

# Beta Diversity: Inter-Community Difference



Brendan J. Kelly, MD, MS

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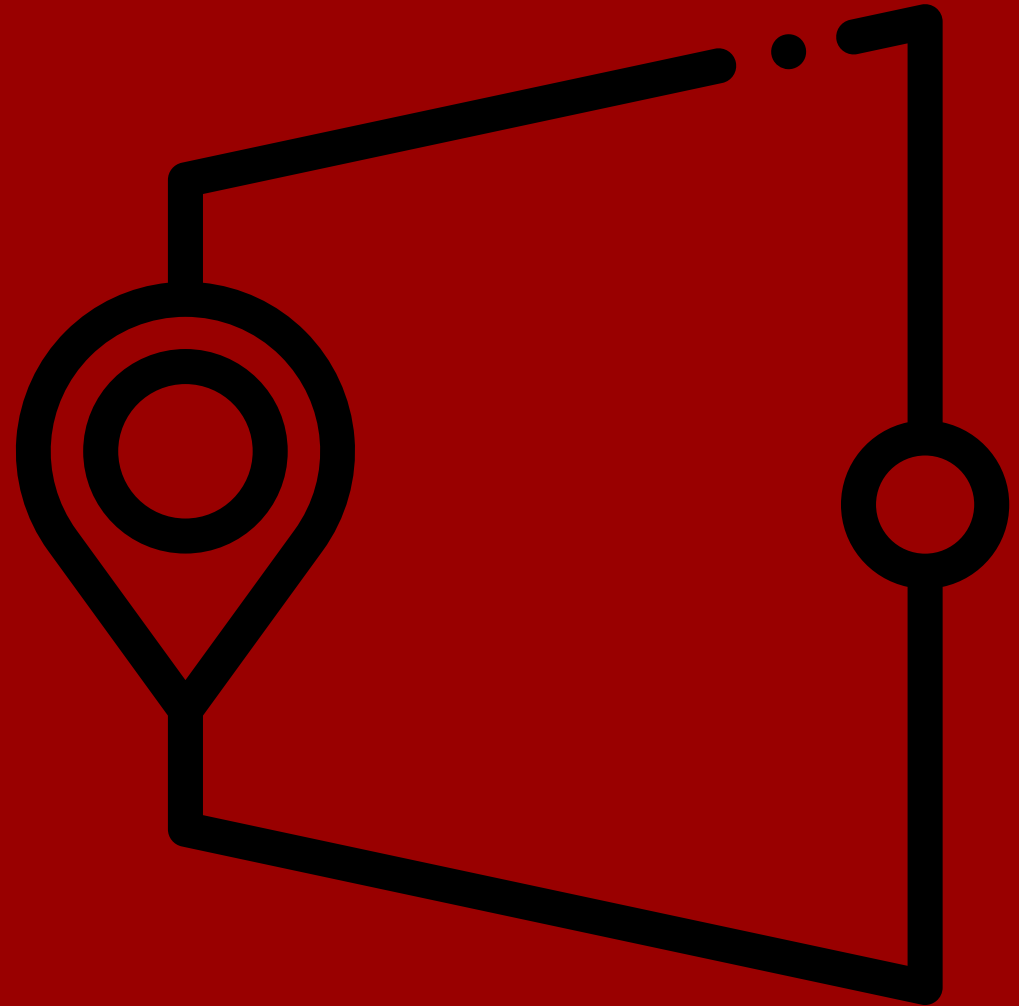
**Beta diversity**

**Pairwise distances**

**Principal coordinates**

**PERMANOVA**

**R's vegan package**



# Beta diversity



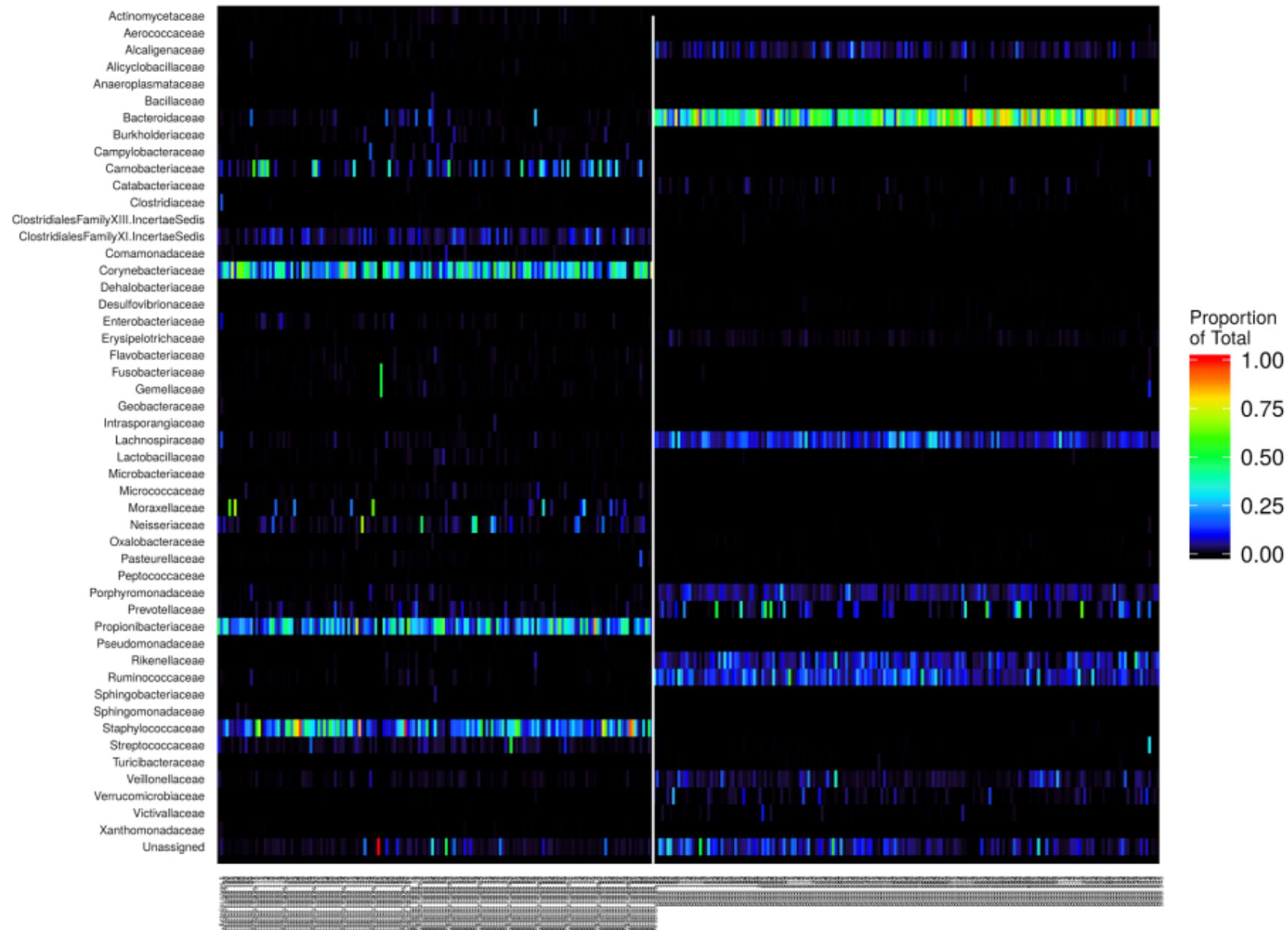
# High Dimensional Microbiome Data

##	700013549	700014386	700014403	700014409	700014412	700014415
## OTU_97.1	0	0	0	0	0	0
## OTU_97.10	0	0	6	4	1	5
## OTU_97.100	0	0	133	7	1	4
## OTU_97.1000	0	0	0	0	0	0
## OTU_97.10000	0	0	0	0	0	0
## OTU_97.10001	0	0	0	0	0	1
## OTU_97.10002	0	0	0	0	0	0
## OTU_97.10003	0	0	0	0	0	0
## OTU_97.10004	0	0	0	0	0	0
## OTU_97.10005	0	0	0	0	0	0
## OTU_97.10006	0	0	0	0	0	0
## OTU_97.10007	0	0	0	0	0	0
## OTU_97.10008	0	1	0	0	0	0
## OTU_97.10009	0	0	1	0	0	0
## OTU_97.1001	0	0	0	0	0	0
## OTU_97.10010	0	0	0	0	0	0

# High Dimensional Microbiome Data

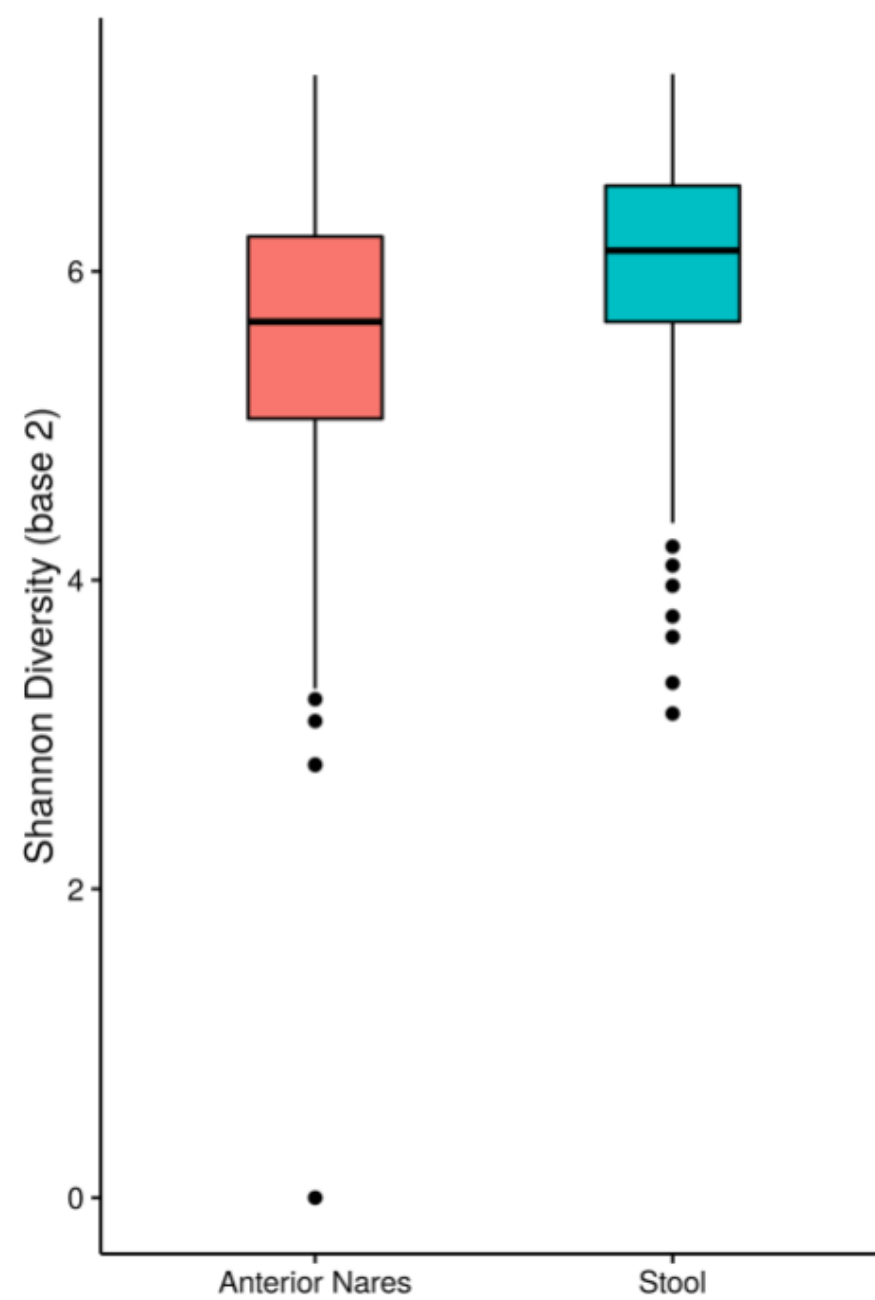
- How to deal with high-dimensional microbiome data?
- **Descriptive (e.g., heatmaps and stacked barplots)**
- Test a priori hypotheses regarding specific OTUs/taxa
- Reduce dimensions:
  - single summary statistic (alpha diversity)
  - pairwise distances (beta diversity) with PCoA or PERMANOVA
  - community types (mixture modeling)

