Download the genome file of Fusarium graminearum from ensembl genome

```
In [2]: ! wget ftp://ftp.ensemblgenomes.org/pub/release-51/fungi/fasta/fusarium_graminearum
       --2024-04-14 18:07:43-- ftp://ftp.ensemblgenomes.org/pub/release-51/fungi/fasta/fus
       arium_graminearum/dna/Fusarium_graminearum.RR1.dna.toplevel.fa.gz
                 => 'fusa_genome.fasta.gz'
       Resolving ftp.ensemblgenomes.org (ftp.ensemblgenomes.org)... 193.62.193.161
       Connecting to ftp.ensemblgenomes.org (ftp.ensemblgenomes.org)|193.62.193.161|:21...
       connected.
       Logging in as anonymous ... Logged in!
       ==> SYST ... done. ==> PWD ... done.
       ==> TYPE I ... done. ==> CWD (1) /pub/release-51/fungi/fasta/fusarium_graminearum/d
       na ... done.
       ==> SIZE Fusarium_graminearum.RR1.dna.toplevel.fa.gz ... 11624507
       ==> PASV ... done.
                           ==> RETR Fusarium_graminearum.RR1.dna.toplevel.fa.gz ... done.
       Length: 11624507 (11M) (unauthoritative)
       Fusarium_graminearu 100%[========>] 11.09M 6.34MB/s
       2024-04-14 18:07:47 (6.34 MB/s) - 'fusa_genome.fasta.gz' saved [11624507]
        the size and format of the downloaded file
In [3]: !du -h fusa_genome.fasta.gz
       12M
               fusa_genome.fasta.gz
        Decompress this file while keeping the compressed version as wel
In [4]: ! gunzip -c fusa_genome.fasta.gz > fusa_genome.fasta
In [5]: | ! du -h fusa_genome.fasta.gz fusa_genome.fasta
       12M
               fusa_genome.fasta.gz
       37M
               fusa_genome.fasta
        Identify the sequences present in this file along with their identifiers
In [6]: ! grep '>' fusa_genome.fasta && echo "Number of sequences : $(grep -c '>' fusa_geno
       >1 dna:chromosome chromosome:RR1:1:1:11760891:1 REF
       >2 dna:chromosome chromosome:RR1:2:1:8997558:1 REF
       >3 dna:chromosome chromosome:RR1:3:1:7792947:1 REF
       >4 dna:chromosome chromosome:RR1:4:1:9395062:1 REF
       >Mt dna:chromosome chromosome:RR1:Mt:1:95638:1 REF
       >HG970330 dna:supercontig supercontig:RR1:HG970330:1:5846:1 REF
       Number of sequences: 6
        The size of the genome of Fusarium graminearum
          grep -v '>' fusa_genome.fasta | tr -d '\n' | wc -c
```

38047942

The number of A/T/G/C bases in the genome

```
In [8]: ! grep -c '[ATCG]' fusa_genome.fasta
       634136
In [9]:
        %%bash
        echo "Retrieve the number of A/T/G/C bases from the genome and deduce the overall G
        # Calculate the total number of nucleotides
        Total_Nucleotide=$(grep -v '>' fusa_genome.fasta | tr -d '\n' | wc -c)
        # Calculate the number of G bases
        Number_of_Ng=$(grep -v '>' fusa_genome.fasta | tr -d -c 'G' | wc -c)
        # Calculate the number of C bases
        Number_of_Nc=$(grep -v '>' fusa_genome.fasta | tr -d -c 'C' | wc -c)
        # Calculate the sum of G and C counts
        GC_sum=$((Number_of_Ng + Number_of_Nc))
        # Calculate the overall GC content
        GC_content=$(bc -1 <<< "scale=2; ($GC_sum / $Total_Nucleotide) * 100")
        echo "The overall GC content of Fusarium graminearum is: $GC content%"
```

Retrieve the number of A/T/G/C bases from the genome and deduce the overall GC conte nt of Fusarium graminearum.

The overall GC content of Fusarium graminearum is: 48.00%

We want to use the restriction enzyme BamH1, whose sequence is GGATCC. To guide the insertion of the reporter gene into this site.

The potential restriction sites are located in the genome.

```
In [10]: %%bash
    potential_restriction=$(grep --only-matching GGATCC fusa_genome.fasta | wc -1)
    echo $potential_restriction

6706

The distance between two BamH1 sites (in Kb) "average"

In [11]: %%bash
    dd=$(expr $Total_Nucleotide / $potential_restriction )
    echo "$dd"

/
```

Download the structural annotation of Fusarium graminearum

```
In [12]: ! wget ftp://ftp.ensemblgenomes.org/pub/release-51/fungi/gff3/fusarium_graminearum/
```

