

## Module 2: Western Blot

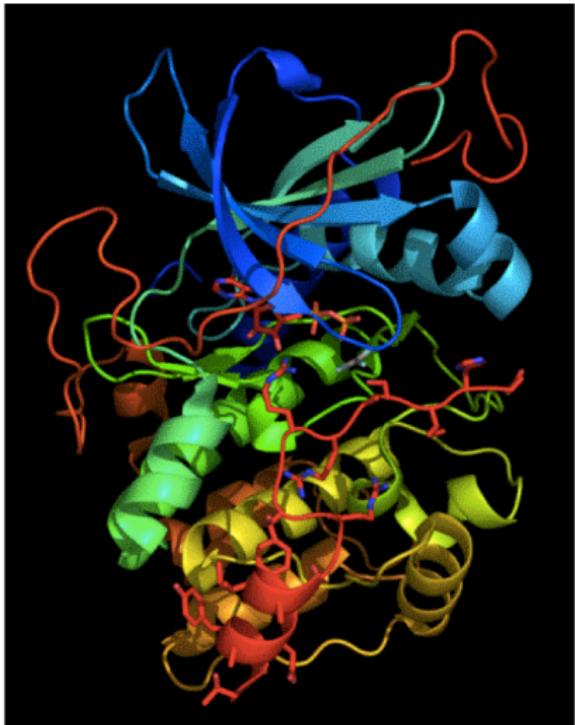
BMES Cell Team

Fall 2020



# Outline

- Protein Basics
- What is a Western Blot?
- Western Blot Protocol
  - BCA Assay
  - SDS Gel Electrophoresis
  - Immunoblotting
- Western Blot Video
- Pipetting basics



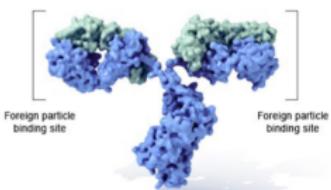
# Proteins

- **Definition:** Proteins are macromolecules made of amino acids.

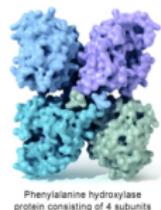
→ Chains of amino acids make up proteins

- Proteins have a wide structural range → large functional range
- Key players in organism's metabolic & regulatory activity

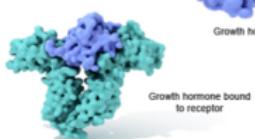
Immunoglobulin G (IgG)



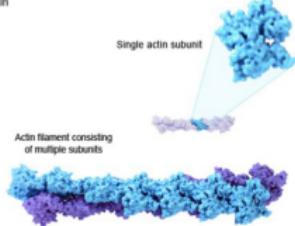
Phenylalanine hydroxylase



Growth hormone



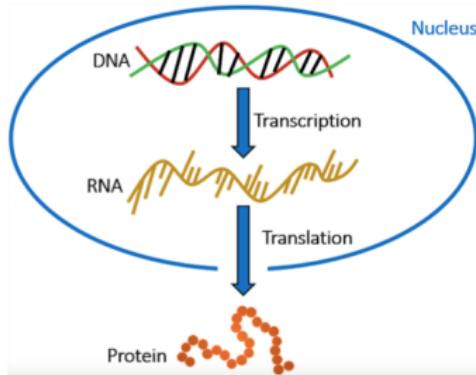
Actin



# What is Western Blotting?

- **Definition:** A **Western Blot** separates and identifies target proteins.

- Two stage procedure confirms protein presence and quantifies target
  - Proteins separated by size through gel electrophoresis
  - Target proteins visualized through immunostaining

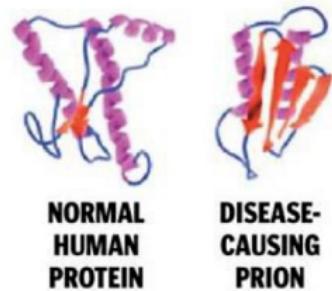


**S N O W**  
**D R O P**

S - SOUTHERN	- DNA	- D
N - NORTHERN	- RNA	- R
O - OOOOOOOO	- OOOO	- O
W - WESTERN	- PROTEIN	- P

# Western Blot Utility

- Understand mechanisms of cell behavior
  - Protein's primary function in organism
- Probe for a specific disease
  - **Viral:**
    - HIV
  - **Bacterial:**
    - Melioidosis
  - **Prion:**
    - Creutzfelt-Jakob disease



