multimark: an R package for analysis of capture-recapture data consisting of multiple "non-invasive" marks

Brett T. McClintock¹

National Marine Mammal Laboratory

Alaska Fisheries Science Center NOAA National Marine Fisheries Service Seattle, Washington, U.S.A. ¹Email: brett.mcclintock@noaa.gov

RUNNING HEAD: multimark mark-recapture package

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1 Summary

- 2 1. I describe an open source R package, multimark, for estimation of survival and
- abundance from capture-mark-recapture data consisting of multiple "non-invasive"
- 4 marks. Non-invasive marks include natural pelt or skin patterns, scars, and ge-
- 5 netic markers that enable individual identification in lieu of physical capture, and
- 6 thus apply to any species that can be individually identified from visual or genetic
- ⁷ sampling surveys. multimark provides a means for combining and jointly analyz-
- 8 ing encounter histories from multiple non-invasive sources that otherwise cannot be
- 9 reliably matched (e.g. left- and right-sided photos of bilaterally asymmetrical indi-
- 10 viduals).
- 2. multimark is currently capable of fitting open population Cormack-Jolly-Seber
- ₂ (CJS) and closed population abundance models with up to two mark types using
- Bayesian Markov chain Monte Carlo (MCMC) methods. Although originally moti-
- vated by the challenges posed by multiple non-invasive marks, multimark can also
- be used for Bayesian analyses of conventional capture-recapture data consisting of a
- 16 single mark type.
- 3. Some package features include: (i) general model specification using formulas al-
- 18 ready familiar to most R users, (ii) ability to include temporal, behavioural, age, co-
- hort, and individual heterogeneity effects in detection and survival probabilities, (iii)
- 20 improved MCMC algorithm that is computationally faster and more efficient than
- 21 previously proposed methods, (iv) Bayesian multimodel inference using reversible
- 22 jump MCMC, and (v) data simulation capabilities for power analyses and assessing
- 23 model performance.

4. I demonstrate use of multimark using left- and right-sided encounter histories for bobcats (*Lynx rufus*) collected from remote single-camera stations in southern California. In this example, there is evidence of a behavioural effect (i.e. trap "happy" response) that is otherwise indiscernible using traditional single-sided analyses.

5. The package will be most useful to ecologists seeking stronger inferences by combining different sources of mark-recapture data that are difficult (or impossible) to reliably reconcile, particularly with the sparse datasets typical of rare or elusive species for which non-invasive sampling techniques are most commonly employed. Addressing deficiencies in currently available software, multimark also provides a user-friendly interface for performing Bayesian multimodel inference using capture-recapture data consisting of a single conventional mark or multiple non-invasive marks.

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Key-words Bayesian multimodel inference, capture-recapture, Cormack-Jolly-Seber (CJS), latent multinomial, mark-recapture, Markov chain Monte Carlo (MCMC), multiple lists, population size

1 Introduction

Capture-recapture methods historically relied on the physical capture, marking, and recapturing of animals for estimating population abundance and related demographic parameters such as survival (e.g. Williams et al. 2002). More recently, "non-invasive" capture-recapture sampling techniques are becoming commonplace for monitoring animal populations (e.g. Hammond 1990; Lukacs & Burnham 2005; O'Connell et al. 2010). Non-invasive marks can include natural pelt or skin patterns, scars, and

genetic markers that enable individual identification in the absence of physical capture. Capture-recapture methods based on non-invasive marks have been applied to
diverse taxa, including sharks (e.g. Holmberg et al. 2008), reptiles (e.g. Nair et al.
2012), ursids (e.g. Dreher et al. 2007), felids (e.g. Karanth & Nichols 1998; Ruell
et al. 2009), and marine mammals (e.g. Hammond 1990; Wilson et al. 1999; Madon
et al. 2011). While non-invasive capture-recapture methods have many advantages
related to financial cost and animal welfare, they also pose new difficulties such as
animal misidentification (Wright et al. 2009; Yoshizaki et al. 2009; Link et al. 2010;
Morrison et al. 2011) and the complexity of multiple types of marks (Corkrey et al.
2008; Madon et al. 2011; Bonner & Holmberg 2013; McClintock et al. 2013).

Multiple marks can arise from sighting or camera surveys of species with natu-57 ral mark patterns that are bilaterally asymmetrical (e.g. cetaceans, felids) or from multiple sources of non-invasive capture-recapture data being collected concurrently 59 (e.g. faecal DNA sampling and visual surveys). With multiple marks, an encounter history is produced for each individual and mark type, but there is typically no reliable means to match them (unless each mark type is simultaneously observed at least once for every encountered individual). Because the number of unique individuals encountered must be known for standard capture-recapture analyses, the typical approach is to conduct separate analyses for each mark type and compare the results (e.g. Wilson et al. 1999; Berrow et al. 2012; Nair et al. 2012). However, given that sample sizes (and precision) may be considerably reduced, this is not as efficient as conducting an integrated analysis utilizing encounter histories arising from all mark types (McClintock et al. 2013). Additional costs of conducting separate analyses for each mark type include a limited ability to explore models with behavioural or cohort effects, and, for capture-recapture models that condition on first encounter, a forfeiting of information from histories with the (apparent) first encounter occurring on the last sampling occasion. These limitations can be particularly important for the sparse datasets typical of rare and elusive populations for which non-invasive sampling techniques are most commonly employed.

