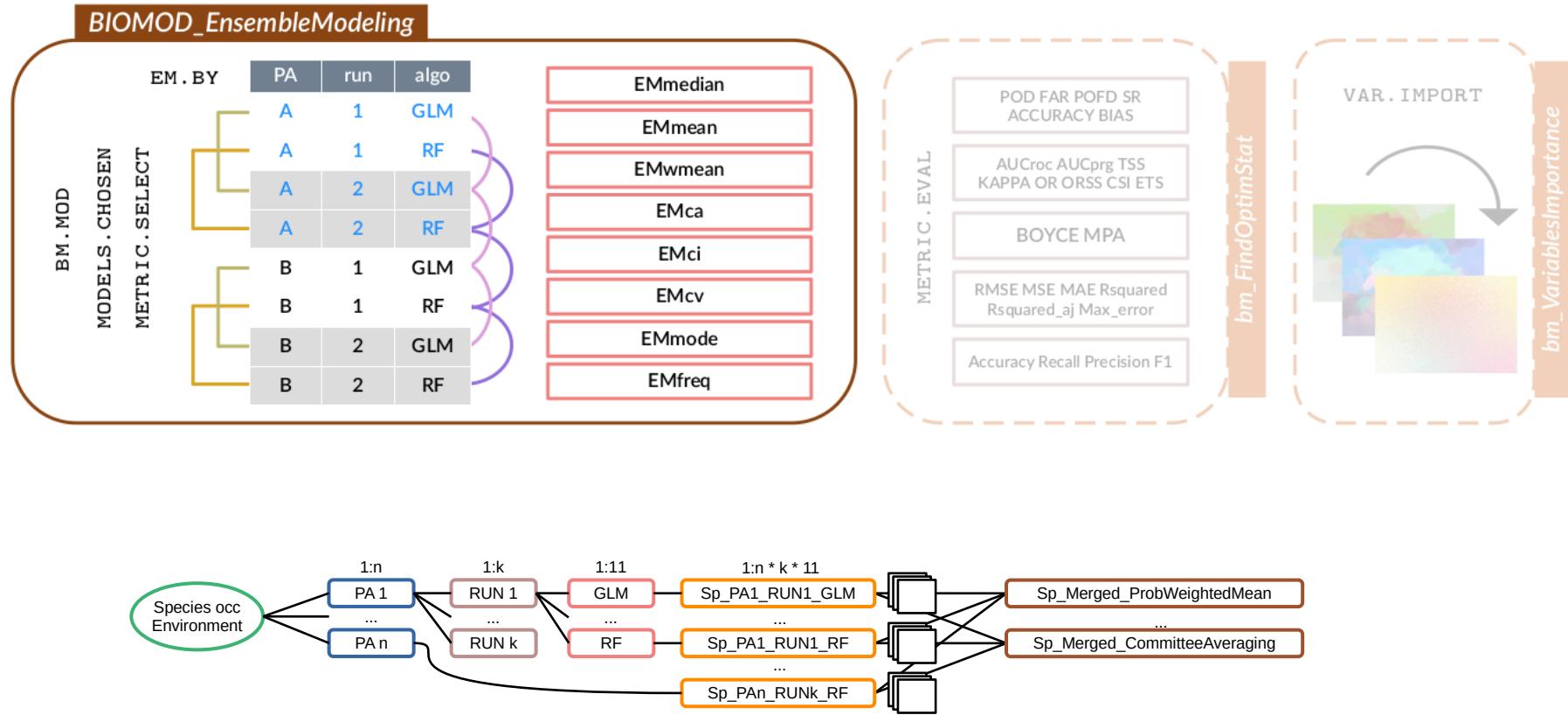


4.a Projecting single models

- » transformation associated to one evaluation metric (one map created for each metric selected)
- » use the threshold maximising the chosen metric



2.b Ensemble models

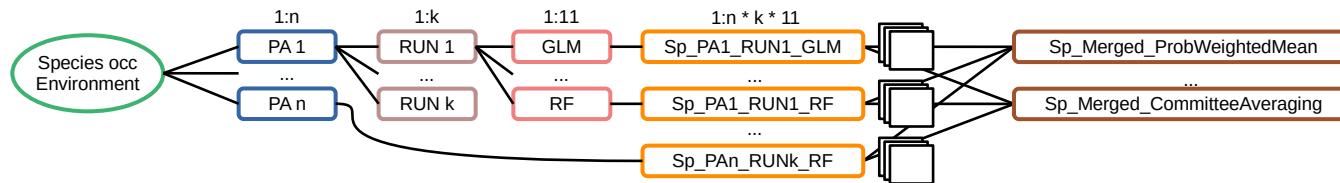
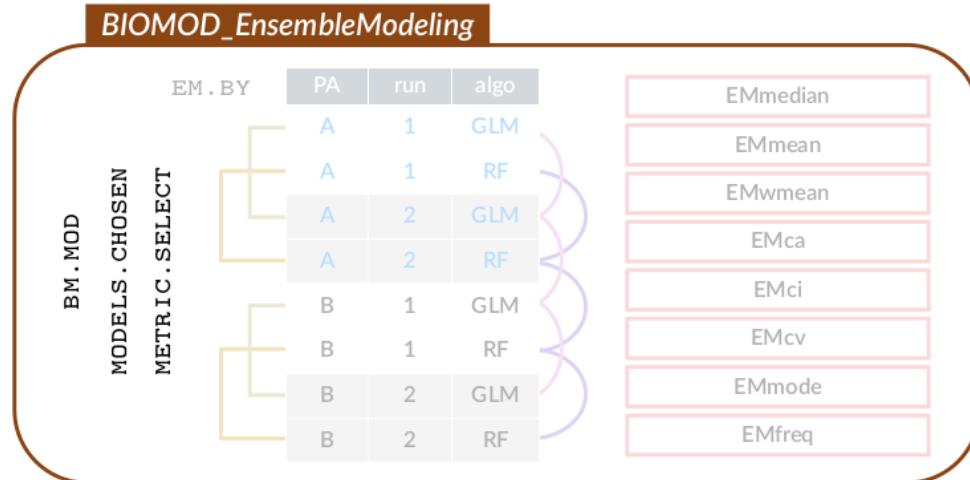


2.b Ensemble models

Step 1 : filter single models

- » **filtering** associated to one evaluation metric
(one set of ensemble models created for each metric selected)

- » use a threshold to keep single models



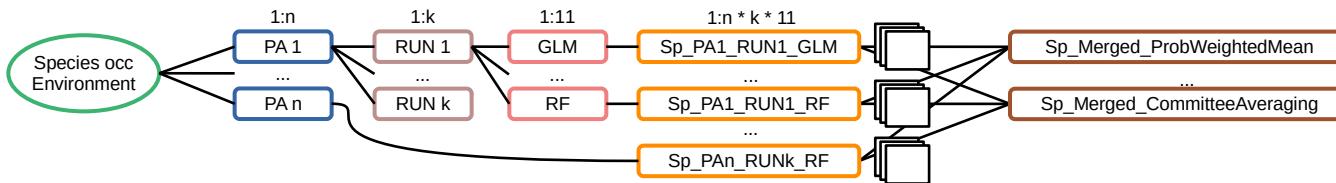
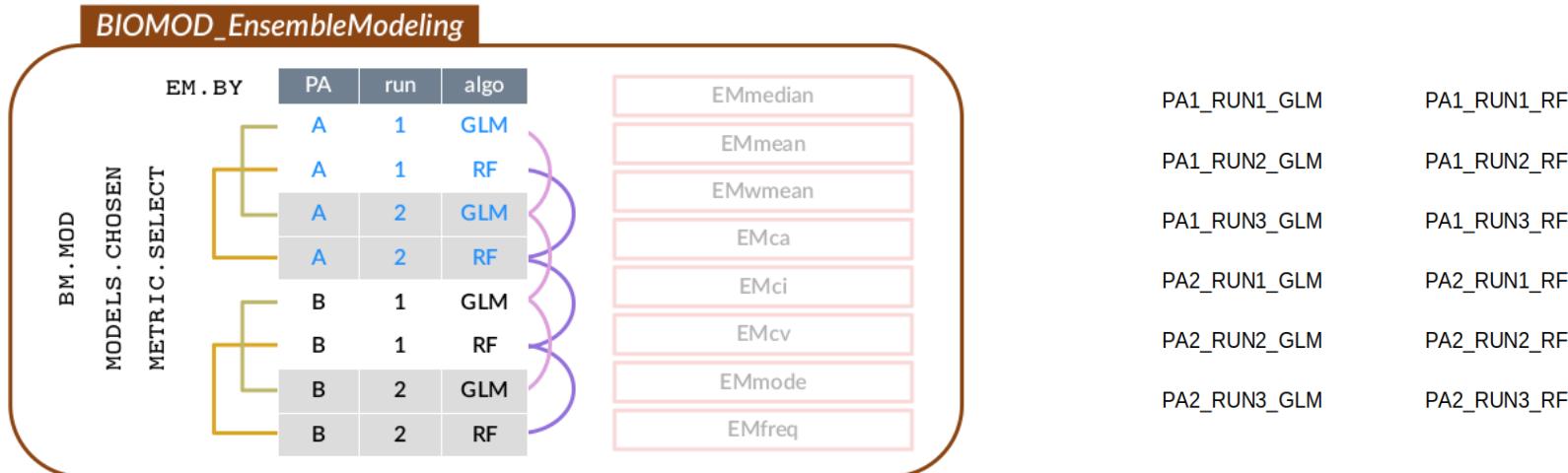
2.b Ensemble models

Step 1 : filter single models

Step 2 : gather single models

» different ways of **combining** single models together :

- *all*
- *algo*
- *PA*
- *PA+algo*
- *PA+run*



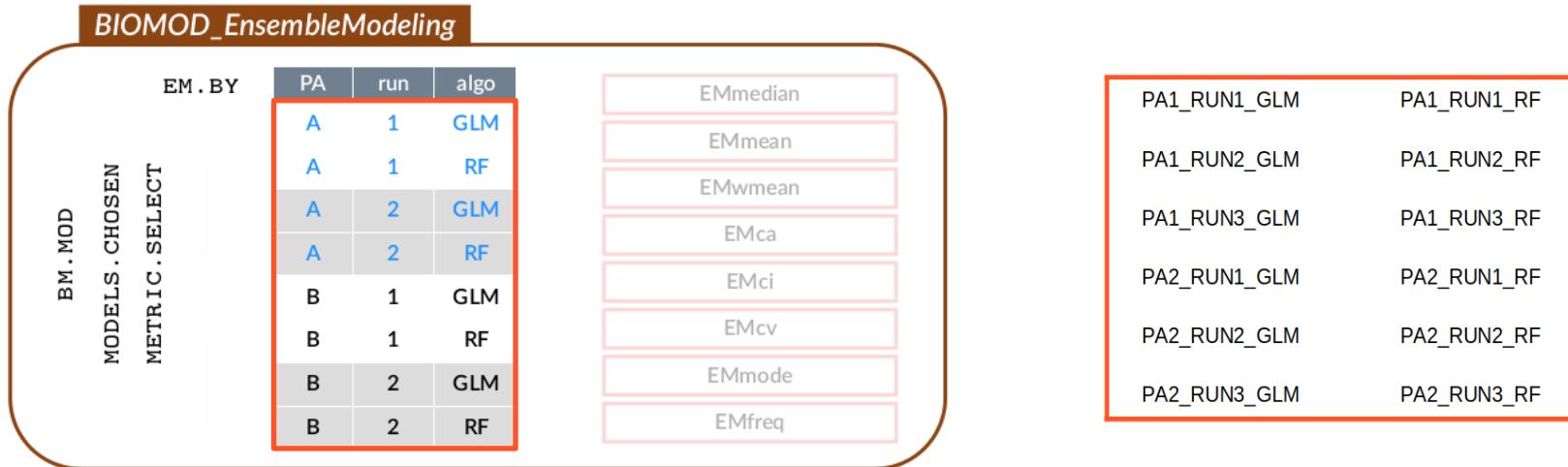
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Step 1 : filter single models