# Basic Protocol for a Western Blot

1. Lyse cells and collect proteins

2. BCA Assay

- Calculate sample protein concentration

3. SDS Gel Electrophoresis

- Separate proteins by size

4. Transfer proteins to membrane

5. Immunoblot target proteins

- Antibody Binding

6. Image membrane

## Western Blot Protocol



Sample extraction



Run Gel



Transfer proteins to membrane



Antibodies incubations and washes



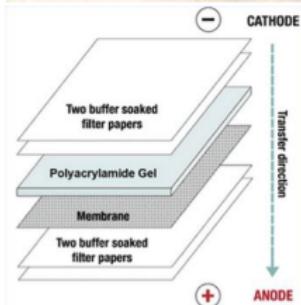
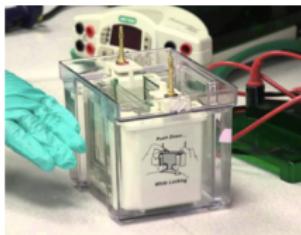
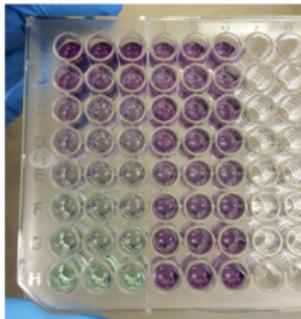
Fail to visualize protein bands



Cry

# Basic Protocol for a Western Blot

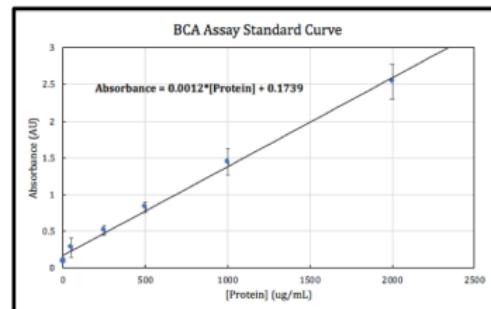
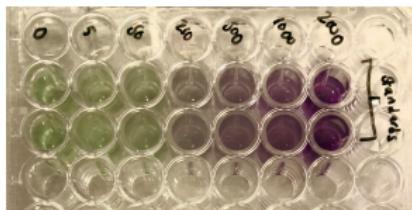
1. Lyse cells and collect proteins
2. BCA Assay
  - Calculate sample protein concentration
3. SDS Gel Electrophoresis
  - Separate proteins by size
4. Transfer proteins from gel to membrane
5. Immunoblot target proteins
  - Antibody Binding
6. Image membrane



# BCA Assay

- **Definition:** A bicinchoninic acid assay (**BCA Assay**) uses absorbance readings to determine total protein concentration in a sample.

- First, create a **standard curve**
  - Concentration of protein in each well is known
- Then, add experimental samples
  - Concentration of protein is unknown
- Using standard curve absorbance readings, correlate protein concentration to absorbance
- Use standard curve to calculate experimental protein concentration



# How does a BCA Assay work?

- Step 1: Biuret Reaction

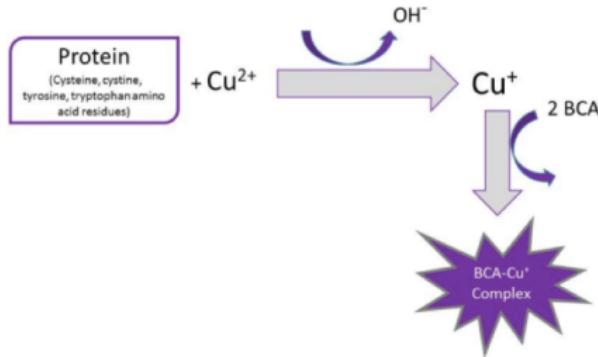
- **Green** cupric Cu<sup>2+</sup> in BCA reagent binds to sample protein  
→ reduction to cuprous Cu<sup>1+</sup>