Based on the latent multinomial model of Link et al. (2010), Bonner & Holmberg 76 (2013) and McClintock et al. (2013) recently developed methods for performing integrated analyses of capture-recapture data consisting of multiple non-invasive marks. However, to my knowledge, their approaches have yet to be applied by practitioners. This is certainly not due to a lack of appropriate data (e.g. Wilson et al. 1999; Holmberg et al. 2008; Madon et al. 2011; Berrow et al. 2012; Nair et al. 2012), and is likely 81 attributable to the mathematical and computational complexity of the models, as 82 well as a lack of user-friendly software for implementing them. Generalized software 83 for performing Bayesian multimodel inference with capture-recapture data has also been lacking, thereby leaving these procedures largely inaccessible to non-statisticians (e.g. Brooks et al. 2000; Durban & Elston 2005; King & Brooks 2008; Royle 2008; McClintock et al. 2013). These software needs were the motivation for multimark, an R (R Core Team 2013) package for Bayesian analysis of capture-recapture data consisting of multiple non-invasive marks.

After providing some additional background on capture-recapture with multiple marks, I briefly describe the models implemented in multimark. These currently include open population Cormack-Jolly-Seber (CJS) and closed population abundance models (e.g. Williams et al. 2002) with up to two mark types. Although originally motivated by the challenges posed by multiple non-invasive marks, multimark can

also be used for analyses of conventional capture-recapture data consisting of a single mark type. Using real and simulated data for illustration, I provide an overview of the workflow for the package and a new analysis of left- and right-sided encounter histories for bobcats (*Lynx rufus*) collected from remote single-camera stations in southern California. Additional information, including help files, data, examples, and package usage is available by downloading the multimark package from CRAN (http://cran.r-project.org) or github (https://github.com/bmcclintock/multimark). This article describes multimark version 1.3.0.

$_{\scriptscriptstyle 03}$ 2 Description

104 2.1 Background

Capture-recapture data are typically represented by a collection of encounter histo-105 ries $\mathbf{Y} = \{\mathbf{y}_1, \mathbf{y}_2, \dots, \mathbf{y}_n\}$, where each element of $\mathbf{y}_i = (y_{i,1}, y_{i,2}, \dots, y_{i,T})$ indicates 106 whether individual i was detected $(y_{i,t} = 1)$ or not detected $(y_{i,t} = 0)$ on each 107 of $t=1,\ldots,T$ sampling occasions. Typical analyses then proceed by formulat-108 ing a likelihood conditional on the n unique individuals encountered (e.g. Williams 109 et al. 2002). With two mark types, we instead have $\tilde{\mathbf{Y}}_m = {\{\tilde{\mathbf{y}}_{m_1}, \tilde{\mathbf{y}}_{m_2}, \dots, \tilde{\mathbf{y}}_{m_{n_m}}\}}$ for 110 $m \in \{1, 2\}$, where each element of $\tilde{\mathbf{y}}_{m_i} = (\tilde{y}_{m_{i,1}}, \tilde{y}_{m_{i,2}}, \dots, \tilde{y}_{m_{i,T}})$ indicates individual i was detected $(\tilde{y}_{m_{i,t}} = m)$ or not detected $(\tilde{y}_{m_{i,t}} = 0)$, and n_m is the number of unique individuals encountered for mark type m. We focus on situations where it is difficult (or impossible) to reliably match individuals from \mathbf{Y}_1 and \mathbf{Y}_2 . In this case, 114 although we know $n \leq n_1 + n_2$, n is nevertheless unknown and standard capture-115 recapture analysis methods cannot be reliably used for simultaneous inference using 116 both sources of data.

Depending on the mark types and sampling design, it may sometimes be possible 118 to observe both marks simulateneously within a sampling occasion. In this case, 119 some of the encounter histories from $\tilde{\mathbf{Y}}_1$ and $\tilde{\mathbf{Y}}_2$ can be matched to unique indi-120 viduals with certainty. For example, suppose images were collected during vessel-121 based line transect surveys of surfacing whales, where mark type 1 corresponds 122 to patch patterns on the left side and mark type 2 corresponds to patterns on the right side. If an individual happens to be photographed on both sides si-124 multaneously on at least one sampling occasion, then the true encounter history 125 for this individual would be known (i.e. left- and right-sided images could be 126 matched). This results in an additional set of n_{known} observed encounter histories, 127 $\mathbf{\tilde{Y}}_{known} = \left\{ \mathbf{\tilde{y}}_{known_1}, \mathbf{\tilde{y}}_{known_2}, \dots, \mathbf{\tilde{y}}_{known_{n_{known}}} \right\}$, consisting of histories that are known 128 with certainty (Table 1). 129 In essence, multimark facilitates the joint analysis of type 1 $(\tilde{\mathbf{Y}}_1)$, type 2 $(\tilde{\mathbf{Y}}_2)$, 130 and known encounter histories $(\tilde{\mathbf{Y}}_{known})$ while accounting for uncertainty in the num-131 ber of unique individuals encountered using extensions of the methodology proposed 132 by Bonner & Holmberg (2013) and McClintock et al. (2013). While the mathematical 133 and computational details are generally of little interest to ecologists, multimark per-134 forms these operations in the background and requires only simple data formatting 135

2.2 Models

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multimark currently includes open population Cormack-Jolly-Seber (CJS) and closed population abundance models (e.g. Williams *et al.* 2002). These Bayesian implementations are similar in spirit to the CJS model of Royle (2008) and the abundance

and model specification formulas familiar to most R users.

Table 1. Latent encounter histories \mathbf{y} and the recorded histories $(\tilde{\mathbf{y}}_1, \tilde{\mathbf{y}}_2, \tilde{\mathbf{y}}_{known})$ they generate for T=2 sampling occasions and two mark types, where $\mathbf{y}=(y_1,y_2)$ for $y_t \in \{0,1,2,3,4\}$. Latent encounter histories are indexed by $j=1+\sum_{t=1}^T y_t 5^{T-t}$, where the encounter types indicate non-detection $(y_t=0)$, type 1 encounter $(y_t=1)$, type 2 encounter $(y_t=2)$, non-simultaneous type 1 and type 2 encounter $(y_t=3)$, and simultaneous type 1 and type 2 encounter $(y_t=4)$. If simultaneous encounters are possible, these result in some \mathbf{y} being completely observable (as indicated by $\tilde{\mathbf{y}}_{known}$).

