Understanding BEAST2 Results:

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Here's the fun part.

Process the Analysis:

- 1. Open Tracer .
- Drag the log file (cpacific-ctmc-strict.log) into the "Traces" section (or File-> Import Trace File).

Step 1. Look for unstable traces.

Because MCMC is a sequential sampling process, we need to worry that our samples are independent enough for our inferences to be statistically robust.

Look at the ESS column – this is the Effective Sample Size, which is a measure of how independent each sample from the MCMC chain is. We want each one to be large. If the ESS is low then it probably means that the parameter is poorly estimated and we shouldn't rely on that parameter: Perhaps the analysis hasn't run long enough, or perhaps we need to put more weight on that parameter in the analysis so it gets modified more often see: http://beast.bio.ed.ac.uk/increasing-esss

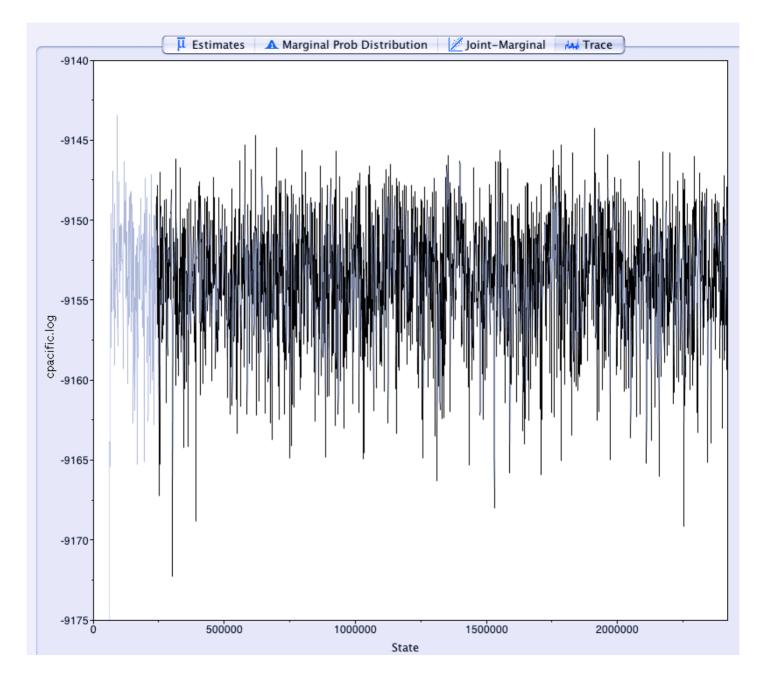
Tracer will highlight anything that's below 200. In the figure below the ESS for gamma_shape_s is slightly low.

[] Look at your data, is there anything with a low ESS?

Statistic	Mean	ESS	T
posterior	-9153.71	1567	R
likelihood	-9163	1669	R
prior	10.027	832	R
treeLikelihood.cpacific	-9163	1669	R
TreeHeight.t:cpacific	0.524	444	R
mutationRate.s:cpacific	1	-	R
gamma_shape_s	2.452	193	R
YuleModel.t:cpacific	12.479	510	R
birthRate.t:cpacific	5.64	2043	R
freqParameter.s:cpac	0.814	315	R
freqParameter.s:cpac	0.186	315	R

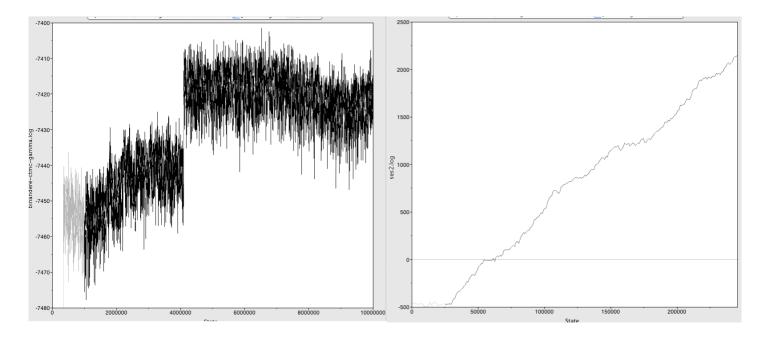
Step 2. Inspect the traces.

Click the Trace tab and go through all the traces of each parameter/statistic from the analysis. You want to make sure they look like this:



... that is, they're consistently estimating a single value (here something around -9153), and not trending up or down.

Here for example, are a couple of bad traces:



... The left one shows a jump in the parameter value. To solve this, increase the burn-in to after the jump (>60,000,000 generations here) and hope you've run the analysis long enough to still get good ESSes in the remainder.

... The right one shows a parameter that's still stabilising. You need to run this analysis a lot longer. Thankfully BEAST2 can resume analyses so you don't need to start from scratch.

[] Are all your traces ok?

[] If not, what are some of the problems you can see?

Step 3. Figure out how much burn-in:

Because of the way MCMC works, we generally start out with terrible parameter estimates based on random guessing and initial settings, and slowly converge towards good estimates (jargon: "reach stationarity").

The samples we take from the period before we reach stationarity is called <code>burn-in</code>. We want to throw these away as they're not good estimates of the real parameter values.