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```
--2024-04-14 18:07:50-- ftp://ftp.ensemblgenomes.org/pub/release-51/fungi/gff3/fusa
        rium_graminearum/Fusarium_graminearum.RR1.51.chr.gff3.gz
                   => 'fusa annot.gff3.gz'
        Resolving ftp.ensemblgenomes.org (ftp.ensemblgenomes.org)... 193.62.193.161
        Connecting to ftp.ensemblgenomes.org (ftp.ensemblgenomes.org)|193.62.193.161|:21...
        connected.
        Logging in as anonymous ... Logged in!
        ==> SYST ... done.
                           ==> PWD ... done.
        ==> TYPE I ... done. ==> CWD (1) /pub/release-51/fungi/gff3/fusarium_graminearum ..
        . done.
        ==> SIZE Fusarium_graminearum.RR1.51.chr.gff3.gz ... 2010033
        ==> PASV ... done.
                            ==> RETR Fusarium_graminearum.RR1.51.chr.gff3.gz ... done.
        Length: 2010033 (1.9M) (unauthoritative)
        Fusarium graminearu 100%[========>] 1.92M 2.14MB/s
                                                                            in 0.9s
        2024-04-14 18:07:52 (2.14 MB/s) - 'fusa_annot.gff3.gz' saved [2010033]
         The features described in this GFF3 file
In [13]: ! zcat fusa_annot.gff3.gz | grep -v '#'|cut -f 3|sort|uniq
        CDS
        RNase_MRP_RNA
        RNase_P_RNA
        SRP_RNA
        biological_region
        chromosome
        exon
        five_prime_UTR
        gene
        lnc_RNA
        mRNA
        ncRNA_gene
        rRNA
        snRNA
        snoRNA
        tRNA
        three_prime_UTR
         Types of different features are you counting
In [14]: |! zcat fusa_annot.gff3.gz | grep -v '#'|cut -f 3|sort|uniq|wc -1
        17
         The genes annotated on this genome
In [15]: ! zcat fusa_annot.gff3.gz|grep -w gene|grep 'ID=gene'|wc -l
        14898
```

Genes and mRNA annotated on each of the chromosomes

```
! zcat fusa_annot.gff3.gz|grep -w gene|grep 'biotype=protein_coding'|cut -f1,3|sort
  4390 1
                gene
  4390 1
                mRNA
  3648 2
                gene
  3648 2
                mRNA
  3085 3
                gene
  3085 3
                mRNA
  3022 4
                gene
  3022 4
                mRNA
```

The BED format is a simpler alternative to the GFF format for describing genomic objects, consisting of 4 columns separated by tabs:

```
* Column 1: chromosome
```

- * Column 2: start of the genomic element described
- * Column 3: stop of the genomic element
- * Column 4: identifier or name of the genomic element

We want to extract from the previous GFF file the lines corresponding to genes and convert this information to BED format. This file should be sorted:

- * by chromosome
- * by coordinate on the chromosome.

For this, I will use combinations of grep/cut/sed/sort commands and generate the file fusa_genes.bed.

```
In [26]:
        ! zcat fusa_annot.gff3.gz | fgrep -v '#' | cut -f 1,4,5,9 | fgrep 'ID=gene' | cut -
In [27]: ! head -n5 fusa_genes.bed
               6089
                       11000 FGRAMPH1_01G00001
        1
        1
               11394
                       12168 FGRAMPH1_01G00003
                       19668 FGRAMPH1_01G00005
        1
               19069
               21463 22193 FGRAMPH1 01G00007
        1
               23519
                       25108
                               FGRAMPH1_01G00009
```

Add a prefix 'FusaChrom' to the chromosome names using sed directly in the file fusa_genes.bed.

```
In [28]: ! sed -i 's/^/FusaChrom/' fusa_genes.bed
         ! head -n5 fusa_genes.bed
        FusaChrom1
                         6089
                                 11000
                                         FGRAMPH1_01G00001
        FusaChrom1
                         11394
                                 12168
                                         FGRAMPH1_01G00003
        FusaChrom1
                         19069
                                 19668
                                         FGRAMPH1_01G00005
                                         FGRAMPH1 01G00007
        FusaChrom1
                         21463
                                 22193
                                 25108
        FusaChrom1
                         23519
                                         FGRAMPH1 01G00009
```

Retrieve the gene sizes in a file gene_size.tab (column1: gene name, column2: gene size), using the created BED file, and calculate the gene sizes with awk.

```
In [30]: ! awk 'BEGIN{OFS="\t"} {genesize=$3-$2+1;print $4,genesize}' fusa_genes.bed > gene_
```

```
FGRAMPH1_01G00001 4912
FGRAMPH1_01G00003 775
FGRAMPH1_01G00005 600
FGRAMPH1_01G00007 731
FGRAMPH1_01G00009 1590
```

Calculate the average size of the genes

The average size of the genes : 1686.88

Genes larger than 5kb are considered particularly large. I will count how many are equal to or larger than 5kb, and how many are smaller than 5kb

Found 14542 genes smaller than 5kb and 356 genes larger than 5kb

I wish to extract the coordinates of promoters for these genes (2000 bp upstream of the genes). I'll create a file named "fusa_prom.bed" containing the coordinates of the promoters of these genes, and we'll name these promoters using the gene name followed by the suffix_prom.

```
4089
FusaChrom1
                        6088
                                 FGRAMPH1_01G00001_prom
FusaChrom1
                9394
                        11393
                                 FGRAMPH1_01G00003_prom
FusaChrom1
                17069
                        19068
                                 FGRAMPH1 01G00005 prom
FusaChrom1
                19463
                        21462
                                 FGRAMPH1_01G00007_prom
FusaChrom1
                21519
                        23518
                                 FGRAMPH1_01G00009_prom
```