Many ways to study the genomic basis of adaptation ... Or all you can think about to study adaptation with genomic tools...

> Claire Mérot & Anna Tigano Physalia Courses September 2020

# Analytical approaches

GWAS Comparative genomics Experimental evolution Transcriptomics QTL mapping **Epigenetics** Population genomics

# Plenty of approachs

- Local adaptation / population genomics
- Other statistics
- Comparative genomics
- Trait-focused genetics/genomics
- GWAS
- QTL
- Multi-omics
- Transcriptomics
- Epigenomics
- Proteomics

- Functions and experiments
- Experimental evolution / experimental selection
- Candidate genes and pathways
- Common garden
- Gene-editing

#### Site frequency spectrum:

- Tajima's D: tests for excess of low or intermediate frequency alleles
- Distribution of fitness effects (DFE)

## Signatures of selective sweeps:

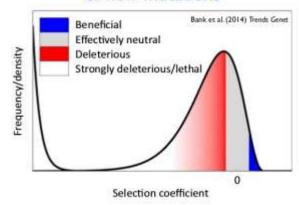
- SweepFinder
- Extended haplotype homozygosity (iHS)

# Inference of Distribution of Fitness Effects and Proportion of Adaptive Substitutions from Polymorphism Data

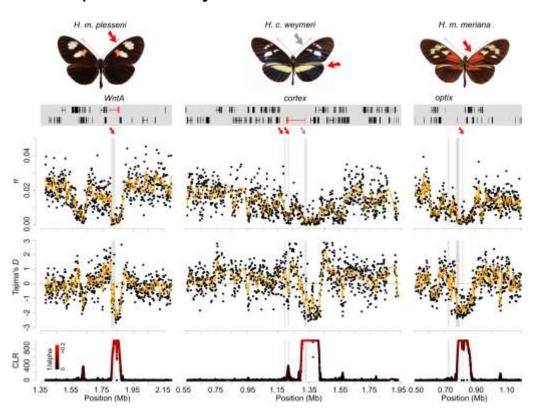
Tataru et al GENETICS November 1, 2017 vol. 207 no. 3 11031119; https://doi.org/10.1534/genetics.117.300323

#### polyDFEv2.0

# Distribution of fitness effects (DFE) of new mutations



## Sweep finder, Tajima's D



# Targeted sequence capture towards colour loci

- -> sweepFinder
- -> low pi (diversity),
- -> Low Tajima's D

Moest M, Van Belleghem SM, James JE, Salazar C, Martin SH, Barker SL, et al. (2020) Selective sweeps on novel and introgressed variation shape mimicry loci in a butterfly adaptive radiation. PLoS Biol 18(2): e3000597. https://doi.org/10.1371/journal.pbio.3000597

G12

## Haplotypes

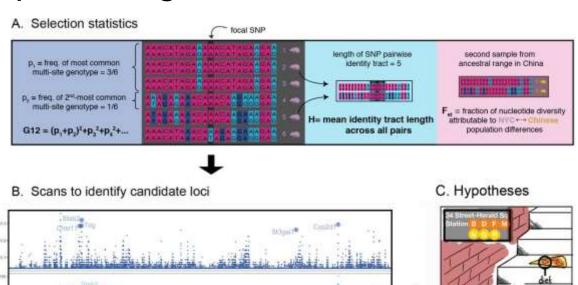




A signal of adaptation in New York City rats?

Harpak, A., Garud, N., Rosenberg, N. A., Petrov, D. A., Combs, M., Pennings, P. S., & Munshi-South, J. (2020). Genetic Adaptation in New York City Rats. *BioRxiv*.