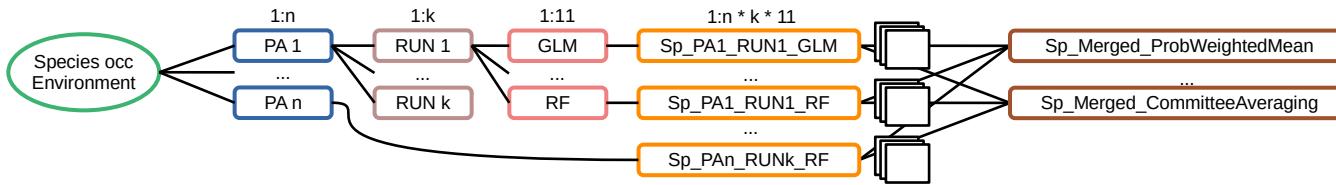
Step 2 : gather single models

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- ◆ *all*
 - ◆ *algo*
 - ◆ *PA*
 - ◆ *PA+algo*
 - ◆ *PA+run*



All models

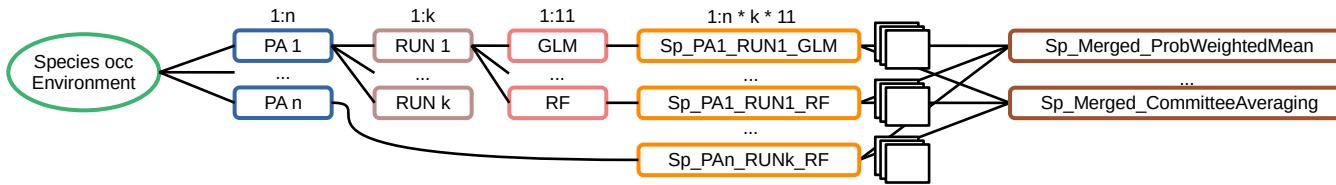
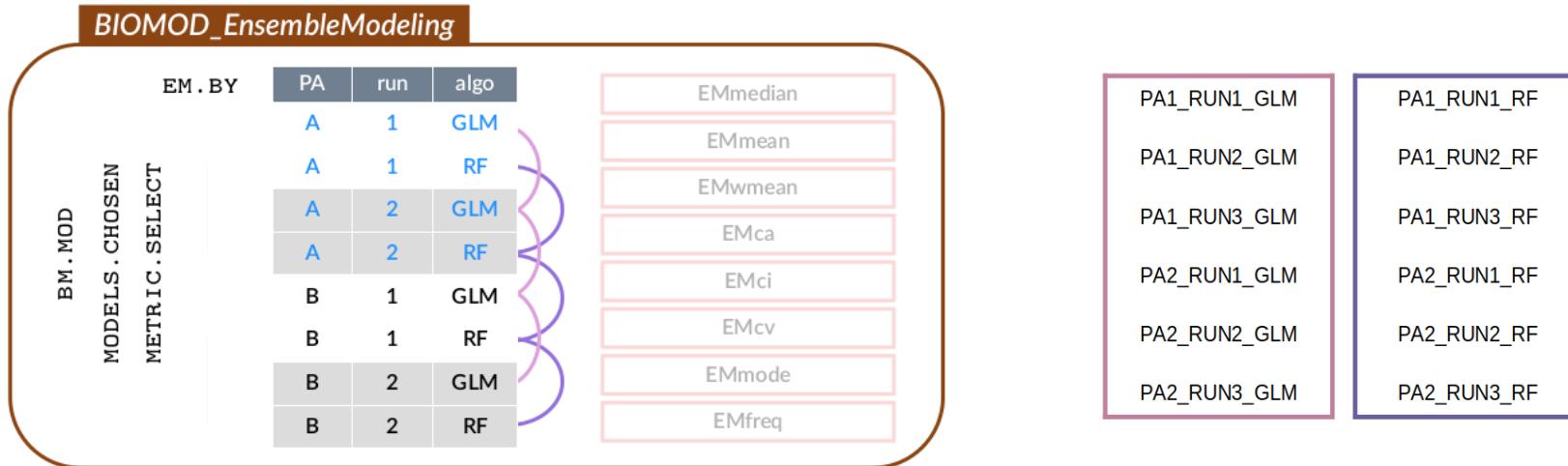


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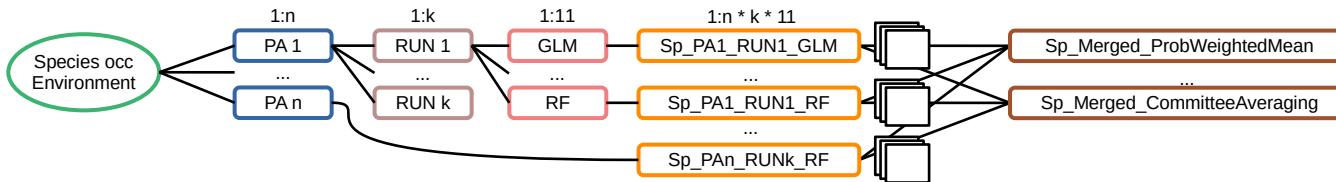
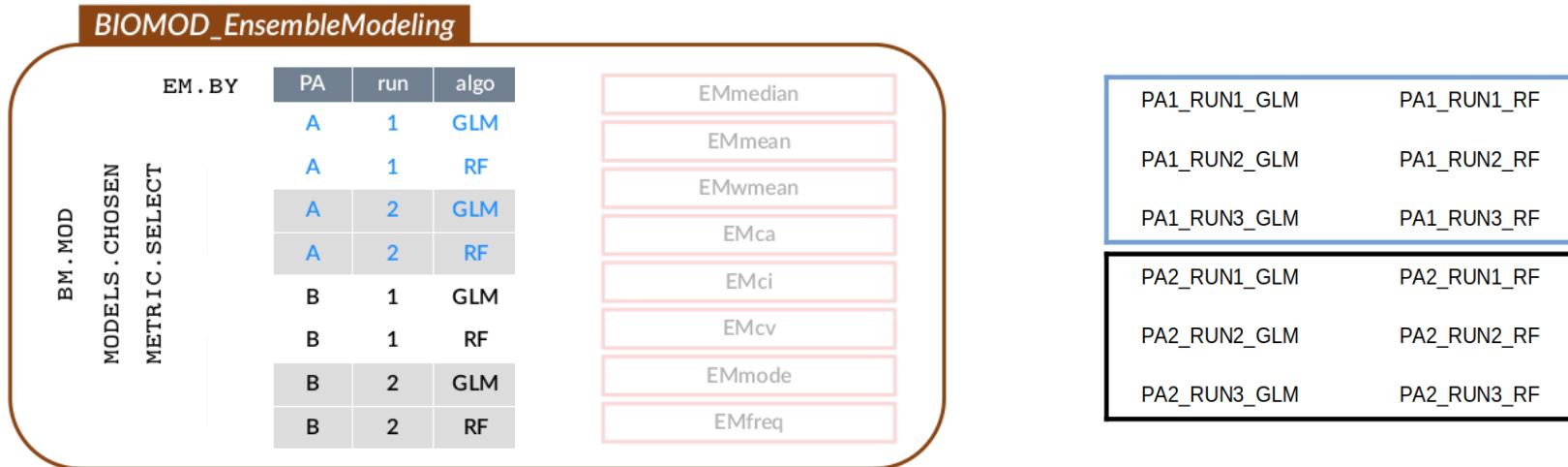
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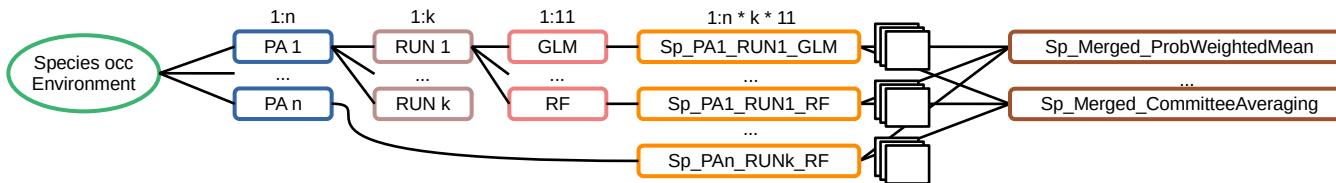
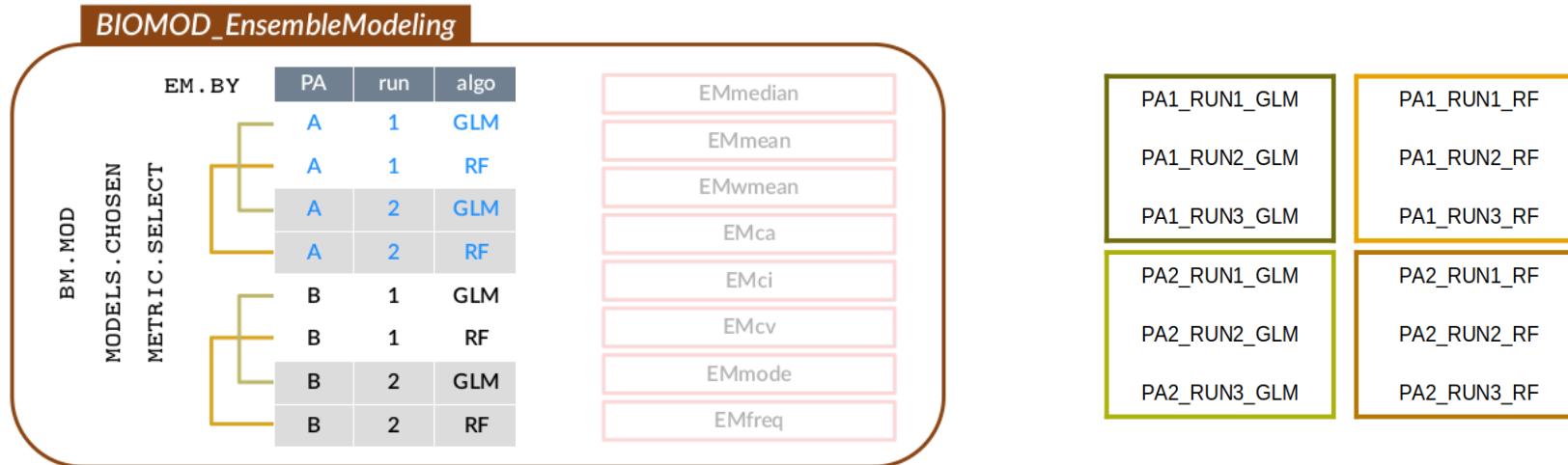


