

Package ‘SpectroX’

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Type Package

Title Proteotypic peptide selection and SRM/PRM/HRM panel creation

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Description Select proteotypic peptides based on full proteom analysis MaxQuant 1.6 search results or create SRM/PRM/HRM panel/library from MaxQuant 1.6 search results.

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Encoding UTF-8

LazyData true

Suggests testthat

RoxygenNote 6.0.1

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```
barplotPeptidesPerProtein
```

plot adj. intensity vs peptide (per protein) barplot

Description

plot adj. intensity vs peptide (per protein) barplot

Usage

```
barplotPeptidesPerProtein(df, ptmRegExp = NA, pepLenTrunc = 12,
  pepLabCex = 0.7, rankingMetric = "rankingMetric", ...)
```

Arguments

df	data.frame
ptmRegExp	default NA, higliht modified peptides in red (do not highlight labels)
pepLenTrunc	integer AFADAMEVIPSTLAENAGLNPISTVTELR -> AFADAMEVIPSTLAE..
pepLabCex	default 0.7
rankingMetric	character

Note

No note

References

NA

Examples

```
print("No examples")
```

```
barplotPetideCountPerProtein
```

plot adj. intensity vs peptide (per protein) barplot

Description

plot adj. intensity vs peptide (per protein) barplot

Usage

```
barplotPetideCountPerProtein(spectralLibrary, protLabCex = 0.9,
  acLenTrunc = 12, col = "blue", cex.axis = 1.25, cex.lab = 1.25, ...)
```

Arguments

spectralLibrary	data.frame
protLabCex	default 0.9
acLenTrunc	integer default 12 "SOMEVERYLONGAC" -> "SOMEVERY.."
ptmRegExp	default NA, highlight modified peptides in red (do not highlight labels)

Note

No note

References

NA

Examples

```
print("No examples")
```

```
createComplementaryIsotopeLibrary
```

Create complementary isotope (Arg, Lys H/L) spectral library

Description

Create complementary isotope (Arg, Lys H/L) spectral library

Usage

```
createComplementaryIsotopeLibrary(sl)
```

Arguments

sl	tibble or data frame
----	----------------------

Details

No details

Value

data.frame

Note

No note

References

NA

Examples

```
print("No examples")
```

createLibrarySpectrum	<i>Parse spectrum framgment match information listed in MaxQuant 'Identifications' results</i>
-----------------------	--

Description

Parse spectrum framgment match information listed in MaxQuant 'Identifications' results

Usage

```
createLibrarySpectrum(psm)
```

Arguments

psm	a row of MaxQuant 'Identifications' data.frame
-----	--

Details

No details

Value

list with attributes intensity, ionType, charge, mz, isNL

Note

No note

References

NA

Examples

```
print("No examples")
```

createSpectralLibrary *Create Spectral Library*

Description

Create Spectral Library

Usage

```
createSpectralLibrary(tb, minFragNb = 3, minNbTransitions = 5,  
  maxNbTransitions = 5, minBasePeakFraction = 0, ionTypeFilter = c("a",  
  "b", "x", "y"), includeNL = T, rankingMetric = "adjustedIntensitySum")
```

Arguments

tb	tibble maxQuant spectrum level search results
minFragNb	min frag number default 3 (i.e. b3 and y3 will be kept)
minNbTransitions	minimum number of fragments
minBasePeakFraction	minimum intensity fraction of base peak (most intense kept fragment)
ionTypeFilter	selected ion type default a,b,x,y
includeNL	include neutral loss peaks default TRUE
rankingMetric	character. column name, used for ranking peptide, default "adjustedIntensity-Sum", c("adjustedIntensitySum","psmScore","precApexIntensity")

Details

No details

Value

data.frame

Note

No note

References

NA

Examples

```
print("No examples")
```

digestProteome	<i>Get peptide candidates per protein</i>
----------------	---

Description

Get peptide candidates per protein

Usage

```
digestProteome(proteins, proteaseRegExp = "[KR](?!P)", dispProgressBar = T,  
  peptideLengthRange = c(6, 21), nbMisleavages = 0,  
  exclusivePeptides = T, trimAC = T)
```

Arguments

proteins	list
proteaseRegExp	character '[KR](?!P)' - trypsin
dispProgressBar	T/F
peptideLengthRange	integer vector default c(6,21)
nbMisleavages	0
exclusivePeptides	TRUE/FALSE get exclusive peptides only i.e. peptides mapping to a single protein. default T
trimAC	TRUE/FALSE default TRUE splP62258 I433E_HUMAN -> P62258

Value

data.frame peptide,protein

Note

No note

References

NA

Examples