https://doi.org/10.1101/2020.02.07.938969



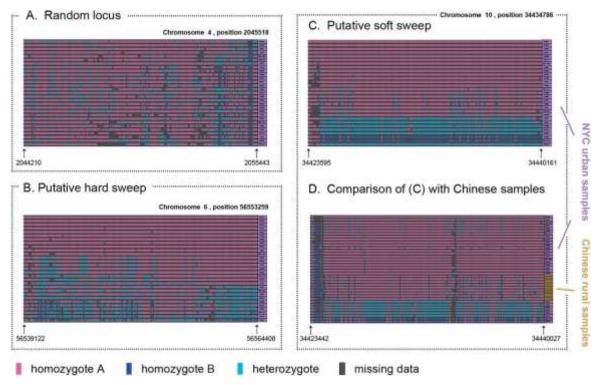
Methods to find hard sweep and soft sweeps based on haplotype + asymptotic MK test

position on chromosome 7 (Mbp)

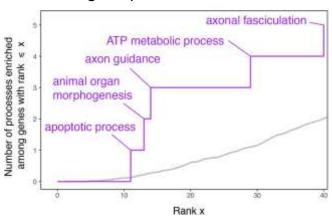
metakolism

lacovigtion

#### Sweep signal in haplotypes



#### Biological processes in outlier loci



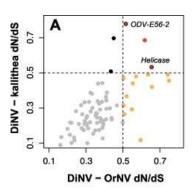
Harpak, A., Garud, N., Rosenberg, N. A., Petrov, D. A., Combs, M., Pennings, P. S., & Munshi-South, J. (2020). Genetic Adaptation in New York City Rats. *BioRxiv*. https://doi.org/10.1101/2020.02.07.938969

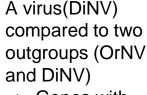
# Population genomics/ comparative genomics

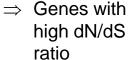
## Comparison to outgroup:

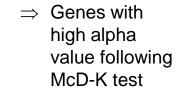
- ω = dN/dS ratio of the number of nonsynonymous to synonymous sites between target species and an outgroup
- McDonald Kreitman test: variation in rate of evolution across the genome and fixed vs. polymorphic sites
- Fay and Wu's H: Test of neutrality by comparing frequency of derived vs.
   ancestral alleles

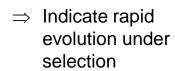
asymptoticMK: A Web-Based Tool for the Asymptotic McDonald–Kreitman Test Benjamin C. Haller and Philipp W. Messer G3: GENES, GENOMES, GENETICS May 1, 2017 vol. 7 no. 5 1569-1575; https://doi.org/10.1534/g3.117.039693

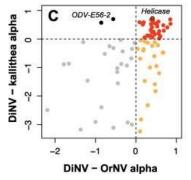










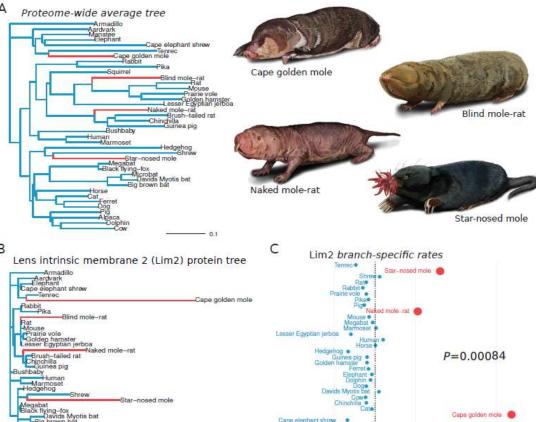


Hill & Unckless, 2018 https://doi.org/10.1016/j.meegid.2017.11.013 Comparative genomics

# Study adaptive convergence across species

 What are the genes that evolved at a more rapid rate in organisms adapted to similar ecological conditions? That evolved comparable traits?

Partha, R.,... & Clark, N. L. (2017). Subterranean mammals show convergent regression in ocular genes and enhancers, along with adaptation to tunneling. *Elife*, *6*, e25884. https://doi.org/10.7554/eLife.25884.001



relative evolutionary rate

Some local-adaptation/pop genomics approaches are also based on traits when contrasting populations with different phenotypes, morphotypes within a species or ecotypes...