# Anterior Nares vs Stool



# High Dimensional Microbiome Data

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# High Dimensional Microbiome Data

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- Descriptive (e.g., heatmaps and stacked barplots)
- Test a priori hypotheses regarding specific OTUs/taxa
- **Reduce dimensions:**
  - single summary statistic (alpha diversity)
  - **pairwise distances (beta diversity) with PCoA or PERMANOVA**
  - community types (mixture modeling)

# Beta Diversity as Dimension Reduction

- Summarize each sample's relationship to other samples:
  - pairwise distances
  - OTU table → square matrix
- Many beta diversity metrics:
  - just counts versus counts + phylogeny
  - weighted versus unweighted

# Pairwise distances



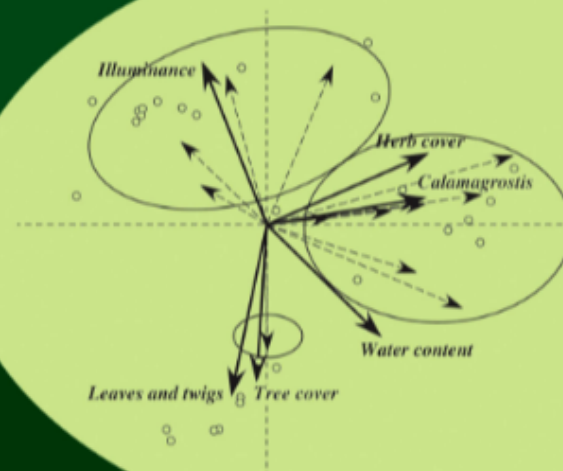


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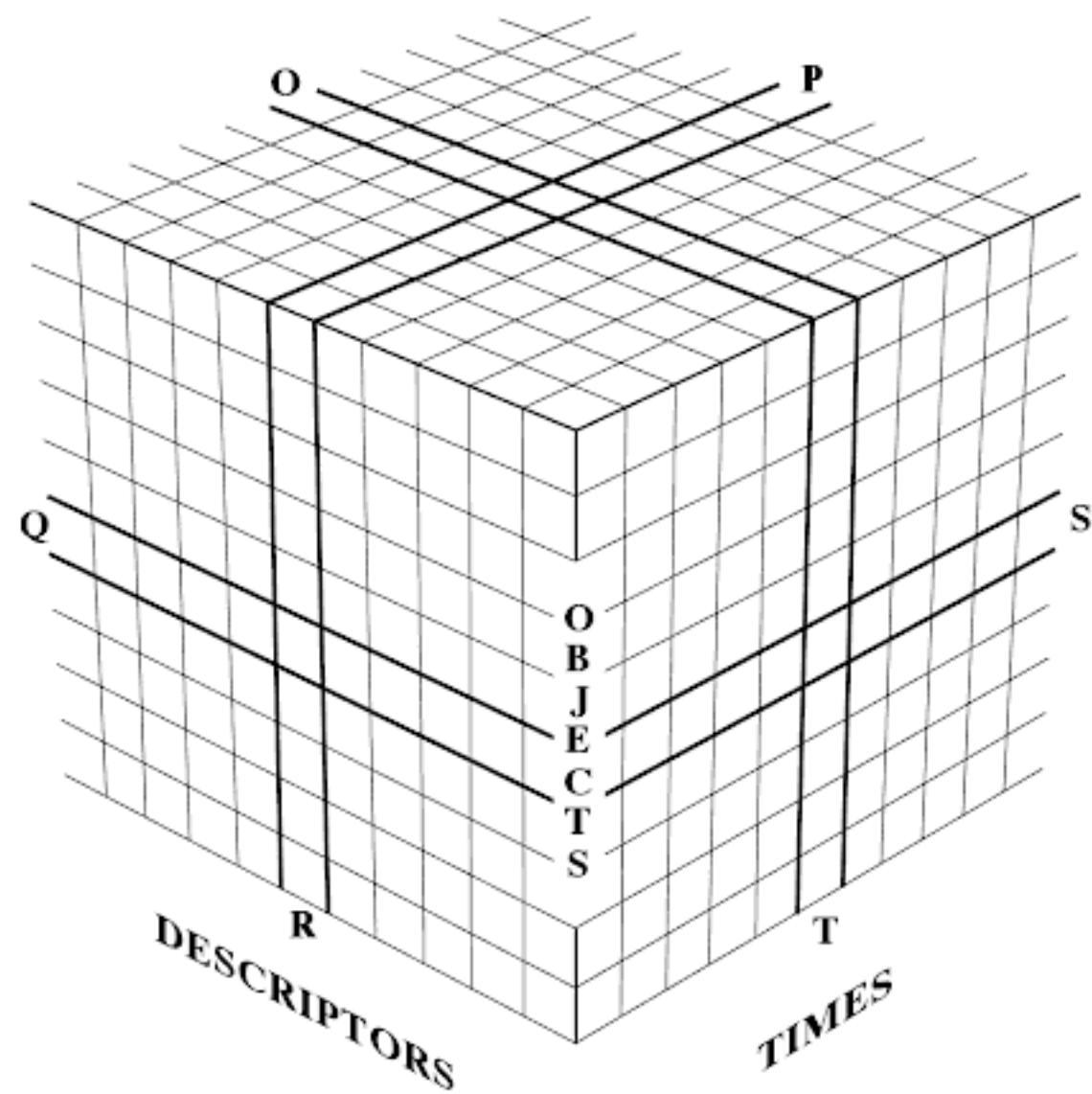
# Numerical Ecology

Pierre Legendre  
Louis Legendre



# What's in a distance?

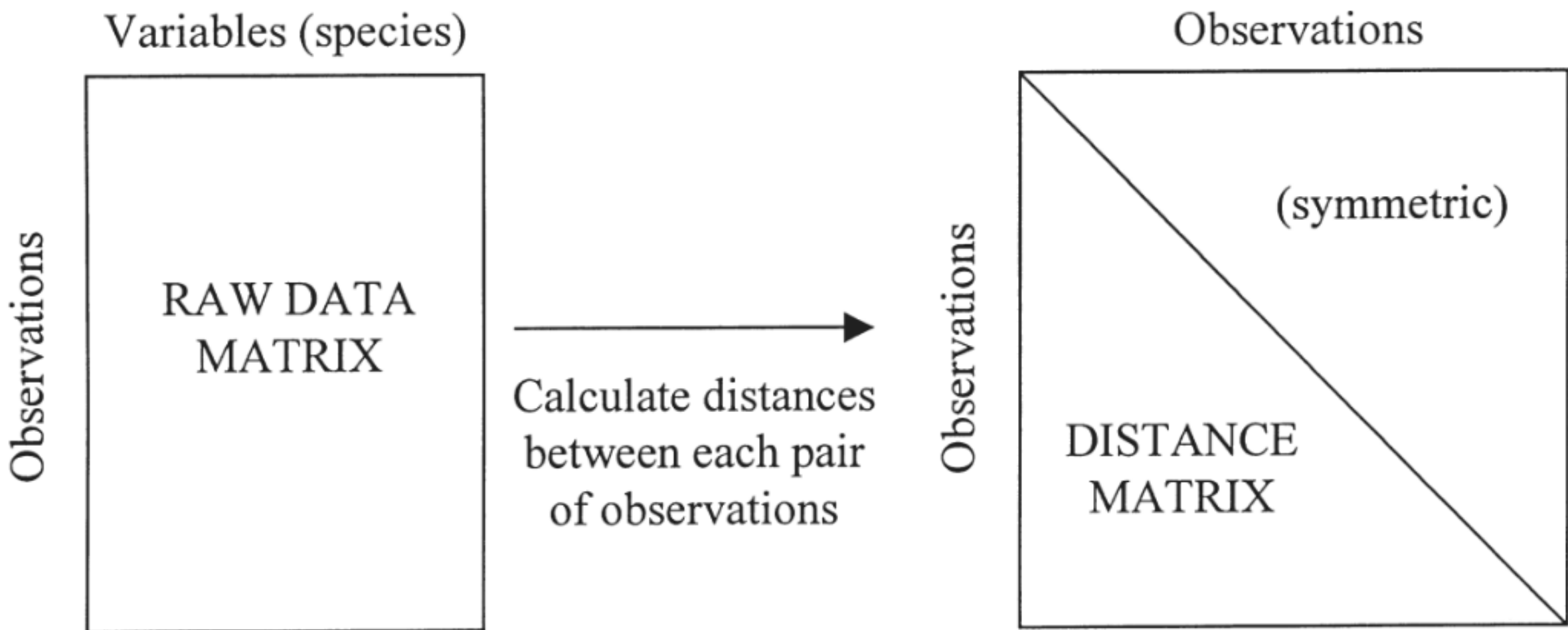
- “The most usual approach to assess the resemblance among objects or descriptors is to first condense all (or the relevant part of) the information available in the ecological data matrix (Section 2.1) into **a square matrix of association** among the objects or descriptors (Section 2.2). In most instances, the association matrix is symmetric.”
- Compare variable-variable: “R-mode” (like Pearson’s  $r$  coefficient)
- Compare object-object: “Q-mode”
- Six modes of analysis if incorporate time series (Cattell 1966)



**Figure 7.1** The three-dimensional data box (objects  $\times$  descriptors  $\times$  times). Adapted from Cattell (1966).

# What's in a distance?

- "... association will be used as a general term to describe any measure or coefficient used to quantify the **resemblance or difference** between objects or descriptors, as proposed by Orlóci (1975)."
- Q-mode studies:
  - similarity coefficients (identical = 1)
  - distance (or dissimilarity) coefficients (identical = 0)



