HEMATOXYLIN HARRIS

IVD *In vitro* diagnostic medical device CE

Modified hematoxylin acc. to Harris for nuclear staining
Reagent for strong, regressive staining in histopathology

INSTRUCTIONS FOR USE

REF Product code: 01HEMH100 (100 ml) 01HEMH110 (10x100 ml) 01HEMH500 (500 ml) 01HEMH1000 (1000 ml) 01HEMH2500 (2500 mL)

Introduction:

Hematoxylin H is a well-known formulation of hematoxylin used in histopathology for a more precise nuclear cell staining. Hematoxylin acc. to Harris is applied using a regressive method in a routine hematoxylin and eosin (HE) staining in histology.
Hematoxylin is extracted from logwood (*Haematoxylon campechianum L.*). Hematoxylin oxidizes to hematein and binds with metal ions (mordants), hematein turns into irreplaceable nuclear color. Positively charged hematein-mordant complex then binds with negatively charged phosphate ions of the DNA's nucleus, creating characteristic blue coloration. The original Hematoxylin acc. to Harris formula is oxidized with mercury oxide. However, Histo-Line version of Hematoxylin acc. to Harris does not contain mercury oxide because of its toxicity; environment-friendly sodium or potassium iodate is used instead. Hematoxylin H stains nuclear membrane, nucleoplasm and nucleolus exceptionally well.

Product description:

- **HEMATOXYLIN H** - Reagent for regressive nuclear staining in histopathology. Contains optimally oxidized hematoxylin with sodium iodate, aluminum ions and antioxidants.

Other slides and reagents that may be used in staining:

- Fixatives such as Histo-Line neutral buffered formaldehyde solutions: Formaldehyde NB 4%, Formaldehyde NB 10%
- Dehydrating/rehydrating agent, such as Histo-Line alcohol solutions: Histanol 70, Histanol 80, Histanol 95 and Histanol 100
- Clearing agents, such as xylene or a substitute, such as Citro Histoclear New agent on the aliphatic hydrocarbons basis
- Infiltration and fitting agent, such as Histo-Line granulated paraffin Lab-O-Wax Plus 56/58
- High-quality glass slides for use in histopathology and cytology, such as SUPER GRADE or one of more than 30 models of Histo-Line glass slides
- Differentiation agent, such as Histo-Line Acid alcohol
- Bluing agents, such as Scott's solution or Bluing reagent
- Covering agents for microscopic sections and mounting cover glass, such as Histo-Line HistoMount, HistoMount High, HistoMount M, HistoMount New, HistoMount New Low, HistoMount DPX, HistoMount DPX High, HistoMount DPX Low, HistoMount DPX Low Eco, HistoMount C, HistoMount Aqua, Canada Balsam
- Cover glass, dimensions range from 18x18mm to 24x60mm
- Counterstaining reagents, such as Histo-Line eosin solutions:

Preparing histological sections for staining

- Fix the tissue sample tightly (4% NB Formaldehyde, 10% NB Formaldehyde), rinse with water and dehydrate through series of ascending alcohol solutions (Histanol 70, Histanol 80, Histanol 95 and Histanol 100)
- Clear the sample with intermedium; in xylene or in a xylene substitute (Citro Histoclear)
- Infiltrate and fit the sample in paraffin Lab-O-Wax Plus 56/58
- Cut the paraffin block to 4-6 µm slices and place them on a glass slide

Hematoxylin and eosin (HE) staining procedure, progressive

1.	Deparaffinize the section in xylene or in a xylene substitute (Citro Histoclear)	3 exchanges, 2 min each
2.	Rehydrate using 100% alcohol (Histanol 100)	2 exchanges, 5 and 3 min
3.	Rehydrate using 95% alcohol (Histanol 95)	2 min
4.	Rehydrate in distilled (demi) water	2 min
5.	Stain using Hematoxylin H	3-5 minutes
	Note: In the case of subsidence in the solution or a formation of metallic glow on the surface, reagent should be filtrated before use	
6.	Immerse the section in distilled/demineralized water until dye is no longer being released from the preparation	
7.	Bluing using Scott's solution or Bluing reagent	1 min
	Note: End the process of bluing after the nuclei turn blue	
8.	Stain with one of eosin contrast solutions until the section is optimally stained	15 seconds - 2 minutes
	Note: Staining the sections in eosin alcoholic solutions causes intensive eosinophil color to show much faster (in under 15 seconds' time). Exposition time for Eosin Y aqueous solutions is 2 min and 90 seconds, respectively.	
9.	Rinse under tap water	2 min
10.	Dehydrate using 95% alcohol (Histanol 95)	2 exchanges, 10-15 dips
11.	Dehydrate using 100% alcohol (Histanol 100)	3 exchanges, 10-15 dips
12.	Clear the section in xylene or in a xylene substitute (Citro Histoclear)	2 exchanges, 2 min each

Immediately after clearing apply an appropriate BioMount medium for covering/mounting on the section. If Citro Histoclear xylene was used, use one of Histo-Line mounting xylene-based media (BioMount, BioMount High, BioMount M, BioMount DPX, BioMount C, or universal BioMount New). If Citro Histoclear or xylene substitute was used, the appropriate covering agent is BioMount New. Cover the section with a cover glass.