\overline{j}	y	$ ilde{\mathbf{y}}_1$	$ ilde{\mathbf{y}}_2$	$ ilde{\mathbf{y}}_{known}$
$\frac{j}{1}$	00			
2	01	01		
3	02		02	••
4	03	01	02	
5	04			04
6	10	10		
7	11	11		
8	12	10	02	
9	13	11	02	
10	14			14
11	20		20	
12	21	01	20	
13	22		22	
14	23	01	22	
15	24			24
16	30	10	20	
17	31	11	20	
18	32	10	22	
19	33	11	22	
20	34			34
21	40			40
22	41			41
23	42			42
24	43			43
25	44			44

model of King et al. (2015). Given the latent encounter histories (\mathbf{Y}) that generated the observed encounter histories ($\tilde{\mathbf{Y}}_1, \tilde{\mathbf{Y}}_2, \tilde{\mathbf{Y}}_{known}$), the likelihood for the CJS model with two mark types is

$$[\mathbf{Y} \mid \mathbf{p}, \boldsymbol{\delta}, \alpha, \boldsymbol{\phi}, \mathbf{Q}] \propto \prod_{i=1}^{n} \prod_{t=C_i+1}^{T} \pi_{i,t}$$
 (1)

$$\pi_{i,t} = \begin{cases} (1 - p_{i,t}) \, \phi_{i,t-1} q_{i,t} + (1 - \phi_{i,t-1}) \, (1 - q_{i,t}) & \text{if } y_{i,t} = 0 \text{ and } q_{i,t-1} = 1 \\ p_{i,t} \delta_1 \phi_{i,t-1} & \text{if } y_{i,t} = 1 \\ p_{i,t} \delta_2 \phi_{i,t-1} & \text{if } y_{i,t} = 2 \\ p_{i,t} \, (1 - \delta_1 - \delta_2) \, (1 - \alpha) \, \phi_{i,t-1} & \text{if } y_{i,t} = 3 \\ p_{i,t} \, (1 - \delta_1 - \delta_2) \, \alpha \phi_{i,t-1} & \text{if } y_{i,t} = 4 \\ 1 & \text{otherwise} \end{cases}$$

where $y_{i,t} = 0$ indicates a non-detection for individual i on occasion t, $y_{i,t} = 1$ indicates a type 1 encounter, $y_{i,t} = 2$ indicates a type 2 encounter, $y_{i,t} = 3$ indicates 145 a non-simultaneous type 1 and type 2 encounter, $y_{i,t} = 4$ indicates a simultane-146 ous type 1 and type 2 encounter, $C_i \in \{1, ..., T\}$ is the time of first capture for 147 individual i, $p_{i,t}$ is the detection probability for individual i during sampling occa-148 sion t, δ_m is the conditional probability of a type m encounter (given detection), 149 α is the conditional probability of a simultaneous type 1 and type 2 encounter 150 (given both mark types detected), $\phi_{i,t-1}$ is the survival probability between times 151 t-1 and t, and $q_{i,t}$ is an indicator for whether individual i was alive $(q_{i,t}=1)$ or 152 not $(q_{i,t} = 0)$ during sampling occasion t. With T = 3, we for example have cell 153 probabilities $\pi_i = \prod_{t=C_i+1}^T \pi_{i,t} = (1-p_{i,3}) \phi_{i,2} q_{i,3} + (1-\phi_{i,2}) (1-q_{i,3})$ for latent en-154 counter history 020, $\pi_i = p_{i,2}\delta_1\phi_{i,1}p_{i,3}\delta_2\phi_{i,2}$ for latent encounter history 412, $\pi_i =$ 155 $(1 - p_{i,2}) \phi_{i,1} p_{i,3} (1 - \delta_1 - \delta_2) (1 - \alpha) \phi_{i,2}$ for history 103, and $p_{i,3} (1 - \delta_1 - \delta_2) \alpha \phi_{i,2}$ 156 for history 034. 157

For added flexibility, p and ϕ are modeled using the probit link function:

$$\Phi\left(p_{i,t}\right) = \mathbf{x}_t^{p'} \boldsymbol{\beta}^p + z_i^p$$

$$\Phi\left(\phi_{i,t}\right) = \mathbf{x}_{t}^{\phi'} \boldsymbol{\beta}^{\phi} + z_{i}^{\phi}$$

where $\Phi()$ the cumulative distribution function of the standard normal density, \mathbf{x}_t^p and \mathbf{x}_t^{ϕ} are row t of the design matrices for p and ϕ , $\boldsymbol{\beta}^p$ and $\boldsymbol{\beta}^{\phi}$ are the corresponding 159 regression coefficients, and $z_{i}^{p} \sim \mathcal{N}\left(0, \sigma_{z^{p}}^{2}\right)$ and $z_{i}^{\phi} \sim \mathcal{N}\left(0, \sigma_{z^{\phi}}^{2}\right)$ are individual-level 160 effects that respectively allow for individual heterogeneity in detection and survival 161 probability. Thus, while exploring the feasible set of latent encounter histories (Y), 162 the parameters and latent variables to be estimated by multimark include β^p , β^{ϕ} , 163 $\boldsymbol{\delta}$, α , \mathbf{Q} , \mathbf{z}^p , \mathbf{z}^{ϕ} , $\sigma_{z^p}^2$, and $\sigma_{z^{\phi}}^2$. 164 The probit link is implemented for CJS models in multimark because it facilitates 165 a Gibbs sampler in the spirit of Albert & Chib (1993) and Laake et al. (2013). The probit link is very similar to the logit link, but the logit link has slightly fatter tails and is interpretable in terms of log-odds. I note that this model reduces to that of 168

when $\delta_1=1$ and $\delta_2=0$ for $y_{i,t}\in\{0,1\}.$ Similarly, the likelihood for the closed population abundance model with two mark types is