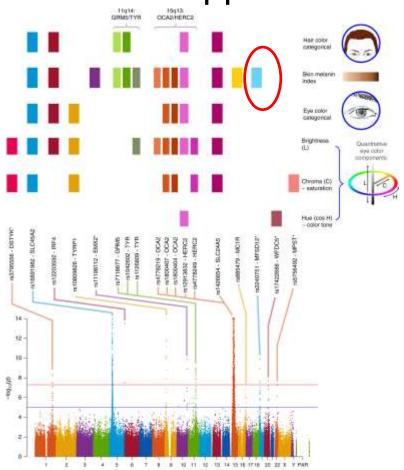
- ⇒ Need to characterize phenotypes and/or adaptative traits
- ⇒ Quantifying phenotypes can be complex (physiology, morphology, etc)

#### **GWAS**

#### Principle:

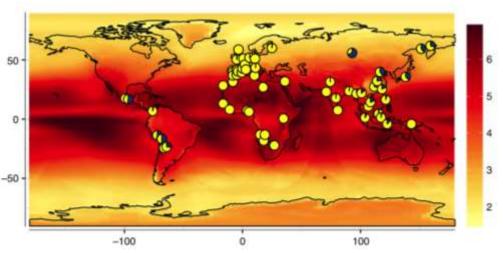
- many individuals (hundreds to thousands)
- A trait is quantified for all individuals (case-control, quantitative)
- Many markers (usually SNP-array or Whole-genome)
- ⇒ Locus-by-locus association test
- ⇒ Polygenic scores

Note many warning of the need to account for population structure



#### **GWAS**

6357 individuals 730,525 SNPs pigmentation of skin and eyes



Adhikari, K., Mendoza-Revilla, J., Sohail, A. *et al.* A GWAS in Latin Americans highlights the convergent evolution of lighter skin pigmentation in Eurasia. *Nat Commun* **10**, 358 (2019). https://doi.org/10.1038/s41467-018-08147-0

**QTL** (quantitative trait loci)

QTL analyses are usually based on family-design with crosses (or recombinant lines or pedigree) between individuals with the phenotype of interest.

Linkage maps can be build with genetic markers - medium density is ok so RAD-seq is well-suited for that.

Then, the association between genotype and phenotype are tested in the progeny

**QTL** (quantitative trait loci)

Huber, B., Whibley, A., Poul, Y. *et al.* Conservatism and novelty in the genetic architecture of adaptation in *Heliconius* butterflies. *Heredity* **114**, 515–524 (2015). https://doi.org/10.1038/hdy.2015.22

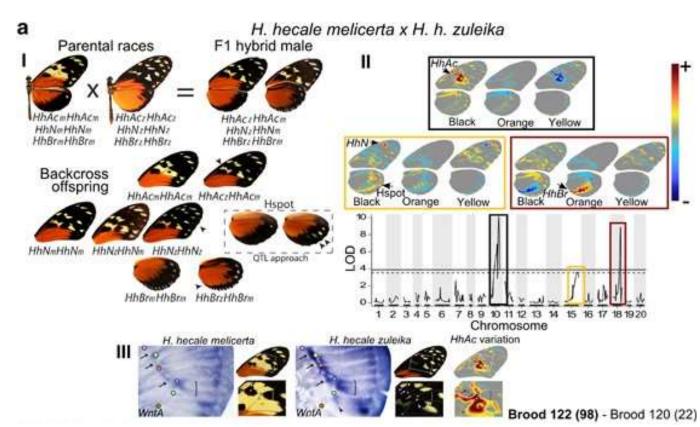
Targeted crosses and back-crosses

RAD-seq

Image analysis for colour pattern

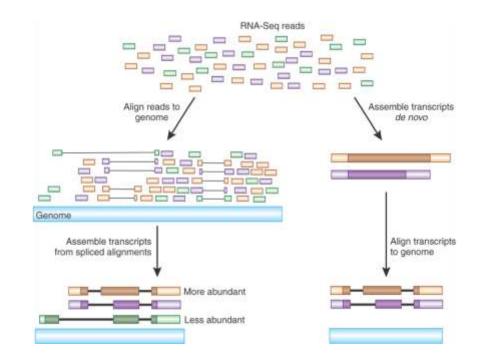
Multivariate associations

+ in situ hybridiation of WntA transcripts

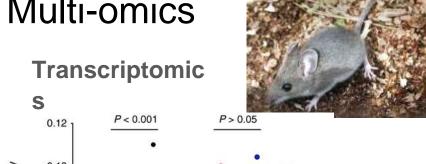


## **Transcriptomics**

- Quantify RNA expression
- Can be done on the wholeorganism or specific tissue
- qPCR on genes of interest or RNAseq of all transcripts
- Need to build a transcriptome (collection of transcripts)

