Molecular Weight Estimation



Bulletin 6210

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Run the standards and samples on an SDS-PAGE gel. Process the gel with the desired stain and then destain to visualize the protein bands. Determine the R_f graphically or using Quantity One[®] analysis software (or equivalent).

- Using a ruler, measure the migration distance from the top of the resolving gel to each standard band and to the dye front.
- For each band in the standards, calculate the R_f value using the following equation:

 R_f = migration distance of the protein/migration distance of the dye front
- Repeat this step for the unknown bands in the samples.
- Use a graphing program, plot the log (MW) as a function of R_f.
- Generate the equation y = mx + b, and solve for y to determine the MW of the unknown protein.

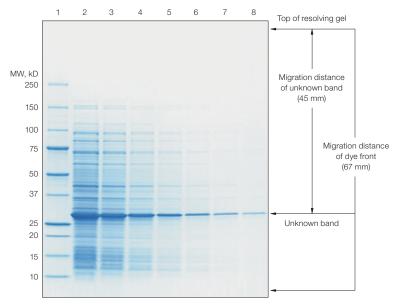


Fig. 1. Example showing MW determination of an unknown protein.

Lane 1, 10 µl of Precision Plus Protein™ unstained standards; lanes 2–8, a dilution series of an *E. coli* lysate containing a hypothetical unknown protein (GFP). Proteins were separated by SDS-PAGE in a Criterion™ 4–20% Tris-HCl gel and stained with Bio-Safe™ Coomassie stain. Gel shown is the actual size.

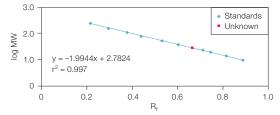


Fig. 2. Determining the MW of an unknown protein by SDS-PAGE. A standard curve of the log (MW) versus $R_{\rm f}$ was generated using the Precision Plus Protein standards from Figure 1. The strong linear relationship (${\rm r}^2 > 0.99$) between the proteins' MW and migration distances demonstrates exceptional reliability in predicting MW.



This is an excerpt from Bio-Rad's comprehensive Electrophoresis Guide (Bulletin 6040).



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