2.b Ensemble models

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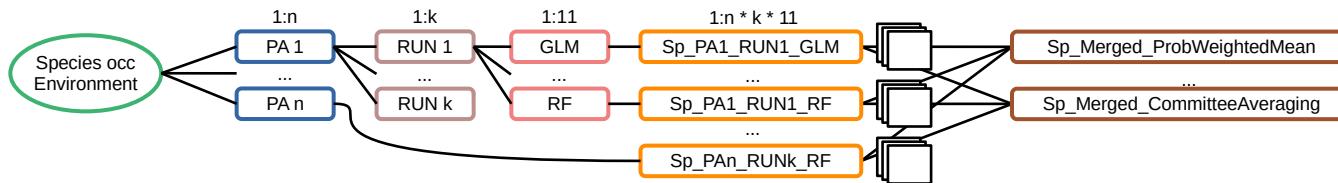
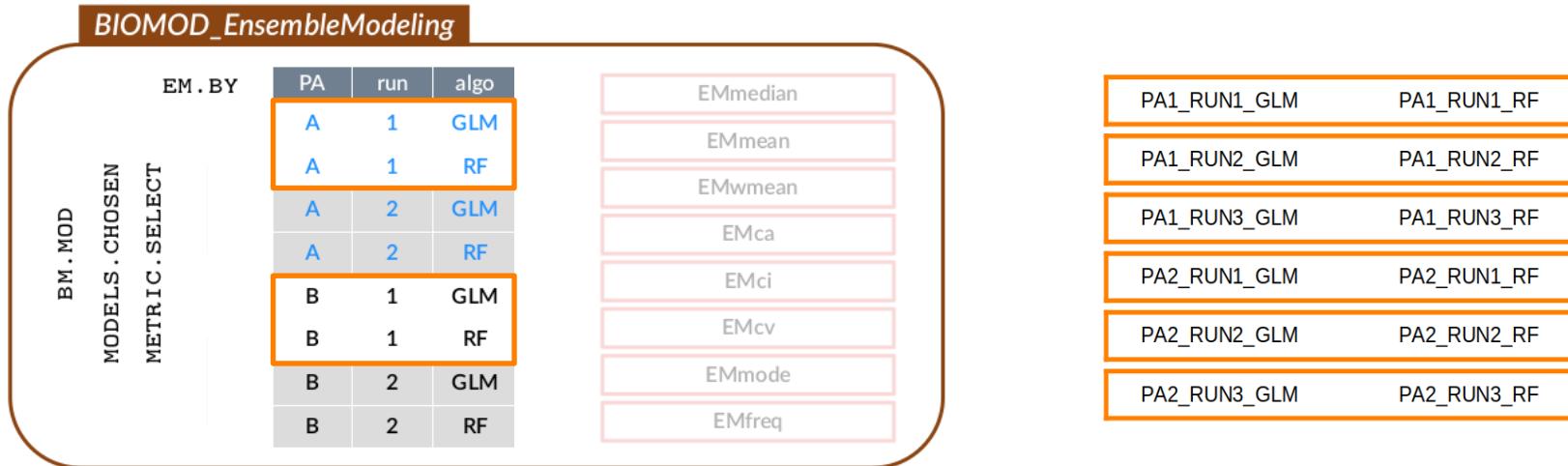
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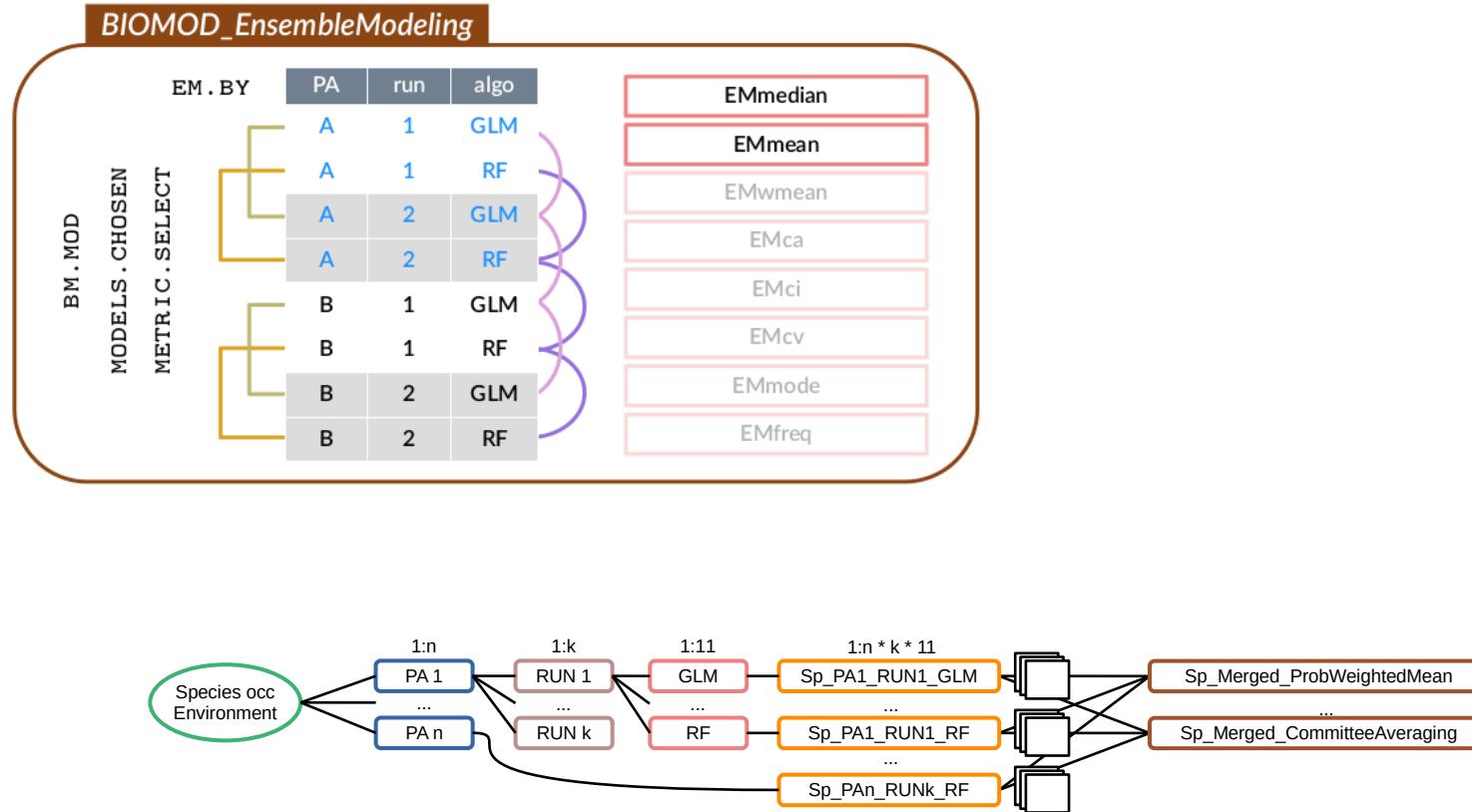
2.b Ensemble models

- » « simple » ensemble models : **mean** or **median**

Step 1 : filter single models

Step 2 : gather single models

Step 3 : build ensemble models



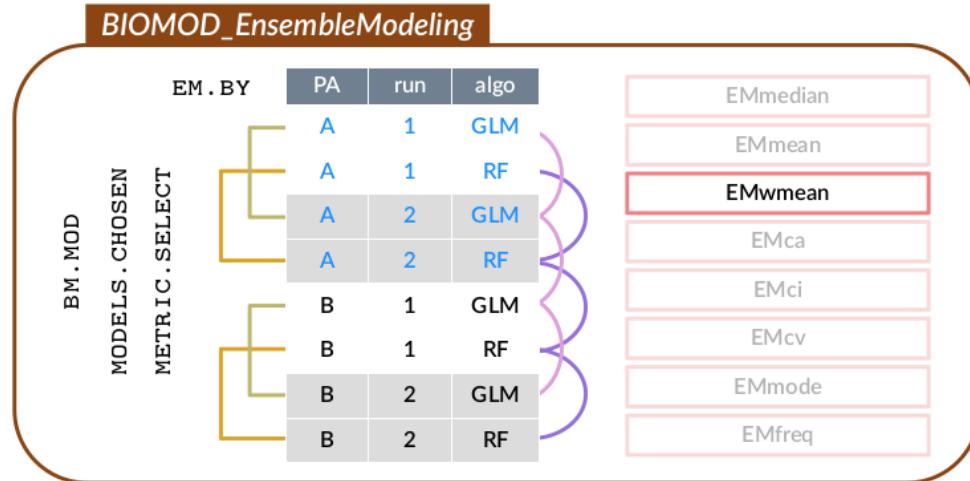
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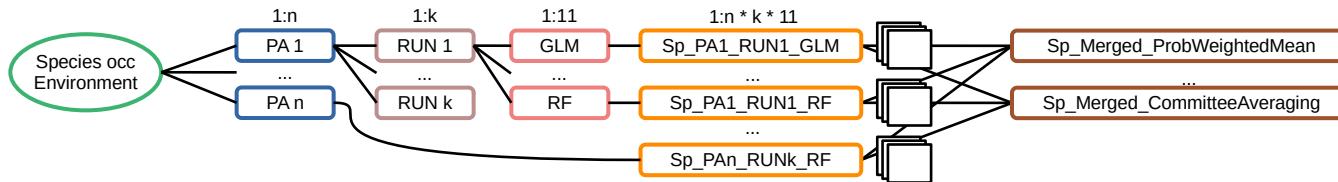
Step 2 : gather single models

Step 3 : build ensemble models

- » « simple » ensemble models : mean or median
- » « complex » ensemble models :
 - probability weighted mean



