R for cytometry - phenograph and spade

Nello Blaser

Department of Mathematics, University of Bergen

March 16th 2018

Download data from cytobank

```
# load CRAN package
require(CytobankAPI)
# Authenticate via authentication token
cyto session <- authenticate(site="cellmass",
                             auth token="xxx")
# specify experiment id
experiment id <- 12
# specify data directory
data dir <- "fcs from cytobank"
dir.create(data dir, recursive = TRUE)
# list fcs files
files <- fcs_files.list(cyto_session,
                        experiment_id = experiment_id)
```

Download data from cytobank

```
# download fcs files
downloaded_zip <- fcs_files.download_zip(
   UserSession = cyto_session,
   experiment_id = experiment_id,
   fcs_files = files[, "id"],
   directory = data_dir,
   timeout = 60*nrow(files))
unzip(downloaded_zip, exdir = data_dir)</pre>
```

Read fcs files

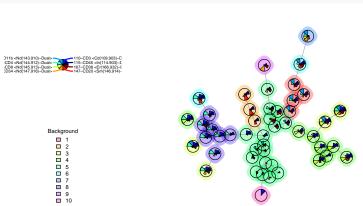
Transform data

```
# decide on lineage markers
pData(fset[[1]]@parameters)
lineage_markers <- c(</pre>
  "In(114.903)-Dual", "Cd(109.903)-Dual",
  "Nd(143.910)-Dual", "Nd(144.912)-Dual",
  "Nd(145.913)-Dual", "Nd(147.916)-Dual",
  "Sm(146.914)-Dual", "Er(166.932)-Dual")
# asinh transform
fset_lineage <- fsApply(fset, function(ff){</pre>
  ff <- ff[, lineage_markers]</pre>
  exprs(ff) <- asinh(exprs(ff)/5)
  ff
})
```

FlowSOM

```
require(FlowSOM)
# Set SOM parameters
flowSOM_metaClusters <- 10
flowSOM xdim <- 7
flowSOM ydim <- 7
flowSOM_seed <- 20180309 # for reproducible results
# run self-organizing maps
fsom <- FlowSOM(fset lineage,
                colsToUse = lineage markers,
                xdim=flowSOM xdim,
                ydim=flowSOM_xdim,
                nClus = flowSOM_metaClusters,
                seed = flowSOM seed)
```

Plotting with FlowSOM



Subsample

```
# set a seed for reproducible results
set.seed(20180308)
# extract data
concatinated_dta <- fsApply(fset_lineage, exprs)
# subsample
subsample <- sample(1:nrow(concatinated_dta), 1000)
# subsampled data
subsampled_dta <- concatinated_dta[subsample,]</pre>
```

Phenograph

```
## devtools::install_github("JinmiaoChenLab/Rphenograph")
require (Rphenograph)
# Set spade parameters
phenograph neighbors <- 50
# run phenograph
pheno <- Rphenograph(subsampled_dta[, lineage_markers],</pre>
                     k = phenograph_neighbors)
##
     Finding nearest neighbors...DONE ~ 0.01 s
##
     Compute jaccard coefficient between nearest-neighbor :
##
     Build undirected graph from the weighted links...DONE
##
     Run louvain clustering on the graph ... DONE ~ 0.08 s
##
     Return a community class
     -Modularity value: 0.7380464
##
     -Number of clusters: 8
##
pheno_cluster <- factor(membership(pheno[[2]]))</pre>
```

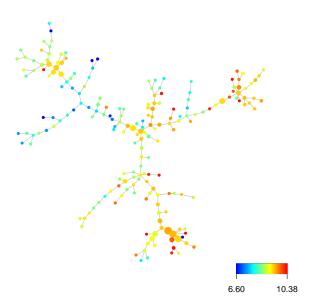
SPADE

```
require(spade)
# prepare to run spade
result_dir <- "spade_results"</pre>
dir.create(result_dir)
n_spade_clusters <- 200
# run spade
SPADE.driver(files,
              out_dir = result_dir,
              cluster_cols = lineage_markers,
             k = n spade clusters)
```

Plot SPADE

Plot SPADE

Marrow1_01_Basal1
Coeff. of Variation of absoluteEventNumber



t-SNE

```
# load tSNE package
require(Rtsne)
# Run tSNE
tsne_out <- Rtsne(subsampled_dta[, lineage_markers])</pre>
```

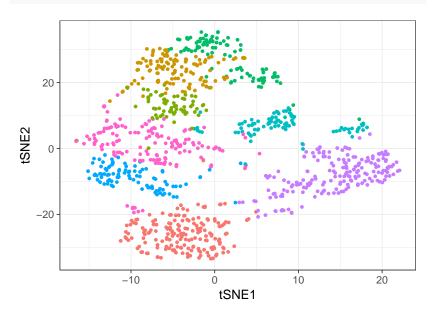
Combine results

```
# som results
fsom_cluster <- fsom$FlowSOM$map$mapping[subsample, 1]
fsom_meta_cluster <- fsom$metaclustering[fsom_cluster]</pre>
# combined results
result <- data.frame(
 tSNE1 = tsne out\$Y[, 1],
  tSNE2 = tsne out\$Y[, 2],
  fsom cluster = factor(fsom cluster),
  fsom meta cluster = fsom meta cluster,
  pheno cluster = factor(pheno cluster))
```

Plot

Plot

p_phenograph



Prepare data update

```
# numeric results
require(dplyr)
cols <- c("tSNE1", "tSNE2", "pheno_cluster",</pre>
            "fsom cluster", "fsom meta cluster")
result <- dplyr::mutate_at(result, cols, as.numeric)</pre>
# files and sample ids
result$filenames <-
  rep(sampleNames(fset),
      fsApply(fset, nrow, use.exprs = TRUE))[subsample]
result$sample id <- subsample
result$sample fid <-
  apply(sapply(c(0, cumsum(fsApply(fset, nrow,
                                    use.exprs = TRUE))).
             function(x) subsample - x), 1,
      function(y) min(y[y>0]))
```

Update data

```
update ff <- function(ff, res data, cols){
  rd <- res data[res data$filenames == identifier(ff), ]
  ff@exprs <- cbind(ff@exprs[rd$sample_fid, ],
                    as.matrix(rd[, cols]))
  ff@parameters@data <- rbind(
    ff@parameters@data,
    data.frame(
      name = cols,
      desc = "R cols",
      range = apply(rd[, cols], 2, function(x)
        diff(range(x))),
      minRange = apply(rd[, cols], 2, min),
      maxRange = apply(rd[, cols], 2, max),
      row.names =
        paste0("$P", nrow(ff@parameters@data) +
                 1:length(cols))))
  ff
```

Update data

Writing FCS file

Uploading files to cytobank

```
# create new experiment
new_experiment_name <- "test_clustered"</pre>
new experiment <- experiments.new(</pre>
  cyto session,
  experiment name = new experiment name,
  purpose = "testing")
# upload files
fcs_files.upload_zip(cyto_session,
                      experiment_id = new_experiment$id,
                      file_path = zip_filename,
                      timeout = 720)
```