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Laake et al. (2013) for conventional capture-recapture data with a single mark type

$$[\mathbf{Y} \mid \mathbf{p}, \boldsymbol{\delta}, \alpha, N] \propto \frac{1}{(p^*)^n} \prod_{i=1}^n \prod_{t=1}^T \pi_{i,t} \times \text{Binomial}(n; N, p^*)$$
 (2)

$$\pi_{i,t} = \begin{cases} (1 - p_{i,t}) & \text{if } y_{i,t} = 0\\ p_{i,t}\delta_1 & \text{if } y_{i,t} = 1\\ p_{i,t}\delta_2 & \text{if } y_{i,t} = 2\\ p_{i,t} \left(1 - \delta_1 - \delta_2\right) \left(1 - \alpha\right) & \text{if } y_{i,t} = 3\\ p_{i,t} \left(1 - \delta_1 - \delta_2\right) \alpha & \text{if } y_{i,t} = 4\\ 1 & \text{otherwise} \end{cases}$$

where N is the population size, and p^* is the probability that a randomly selected individual is detected at least once. As before, this model reduces to that for conventional capture-recapture data with a single mark type when $\delta_1=1$ and $\delta_2=0$ for $y_{i,t}\in\{0,1\}$. Returning to Table 1 with T=2, we for example have cell probabilities $\pi_i=\prod_{t=1}^T\pi_{i,t}=(1-p_{i,1})\,p_{i,2}\delta_1$ for latent encounter history 01, $\pi_i=p_{i,1}\delta_2\,(1-p_{i,2})$ for history 20, $\pi_i=p_{i,1}\delta_2 p_{i,2}\,(1-\delta_1-\delta_2)\,(1-\alpha)$ for history 23, and $\pi_i=p_{i,1}\,(1-\delta_1-\delta_2)\,\alpha p_{i,2}\,(1-\delta_1-\delta_2)\,(1-\alpha)$ for history 43.

For closed population models, p is modeled using the logit link function:

$$logit (p_{i,t}) = \mathbf{x}_t^{p'} \boldsymbol{\beta}^p + z_i^p$$

such that

$$p^* = 1 - \int_{-\infty}^{\infty} \prod_{t=1}^{T} \left(1 - \frac{1}{1 + \exp\left(-\left(\mathbf{x}_t^{p'}\boldsymbol{\beta}^p + z^p\right)\right)} \right) \mathcal{N}\left(z^p; 0, \sigma_{z^p}^2\right) dz^p$$

is the probability of being detected at least once after accounting for individual heterogeneity in p (note that $p^* = 1 - \prod_{t=1}^T \left(1 - \operatorname{logit}^{-1}\left(\mathbf{x}_t^{p'}\boldsymbol{\beta}^p\right)\right)$ when $\sigma_{z^p}^2 = 0$). The parameters and latent variables to be estimated therefore include $\boldsymbol{\beta}^p$, $\boldsymbol{\delta}$, α , N, \mathbf{z}^p , and $\sigma_{z^p}^2$. Although a Gibbs sampler has been proposed for closed population models using the probit link and a complete data likelihood (McClintock *et al.* 2014), this does not apply to the "semi-complete" data likelihood in Eq. 2 (hence the traditional

Table 2. Summary of three different types of multiple-mark data. The data differ in terms of the latent encounter types (y_t) that are possible based on the conditional probability of a simultaneous type 1 and type 2 encounter, $\alpha = \Pr(y_t = 4|y_t = 3 \text{ or } y_t = 4)$.

Data type	y_t	Constraints
"never"	$\{0, 1, 2, 3\}$	$\alpha = 0$
"sometimes"	$\{0, 1, 2, 3, 4\}$	$0 < \alpha < 1$
"always"	$\{0, 1, 2, 4\}$	$\alpha = 1$

logit link is used). The primary utility of multimark is finding the set of latent encounter histories that are feasible given the observed encounter histories (sensu Link et al. 2010; Bonner & Holmberg 2013; McClintock et al. 2013, 2014). Given a feasible set of latent encounter histories (Y), fitting capture-recapture models such as Eqs. 1 or 2 is relatively straightforward.

3 Workflow

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3.1 Multiple non-invasive marks

193 3.1.1 Data formatting

There are three types of multiple-mark data that can be analyzed with multimark. 194 These are the "never", "sometimes", and "always" data types, and they are named 195 based on their respective probabilities of a simultaneous type 1 and type 2 encounter 196 (Table 2). An example of the "never" data type is provided with multimark and 197 includes 23 left-sided ($\tilde{\mathbf{Y}}_1$) and 23 right-sided ($\tilde{\mathbf{Y}}_2$) encounter histories for bobcats 198 (Lynx rufus) collected from remote single-camera stations in southern California over 199 T=8 sampling periods between July 2006 and January 2007 (McClintock et al. 2013; 200 Alonso et al. 2015). 201

multimark expects observed encounter history data to be a matrix with rows

corresponding to individuals and columns corresponding to sampling occasions. Because the bobcat data were collected from single-camera stations, simultaneous leftand right-sided encounters were not possible; hence $\alpha = 0$ and the rows consist of
either 0's and 1's or 0's and 2's:

- > library(multimark)
- > data(bobcat)
- > head(bobcat)

	occ1	occ2	occ3	occ4	occ5	occ6	occ7	occ8
ID2	0	0	0	0	0	1	1	0
ID3	0	0	1	0	1	0	0	0
ID4	0	0	0	0	1	0	0	0
ID6	1	0	0	0	0	0	0	0
ID7	0	0	1	0	0	0	0	1
ID8	0	1	0	0	0	0	0	0

> tail(bobcat)

	occ1	occ2	occ3	occ4	occ5	occ6	occ7	occ8
ID49	0	0	2	0	0	0	0	0
ID50	0	0	2	0	0	0	0	0
ID51	0	0	0	2	0	0	0	0
ID52	0	0	0	0	2	0	0	0
ID53	0	0	0	0	0	2	0	0
ID54	0	0	0	0	0	0	2	0

The ordering of the rows is unimportant; the package automatically recognizes which histories belong to $\tilde{\mathbf{Y}}_1$, $\tilde{\mathbf{Y}}_2$, and, if applicable, $\tilde{\mathbf{Y}}_{known}$.