- Step 2: BCA and Copper Chelation

- 2 BCA molecules bind to Cu<sup>1+</sup>  
→ **purple** chelated complex

- Step 3: Measure Absorbance

- Purple complex absorbs maximally at 562 nm
  - Absorbance  $\propto$  # purple complexes  $\propto$  # peptides

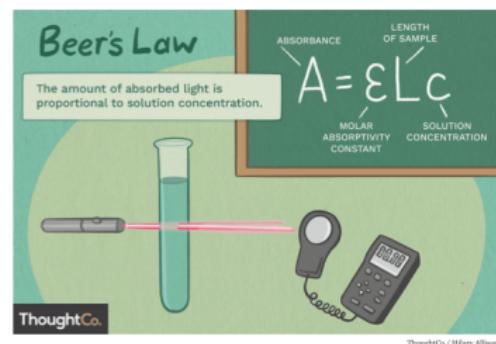


# Beer's Law

- **Definition:** Beer's Law relates a sample's absorbance reading to total protein concentration.

$$A = \epsilon \cdot L \cdot C$$

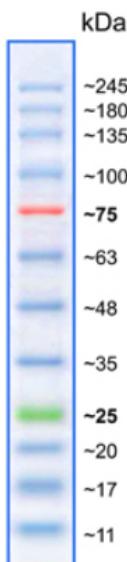
- $A$  = absorbance reading from plate reader
- $\epsilon$  = molar absorptivity constant
- $L$  = path length
- $C$  = protein concentration
  - As  $\epsilon$  and  $L$  are constant, there is a linear relationship between absorbance and protein concentration ( $A \propto C$ )



# SDS-PAGE

- **Definition:** Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (**SDS-PAGE**) separates proteins by size.

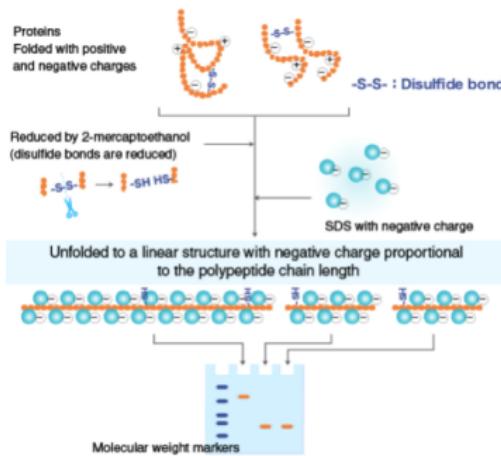
- First, load the protein ladder at the two ends of the well
  - Set of standards that allow us to estimate protein size
  - Dalton (Da) = atomic mass unit
  - $kDa = 1000 \text{ Da}$
- Then, load your sample into the central wells
- Run the gel and use the ladder to estimate protein size



# How does SDS-PAGE work?

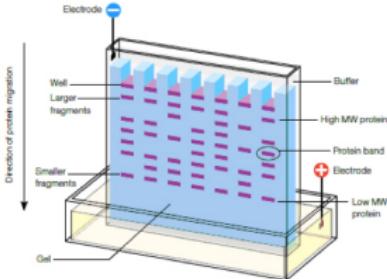
## Key Reagents

- Beta-mercaptopethanol ( $\beta$ ME)
  - Reduces disulfide bonds in protein → disruption covalent bonds
  - Protein linearization
- Sodium Dodecyl Sulfate (SDS)
  - Anionic detergent that binds to protein side chains → disruption noncovalent bonds → protein denaturation
  - Coats denatured protein in uniform negative charge
  - Charge of protein  $\propto$  length of protein



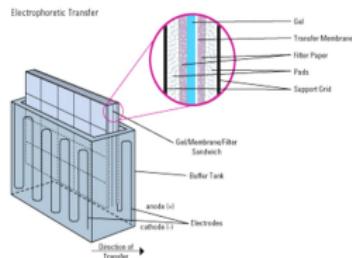
# How does SDS-PAGE work?