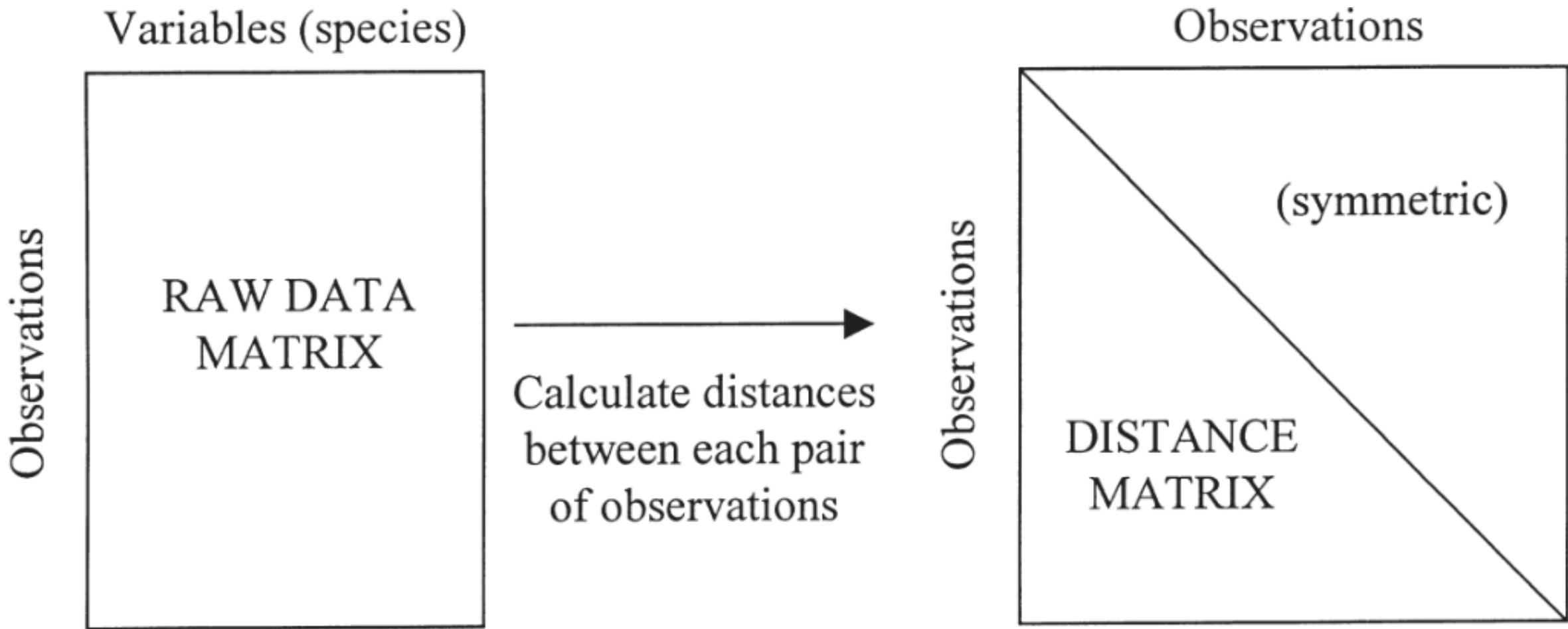


# OTU Table: OTUs x Specimens

##	700013549	700014386	700014403	700014409	700014412	700014415
## OTU_97.1	0	0	0	0	0	0
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## OTU_97.10003	0	0	0	0	0	0
## OTU_97.10004	0	0	0	0	0	0
## OTU_97.10005	0	0	0	0	0	0
## OTU_97.10006	0	0	0	0	0	0
## OTU_97.10007	0	0	0	0	0	0
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# OTU Table: Specimens x OTUs

##	OTU_97.1	OTU_97.10	OTU_97.100	OTU_97.1000	OTU_97.10000	OTU_97.10001
## 700013549	0	0	0	0	0	0
## 700014386	0	0	0	0	0	0
## 700014403	0	6	133	0	0	0
## 700014409	0	4	7	0	0	0
## 700014412	0	1	1	0	0	0
## 700014415	0	5	4	0	0	1
## 700014418	0	2	0	0	0	0
## 700014421	0	3	25	0	0	0
## 700014424	0	1	5	0	0	0
## 700014427	0	1	0	0	0	0
## 700014430	0	6	0	0	0	0
## 700014445	0	0	0	0	0	0
## 700014501	0	2	1	0	0	0
## 700014515	0	0	0	0	0	0
## 700014516	0	0	0	0	0	0
## 700014517	0	0	0	0	0	0



# Distance Metrics for Beta Diversity

- Just counts versus counts + phylogeny:
  - Jaccard:  $J(A, B) = \frac{A \cap B}{A \cup B}$  &  $d_J(A, B) = 1 - J(A, B)$
  - UniFrac: fraction of unique branch length in tree
- Weighted versus unweighted:
  - weighted: counts matter
  - unweighted: binary (presence-absence)

# The "Double Zero" Problem

- “The proportion of zeros in community composition data generally increases with the variability in environmental conditions among the sampling sites. If sampling has been conducted along one or several environmental axes, the species present are likely to differ at least partly from site to site. **Including double zeros in the comparison between sites would result in high values of similarity for the many pairs of sites holding only a few species**, these pairs presenting many double zeros; this would not provide a correct ecological assessment of the situation.”

# The "Double Zero" Problem

- “Because double zeros are not informative, their interpretation generates **the double zero problem: is the value of an association coefficient affected by inclusion of double zeros in its calculation?** When choosing an association coefficient, ecologists must pay attention to the interpretation of double zeros: except in very limited cases (e.g. controlled experiments involving very few species and with small uncontrolled ecological variation), it is preferable to draw no ecological conclusion from the simultaneous absence of a species at two sites.... In numerical terms, this means to **skip double zeros when computing similarity or distance coefficients** using species presence-absence or abundance data.”

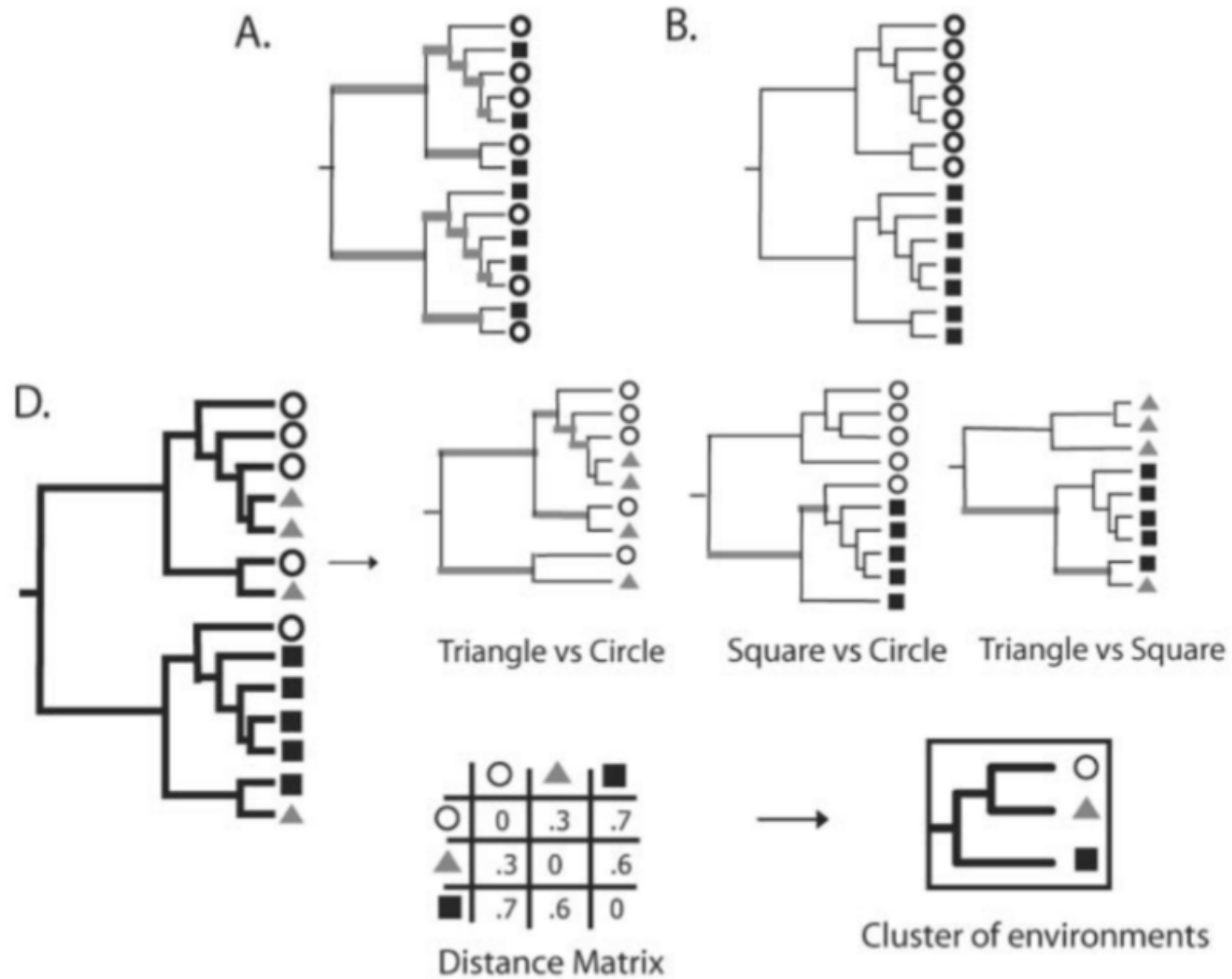
# UniFrac

- UniFrac measures the distance between communities based on the lineages they contain.
- Satisfies the technical requirements of a distance metric:
  - always positive
  - transitive
  - satisfies the triangle inequality
- Can thus be used with standard multivariate statistics (e.g., UPGMA, clustering, and **PCoA**).

# UniFrac

- UniFrac “exploits the different degrees of similarity between sequences”:
  - “the unique fraction metric, or UniFrac, measures the phylogenetic distance between sets of taxa in a phylogenetic tree as the fraction of the branch length of the tree that leads to descendants from either one environment or the other, but not both”
  - “captures the total amount of evolution that is unique to each state, presumably reflecting adaptation to one environment that would be deleterious in the other” (designed to be based on rRNA)





**Table 7.2** Some properties of distance coefficients calculated from the similarity coefficients presented in Section 7.3. These properties (from Gower & Legendre, 1986), which will be used in Section 9.3, strictly apply when there are no missing data.

Similarity coefficient	$D = 1 - S$ metric, etc.	$D = 1 - S$ Euclidean	$D = \sqrt{1 - S}$ metric	$D = \sqrt{1 - S}$ Euclidean
$S_1 = \frac{a + d}{a + b + c + d}$ (simple matching; eq. 7.1)	metric	No	Yes	Yes
$S_2 = \frac{a + d}{a + 2b + 2c + d}$ (Rogers & Tanimoto; eq. 7.2)	metric	No	Yes	Yes
$S_3 = \frac{2a + 2d}{2a + b + c + 2d}$ (eq. 7.3)	semimetric	No	Yes	No
$S_4 = \frac{a + d}{b + c}$ (eq. 7.4)	nonmetric	No	No	No
$S_5 = \frac{1}{4} \left[ \frac{a}{a+b} + \frac{a}{a+c} + \frac{d}{b+d} + \frac{d}{c+d} \right]$ (eq. 7.5)	semimetric	No	No	No
$S_6 = \frac{a}{\sqrt{(a+b)(a+c)}} \frac{d}{\sqrt{(b+d)(c+d)}}$ (eq. 7.6)	semimetric	No	Yes	Yes
$S_7 = \frac{a}{a+b+c}$ (Jaccard; eq. 7.10)	metric	No	Yes	Yes
$S_8 = \frac{2a}{2a+b+c}$ (Sørensen; eq. 7.11)	semimetric	No	Yes	Yes
$S_9 = \frac{3a}{3a+b+c}$ (eq. 7.12)	semimetric	No	No	No
$S_{10} = \frac{a}{a+2b+2c}$ (eq. 7.13)	metric	No	Yes	Yes
$S_{11} = \frac{a}{a+b+c+d}$ (Russell & Rao; eq. 7.14)	metric	No	Yes	Yes
$S_{12} = \frac{a}{b+c}$ (Kulczynski; eq. 7.15)	nonmetric	No	No	No

# Beta Diversity: Which Distance Metric?

- Why use Jaccard? UniFrac?
- Why use weighted? Unweighted?