The multimark function *processdata()* performs all additional data formatting.

The basic inputs are the matrix of observed encounter histories (*Enc.Mat*) and the
data type (*data.type*):

> bobcatsetup <- processdata(Enc.Mat=bobcat,data.type="never")

This creates an object of class multimarksetup that includes everything needed for model fitting and further analysis. In particular, processdata() calculates all of the necessary ingredients for identifying the feasible set of latent encounter histories (for technical details see Bonner & Holmberg 2013; McClintock et~al.~2013). There is also a feature enabling designation of individual encounter histories as known with certainty despite no simultaneous type 1 and type 2 detections (i.e. $y_{i,t} \neq 4 \forall t$), a situation that can arise from a previous physical capture or concurrent telemetry study (e.g. McClintock et~al.~2013).

$_{220}$ 3.1.2 Model fitting

- The package currently includes functions multimarkCJS() and multimarkClosed()
 for fitting CJS and closed population models, respectively, with two mark types.
 Use of these functions is perhaps best explained by example. To fit a simple closed
- $_{\rm 224}$ $\,$ population model assuming constant detection probability using the default settings:
 - > bobcat.dot <- multimarkClosed(mms=bobcatsetup,
 + mod.p=~1)</pre>
- Equivalently, Enc. Mat and data.type can be provided in lieu of the mms argument.
- 226 In this case, processdata() is called from within multimarkClosed():
 - > bobcat.dot <- multimarkClosed(Enc.Mat=bobcat,data.type="never",
 + mod.p=~1)</pre>
- This creates a list, bobcat.dot, containing the MCMC output for the model
- (bobcat.dot\$mcmc). The MCMC output is of class mcmc, which should be familiar
- 229 to users of the R package coda (Plummer et al. 2006):
 - > summary(bobcat.dot\$mcmc)

Iterations = 2001:12000
Thinning interval = 1
Number of chains = 1
Sample size per chain = 10000

1. Empirical mean and standard deviation for each variable, plus standard error of the mean:

```
MeanSDNaive SE Time-series SEpbeta[(Intercept)]-1.31570.242840.00242840.016369N35.60935.082350.05082350.250575delta_10.40190.074490.00074490.007738delta_20.41820.074420.00074420.007939
```

2. Quantiles for each variable:

```
2.5%
                                25%
                                         50%
                                                 75%
                                                        97.5%
pbeta[(Intercept)] -1.8037 -1.4794 -1.3152 -1.1486 -0.8360
N
                    28.0000 32.0000 35.0000 39.0000 47.0000
delta_1
                     0.2576
                             0.3505
                                      0.4024
                                              0.4525
                                                       0.5457
                     0.2750
                             0.3666
                                     0.4186
                                              0.4700
delta_2
                                                       0.5622
```

> coda::effectiveSize(bobcat.dot\$mcmc)

delta_2	delta_1	N	<pre>pbeta[(Intercept)]</pre>
87.86513	92.66564	411.38952	220.09308

- Here we can see posterior summaries for the default monitored parameters $(\beta^p, N, \delta_1, \delta_2)$.
- Based on the effective sample sizes, it's clear that the default chain length is inade-
- quate for this example; a typical "rule of thumb" is effective sample sizes > 4000 for
- 233 all quantities of interest.
- Other common models for detection probability can be easily specified using linear
- model formulas for mod.p, including shorthands for time variation (mod.p = time),
- temporal trends $(mod.p = \tilde{r} Time)$, behavioural response to first capture $(mod.p = \tilde{r}c)$,
- ²³⁷ and individual heterogeneity ($mod.p = \tilde{h}$). Additive or interaction terms can be in-

cluded (e.g. $mod.p = \tilde{t}ime + c + h$, $mod.p = \tilde{T}ime + I(Time^2)$, $mod.p = \tilde{t}ime * c$). Userspecified temporal covariates in detection probability can also be used:

The *covs* argument is a data frame used to enter discrete- or continuous-valued temporal covariates, and *parms* specifies the parameters to monitor.

There are currently two options for specifying models for the conditional probabilities of type 1 and type 2 encounters (δ), the default $mod.delta=\tilde{}type$ (i.e. $\delta_1 \neq \delta_2$), and $mod.delta=\tilde{}1$ (i.e. $\delta_1 = \delta_2$). The constraint $\delta_1 = \delta_2$ will often be reasonable when type 1 and type 2 encounters arise from a very similar process, such as with left- and right-sided images (see Example). However, when type 1 and type 2 encounters arise from very different processes (e.g. faecal DNA and visual surveys), then specifying $\delta_1 \neq \delta_2$ is likely a model deserving consideration.

There are many additional arguments for specifying the number (*nchains*) and length (*iter*) of chains, including burn-in and adaptive periods. For potential improvements in mixing, the number of "moves" used to update the feasible set of latent encounter histories at each iteration can be user specified (*maxnumbasis*; see Appendix S1). The default priors are "uninformative", but user-specified priors can be used for each parameter. Initial values can be automatically generated or user specified for each parameter.