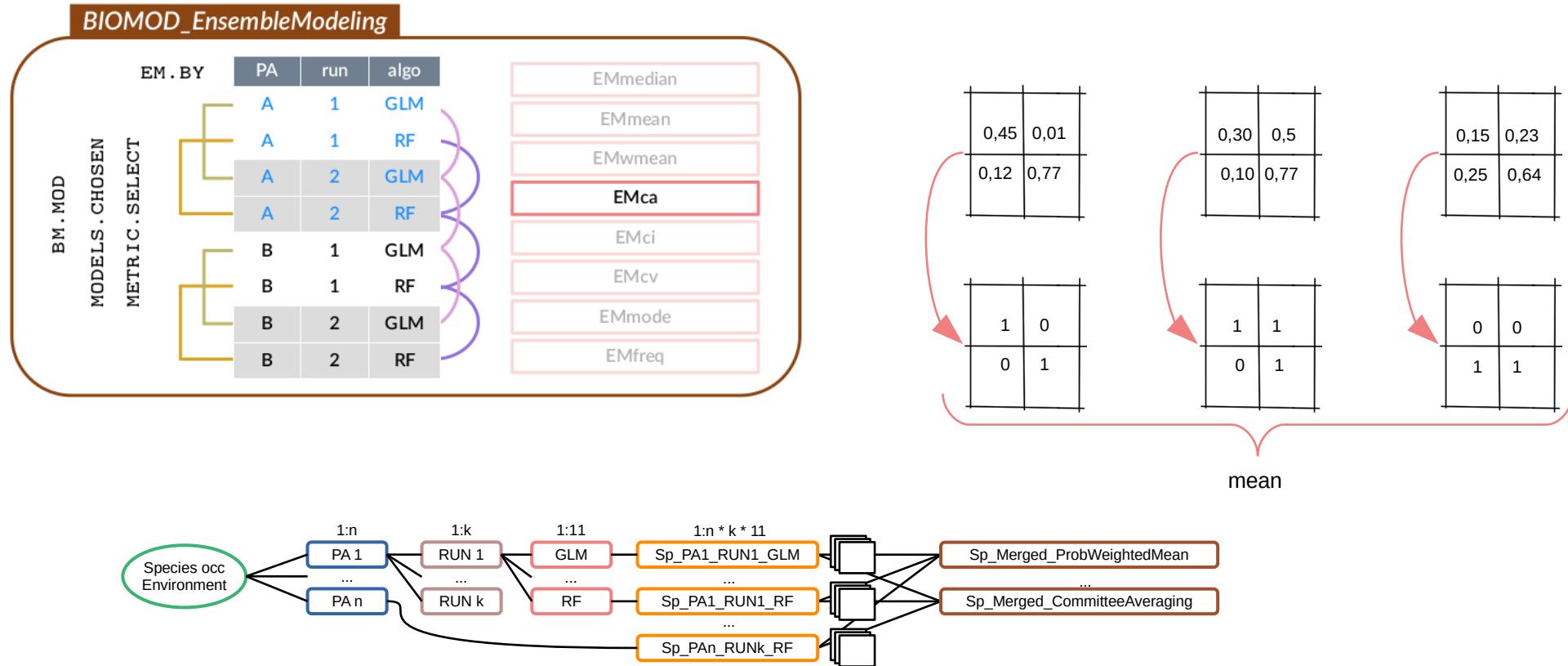
$$W1 * \begin{pmatrix} 0,45 & 0,01 \\ 0,12 & 0,77 \end{pmatrix} + W2 * \begin{pmatrix} 0,30 & 0,5 \\ 0,10 & 0,77 \end{pmatrix} + W3 * \begin{pmatrix} 0,15 & 0,23 \\ 0,25 & 0,64 \end{pmatrix}$$



2.b Ensemble models

- Step 1 : filter single models
- Step 2 : gather single models
- Step 3 : build ensemble models

- » « simple » ensemble models : mean or median
- » « complex » ensemble models :
 - probability weighted mean
 - **committe averaging**



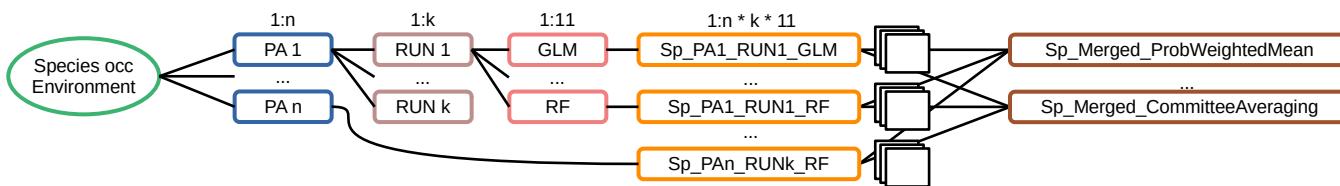
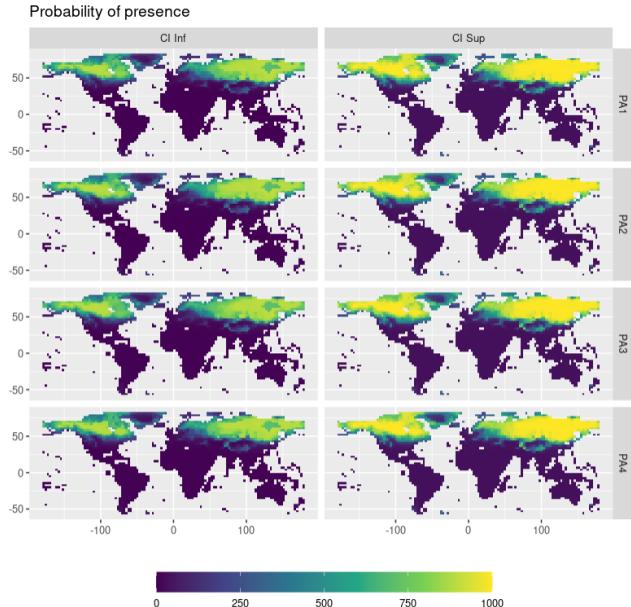
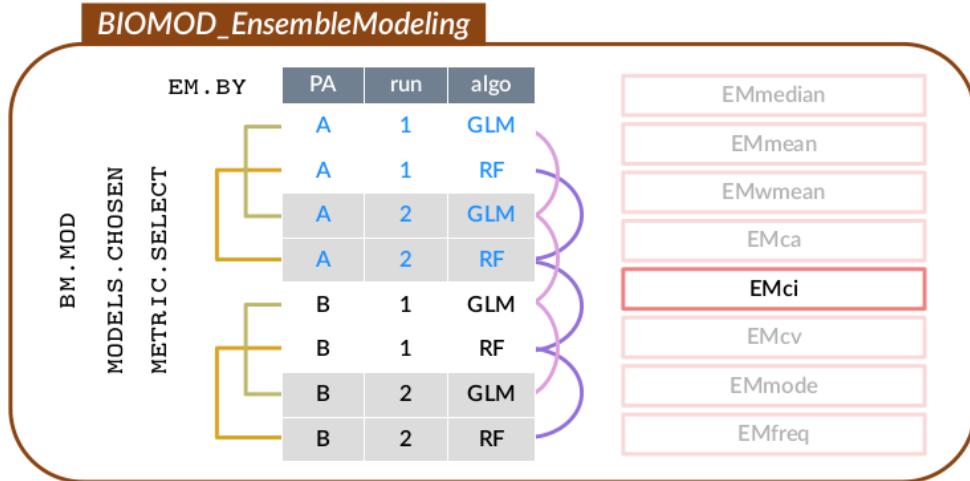
2.b Ensemble models

Step 1 : filter single models

Step 2 : gather single models

Step 3 : build ensemble models

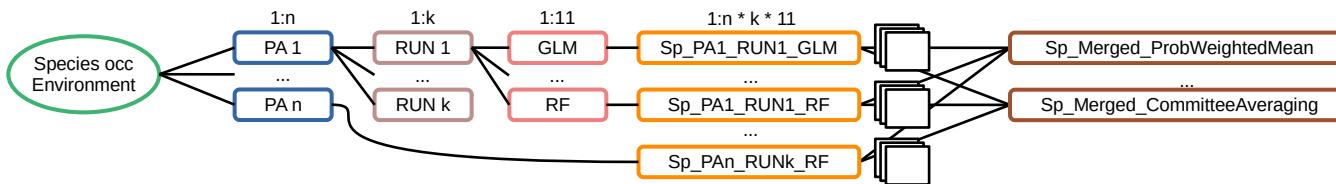
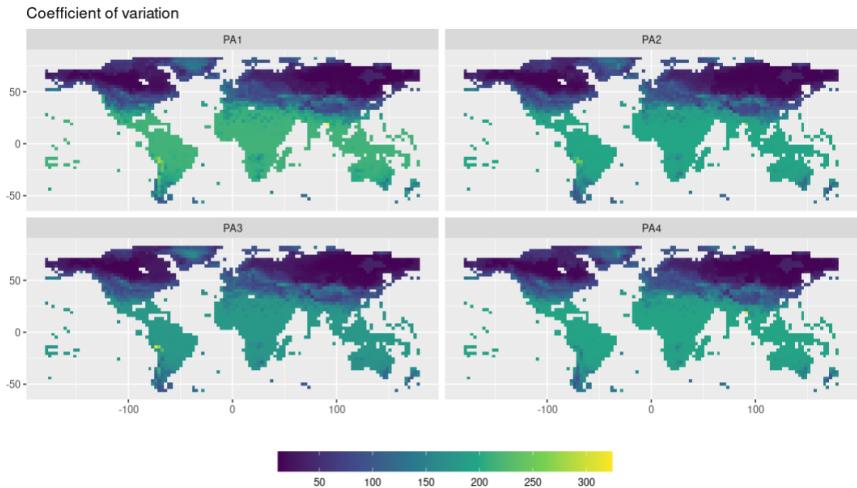
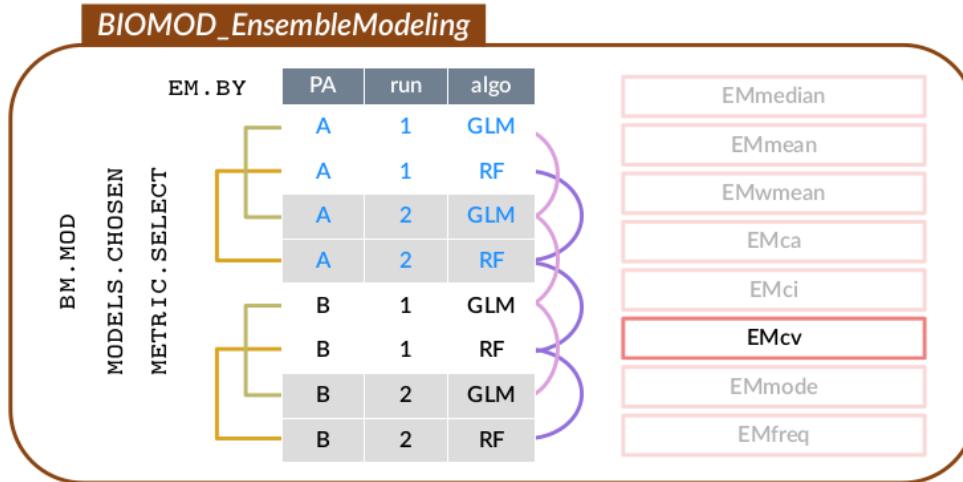
- » « simple » ensemble models : mean or median
 - » « complex » ensemble models :
 - probability weighted mean
 - committee averaging
 - » « exploratory » ensemble models :
 - **confidence intervals** or **coefficient of variation**



2.b Ensemble models

- Step 1 : filter single models
- Step 2 : gather single models
- Step 3 : build ensemble models

- » « simple » ensemble models : mean or median
- » « complex » ensemble models :
 - probability weighted mean
 - committee averaging
- » « exploratory » ensemble models :
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2.b Ensemble models

Step 1 : filter single models

Step 2 : gather single models

Step 3 : build ensemble models

- » « simple » ensemble models : mean or median

- » « complex » ensemble models :