# Principal Coordinates



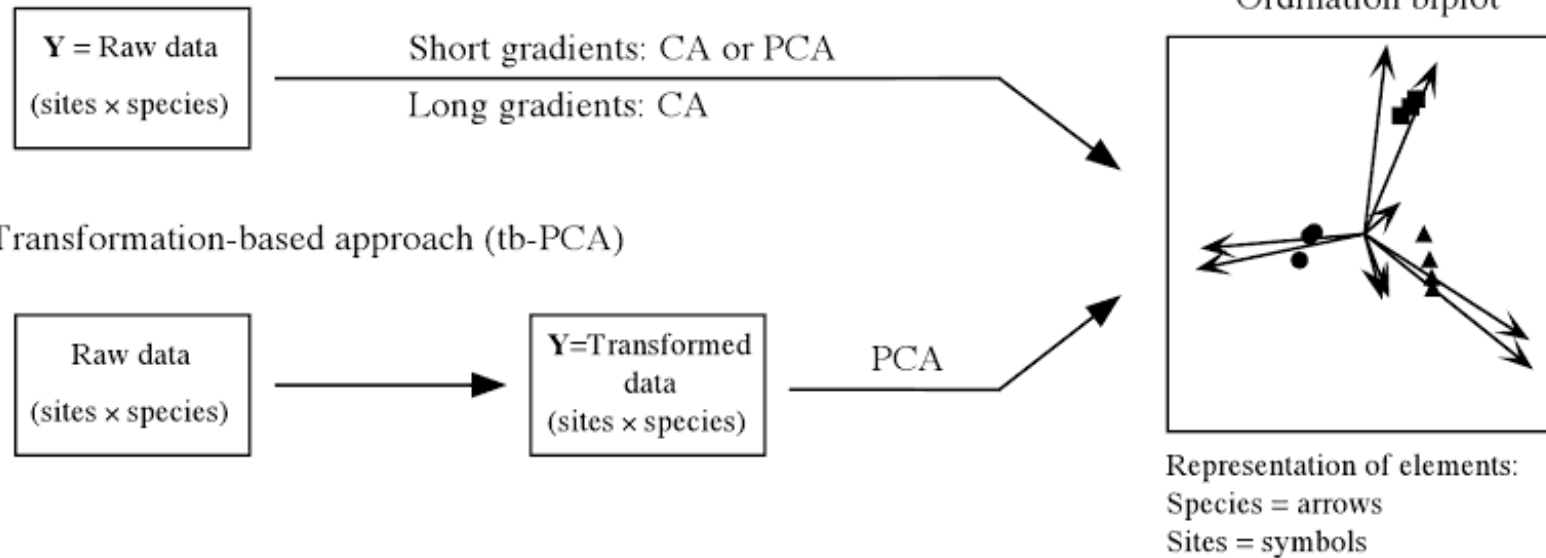
# Original Descriptors → PCA

- PCA: principal component analysis
  - rigid rotation for successive directions of maximum variance
  - lots of restrictions (Euclidean)
  - but allows projection of original descriptors in PCA space

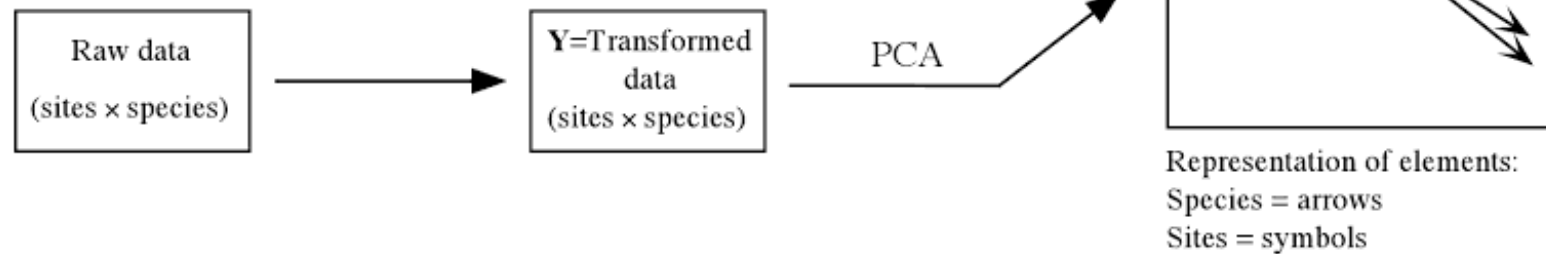
# Pairwise Distances → PCoA

- PCoA: principal coordinate analysis
  - any metric distance, even if non-Euclidean
  - like PCA, eigenvalue decomposition (maximum variance) but mediated by distance function (no original descriptors)
  - unlike PCA, does not allow projection of original descriptors in reduced-dimension space

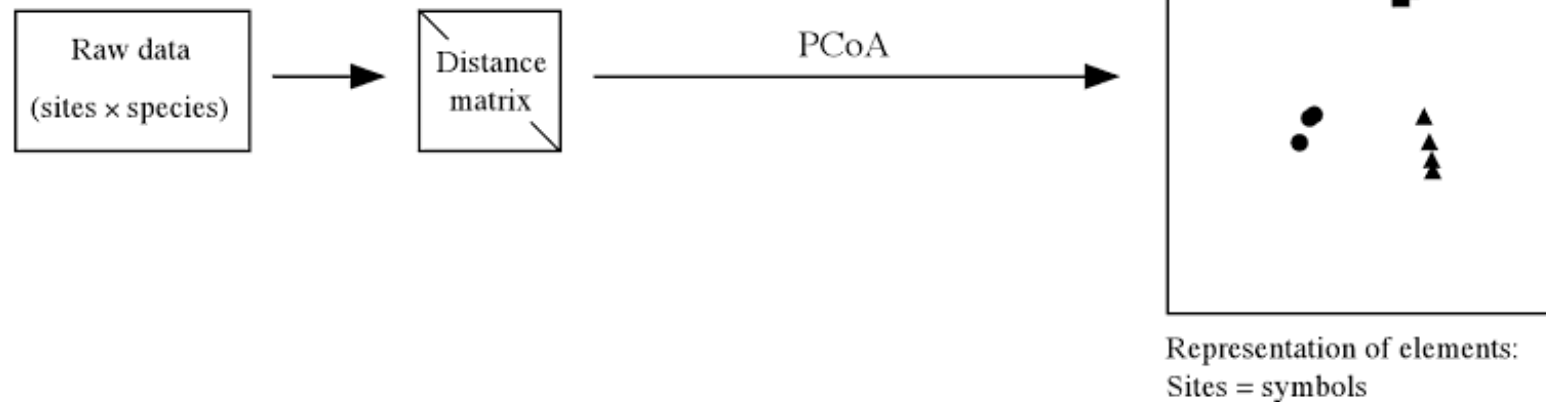
(a) Classical approach



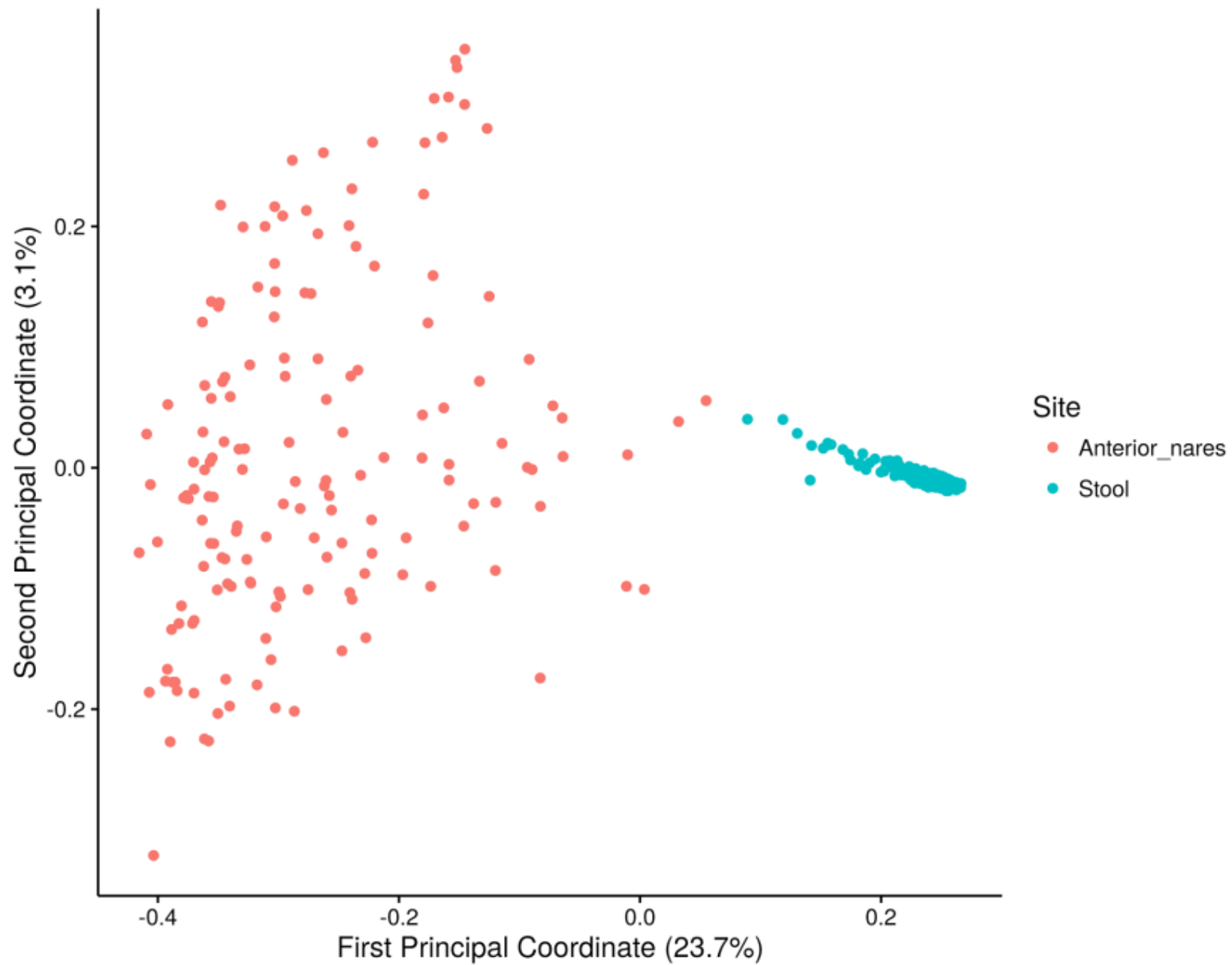
(b) Transformation-based approach (tb-PCA)



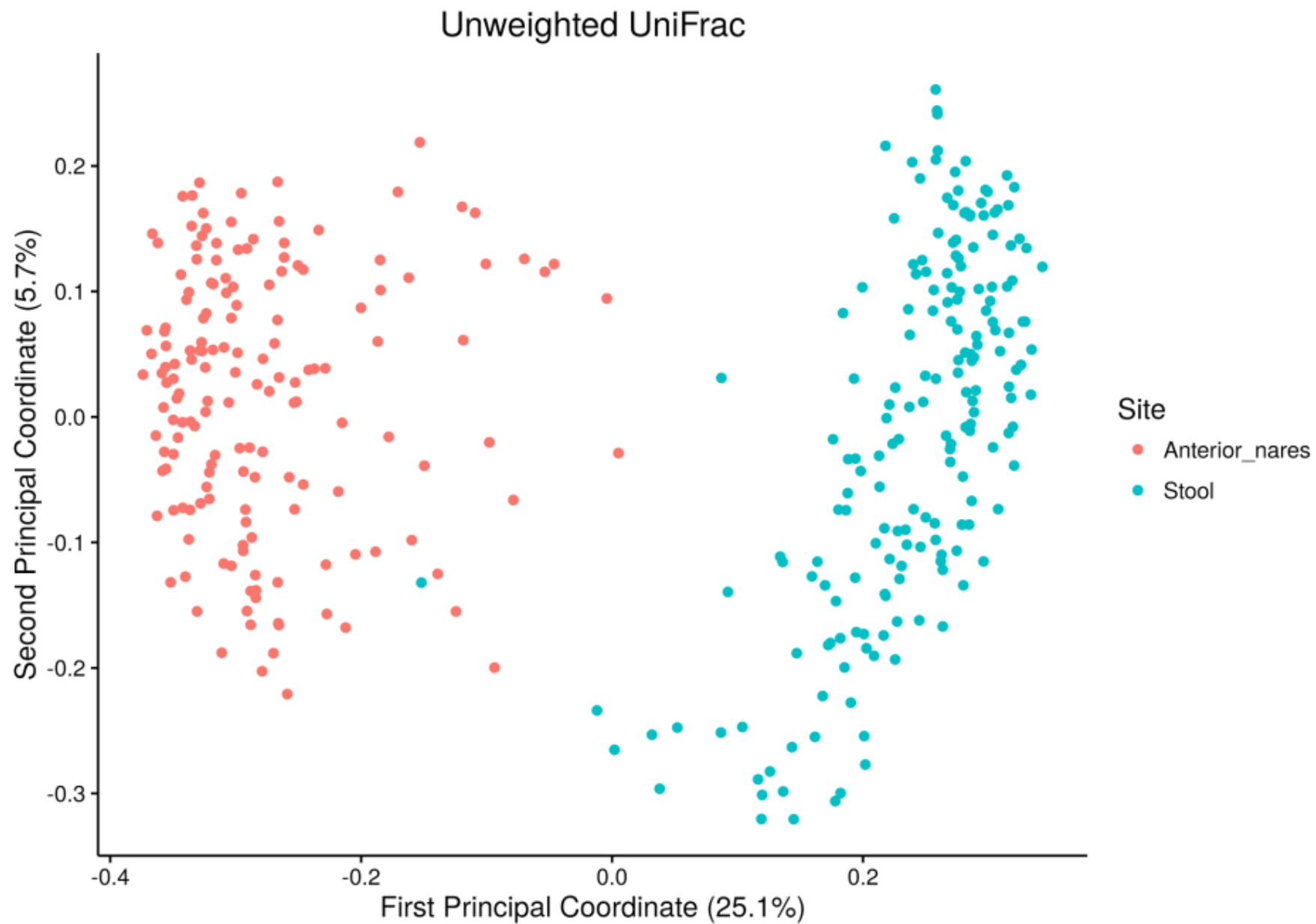
(c) Distance-based approach (PCoA)