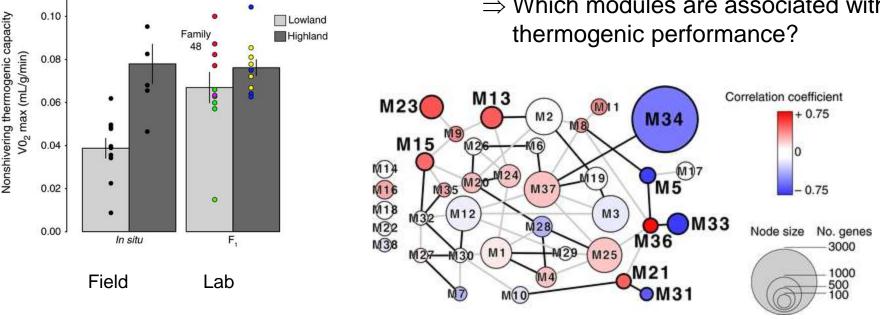


- ⇒Compare expression levels between two groups (differential expression)
- ⇒Find co-expressed genes
- ⇒eQTL if linked to phenotypes
- ⇒Allele-specific expression, isoform expression



RNA seq of brown adipose tissu 19 samples

- ⇒ Genes co-expression network
- ⇒ Which modules are associated with thermogenic performance?



Velotta, J.P., Jones, J., Wolf, C.J. and Cheviron, Z.A. (2016), Transcriptomic plasticity in brown adipose tissue contributes to an enhanced capacity for nonshivering thermogenesis in deer mice. Mol Ecol, 25: 2870-2886. doi:10.1111/mec.13661



## **Transcriptomics**

## ⇒ Enrichment analysis for GO terms

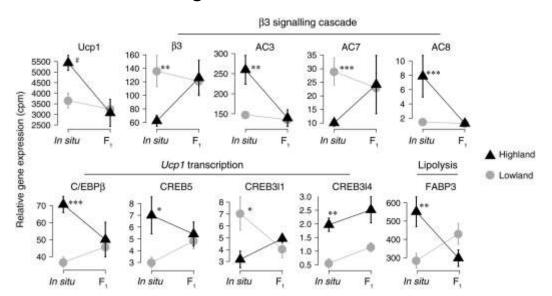
GO term	Description	P	9	Enrichment
GO:0001525	Angiogenesis	2.42E-09	4.37E-06	3.67
GO:0048514	Blood vessel morphogenesis	8.13E-05	1.03E-02	4.01
GO:0007219	Notch signalling pathway	8.61E-05	1.07E-02	3.7
GO:0043547	Positive regulation of GTPase activity	4.66E-07	2.19E-04	2.76
GO:1904018	Positive regulation of vasculature development	1.05E-04	1.27E-02	3.41
GO:0045765	Regulation of angiogenesis	1.25E-07	8.31E-05	3.68
GO:0070372	Regulation of ERK1 and ERK2 cascade	6.79E-05	9.15E-03	2.93
GO:0048729	Tissue morphogenesis	2.94E-07	1.69E-04	2.82
GO:0001570	Vasculogenesis	4.13E-04	3.74E-02	3.94

Velotta, J.P., Jones, J., Wolf, C.J. and Cheviron, Z.A. (2016), Transcriptomic plasticity in brown adipose tissue contributes to an enhanced capacity for nonshivering thermogenesis in deer mice. Mol Ecol, 25: 2870-2886. doi:10.1111/mec.13661