The function multimarkCJS() works in exactly the same fashion, with the only notable difference being specification of models for ϕ (in addition to p and δ). Al-

though CJS-specific data are not included with multimark, data can be simulated

using the simdataCJS() function (or simdataClosed() for closed populations):

```
> CJSdata <- simdataCJS(N=100,noccas=7,pbeta=-0.25,phibeta=1,delta_1=0.2,
+ delta_2=0.5,alpha=0.5,sigma2_zphi=0.25,data.type="sometimes")
> Enc.Mat <- CJSdata$Enc.Mat
> head(Enc.Mat)
```

```
[,1] [,2] [,3] [,4] [,5] [,6] [,7]
[1,]
         2
                    4
                          0
                                0
               0
                                      0
[2,]
         1
                    0
                          0
                                0
                                           0
[3,]
                                           0
         1
              0
                    0
                          0
                                0
                                      0
[4,]
               3
         4
                    0
                          0
                                0
                                           0
[5,]
               0
                    0
                          0
                                0
                                      0
                                           0
         1
                                2
                                      2
                                           4
[6,]
                          0
```

```
> CJSsetup <- processdata(Enc.Mat=Enc.Mat,data.type="sometimes")</pre>
```

```
> CJS.dot.h <- multimarkCJS(mms=CJSsetup,</pre>
```

```
+ mod.p=~1,mod.delta=~type,mod.phi=~h,
+ parms=c("pbeta","delta","alpha","phibeta","sigma2_zphi"),
+ nchains=2,iter=45000,burnin=5000)
```

> summary(CJS.dot.h\$mcmc)

```
Iterations = 5001:45000
Thinning interval = 1
Number of chains = 2
Sample size per chain = 40000
```

1. Empirical mean and standard deviation for each variable, plus standard error of the mean:

```
SD Naive SE Time-series SE
pbeta[(Intercept)]
                     -0.1454 0.13339 0.0004716
                                                      0.004308
phibeta[(Intercept)] 1.3497 0.27133 0.0009593
                                                      0.010345
alpha
                      0.4120 0.11551 0.0004084
                                                      0.003051
                      0.0296 0.05728 0.0002025
sigma2_zphi
                                                      0.003003
delta_1
                      0.1964 0.04481 0.0001584
                                                      0.001330
delta_2
                      0.6095 0.05178 0.0001831
                                                      0.001139
```

2. Quantiles for each variable:

```
2.5%
                                       25%
                                                 50%
                                                           75%
                                                                97.5%
pbeta[(Intercept)]
                      -0.395050
                                 -0.238821 -0.14899 -0.05648 0.1237
phibeta[(Intercept)]
                       0.911421
                                  1.156246
                                             1.31792
                                                      1.50971 1.9680
alpha
                       0.206701
                                  0.328765
                                             0.40488
                                                      0.48814 0.6552
                       0.002641
                                             0.01301
sigma2_zphi
                                  0.006682
                                                      0.02896 0.1638
delta_1
                       0.114693
                                  0.164896
                                             0.19448
                                                      0.22568 0.2885
                       0.505645
                                  0.575027
                                             0.61053
                                                      0.64510 0.7080
delta_2
```

An additional feature for multimarkCJS() is simple specification of "age" and cohort effects for $p \pmod{p}$ age and mod.p = `cohort) and $\phi \pmod{phi} = \text{`age}$ and mod.phi = `cohort), which can be useful for investigating structure related to time since first capture and time of initial capture, respectively. These variables by default include a level for each unique age or cohort, but they can be binned to reduce the number of levels using additional arguments.

66 3.2 Single mark type

For conventional capture-recapture data consisting of a single mark type, encounter 267 histories are formatted the same way but now consist solely of 1's (detections) and 268 0's (non-detections). The package currently includes the functions markCJS() and 269 markClosed() for fitting conventional CJS and closed population models, respec-270 tively. These functions are essentially wrappers that "trick" multimarkCJS() and 271 multimarkClosed() to fit models with a single mark type. The functions simdat-272 aCJS() and simdataClosed() can also be used to simulate encounter history data 273 with a single mark type by setting the arguments $delta_1=1$ and $delta_2=0$. For 274 example, to simulate CJS data and fit a model with constant detection probability 275 and individual heterogeneity in survival:

There are fewer arguments for markCJS() and markClosed() because there is only one mark type (e.g. the arguments mms and mod.delta are no longer necessary), but the remaining arguments are specified exactly as for multimarkCJS() and multimarkClosed().

$_{281}$ 3.3 Further analysis

While the coda package can be used to summarize, plot, and assess convergence of MCMC samples from markClosed(), multimarkClosed(), markCJS(), and multimarkCJS(), several additional functions are available for further analysis of model output. Because link functions are used for p and ϕ , the functions getprobsClosed() and getprobsCJS() provide estimates on the real scale. For example, we can compare the probabilities of capture (p) and recapture (c) when there is a behavioural response to first capture (i.e. $mod.p=\tilde{\ }c)$:

```
> bobcat.c <- multimarkClosed(mms=bobcatsetup,mod.p=~c)
> pc <- getprobsClosed(bobcat.c)
> summary(pc[,c("p[1]","c[2]")])

Iterations = 2001:12000
Thinning interval = 1
Number of chains = 1
Sample size per chain = 10000
```

1. Empirical mean and standard deviation for each variable, plus standard error of the mean:

```
Mean SD Naive SE Time-series SE p[1] 0.1406 0.05481 0.0005481 0.004992 c[2] 0.2510 0.05481 0.0005481 0.004678
```

2. Quantiles for each variable:

```
2.5% 25% 50% 75% 97.5% p[1] 0.05674 0.09906 0.1338 0.1750 0.2673 c[2] 0.15111 0.21231 0.2487 0.2876 0.3642
```

Here, p[1] and c[2] refer to the probabilities of capture and recapture at times t = 1 and t = 2, respectively.