- Polyacrylamide (PA)
  - Water soluble polymer
  - 3D networks of polyacrylamide → porous gel
  - Smaller proteins can travel faster through the porous gel
- Gel Electrophoresis (GE)
  - When placed in an electric field, the negatively charged proteins will migrate toward the positive electrode
  - Since smaller proteins can travel faster through a porous gel, loading protein samples into a gel then creating an electric field around the gel **separates proteins by size**



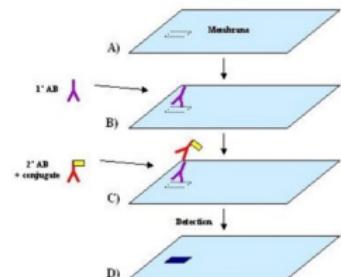
# Protein Transfer

- Polyacrylamide gel → nitrocellulose membrane
  - Antibodies cannot bind to proteins when they are on the gel
  - Must transfer proteins onto a nitrocellulose membrane and retain the gel electrophoresis size sorting
- Transfer proteins using **electroblotting**
  - Align the gel and the membrane
  - Use an electric current to pull negatively charged proteins toward a positively charged anode and onto the membrane

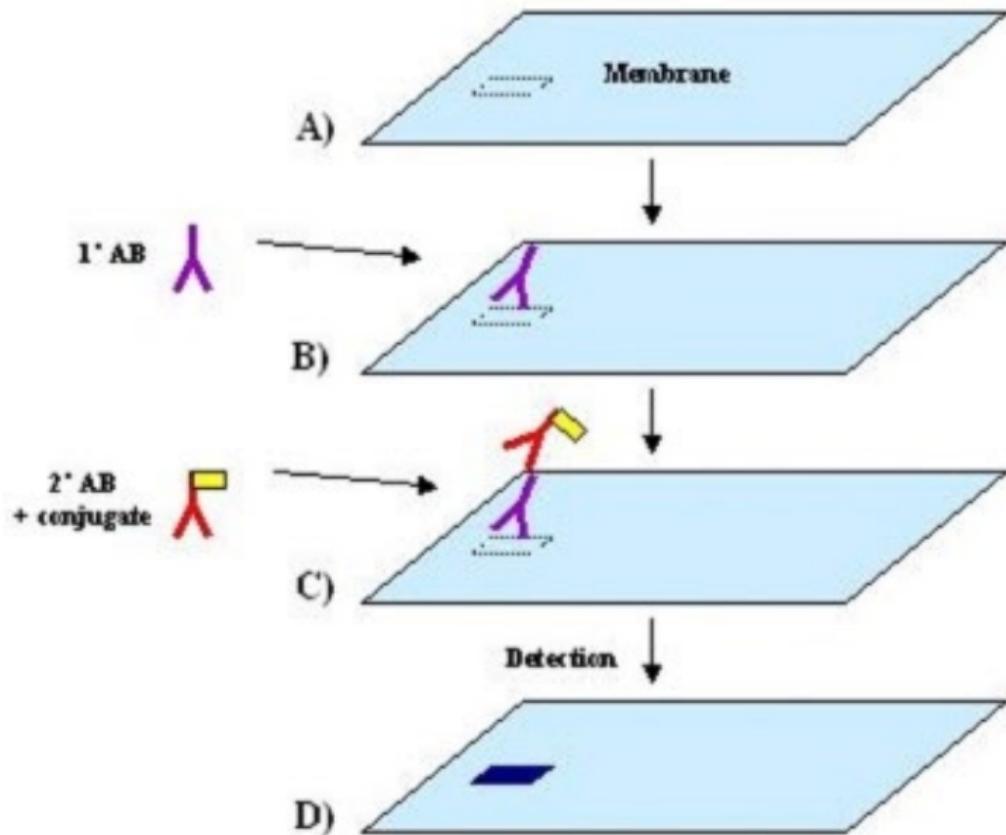


# Immunoblotting (What Makes it a Western Blot!)

- **Definition:** Immunoblotting uses antibodies to identify proteins.
- Antibodies are proteins in the immune system that target specific antigens
- Primary Antibody: binds to target protein
  - Loading Control: Actin (constitutively expressed in all cells)
- Secondary antibody: binds to primary antibody and amplifies the signal
  - Primary and secondary antibodies must be from a different species than the target protein
    - If not, will have non-specific binding