To identify how much burn-in we need, we look at where the traces all stabilise. A common choice is 10% (it should be the default in Tracer). Is this enough for our data? You can change it in the top left of the Tracer window.

[] What burn-in do you think is appropriate here?

Step 4. Check out interesting parameters:

Let's see how old the age of Eastern Polynesian is. Remember that there's a debate between the 'short'

and 'long' chronologies (~800-1000 years vs ~1000-1200 years ago).

Select the mrcatime (EastPolynesian) Parameter and choose the Estimates tab. This is the time to the Most Recent Common Ancestor (MRCA) of the East Polynesian group.

This will show you a nice distribution and details about the age of East Polynesian according to these data and model. The key bits of information are the mean and 95% Highest Posterior Density Interval (HPD) – the Bayesian equivalent of a Confidence Interval.

The mean age of Eastern Polynesian is – in my analysis – about 1283 years, with a 95% H.P.D. of 1153-1414 years.

Your turn. Let's see how old we are estimating Central Pacific to be. The archaeological record suggests that the settlement of Fiji happened around 3200 B.P. (Nunn et al '04).

Select the TreeHeight Parameter and choose the Estimates tab.

[] What's the mean age and 95% HPD of Central Pacific?

[] Is this close to the age of around ~3200 B.P.

View the Trees:

Remember that Bayesian phylogenetics produces a posterior tree distribution which contains lots of trees. We can do a couple of things.

Visualise the entire posterior using Densitree:

A densitree (Bouckaert '10) draws each tree in the posterior on top of each other. It's a great way to visualise the whole posterior at once.

Open the tree in densitree (in the BEAST folder) and then load the tree file that you generated (e.g. cpacific-ctmc-strict.trees).

Central Pacific is quite a reticulate language zone with lots of dialect chains and borrowing between languages (e.g. Walworth '12, Wilson '12, Salisbury '10, Gray et al '10). We should expect to see some conflicting signal where different trees show support for different groupings.

[] What regions of high conflicting signal are there?

Construct a summary "maximum clade credibility tree":

The M.C.C. Tree is a good *single* tree representation of the posterior. Here we calculate the product of all

the clade probabilities on each tree in the posterior to find the 'best' one. See http://beast.bio.ed.ac.uk/summarizing-posterior-trees.

- 1. Open TreeAnnotator
- 2. Set Burn-in Percentage to 10 (percent) or whatever you thought appropriate from the Tracer results.
- 3. Select Input Tree File by clicking 'Choose File' and select your tree file (cpacific-ctmc-strict.trees)
- 4. Select the Output File by clicking 'Choose File' and typing a good file name for the resulting trees in. Personally I'd choose something like "cpacific-ctmc-strict.mcct.trees".
- 5. Make the MCC tree.

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[ ] Successfully make an MCC Tree
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Open this file in FigTree . Because this is a Bayesian analysis with time-dated trees, let's turn on a few options.

- 1. Click the checkbox by scale axis to turn on a time-scale.
- 2. Turn on Node Labels. Open the Node Labels section on the left
- 3. Click the check box
- 4. Change the Display value to posterior.
- 5. Change Sig. Digits to 2 to make it easier to read.

These values give us the number of times each of the subgroups was found in all trees in the posterior. It is the strength of support, in these data, given this model, for that subgrouping. We want high numbers – A value of 1.00 means that this subgrouping was always found. Anything below about 0.7 is weak.

]]	What	subgi	coups	have	very	high	support	(=1.00)
[]	Are	there	subg	roups	with	low	support	(<0.70)

Remember that in our data exploration we saw that Mangareva and Nukuoro in particular had lots of missing data.

[]	Do :	Mangareva	and	Nukuoro	behave	here,	or	should	we	remove	them?
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Which is better?

What visualisation should you prefer? It all depends on how well attested your trees are and how much conflicting signal there is. Densitree shows you everything, but by showing you everything it can be hard to see the forest for the trees. On the other hand, the MCC tree shows the majority signal but loses all the more complex history.

As always, don't rely on one tree. If you go on to use comparative methods or do other subsequent things with your trees, make sure you do it across **all** the trees in the posterior and not just one.

Your Turn (if we have time):

Go back to BEAUTi	(remember that yo	ou left it running,	right?) and c	change the clock	k model to a	a relaxed
clock (log normal).						

Make	sure you change the	tracelog an	d treelog	filenames on the	MCMC	tab to something like
сра	cific-ctmc-relaxe	ed.log and c	pacific-ct	mc-relaxed.t	rees .	
[]	Generate an XML	for the rela	axed clock	analysis		
[]	Run the relaxed	clock analys	sis			

If you have time, you could try setting up a *covarion* analysis. The Covarion is a nice model that allows each cognate in our data to switch 'on' and 'off' at different points on the tree (Penny et al. '01). It's often found to be the best fitting model for language data in my experience, however, it can be a bit trickier to stabilise (especially the hidden frequencies which control the on/off switching).

To do this, use the BinaryCovarion template in BEAUTi rather than BinaryCTMC, and ask me for tips.