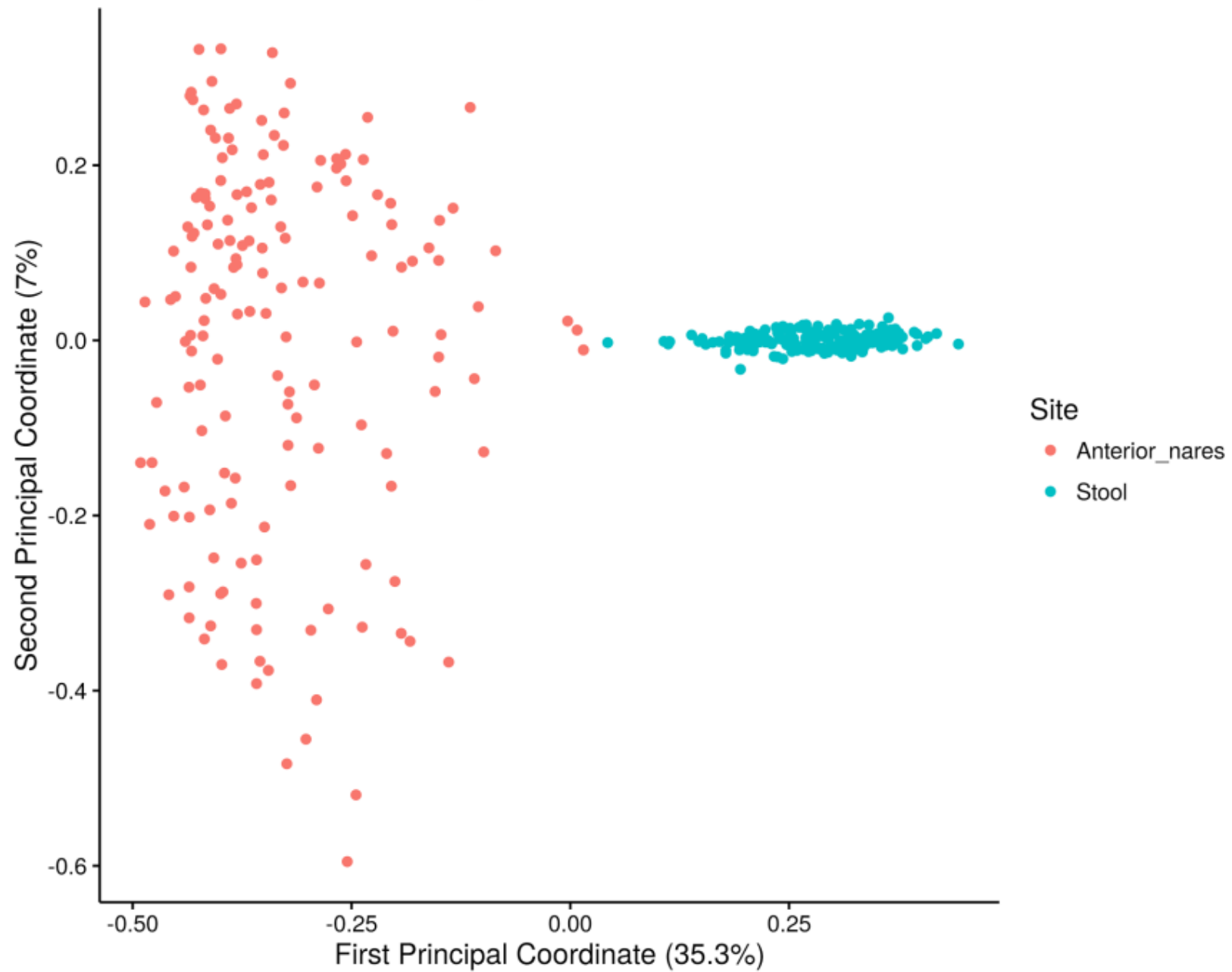
# Weighted UniFrac



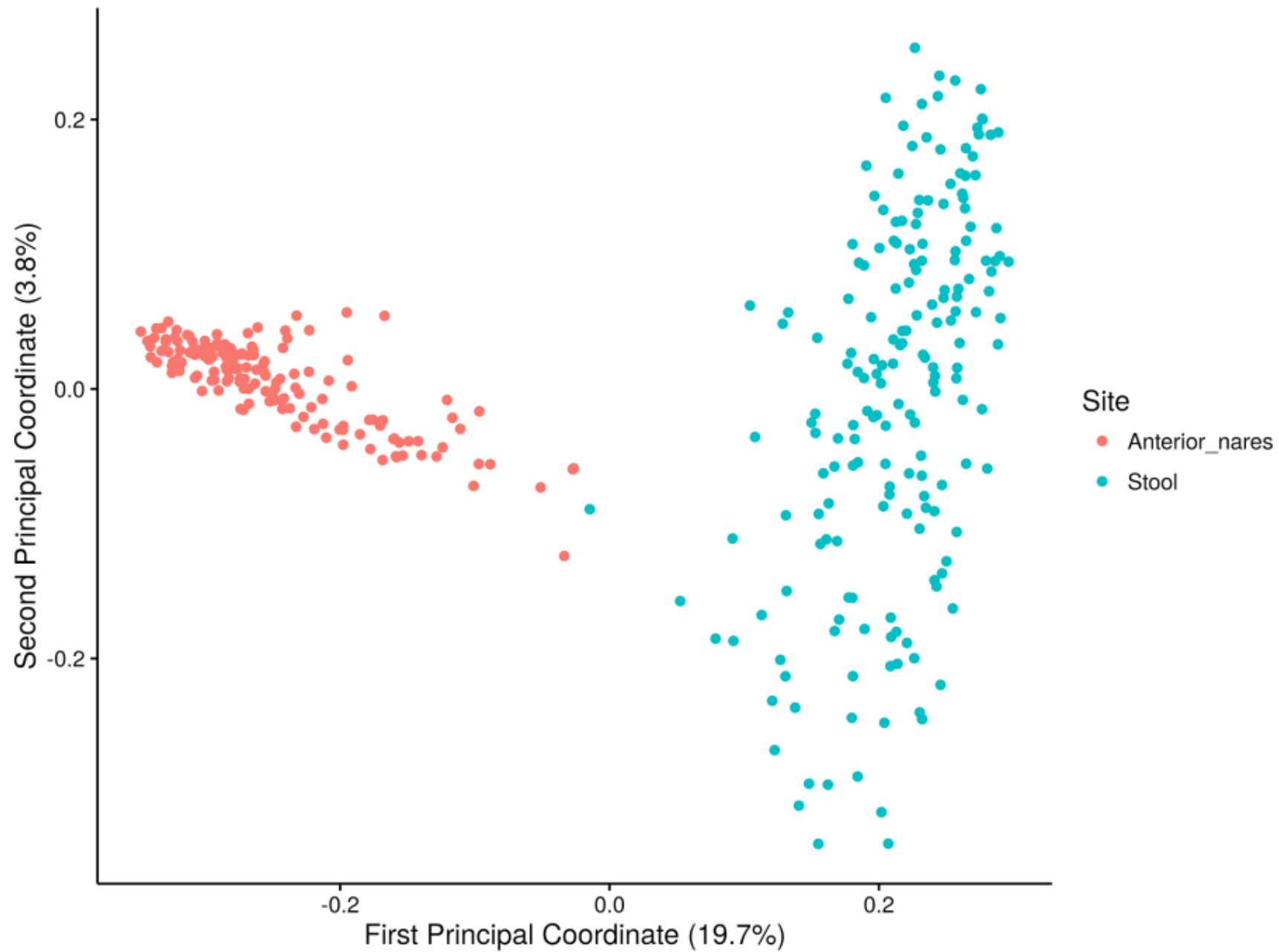




# Weighted Jaccard



# Unweighted Jaccard

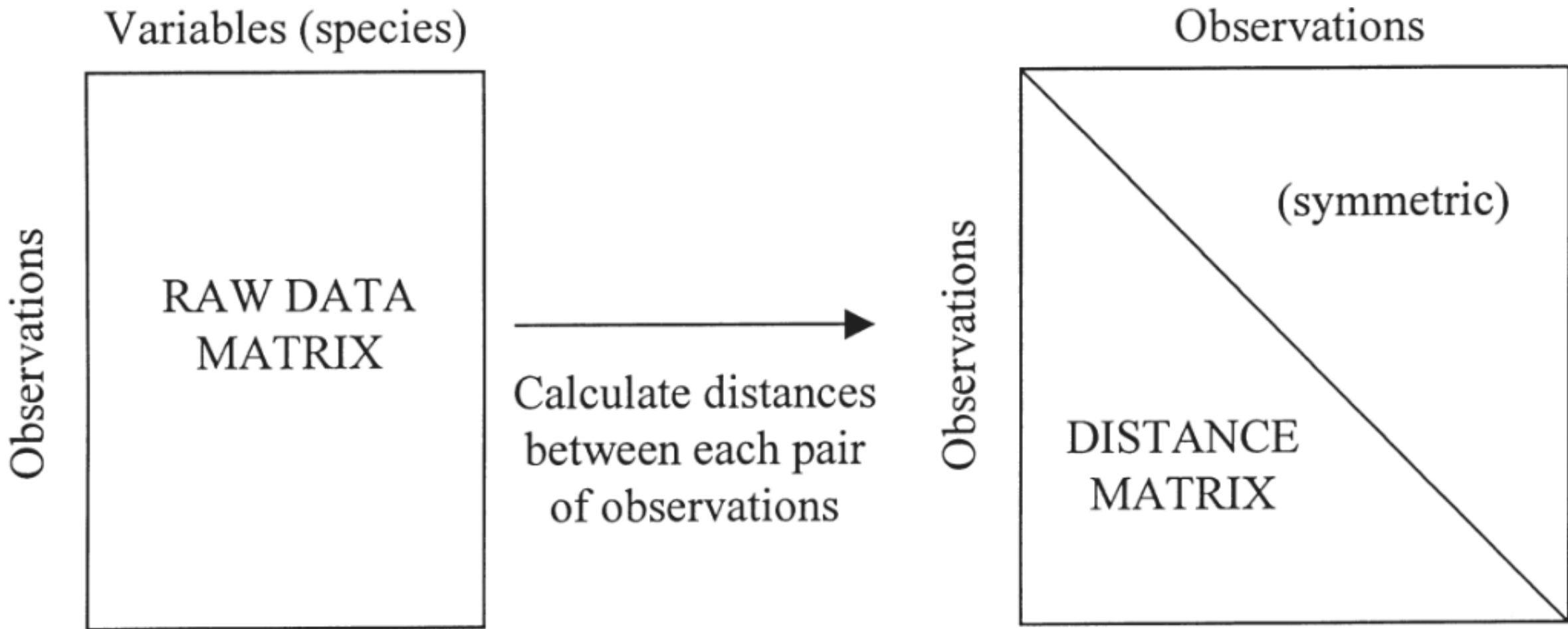


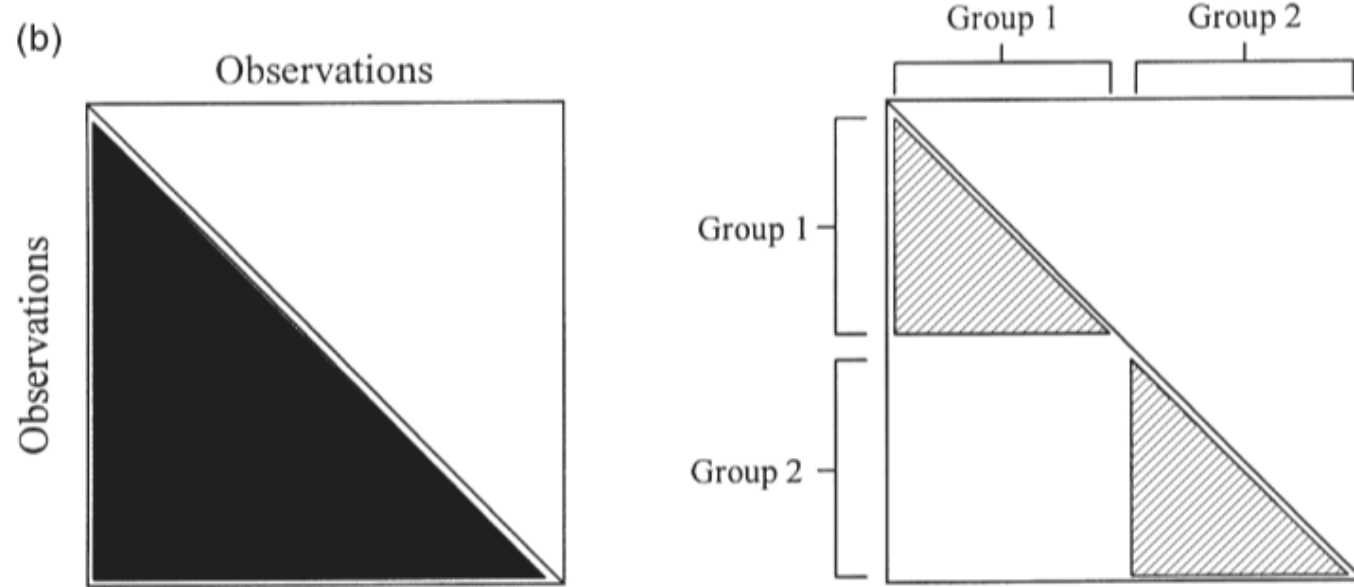


# PERMANOVA

# Pairwise Distances → PERMANOVA

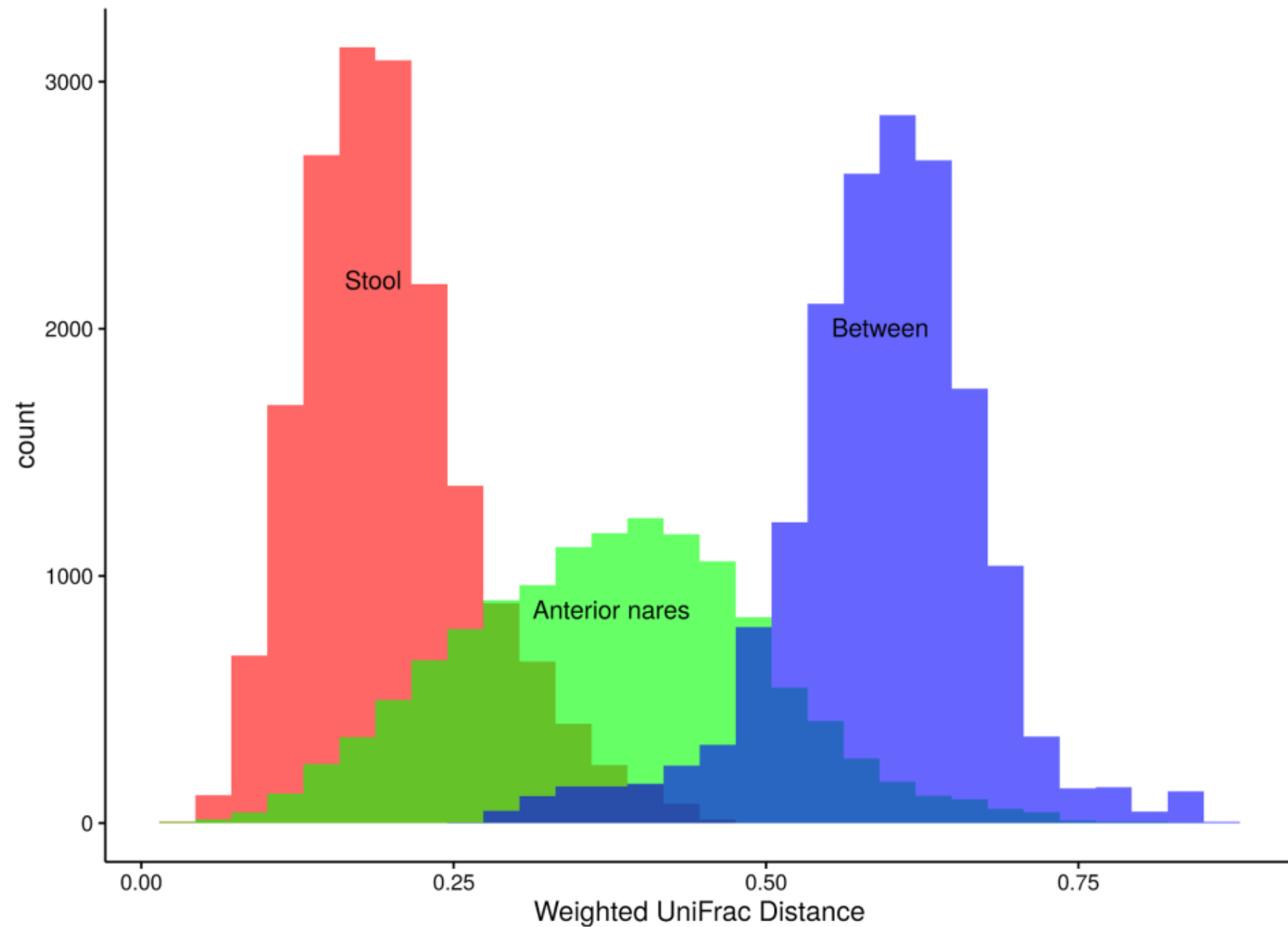
- Pairwise distance matrix can be partitioned by group assignment and ANOVA-like analysis can be applied to detect difference between groups