Based on Barker & Link (2013), Bayesian multimodel inference using reversible jump MCMC is implemented through the functions markClosed(), multimarkClosed(), markCJS(), and multimarkCJS(). Using this approach, models are first run individually and a Gibbs sampler explores the model space using the individual model MCMC output. All that must be provided to the multimodel inference functions is a list containing the output from at least two models. The models must have the same number and length of MCMC chains, and all model parameters must be monitored (this is accomplished by setting parms="all"):

The list bobcat.M includes RJMCMC output (bobcat.M\$rjmcmc) for parameters common to all models (which can be specified using the argument monparms) and

posterior model probabilities (bobcat.M\$pos.prob). Other arguments for multimodelClosed() and multimodelCJS() include prior model probabilities (modprior) and user-specified proposal distributions for moving between models.

$_{\scriptscriptstyle 14}$ 4 Example

I will now provide results from a new closed population analysis of the bobcat data performed in multimark. Previous analyses of these data include McClintock et al. 306 (2013), who performed an integrated analysis but for a limited model set that did not 307 include behavioural or individual effects, and Alonso et al. (2015), who performed 308 standard single-sided analyses that could not investigate behavioural responses to 309 first capture. Using multimark, it is possible to conduct a more complete analysis 310 using both left- and right-sided encounter histories that includes no effects, temporal 311 effects, behavioural effects, and indivdual effects in detection probability. I also 312 investigated two models for δ ($\delta_1 \neq \delta_2$ and $\delta_1 = \delta_2$) because it is reasonable to 313 suspect that the conditional probabilities of left-sided (type 1) and right-sided (type 314 2) encounters are similar. 315

Fitting all possible additive combinations yielded 16 models using the default "non-informative" priors for multimarkClosed():

$$\beta^{p} \sim \mathcal{N}(0, 1.75)$$

$$\boldsymbol{\delta} \sim \begin{cases} \text{Beta}(1, 1) & \text{if } \delta_{1} = \delta_{2} \\ \text{Dirichlet}(1, 1, 1) & \text{if } \delta_{1} \neq \delta_{2} \end{cases}$$

$$z_{i}^{p} \sim \mathcal{N}(0, \sigma_{z^{p}}^{2})$$

$$\sigma_{z^{p}} \sim \text{half-Cauchy}(25)$$

$$N \propto \frac{1}{N}$$

With 2 chains each consisting of 2 million iterations (with thinning every 20 iterations to reduce memory requirements), the simplest models required 12 mins on a computer 319 running Windows 7 (3.4GHz Intel Core i7, 16GB RAM), while the more complicated 320 models including time variation required at most 2 hrs. These relatively fast run times 321 are attributable to multimark's parallel processing of MCMC algorithms written in 322 the C programming language (Kernighan & Ritchie 1988). Bayesian multimodel 323 inference was performed with *multimodelClosed()* using the default equal prior model 324 weights, where 300000 iterations for each chain required 2.6 hrs. The longer run time 325 for multimodelClosed() owes to the number of models and the RJMCMC algorithm 326 being written entirely in R. 327

Models including a positive behavioural response to first capture accounted for 328 0.51 of the posterior model weight, while models including $\delta_1 = \delta_2$ accounted for 329 0.78 of posterior model weight (Table 3). Model-averaged posterior modes were N=330 35 (Highest Posterior Density Interval: 26-101; Fig. 1) for population abundance, 331 p = 0.15 (HPDI: 0.04-0.27) for capture probability, and c = 0.21 (HPDI: 0.07-0.33) 332 for recapture probability. With $\delta_1 = \delta_2 = 0.41$ (HPDI: 0.30-0.50) based on the 333 model with the highest posterior probability, both-sided encounters were relatively 334 infrequent for these data $(1 - \delta_1 - \delta_2 = 0.18; \text{HPDI: } 0.00\text{-}0.39).$ 335

For comparision, I performed conventional left- and right-sided analyses for these data using markClosed() and multimodelClosed(). Because models for δ and behavioural response do not apply, the candidate model set was limited to $mod.p=^{\sim}1$, $mod.p=^{\sim}time$, $mod.p=^{\sim}h$, and $mod.p=^{\sim}time+h$ for these single-sided analyses. As before, the default "non-informative" priors were used, and the length and number of chains, burn-in periods, and adaptive periods were also the same. For the left-side

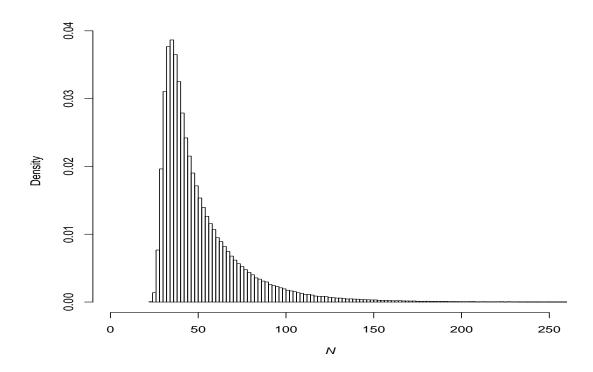


Figure 1. Model-averaged posterior distribution of population abundance (N) for the bobcat data.