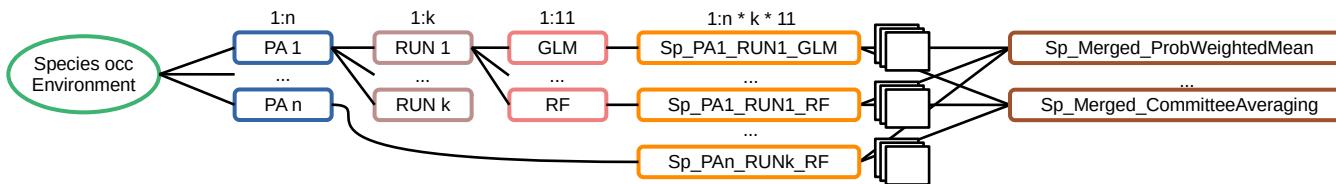
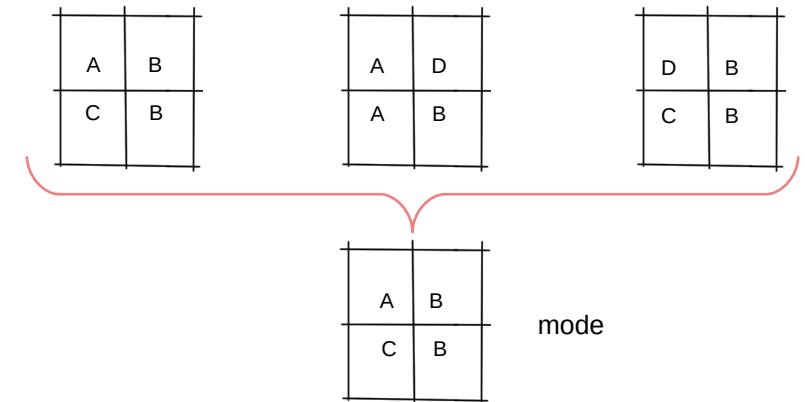
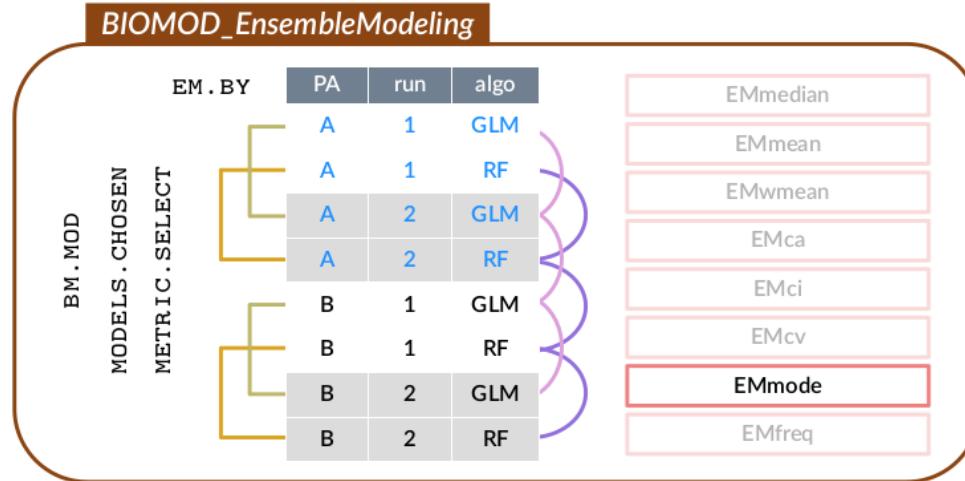
- probability weighted mean
- committee averaging

- » « exploratory » ensemble models :

- confidence intervals or coefficient of variation

- » « multiclass » ensemble models :

- **mode or frequency of the mode**



2.b Ensemble models

Step 1 : filter single models

Step 2 : gather single models

Step 3 : build ensemble models

- » « simple » ensemble models : mean or median

- » « complex » ensemble models :

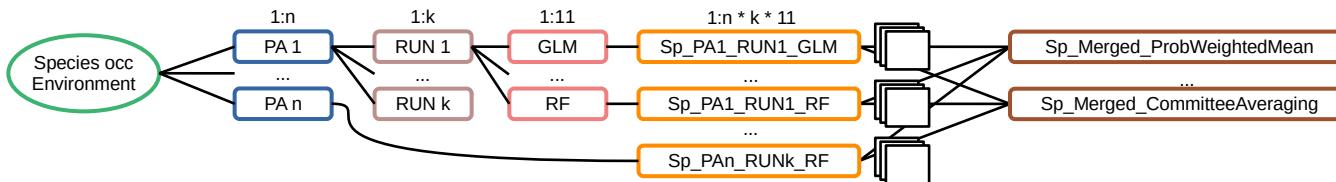
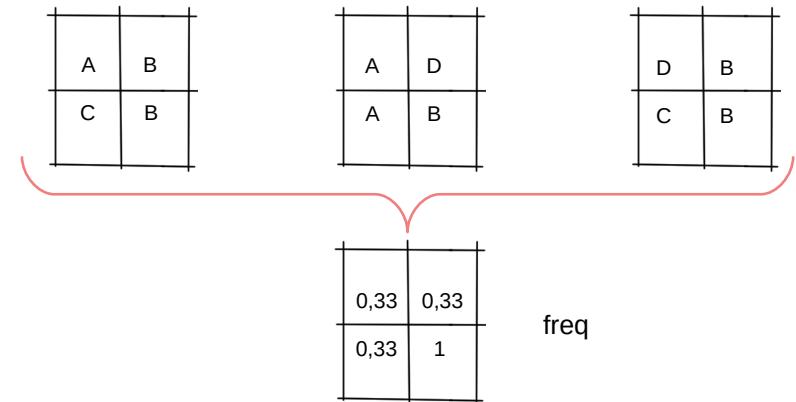
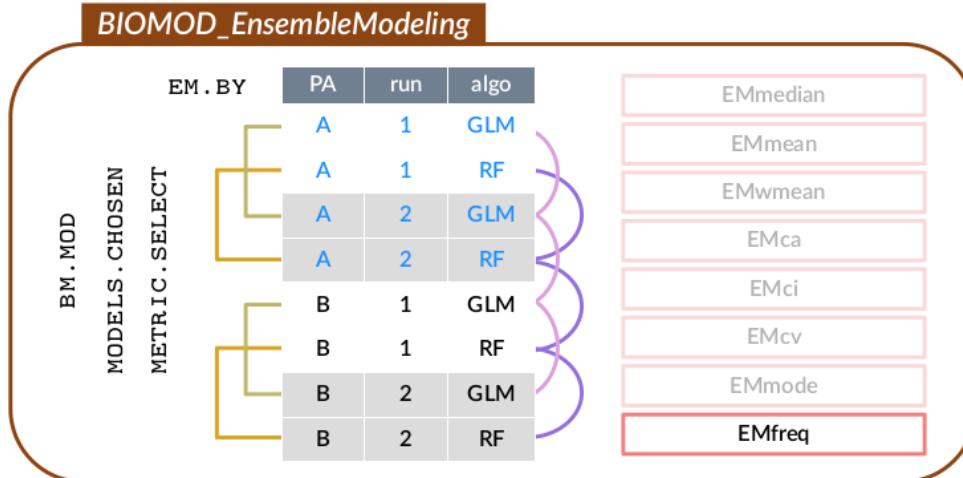
- probability weighted mean
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- » « exploratory » ensemble models :

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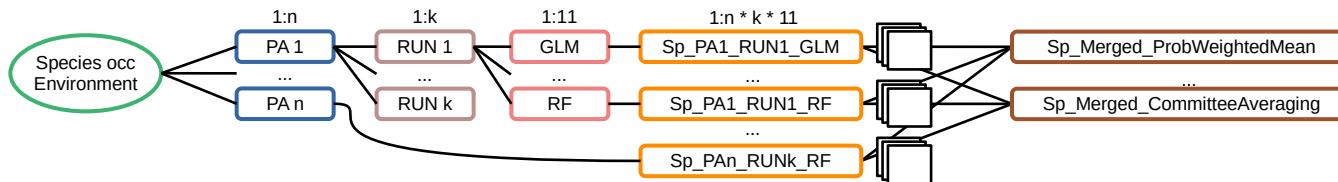
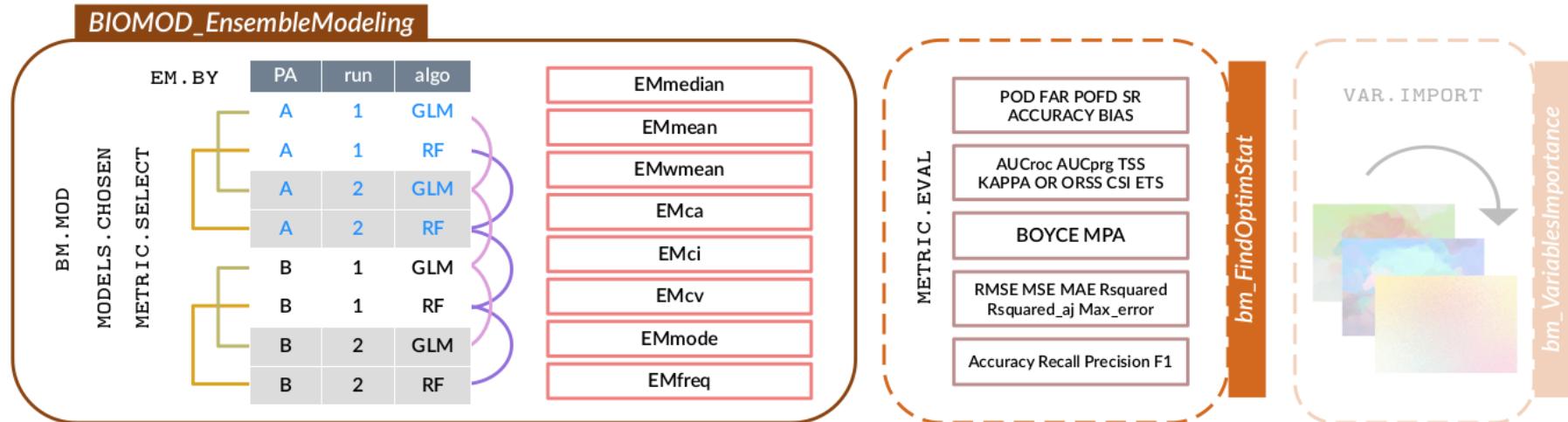
- » « multiclass » ensemble models :