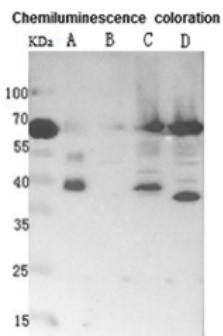
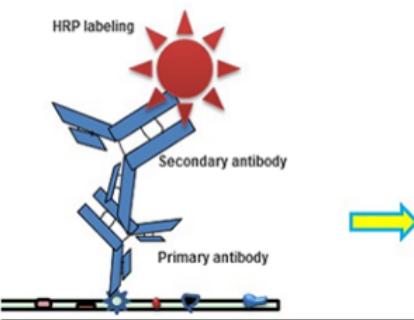
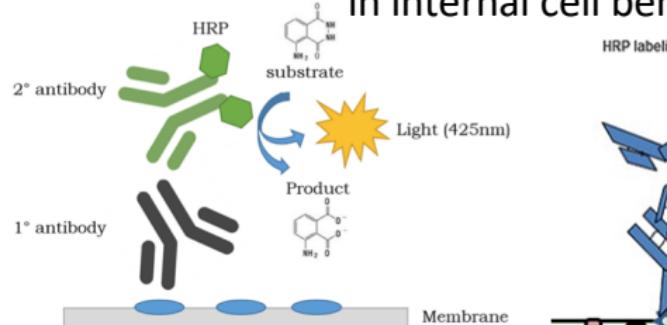
# How does Immunoblotting work?



# Imaging

- Chemiluminescence
  - Chemical reaction between enhanced chemiluminescence (ECL) substrate and horseradish peroxidase (HRP) enzyme conjugated to the secondary antibody
  - Releases energy as light
  - One of the easiest ways to examine proteins involved

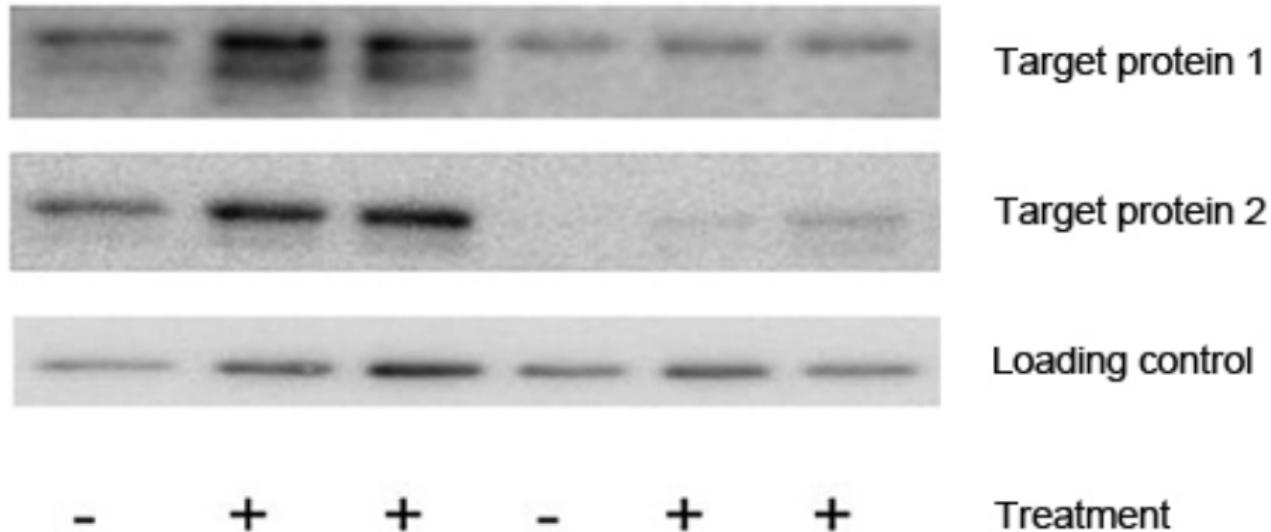
in internal cell behavior and controls



# Interpreting a Western Blot

- Loading control band:
  - should be the same in all samples
  - If loading control is not the same, result is invalid
- Band position on gel:
  - Different sized proteins show up at different heights
  - Larger proteins show up closer to the original well position
- Band intensity:
  - The darker the band, the more protein is present