Model	PMM	N	HPDI	ESS	GRB
$p(^{\sim}c)delta(^{\sim}1)$	0.30	38	27-91	38944	1.00
$p(^{\sim}1)delta(^{\sim}1)$	0.22	33	26-46	54696	1.00
$p(\tilde{h})delta(\tilde{1})$	0.16	46	29-114	11685	1.00
$p(\tilde{c} + h)delta(\tilde{1})$	0.09	50	29 - 145	18544	1.00
$p(\ c)delta(\ type)$	0.09	38	27-90	35054	1.00
$p(^{\sim}1)delta(^{\sim}type)$	0.06	33	26-46	53961	1.00
$p(\tilde{h})delta(\tilde{type})$	0.05	48	29-113	12099	1.00
$p(\tilde{c} + h)delta(\tilde{t}ype)$	0.03	51	29-146	17276	1.00
$p(\tilde{time} + h)delta(\tilde{1})$	0.00	47	28-115	14414	1.00
$p(^c + time + h)delta(^1)$	0.00	45	28-116	21473	1.00
$p(\tilde{time})delta(\tilde{1})$	0.00	33	26 - 45	47781	1.00
$p(^c + time)delta(^1)$	0.00	33	25 - 78	35169	1.00
$p(\tilde{time} + h)delta(\tilde{type})$	0.00	50	29-118	13882	1.00
$p(^c + time + h)delta(^type)$	0.00	46	27 - 115	21337	1.00
$p(\tilde{time})delta(\tilde{type})$	0.00	33	26 - 45	49425	1.00
p(~c + time)delta(~type)	0.00	32	25-78	35360	1.00

Table 3. Posterior model probabilities (PMM) and abundance estimates for the bobcat data. Summaries include posterior modes (N), 95% highest posterior density intervals (HPDI), effective sample sizes (ESS), and Gelman-Rubin-Brooks diagnostics (GRB) for N. Models for detection probability (p) included no effects (~1), behavioural effects (~c), time effects (~time), and individual effects (~h). Models for the conditional probability of a left- or right-sided encounter (delta) included $\delta_1 = \delta_2$ (~1) and $\delta_1 \neq \delta_2$ (~type).

analysis, the constant detection probability model accounted for 0.95 of the posterior model weight, while the individual heterogeneity model accounted for 0.04 of posterior model weight. Model-averaged posterior modes were N=32 (HPDI: 24-52) for population abundance and p=0.12 (HPDI: 0.07-0.19) for capture probability. For the right-side analysis, the constant detection probability model accounted for 0.6 of the posterior model weight and the individual heterogeneity model accounted for 0.39 of posterior model weight. Model-averaged posterior modes were N=33(HPDI: 23-85) for population abundance and p=0.12 (HPDI: 0.04-0.19) for capture

probability. These conflicting results demonstrate the unenviable position one can of-350 ten find oneself when conducting separate analyses for different mark types from the 351 same population. One may be tempted to choose the "most precise" estimate based 352 on the left-side analysis, but the integrated analysis suggests this would considerably 353 underestimate the uncertainty about N. Choosing the "more conservative" rightsided results or averaging the N estimates from the left- and right-sided analyses would also underestimate the uncertainty about N based on the integrated analysis. 356 This discrepancy is likely attributable to the potential behavioural response to first 357 capture identified by the integrated analysis. 358

5 Discussion

I have described some of the key features of multimark, a new R package for the 360 analysis of capture-recapture data consisting of a single conventional mark or multiple 361 non-invasive marks. The package currently includes open population CJS and closed 362 population models, with functions for derived parameters (e.g. ϕ , p) and multimodel inference. It adds to the growing toolbox of freely-available software for the analysis of non-spatial (e.g. White & Burnham 1999; Choquet et al. 2009; Laake 2013; Laake 365 et al. 2013) and spatial (e.g. Gopalaswamy et al. 2012; Efford 2015) capture-recapture 366 data, but it is the first to combine otherwise irreconcilable encounter histories arising 367 from multiple mark types. Although initially developed for integrated analyses of 368 left- and right-sided images for bilaterally asymmetrical species, the package can be 369 used to jointly analyze data arising from any two types of marks. For example, 370 multimark could be used to integrate an analysis of encounter histories arising from 371 genetic (e.g. hair or faecal) and visual (e.g. photo-ID) detections (sensu Madon et al.

2011; but see Bonner 2013). multimark is also the first capture-recapture software to implement generalized Bayesian multimodel inference based on the RJMCMC algorithm proposed by Barker & Link (2013).

Relative to previous applications using multiple marks (Bonner & Holmberg 2013; 376 McClintock et al. 2013), the relatively fast computation times of the package are attributable to its use of "semi-complete" data likelihoods (King et al. 2015), parallel 378 processing, and MCMC algorithms written in C (instead of R). Because parallel pro-379 cessing relies on the parallel package (R Core Team 2013), first-time Windows and 380 OS X users can expect a firewall pop-up dialog box asking if an R process should 381 accept incoming connections. Memory requirements are minimized by condition-382 ing on the observed encounter histories when identifying the feasible set of latent 383 encounter histories. To facilitate better mixing, multimark improves the MCMC 384 algorithms proposed by Bonner & Holmberg (2013) and McClintock et al. (2013, 385 2014) by avoiding latent encounter history proposals with negative frequencies in a 386 manner that requires no proposal tuning (see Appendix S1 for details). 387

Many potentially desirable extensions to multimark are possible. These include 388 a broader suite of capture-recapture models, such as multi-state and robust design 389 models (e.g. Williams et al. 2002). In addition to individual-level heterogeneity, 390 "random effect" distributions for temporal or user-specified covariates could also be 391 incorporated (e.g. Laake et al. 2013). More general modelling formulae for δ and α 392 would allow additional hypotheses related to detection to be explored. The package 393 could also be extended to accommodate >2 mark types and additional link functions. 394 Although many individual covariates tend to be difficult (or impossible) to observe with non-invasive sampling, some (e.g. sex) may be easily discernable for each mark type. For these cases, it would be relatively straightforward to extend multimark

to accommodate individual covariates. Other extensions include spatially-explicit 398

models (e.g. Royle 2015) and allowing for partial overlap in the sampling periods 399

for each mark type. Practitioners interested in such extensions are encouraged to 400

contact the author. 401

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