- **mode** or **frequency of the mode**



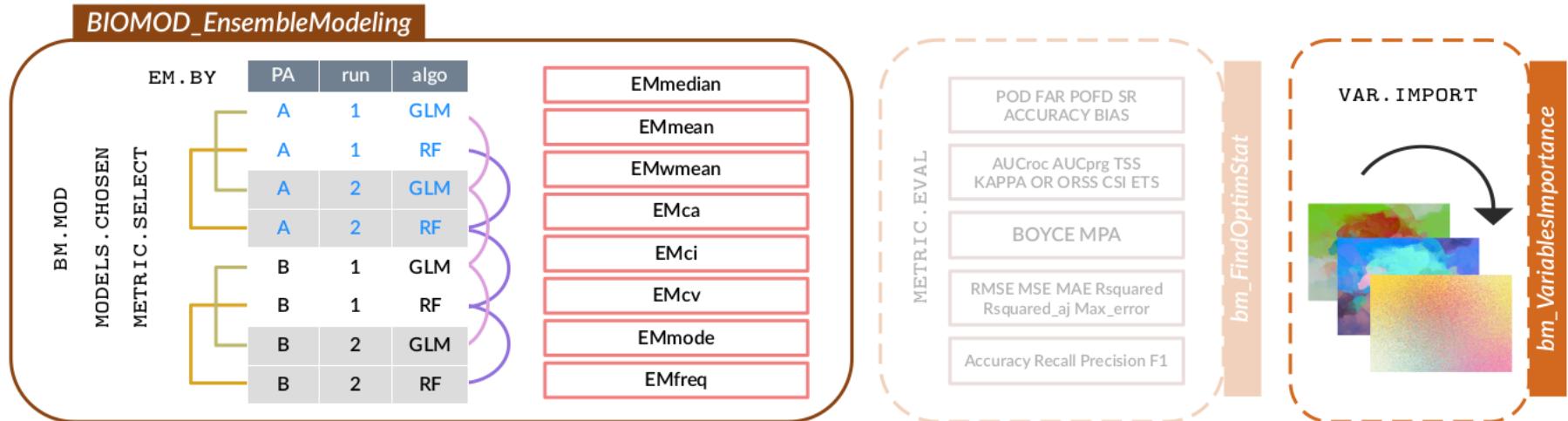
2.b Ensemble models

- » except ROC, all evaluation metrics obtained from contingency table (containing *TP*, *FP*, *TN*, *FN*)
- » require a **binary transformation** :
 - range of thresholds tested
 - keep threshold optimising the evaluation metric



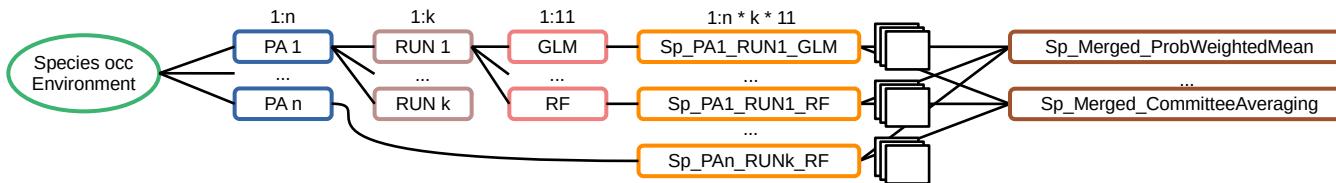
2.b Ensemble models

- » comparison of importance of variables between models
- » **Pearson correlation** between :
 - normal prediction
 - prediction with 1 variable randomised

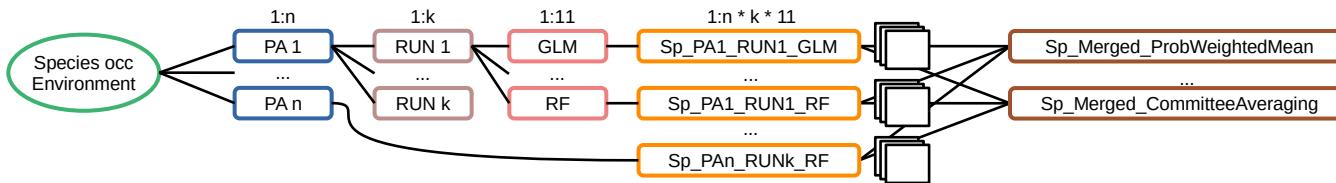
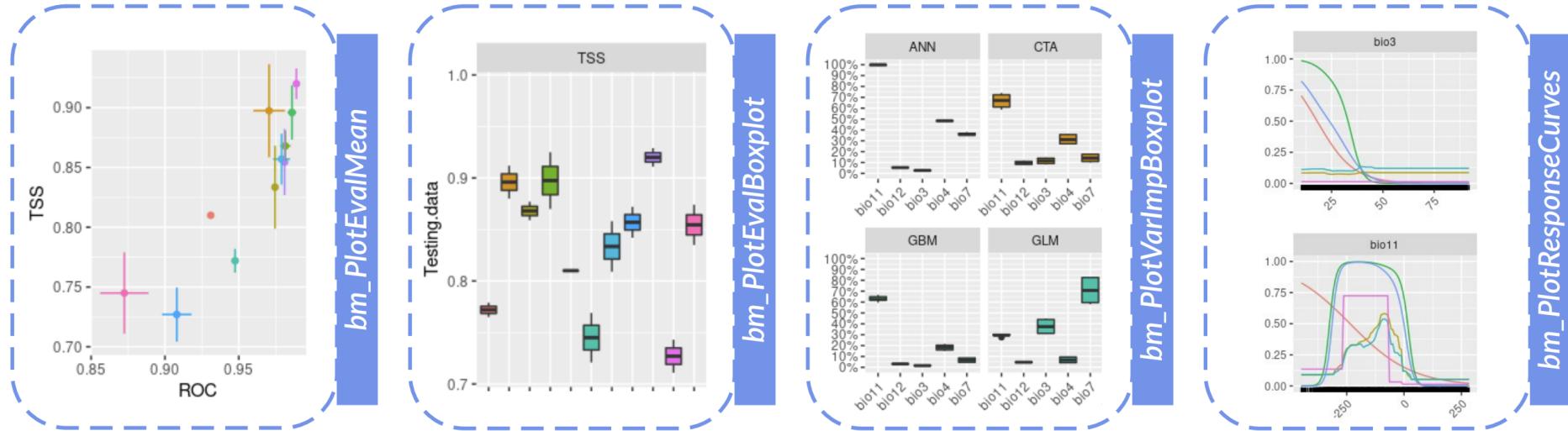


! **IMPORTANT**

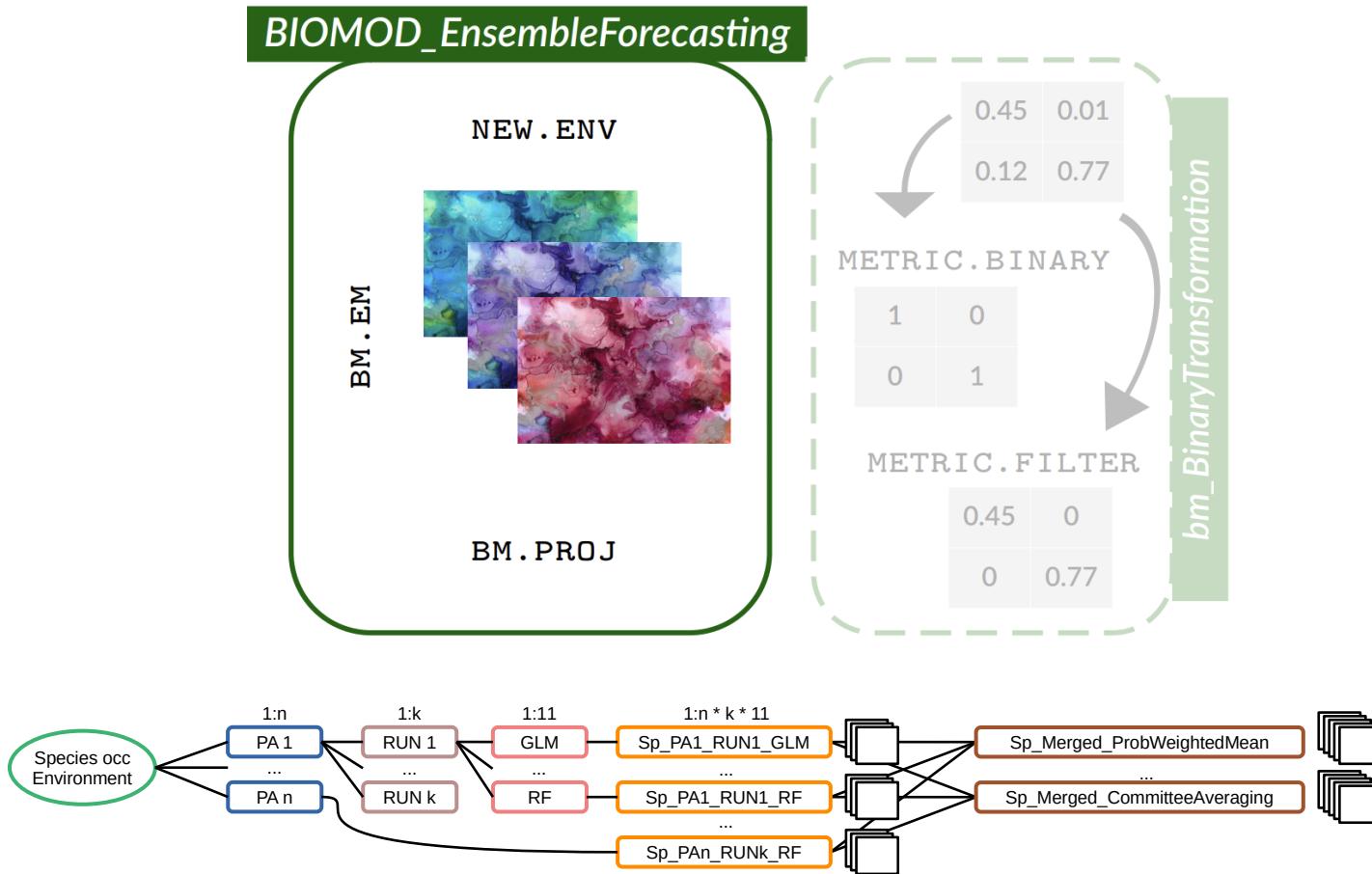
← Takes time ! Has to go through the whole workflow →



