#### R for cytometry - fcs files

Nello Blaser

Department of Mathematics, University of Bergen

February 2<sup>nd</sup> 2018

#### fcs file structures

- ▶ flowFrame: Represents data contained in FCS files. There are three parts:
  - 1. raw measurements
  - 2. annotation for the parameters
  - 3. additional annotation
- flowSet: Container of several flowFrames.

#### reading fcs files

## looking at fcs files

```
# annotation
pData(ff@parameters)
```

##		name			desc	range	${\tt minRange}$	${\tt maxRange}$
##	\$P1	${\tt FSC-H}$			${\tt FSC-H}$	1024	0	1023
##	\$P2	SSC-H			SSC-H	1024	0	1023
##	\$P3	FL1-H			<na></na>	1024	1	10000
##	\$P4	FL2-H			<na></na>	1024	1	10000
##	\$P5	FL3-H			<na></na>	1024	1	10000
##	\$P6	FL1-A			<na></na>	1024	0	1023
##	\$P7	FL4-H			<na></na>	1024	1	10000
##	\$P8	Time	Time	(51.20	sec.)	1024	0	1023

## looking at fcs files

```
# look at data
ff@exprs[1:10, 1:5]
```

```
##
         FSC-H SSC-H
                          FI.1-H
                                     FI.2-H
                                                FI.3-H
           382
##
    [1,]
                  77 259.455272
                                  1.000000
                                            7.566695
    [2,]
           628
                       9.057978 48.260715 10.273508
##
                 280
    [3.]
                 735 537.611747 56.234133
##
          1023
                                            6.915821
    [4,]
##
           373
                 128
                       6.152654 24.144182
                                            2.329097
##
    [5,]
          1023
                1023 259.455272 791.475544 39.241898
    [6,]
##
           489
                 292
                       5.002865
                                 28.902639
                                            3.995421
    [7,]
##
           869
                 453 552.315842
                                  5.935229
                                            4.958068
    [8,]
##
           422
                 136 228.757320
                                  8.896491
                                            2.072079
                                            4.958068
##
    [9,]
           430
                 131 184.342299
                                  5.473703
   [10,]
           364
                 129 220.673407
                                  5.935229
                                             1.910953
##
```

#### Writing fcs files

**Important**: You are responsible that the expressions and the metadata in the new fcs-file correspond to each other!

#### reading multiple fcs files

# looking at flowSet

```
# flowSet
fset
  A flowSet with 3 experiments.
##
##
    column names:
##
    FSC-H SSC-H FL1-H FL2-H FL3-H FL1-A FL4-H Time
# flowFrame
fset[[1]]@exprs[1:3, 1:5]
##
       FSC-H SSC-H
                       FI.1-H FI.2-H
                                         FI.3-H
## [1.] 382 77 259.455272 1.00000 7.566695
## [2,] 628 280 9.057978 48.26071 10.273508
## [3,] 1023 735 537.611747 56.23413 6.915821
```

#### Data from cytobank

```
# load CRAN package
require(CytobankAPI)
# Authenticate via authentication token
cyto session <- authenticate(site="cellmass",
                             auth token="xxx")
# specify experiment id
experiment id <- 123
# specify data directory
data dir <- "fcs from cytobank"
dir.create(data dir, recursive = TRUE)
# list fcs files
files <- fcs_files.list(cyto_session,
                        experiment_id = experiment_id)
```

#### Data from cytobank

```
# download fcs files
downloaded_zip <- fcs_files.download_zip(</pre>
  UserSession = cyto_session,
  experiment_id = experiment_id,
  fcs files = files[, "id"],
  directory = data_dir,
  timeout = 60*nrow(files))
unzip(downloaded zip, exdir = data dir)
# read fcs files
fset <- read.flowSet(paste0(data dir, files))</pre>
```