- **PERMANOVA**: permutational ANOVA (aka, adonis)
  - pseudo F-ratio: conceptually similar but not F-distributed
  - testing by label permutation
  - quantification of effect size by R-squared or omega-squared
  - (the latter a less biased estimator of true effect)





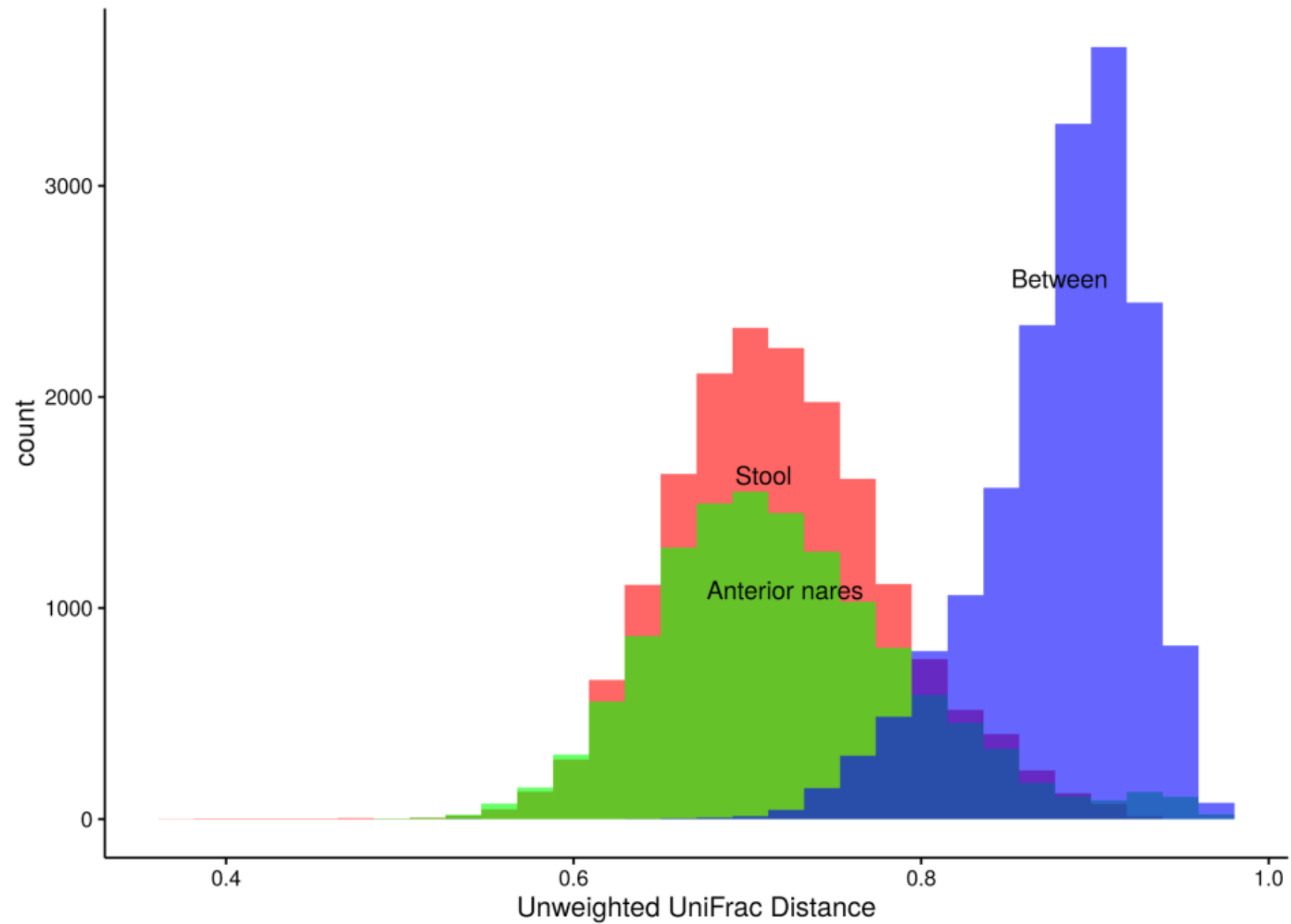
$$F = \frac{SS_A / (a - 1)}{SS_W / (N - a)}$$