```
print("No examples")
```

getCMDLineOptions	<i>Define and User Specified Command Line Options</i>
-------------------	---

Description

Define and User Specified Command Line Options

Usage

```
getCMDLineOptions(version = version)
```

Arguments

version	SpectroX version number
---------	-------------------------

Details

No details

Value

list() cmd line options

Note

No note

References

NA

Examples

```
print("No examples")
```

getEmpiricalIRT	<i>Add iRT metric to data.frame</i>
-----------------	-------------------------------------

Description

Add iRT metric to data.frame

Usage

```
getEmpiricalIRT(tb, fit)
```

Arguments

tb	tibble containing colum labelled "Retention time"
fit	lm object

Details

No details

Value

vector of normalised rt (empirical irt)

Note

No note

References

Using iRT, a normalized retention time for more targeted measurement of peptides, Escher et al. (2012), <http://www.ncbi.nlm.nih.gov/pubmed/22577012>

Examples

```
print("No examples")
```

getFragmentSequence	<i>Get Fragment Sub Sequenece</i>
---------------------	-----------------------------------

Description

Get Fragment Sub Sequenece

Usage

```
getFragmentSequence(peptide = peptide, ionType = ionType,  
  fragmentNb = fragmentNb)
```

Arguments

peptide	character string
ionType	c("a","b","x","y")
fragmentNb	integer

Details

No details

Value

character string

Note

No note

References

NA

Examples

```
print("No examples")
```

getIRTModel	<i>Get linear model predicting iRT as a function retention time (column name "rt")</i>
-------------	--

Description

Get linear model predicting iRT as a function retention time (column name "rt")

Usage

```
getIRTModel(tb)
```

Arguments

tb tibble including column "Retention time" and "Sequence"

Details

Uses Robust Linear Regression. Retention times of at least 3 iRT unique peptides needs to be listed in the input tibble

Value

list including fit rlm object, data.frame peptide,rt,irtRef

Note

No note

References

Using iRT, a normalized retention time for more targeted measurement of peptides, Escher et al. (2012), <http://www.ncbi.nlm.nih.gov/pubmed/22577012>

Examples

```
print("No examples")
```

`getPeptides`*Get Mz Shift of complementary peptide/fragment ion*

Description

Get Mz Shift of complementary peptide/fragment ion

Usage

```
getPeptides(proteinSeq, proteaseRegExp = "[KR](?!P)", nbMiscleavages = 0)
```

Arguments

<code>proteinSeq</code>	protein sequence
<code>proteaseRegExp</code>	protease Regular Expression
<code>nbMiscleavages</code>	default 0
<code>aaSeq</code>	character string
<code>charge</code>	default 1
<code>isHeavy</code>	c(T,F)
<code>annotSpectrum</code>	Annotated Spectrum object

Details

Calculate mass shift of complimentary spectrum.

- PETIDEK (light) -> 8.014199
- PETIDEK (heavy) -> -8.014199
- PETIDER (light) -> 10.008269
- PETIDER (heavy) -> -10.008269

No details

No details

Value

numeric

`annotSpectrum` Annotated Spectrum object

vector of peptides

Note

No note

No note

No note

References

NA

NA

Examples

```
print("No examples")
Created complimentary Heavy/Light annotated spectrum with updates precursor and fragment m/z values.
print("No examples")
Digest protein
print("No examples")
```

`getSearchedModifications`*Get list of variable modifications considered by MaxQuant*

Description

Get list of variable modifications considered by MaxQuant

Usage

```
getSearchedModifications(tb, ignoreArgLysIsoLabel = T)
```

Arguments

<code>ignoreArgLysIsoLabel</code>	default T Do not consider Arg Lys C,N heavy isotope label
<code>df</code>	tibble or data.frame

Details

No details

Value

character vector of modification names (does not return "Arg10", "Lys8" labels)

Note

No note

References

NA

Examples

```
print("No examples")
```

getUserOptions	<i>check and return User Specified Command Line Options</i>
----------------	---

Description

check and return User Specified Command Line Options

Usage

```
getUserOptions(cmdlineOptions)
```

Arguments

list()	command line options
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Details

No details

Value

list() user options

Note

No note

References

NA

Examples