Hematoxylin and eosin (HE) staining procedure, regressive

1.	Deparaffinize the section in xylene or in a xylene substitute (Citro Histoclear)	3 exchanges, 2 min each
2.	Rehydrate using 100% alcohol (Histanol 100)	2 exchanges, 5 and 3 min
3.	Rehydrate using 95% alcohol (Histanol 95)	2 min
4.	Rehydrate in distilled (demi) water	2 min
5.	Stain using Hematoxylin H	8-15 minutes
	Note: In the case of subsidence in the solution or a formation of metallic glow on the surface, reagent should be filtrated before use.	
6.	Immerse the section in distilled or demineralized water until dye is no longer being released from the section	
7.	Differentiate using Acid alcohol	3-10 dips
	Note: This step removes excessive hematoxylin from the nucleus and cytoplasm. Discoloration of the nuclei can occur if the section is treated with the differentiation agent for too long.	
8.	Rinse in distilled water	
9.	Bluing using Scott's solution or Bluing reagent	1 min
	Note: End the process of bluing after the nuclei turn blue	
10.	Stain with one of eosin contrast solutions until the section is optimally stained	15 seconds - 2 minutes
	Note: Staining the sections in eosin alcoholic solutions causes intensive eosinophil color to show much faster (in under 15 seconds' time). Exposition time for eosin aqueous solutions is 2 min and 90 seconds, respectively.	
11.	Rinse under tap water	2 min
12.	Dehydrate using 95% alcohol (Histanol 95)	2 exchanges, 10-15 dips
13.	Dehydrate using 100% alcohol (Histanol 100)	3 exchanges, 10-15 dips
14.	Clear the section in xylene or in a xylene substitute (Citro Histoclear)	2 exchanges, 2 min each

Immediately after clearing apply an appropriate BioMount medium for covering/mounting on the section. If Citro Histoclear xylene was used, use one of Histo-Line mounting xylene-based media (BioMount, BioMount High, BioMount M, BioMount DPX, BioMount C, or universal BioMount New). If Citro Histoclear xylene substitute was used, the appropriate covering agent is BioMount New. Cover the section with a cover glass.

Result:

Nucleus - blue

Cytoplasm, collagen, muscle fibers, erythrocytes - hues of pink (hues of red when staining with Eosin Contrast and Eosin Contrast Plus)

Note

Time periods of staining processes are not entirely standardized and they approximately correspond to clinical and laboratory practical experience. Intensity of staining depends on the period of immersion in the dye. Real staining protocol depends on personal requests and priorities.

Preparing the sample and diagnostics

Use only appropriate instruments for collecting and preparing the samples. Process the samples with modern technology and mark them clearly. Follow the manufacturer's instructions for handling. In order to avoid mistakes, the staining procedure and diagnostics should only be conducted by authorized and qualified personnel. Use only microscope according to standards of the medical diagnostic laboratory.

Safety at work and environmental protection

Handle the product in accordance with safety at work and environmental protection guidelines. Used solutions and out of date solutions should be disposed of as special waste in accordance with national guidelines. Chemicals used in this procedure could pose danger to human health. Tested tissue specimens are potentially infectious. Necessary safety measures for protecting human health should be taken in accordance with good laboratory practice. Act in accordance with signs and warnings notices printed on the product's label, as well as in Histo-Line material safety data sheet.

Storing, stability and expiry date

Keep Hematoxylin H in a tightly closed original package at room temperature. Keep in dry places, do not freeze and avoid exposing to direct sunlight. Date of manufacture and expiry date are printed on the product's label.

References

1. Baker, J.R. (1962): Experiments on the action of mordants. 2. Aluminium-hematein. *Q.J.Microsc. Sci.* p103 493-517.
2. Conn, J. (1977): *Biological Stains*, 9th ed., Baltimore: Williams and Wilkens Co.
3. Harris, H.F. (1898): A new method of "ripening" haematoxylin. *Microsc. Bull.* (Philadelphia) Dec. 47.
4. Harris, H.F. (1900): On the rapid conversion of haematoxylin into haematein in staining reactions. J. *Appl. Microsc.* p3 777-780.

01HEMHX, V10 EN4, 29 June 2016, IŠP/VR

	Refer to the supplied documentation	°C °F	Storage temperature range	Σ	Number of tests in package	REF	Product code	CE	European Conformity
	Refer to supplied instructions		Keep away from heat and sunlight		Valid until	LOT	Lot number		Manufacturer
IVD	For *in vitro* diagnostic use only		Keep in dry place		Caution - fragile				

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