# HMP V1-V3 16S rRNA Amplicon

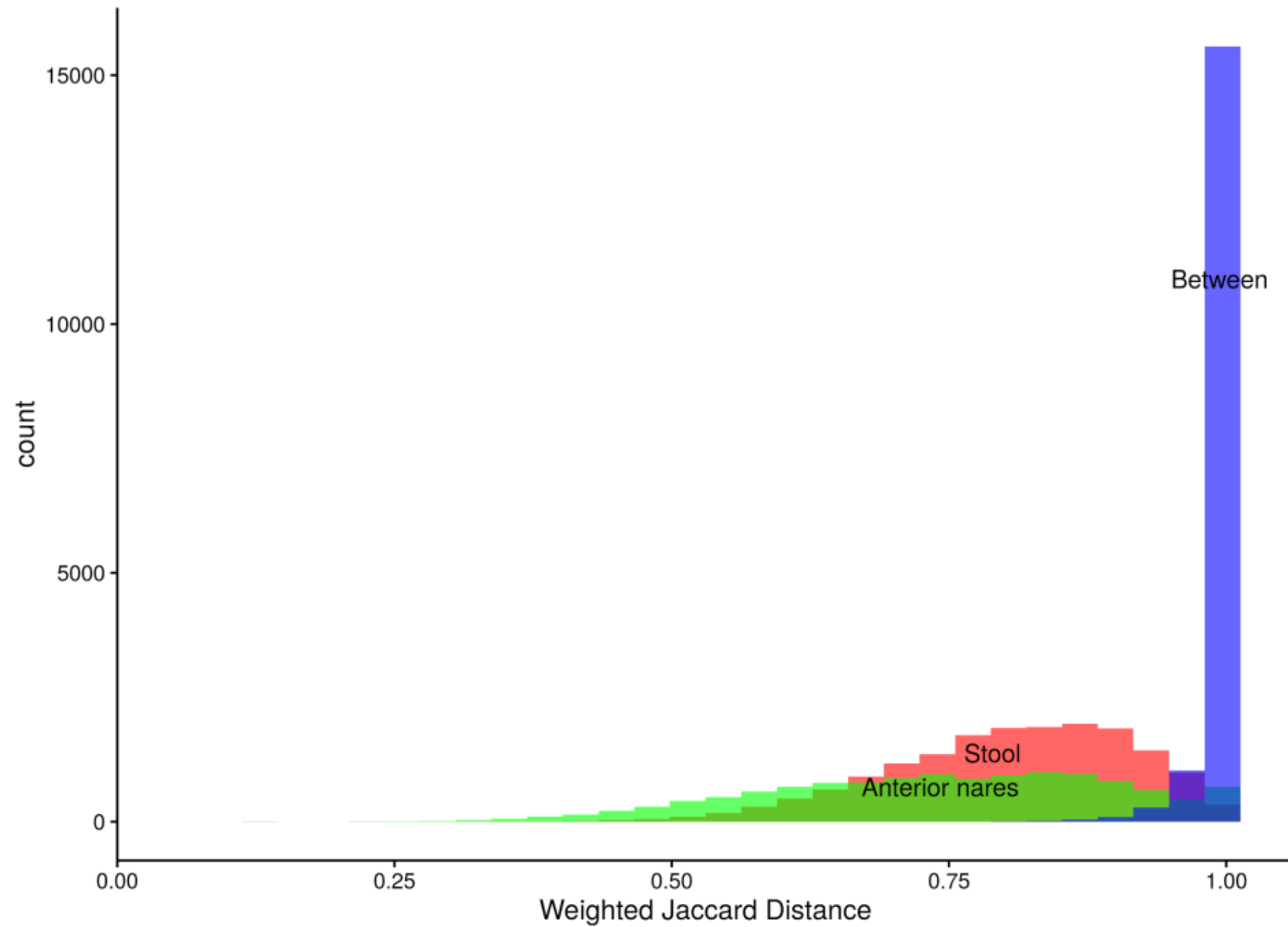




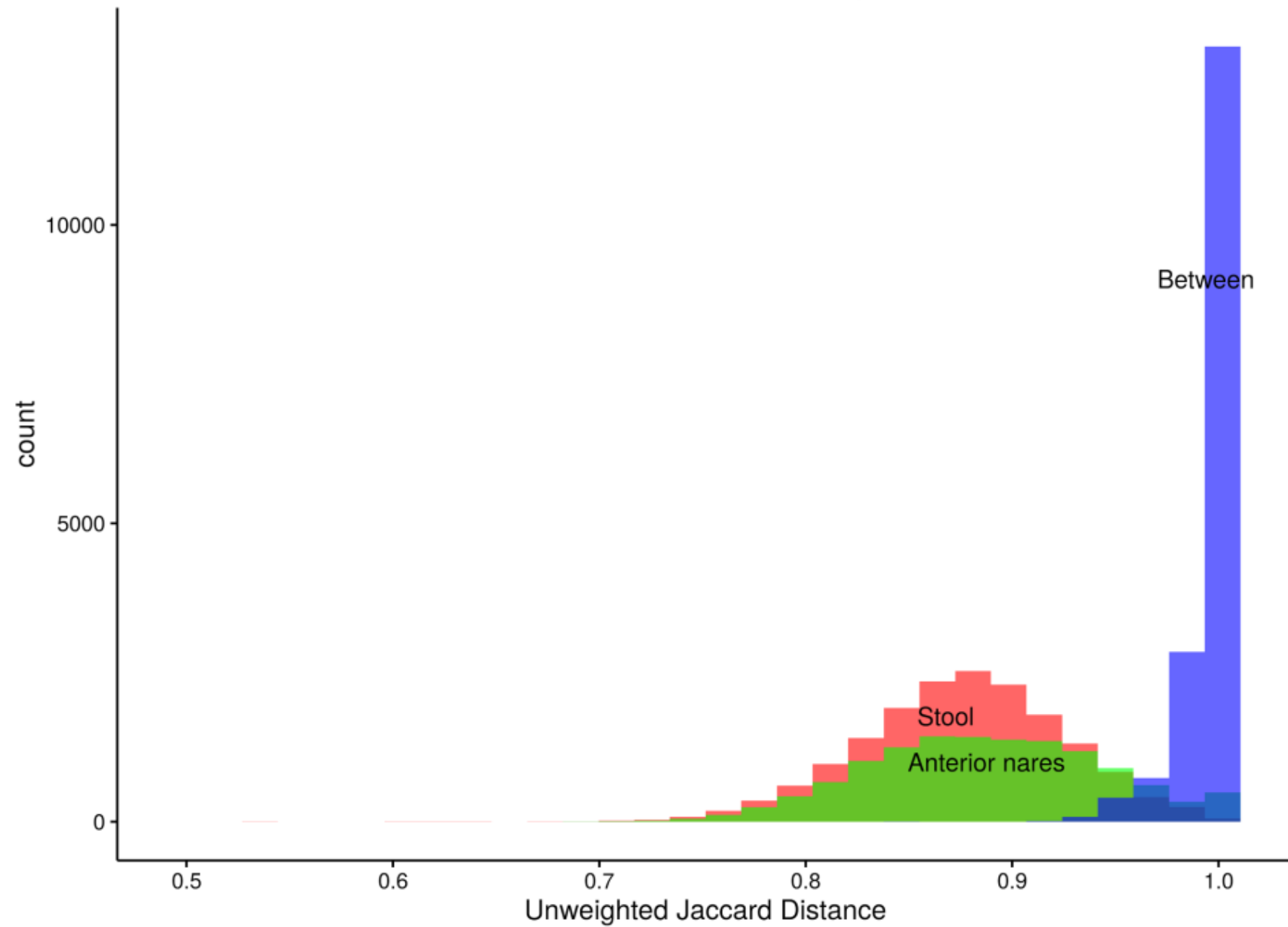
# HMP V1-V3 16S rRNA Amplicon



# HMP V1-V3 16S rRNA Amplicon



# HMP V1-V3 16S rRNA Amplicon



$$R^2 = 1 - \frac{SS_W}{SS_W + SS_A} = \frac{SS_A}{SS_T}$$

$$\omega^2 = \frac{SS_A - (a-1) \frac{SS_W}{N-a}}{SS_T + \frac{SS_W}{N-a}}$$

**Table 1.** Effect sizes observed from various exposures/interventions in studies of various microbiome sampling sites are shown as measured by omega-squared ( $\omega^2$ ) statistics, together with the *P*-values from PERMANOVA test

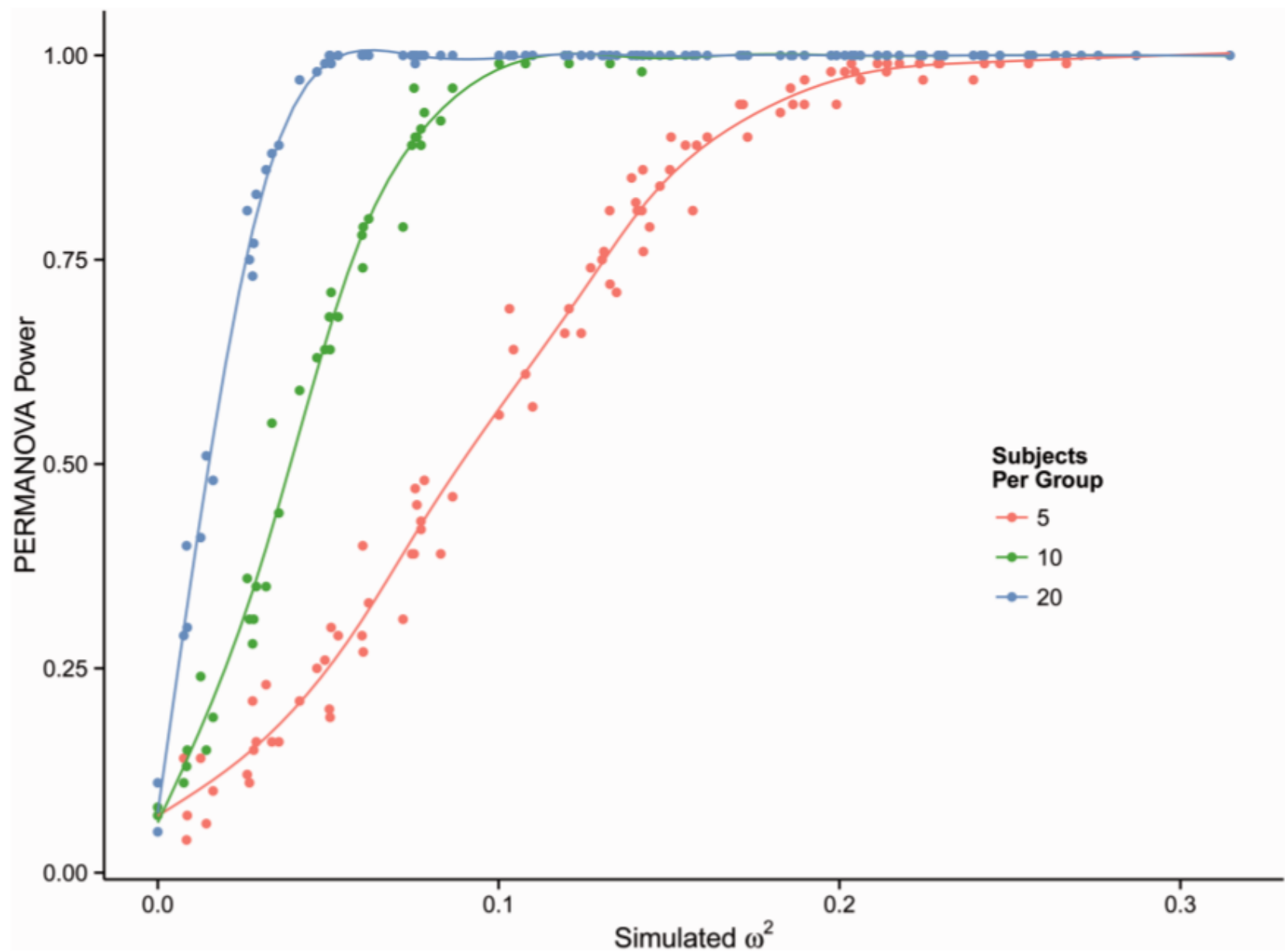
Site	Comparison groups		$\omega^2/P$ -value				Reference
	Control	Exposure	Weighted UniFrac	Unweighted UniFrac	Weighted Jaccard	Unweighted Jaccard	
Nares	Non-smoker (33)	Smoker (29)	0.042/0.001	0.009/0.001	0.023/0.001	0.007/0.001	<a href="#">Charlson <i>et al.</i> (2010)</a>
Oral	Non-smoker (33)	Smoker (29)	0.032/0.001	0.008/0.001	0.024/0.001	0.007/0.001	<a href="#">Charlson <i>et al.</i> (2010)</a>
Gut	Before feeding (10)	After feeding (10)	0.056/0.138	0.013/0.986	0/0.989	0.014/0.985	<a href="#">Wu <i>et al.</i> (2011)</a>
Oral	No azithromycin (42)	Azithromycin (6)	0.063/0.01	0.039/0.001	0.099/0.004	0.032/0.001	<a href="#">Charlson <i>et al.</i> (2012)</a>
Lung	No azithromycin (34)	Azithromycin (6)	0.065/0.005	0.038/0.001	0.019/0.089	0.033/0.001	<a href="#">Charlson <i>et al.</i> (2012)</a>
Skin	Left retroauricular (186)	Right retroauricular (187)	0.000/0.828	0.0001/0.327	0.000/0.986	0.000/1.000	<a href="#">HMP Consortium (2012b)</a>
Human	Anterior nares (161)	Stool (187)	0.567/0.001	0.201/0.001	0.230/0.001	0.117/0.001	<a href="#">HMP Consortium (2012b)</a>

$$R^2 = 1 - \frac{SS_W}{SS_W + SS_A} = \frac{SS_A}{SS_T}$$

$$\omega^2 = \frac{SS_A - (a-1) \frac{SS_W}{N-a}}{SS_T + \frac{SS_W}{N-a}}$$

**Table 1.** Effect sizes observed from various exposures/interventions in studies of various microbiome sampling sites are shown as measured by omega-squared ( $\omega^2$ ) statistics, together with the *P*-values from PERMANOVA test