## **Transcriptomics**

### ⇒ Candidate genes



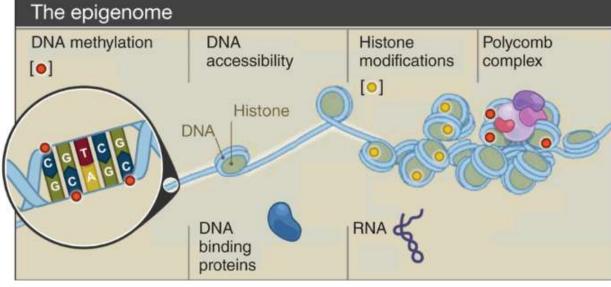
- β3 adrenergic receptor signalling cascade
- *Ucp1* transcription
- lipolysis
- => facilitate non-shivering thermogenesis within brown adipocytes

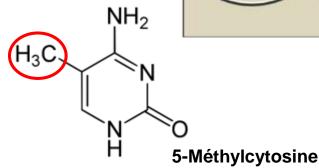
Velotta, J.P., Jones, J., Wolf, C.J. and Cheviron, Z.A. (2016), Transcriptomic plasticity in brown adipose tissue contributes to an enhanced capacity for nonshivering thermogenesis in deer mice. Mol Ecol, 25: 2870-2886. doi:10.1111/mec.13661

## **Epi-genomics**

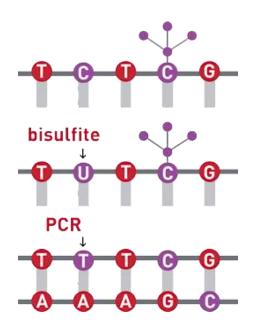
Epigenome is change to the DNA or histones, can be plastic and/or heritable, vary between tissues

The most accessible aspect for studies in ecology and evolution will be DNA methylation





## **Epi-genomics : DNA methylation**



use of bisulfite treatment of DNA before routine sequencing

- ⇒ Unmethylated C are converted into T
- ⇒ Remaining C are the ones that were metylated

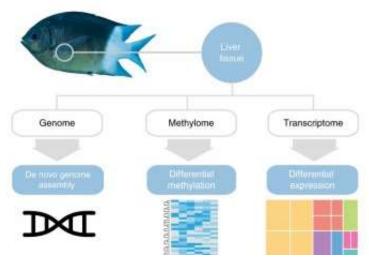
	X	30%	100%	
Studied variation is the % of methylated cytosine at a given position	C T T T T	C C T T T	00000	
	C/T	C/C	C/C	

Cannot look at sites with C/T polymorphism

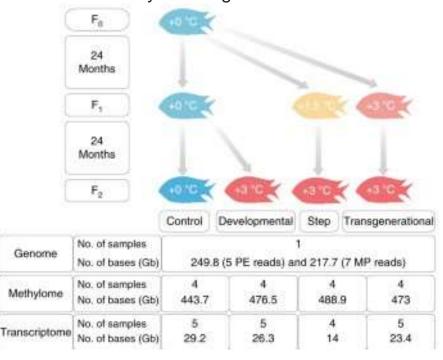
⇒ Variation in methylation level between tissues, between samples, between sexes, between populations, between ecotypes, etc...

## **Epi-genomics : DNA methylation**

- coral reef fish, Acanthochromis polyacanthus
- liver genome, methylomes and transcriptomes

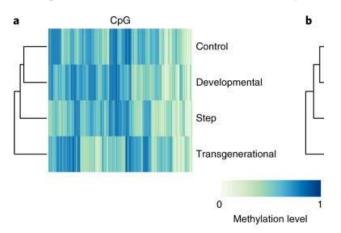


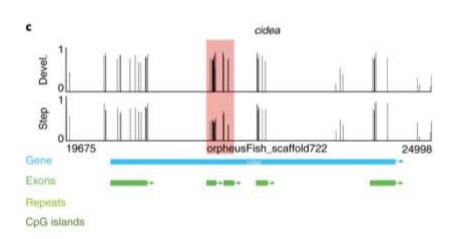
Exposed to current day (+0 °C) or future ocean temperatures (+3 °C) for one generation, two generations and incrementally across generations