3.b Exploring ensemble models

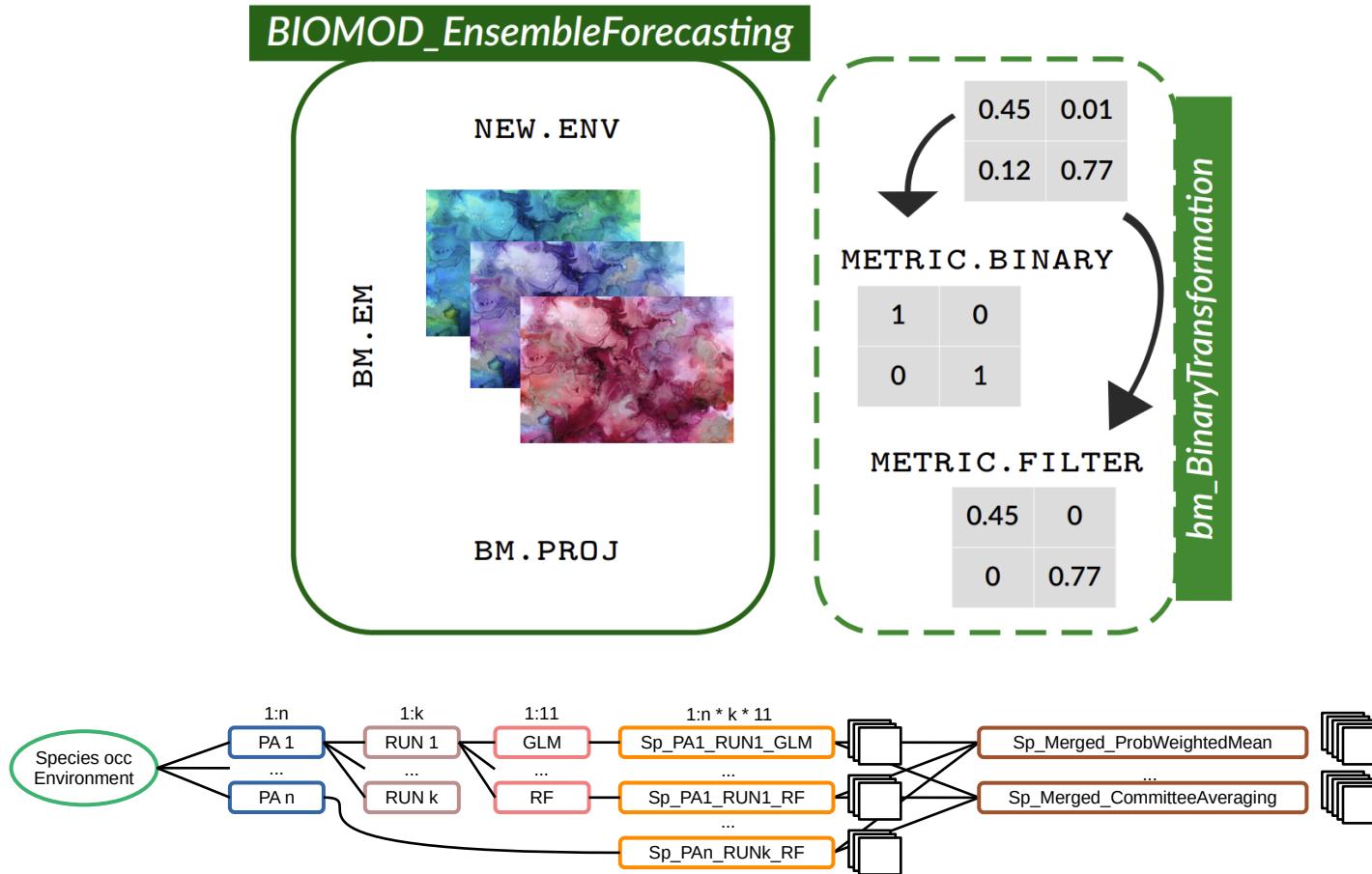


4.b Projecting ensemble models

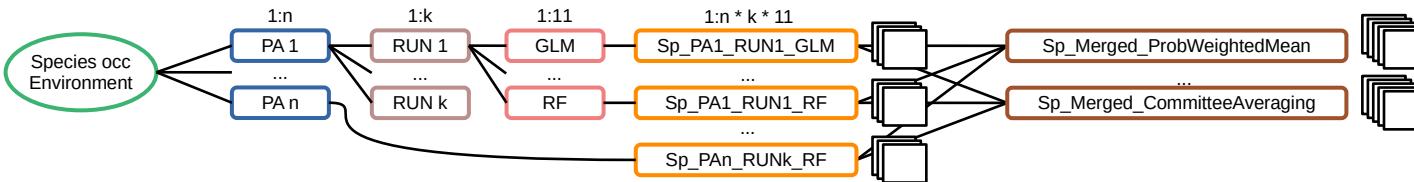
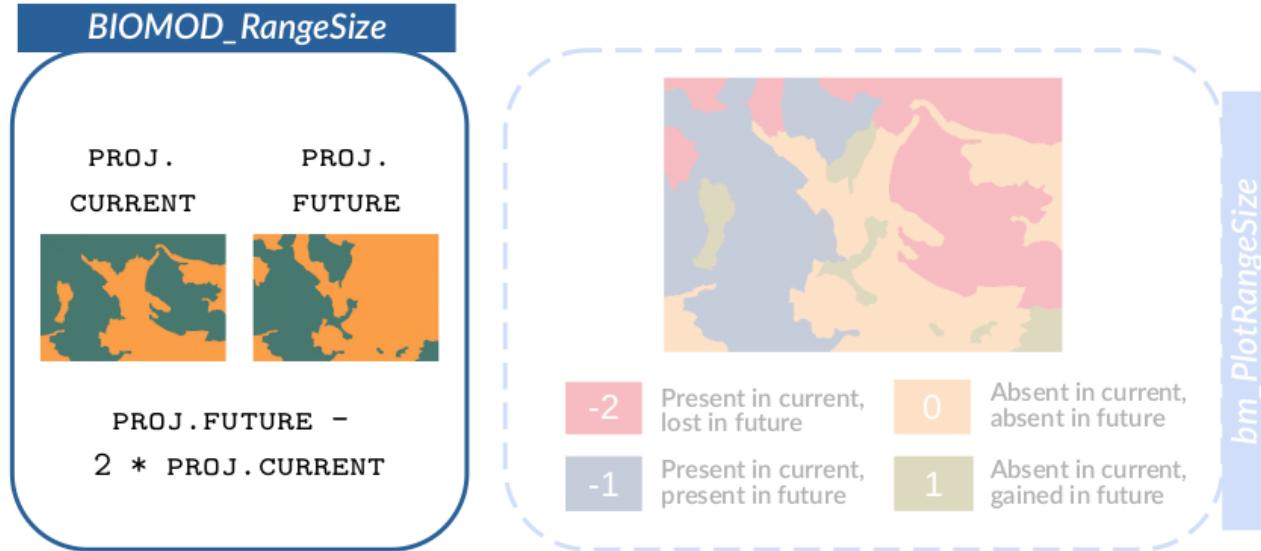


4.b Projecting ensemble models

- » transformation associated to one evaluation metric (one map created for each metric selected)
- » use the threshold maximising the chosen metric

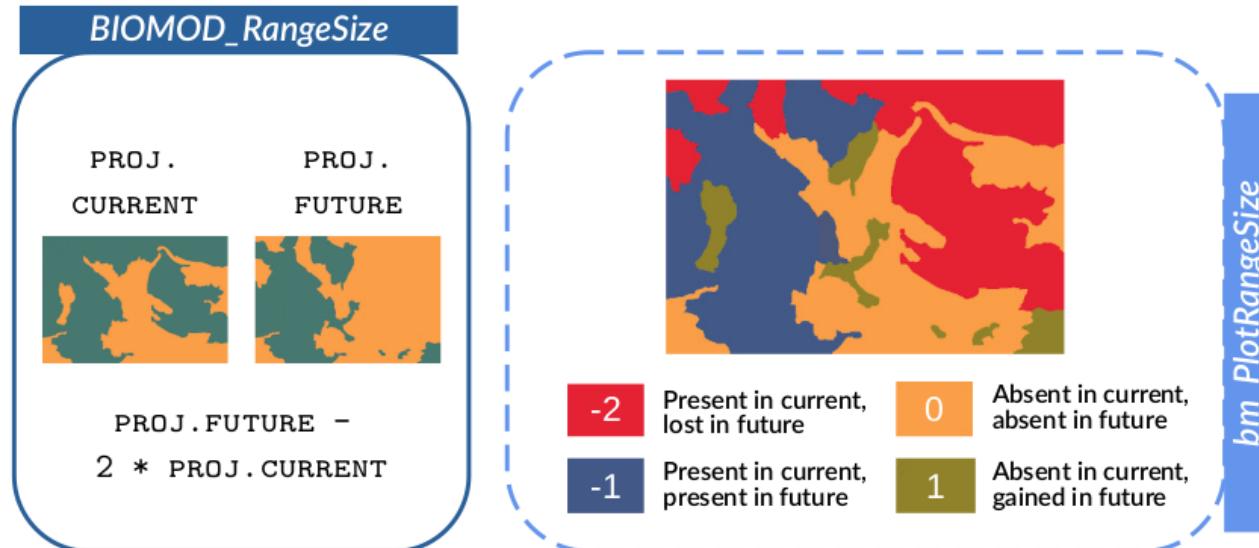


5. Species range change



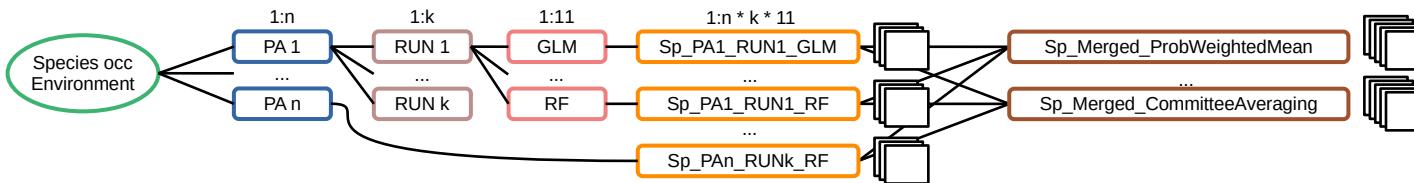
5. Species range change

- » explore **spatially** the difference in predictions
- » provide **summary values** :
 - percentage of loss / gain
 - species range change

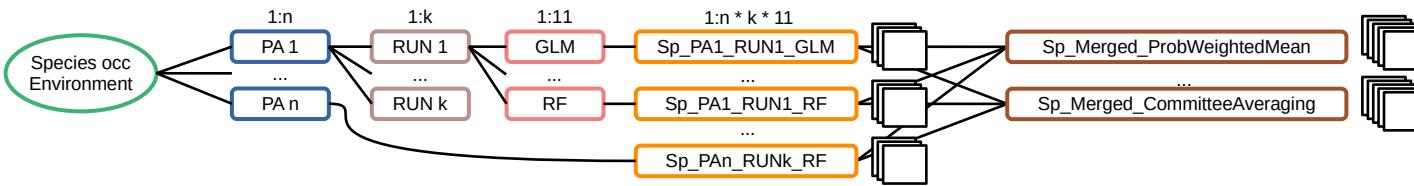
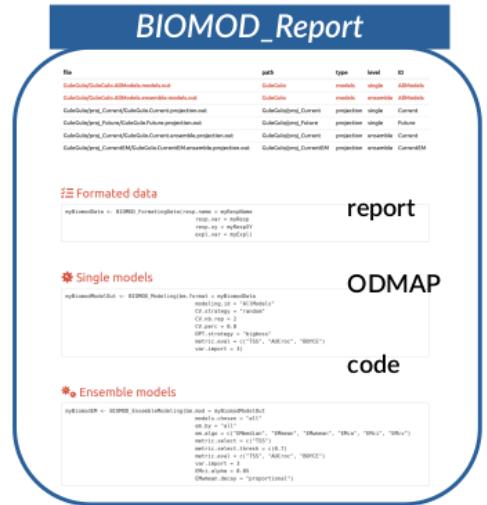


! **IMPORTANT**

Work with *binary maps* (and not predictions between 0 and 1)



6. Report / ODMAP / code

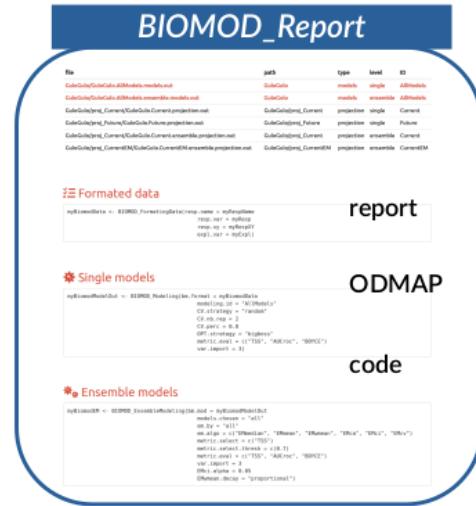


6. Report / ODMAP / code

biomod2 - Modeling GuloGulo - report

Last compiled on 10 décembre, 2025

- Formated data
- Single models
 - Modeling options
 - Evaluation
 - Variables' importance
 - Response curves
- Ensemble models
 - Evaluation
 - Variables' importance
 - Response curves
- Projections



- report : provide all possible prints, summaries, results and plots from a biomod2 object

file	path	type	level	ID
GuloGulo/GuloGulo.AllModels.models.out	GuloGulo	models	single	AllModels
GuloGulo/GuloGulo.AllModels.ensemble.models.out	GuloGulo	models	ensemble	AllModels
GuloGulo/proj_Current/GuloGulo.Current.projection.out	GuloGulo/proj_Current	projection	single	Current
GuloGulo/proj_Future/GuloGulo.Future.projection.out	GuloGulo/proj_Future	projection	single	Future
GuloGulo/proj_Current/GuloGulo.Current.ensemble.projection.out	GuloGulo/proj_Current	projection	ensemble	Current
GuloGulo/proj_CurrentEM/GuloGulo.CurrentEM.ensemble.projection.out	GuloGulo/proj_CurrentEM	projection	ensemble	CurrentEM

Formated data

The GuloGulo species dataset contains 2488 points.