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Human	Anterior nares (161)	Stool (187)	0.567/0.001	0.201/0.001	0.230/0.001	0.117/0.001	HMP Consortium (2012b)



# R's vegan package



# vegan::vegdist()

```
# install.packages("tidyverse")
library(tidyverse)

# install.packages("vegan")
library(vegan)

otu_long <- read_csv(
  "./data/HMP_OTU_table_longformat_stool_nares.csv"
)

otu_long
```

```
## # A tibble: 431,400 x 4
##   otu_id      specimen_id read_count HMPbodysubsite
##   <chr>          <dbl>      <dbl> <chr>
## 1 OTU_97.1      700014718          0 Stool
## 2 OTU_97.10     700014718          0 Stool
## 3 OTU_97.100    700014718          0 Stool
## 4 OTU_97.1000   700014718          0 Stool
## 5 OTU_97.10000  700014718          0 Stool
## 6 OTU_97.10001  700014718          0 Stool
## 7 OTU_97.10002  700014718          0 Stool
## 8 OTU_97.10003  700014718          0 Stool
## 9 OTU_97.10004  700014718          0 Stool
## 10 OTU_97.10005  700014718          0 Stool
## # ... with 431,390 more rows
```



# vegan::vegdist()

```
otu_matrix <- read_rds(
  "./data/HMP_OTU_table_matrix_stool_nares.rds"
)

otu_matrix %>%
  str(vec.len = 2)
```

```
## num [1:43140, 1:10] 0 0 0 0 0 ...
## - attr(*, "dimnames")=List of 2
## ..$ : chr [1:43140] "OTU_97.1" "OTU_97.10" ...
## ..$ : chr [1:10] "700014718" "700014767" ...
```

# vegan::vegdist()

```
otu_matrix <- read_rds(
  "./data/HMP_OTU_table_matrix_stool_nares.rds"
)

otu_matrix %>%
  t() %>% # TRANSPOSE
  str(vec.len = 2)
```

```
## num [1:10, 1:43140] 0 0 0 0 0 ...
## - attr(*, "dimnames")=List of 2
## ..$ : chr [1:10] "700014718" "700014767" ...
## ..$ : chr [1:43140] "OTU_97.1" "OTU_97.10" ...
```

# vegan::vegdist()

```
otu_matrix <- read_rds(
  "./data/HMP_OTU_table_matrix_stool_nares.rds"
)
otu_matrix %>%
  t() %>% #TRANSPOSE
  vegdist(x = .,
    method = "jaccard",
    binary = TRUE) %>%
  str(vec.len=2)
```

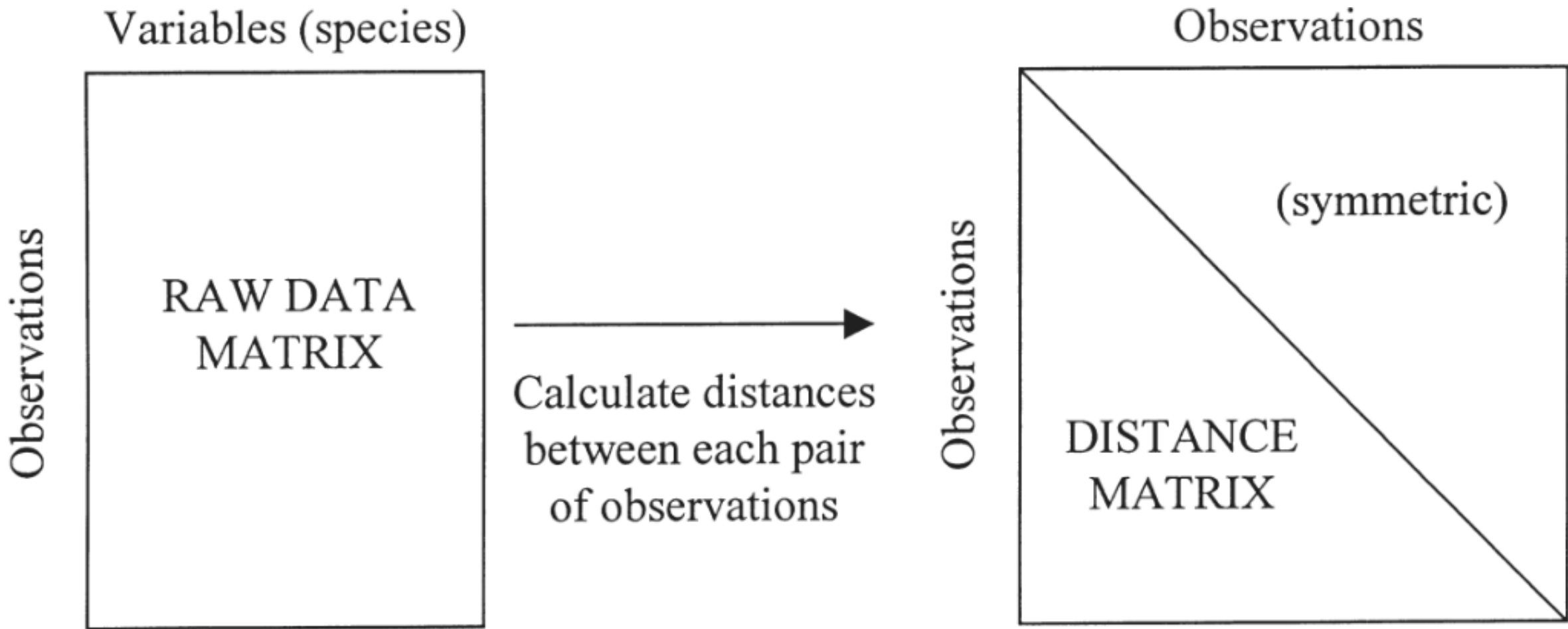
```
## 'dist' num [1:45] 1 0.982 ...
## - attr(*, "Size")= int 10
## - attr(*, "Labels")= chr [1:10] "700014718" "700014767"
## - attr(*, "Diag")= logi FALSE
## - attr(*, "Upper")= logi FALSE
## - attr(*, "method")= chr "binary jaccard"
## - attr(*, "call")= language vegdist(x = ., method = "j
```

# vegan::vegdist()

```
otu_matrix <- read_rds(
  "./data/HMP_OTU_table_matrix_stool_nares.rds"
)

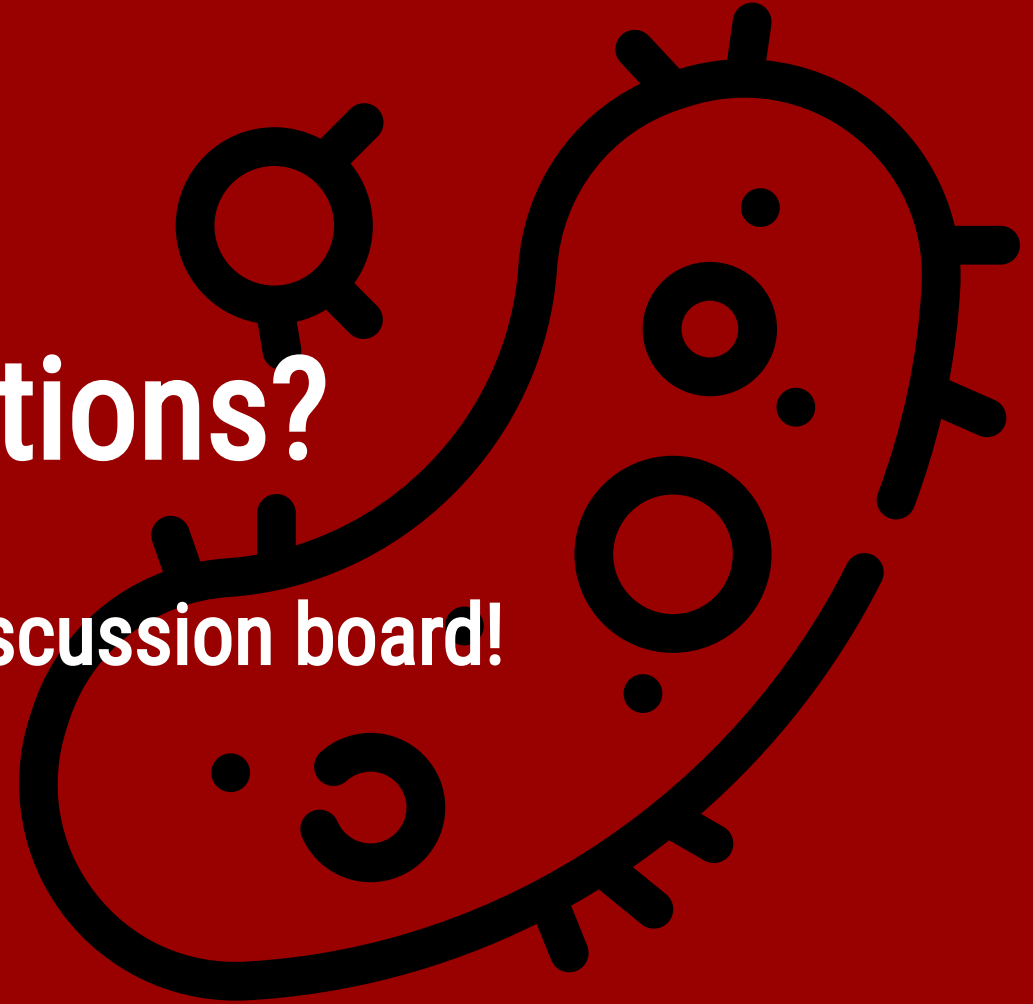
otu_matrix %>%
  t() %>% #TRANSPOSE
  vegdist(x = .,
    method = "jaccard",
    binary = TRUE) %>%
  as.matrix() %>%
  str(vec.len=2)
```

```
## num [1:10, 1:10] 0 1 ...
## - attr(*, "dimnames")=List of 2
## ..$ : chr [1:10] "700014718" "700014767" ...
## ..$ : chr [1:10] "700014718" "700014767" ...
```



# Questions?

Post to the discussion board!



# Thank you!

Slides available: [github.com/bjklab](https://github.com/bjklab)

[brendank@pennmedicine.upenn.edu](mailto:brendank@pennmedicine.upenn.edu)