Ryu, T., Veilleux, H.D., Donelson, J.M. *et al.* The epigenetic landscape of transgenerational acclimation to ocean warming. *Nature Clim Change* **8**, 50 509 (2018). <a href="https://doi.org/10.1038/s41558-018-0159-0">https://doi.org/10.1038/s41558-018-0159-0</a>

## **Epi-genomics : DNA methylation**





2,467 differentially methylated regions (DMRs) and 1,870 associated genes that respond to higher temperatures within and between generations

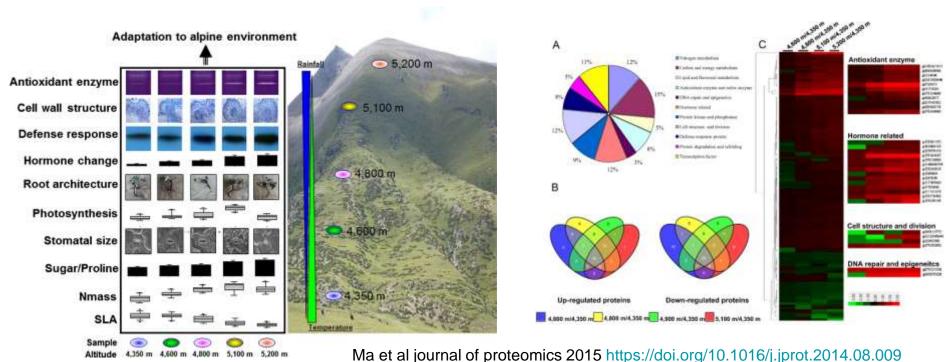
Some genes also show differential expression (but not many of them)

⇒ Association between DNA methylation and transgenerational acclimation to climate change

Ryu, T., Veilleux, H.D., Donelson, J.M. *et al.* The epigenetic landscape of transgenerational acclimation to ocean warming. *Nature Clim Change* **8**, 504–509 (2018). <a href="https://doi.org/10.1038/s41558-018-0159-0">https://doi.org/10.1038/s41558-018-0159-0</a>

#### Proteomics: characterization of all proteins with large-scale mass spectrometry

- ⇒ an analysis of morphology and proteome along an altitudinal gradient
- ⇒ Identify pathways involved in adaptation (hormones, anti-oxydant, epi-genetic regulations, etc.



## **Experimental evolution / Experimental selection**

From a starting population which is usually clonal (in yeast/bacteria) or highly-diverse (diploid species, drosophila, insects, etc), several replicates are kept under controlled conditions (one or several treatments) for a given number of generations (5 to 1 000...)

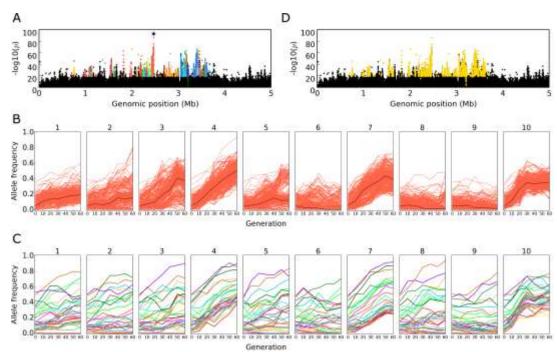
Phenotypes can be regularly measured

#### With genomics:

Usually pool-seq. The change of allelic frequencies is followed at regular intervals or contrasted between the beginning and the end of the experiment

⇒ Powerful way to follow evolution "under our eyes"!