# Interpreting a Western Blot



- Loading control band is the same in all trials
- Greater target protein intensity in wells 2 and 3

# Western Blot Video



 Cell Signaling  
TECHNOLOGY®  

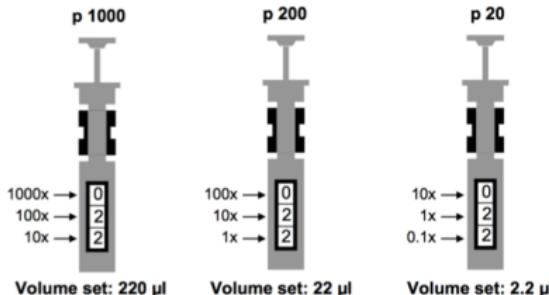

<https://youtu.be/yUstng0npaY>

# Micropipettes

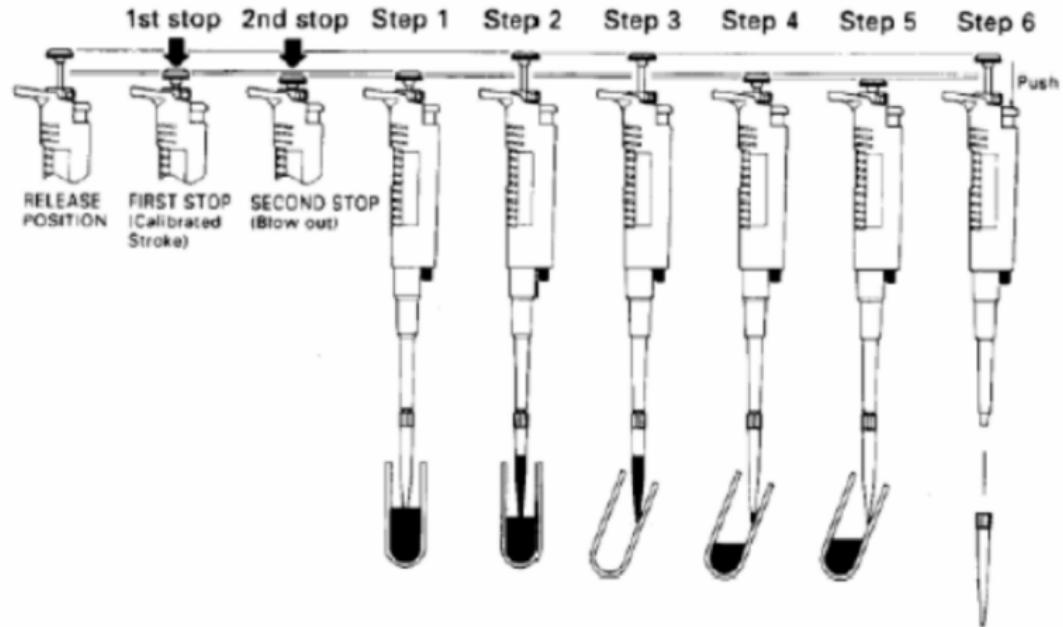
- **Definition:** A **micropipette** is a laboratory instrument used to measure small volumes (on the order of microliters).

- Micropipette sizes
  - P20: 2 – 20  $\mu\text{L}$
  - P200: 20-200  $\mu\text{L}$
  - P1000: 100-1000  $\mu\text{L}$
- Be mindful of which size you are using

Micropipettor: Reading the Volume



# How to use a Micropipette



<https://www.youtube.com/watch?v=TMFeV9h6zEA>