```
print("No examples")
```

parseMaxQuantMSMS	<i>Parse MaxQuant msms.txt</i>
-------------------	--------------------------------

Description

Parse MaxQuant msms.txt

Usage

```
parseMaxQuantMSMS(file, pepCutoff = 0.05, targetPeptides = NA,  
  targetProteins = NA, filterContaminants = T,  
  contaminantRegExp = "^CON_", selectedPTMRegExp = NA,  
  filterNonExclusivePeptides = T, pepLength = c(1, Inf),  
  chargeState = c(1, Inf), label = NA, maxMissedCleavages = 0,  
  keepBestSpectrumOnly = T, requiredPepSeqRegExp = ".", ...)
```

Arguments

<code>file</code>	<code>path</code>
<code>pepCutoff</code>	numeric default 0.05
<code>targetPeptides</code>	character default NA
<code>targetProteins</code>	character default NA
<code>filterContaminants</code>	TRUE
<code>contaminantRegExp</code>	<code>'^CON_'</code>
<code>selectedPTMRegExp</code>	NA
<code>filterNonExclusivePeptides</code>	default TRUE
<code>pepLength</code>	default [1,Inf)
<code>chargeState</code>	default [1,Inf)
<code>label</code>	(Arg10 and Lys8 only filter) options NA (deafault,no filter), 'light','heavy'
<code>keepBestSpectrumOnly</code>	keep top-scoring spectrum only, per peptidoform, default TRUE
<code>requiredSequenceRegExp</code>	dafault '.' (no filter)

Details

No details

Value

data.frame of maxQuant psm level results

Note

No note

References

NA

Examples

```
print("No examples")
```

parseRange	<i>Parse range in format 2:4 and return integer vector. Inthis case 2,3,4</i>
------------	---

Description

Parse range in format 2:4 and return integer vector. Inthis case 2,3,4

Usage

```
parseRange(rangeStr)
```

Arguments

rangeStr	e.g. '2:4'
----------	------------

Details

No details

Value

vectot of integers

Note

No note

References

NA

Examples

```
print("No examples")
```

plotIRTCalibration	<i>Plot IRT calibration curve</i>
--------------------	-----------------------------------

Description

Plot IRT calibration curve

Usage

```
plotIRTCalibration(irtModel, cex = 1.5, col = "blue", pch = 19, ...)
```

Arguments

irtModel	list
----------	------

Details

No details

Note

No note

References

NA

Examples

```
print("No examples")
```

```
proteotypicPeptideExport  
# Export top X peptide per protein (ordered by 'adjustedIntensitySum')  
# and add theoretical peptides ranked by length if fewer than X peptides  
# were identified by MaxQuant
```

Description

Export top X peptide per protein (ordered by 'adjustedIntensitySum') and add theoretical peptides ranked by length if fewer than X peptides were identified by MaxQuant

Usage

```
proteotypicPeptideExport(spectralLibrary, theoPeptides = NA,  
  targetProteins = unique(spectralLibrary$protein),  
  nbPeptidesPerProtein = 5, outFile = paste0(tempdir(), "/tmp.xls"))
```

Arguments

spectralLibrary	tibble
theoPeptides	data.frame 'peptide' 'protein' 'length'
targetProteins	list of protein accession numbers e.g. P62258. default all proteins in spectralLibrary
nbPeptidesPerProtein	number of peptides per protein to be exported
outFile	path to output file default tmp.xls in tmp dir

Note

No note

References

NA

Examples

```
print("No examples")
```

skylineExport	<i>Write skyline compatible xls file</i>
---------------	--

Description

Write skyline compatible xls file

Usage

```
skylineExport(sl, file)
```

Arguments

sl	tibble or data frame
----	----------------------

Details

No details

Note

No note

References

NA

Examples

```
print("No examples")
```

spectroDiveExport	<i>Write SpectroDive compatible xls file</i>
-------------------	--

Description

Write SpectroDive compatible xls file

Usage

```
spectroDiveExport(sl, file)
```

Arguments

sl	tibble or data frame
----	----------------------

Details

No details

Note

No note

References

NA

Examples

```
print("No examples")
```

spectronautExport	<i>Write Spectronaut compatible xls file</i>
-------------------	--

Description

Write Spectronaut compatible xls file

Usage

```
spectronautExport(sl, file)
```

Arguments

sl	tibble or data frame
----	----------------------

Details

No details

Note

No note

References

NA

Examples

```
print("No examples")
```

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