## **Experimental evolution / Experimental selection**



Barghi N, Tobler R, Nolte V, Jakšić AM, Mallard F, Otte KA, et al. (2019) Genetic redundancy fuels polygenic adaptation in *Drosophila*. PLoS Biol 17(2): e3000128. https://doi.org/10.1371/journal.pbio.3000128

10 replicates of a *Drosophila* simulans population to a new temperature regime

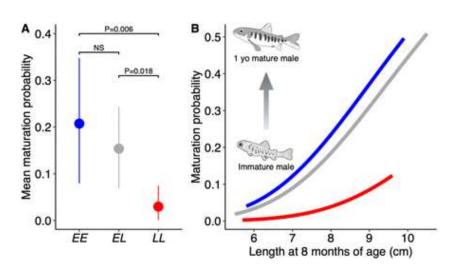
Pool-seq

Convergent responses for several phenotypes

a strong polygenic response (99 selected alleles; mean s = 0.059)

⇒ redundancy : not the same loci contribute to the evolution of the same phenotypes between replicates

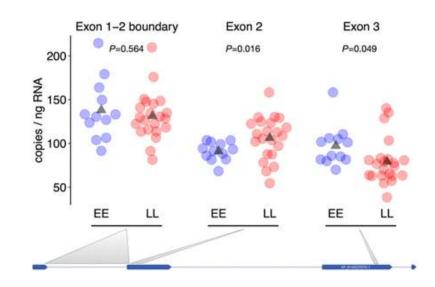
## Common-garden



A *candidate gene* from GWAS for age at maturity in Salmon : vgll3

⇒ Genotype and raise in controlled conditions 656 individuals

Cis-regulatory differences in isoform expression associate with life history strategy variation in Atlantic salmon Jukka-Pekka Verta et al bioRxiv 777300; doi: https://doi.org/10.1101/777300

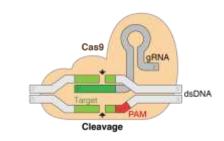


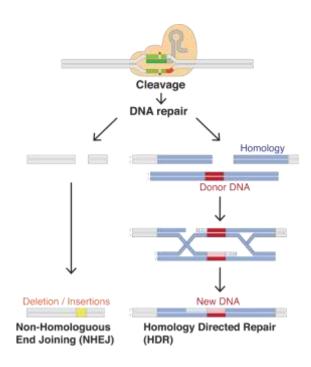
- ⇒ Trancriptomics : complex pattern of differential expression:
  It depends on the exon...
- ⇒ A different isoform expressed in EE and LL : *isoform-specific expression*

## **Gene-editing**

Target a candidate gene to silence it or change its sequence:

- -> used to be possible only in model species
- -> CRISPR-Cas9 is a game-changer



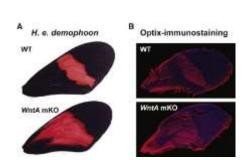


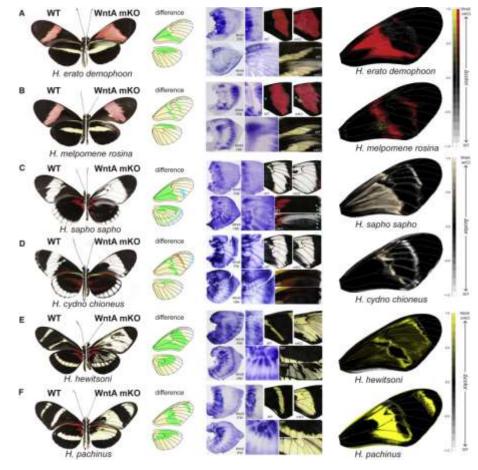
#### **Gene-editing**

Inject eggs and larva with CRISPR-CAs9 to make *WntA* KO mutant

- -> generation 0: 30% with a mutant phenotype (indels in ~80% of PCR *WntA*)
- ⇒ Confirm the relation genotype/phenotype
- ⇒ Compare multiple species
- Matches the area of influence during development

+ antibody immunostaining of transcripts





Concha, C., Wallbank, R. W., Hanly, J. J., Fenner, J., Livraghi, L., Rivera, E. S., ... & Morrison, C. (2019). Interplay between developmental flexibility and determinism in the evolution of mimetic Heliconius wing patterns. *Current Biology*, *29*(23), 3996-4009. <a href="https://doi.org/10.1016/j.cub.2019.10.010">https://doi.org/10.1016/j.cub.2019.10.010</a>

# Studying adaptation... Integrative biology!