The GuloGulo species dataset contains 661 presences and 1827 true absences.

```
=====
BIOMOD.formated.data =====

Modeling directory (dir.name) : /home/gueguema/Documents/_PACKAGES/biomod2

Modeled species (sp.name) : GuloGulo

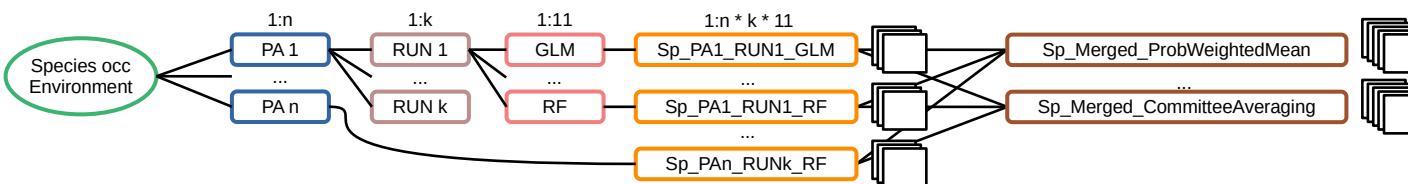
661 presences, 1827 true absences and 0 undefined points in dataset

5 explanatory variables

  bio3      bio4      bio7      bioll      bio12
Min. :10.19  Min. : 72  Min. :54.5  Min. :-447.75  Min. : 0.0278
1st Qu.:21.22 1st Qu.:2641 1st Qu.:186.0 1st Qu.:-184.32 1st Qu.: 276.4931
Median :35.00  Median :6682  Median :306.2  Median : 24.23  Median :562.9308
Mean   :40.29  Mean   :7356  Mean   :310.9  Mean   : -2.64  Mean   :853.5157
3rd Qu.:56.35 3rd Qu.:11752 3rd Qu.:424.6 3rd Qu.: 196.30 3rd Qu.:1200.5915
Max.  :92.00  Max.  :22314  Max.  :718.0  Max.  : 283.00  Max.  :5431.0020
```

dataset	run	PA	Presences	True_Absences	Pseudo_Absences	Undefined
initial	NA	NA	661	1827	0	0

! Evaluation data is missing, plot.eval set to FALSE

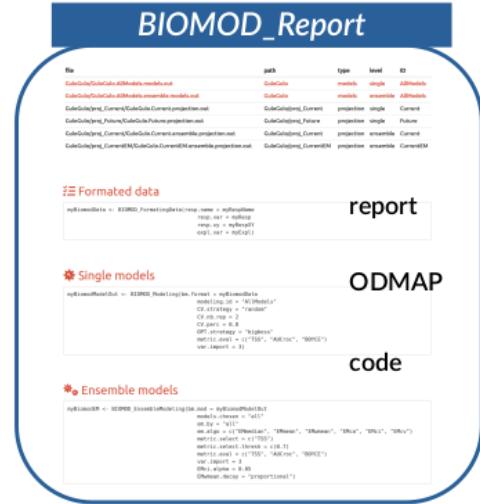


6. Report / ODMAP / code

biomod2 - Modeling GuloGulo - ODMAP

Last compiled on 08 décembre, 2025

- **ODMAP protocol**
 - Overview
 - Data
 - Model
 - Assessment
 - Prediction



» **report** : provide all possible prints, summaries, results and plots from a biomod2 object

» **ODMAP** : pre-fill ODMAP protocol following Zurell et al. 2020 with biomod2 modelling elements

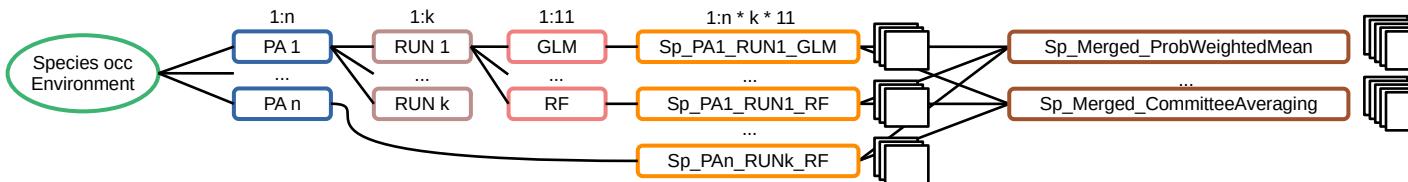
▪ **ODMAP protocol**

Overview

Authorship	
Study title	tobefilledbyuser
Author names	tobefilledbyuser
Contact	tobefilledbyuser
Study link	tobefilledbyuser
Model objective	
Model objective	NA
Target output	tobefilledbyuser
Focal Taxon	
Focal Taxon	GuloGulo
Location	
Location	tobefilledbyuser
Scale of Analysis	
Spatial extent	ext(-180, 180, -57.499992177, 83.500007823)
Spatial resolution	3, 3
Temporal extent	NA
Temporal resolution	NA
Boundary	NA
Biodiversity data	
Observation type	NA
Response data type	presence-absence
Predictors	
Predictor types	NA
Hypotheses	
Hypotheses	tobefilledbyuser
Assumptions	
Model assumptions	tobefilledbyuser
Algorithms	
Modelling techniques	ANN, CTA, FDA, GAM, GBM, GLM, MARS, MAXENT, MAXNET, RF, RFd, SRE, XGBOOST
Model complexity	NA
Model averaging	EMmeanByTSS, EMcaByTSS, EMciInfByTSS, EMciSupByTSS, EMcvByTSS, EMmeanByTSS, EMmedianByTSS,
EMmeanByTSS	
Workflow	
Model workflow	NA
Software	
Software	biomod2 v4.3.4.3
Code availability	NA
Data availability	NA

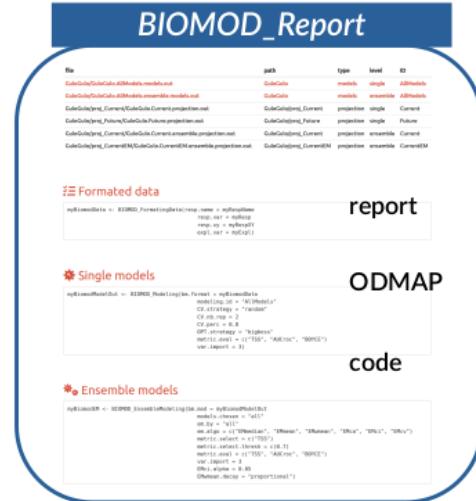


Overview, Data, Model, Assessment and Prediction



6. Report / ODMAP / code

- » **report** : provide all possible prints, summaries, results and plots from a biomod2 object
- » **ODMAP** : pre-fill ODMAP protocol following Zurell et al. 2020 with biomod2 modelling elements
- » **code** : provide command lines used to call main functions



⌘ Formated data

```

myBiomodData <- BIOMOD_FormattingData(resp.name = myRespName
                                         resp.var = myResp
                                         resp.xy = myResXY
                                         expl.var = myExpl)

```

✿ Single models

```

myBiomodModelOut <- BIOMOD_Modeling(bm.format = myBiomodData
#>                                     modeling.id = "AllModels"
#>                                     CV.strategy = "random"
#>                                     CV.nb.rep = 2
#>                                     CV.perc = 0.8
#>                                     OPT.strategy = "bigboss"
#>                                     metric.eval = c("TSS", "AUCroc", "BOYCE")
#>                                     var.import = 3)

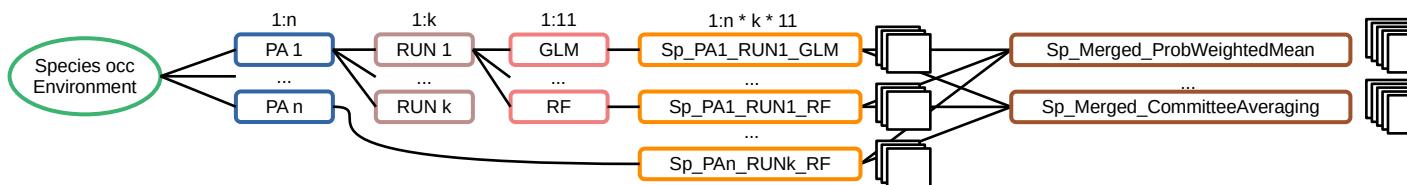
```

✿ Ensemble models

```

myBiomodEM <- BIOMOD_EnsembleModeling(bm.mod = myBiomodModelOut
#>                                         models.chosen = "all"
#>                                         em.by = "all"
#>                                         em.algo = c("EMmedian", "EMmean", "EMwmean", "EMca", "EMci", "EMcv")
#>                                         metric.select.thresh = c(0.7)
#>                                         metric.eval = c("TSS", "AUCroc", "BOYCE")
#>                                         var.import = 3
#>                                         EMci.alpha = 0.05
#>                                         EMwmean.decay = "proportional")

```



**Species distribution modeling,
calibration and evaluation,
ensemble modeling**



Thuiller, W. (2003), *BIOMOD – optimizing predictions of species distributions and projecting potential future shifts under global change*. *Global Change Biology*, 9: 1353-1362. <https://doi.org/10.1046/j.1365-2486.2003.00666.x>

Thuiller, W., Lafourcade, B., Engler, R. and Araújo, M.B. (2009), *BIOMOD – a platform for ensemble forecasting of species distributions*. *Ecography*, 32: 369-373. <https://doi.org/10.1111/j.1600-0587.2008.05742.x>

<https://github.com/biomodhub/biomod2/>

<https://biomodhub.github.io/biomod2/>