

# Package ‘SpectroX’

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**Type** Package

**Title** Proteotypic peptide selection and SRM/PRM/HRM panel creation

**Version** 2.0

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**Description** Select proteotypic peptides based on full proteom analysis MaxQuant 1.6 search results or create SRM/PRM/HRM panel/library from MaxQuant 1.6 search results.

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**Suggests** testthat

**RoxygenNote** 6.0.1

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`barplotPeptidesPerProtein`*plot adj. intensity vs peptide (per protein) barplot*

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**Description**

plot adj. intensity vs peptide (per protein) barplot

**Usage**

```
barplotPeptidesPerProtein(df, ptmRegExp = NA, pepLenTrunc = 12,  
  pepLabCex = 0.7, ...)
```

**Arguments**

<code>df</code>	data.frame
<code>ptmRegExp</code>	default NA, highlight modified peptides in red (do not highlight labels)
<code>pepLenTrunc</code>	integer AFADAMEVIPSTLAENAGLNPISTVTELRL -> AFADAMEVIPSTLAE..
<code>pepLabCex</code>	default 0.7

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

`createComplementaryIsotopeLibrary`*Create complementary isotope (Arg, Lys H/L) spectral library*

---

**Description**

Create complementary isotope (Arg, Lys H/L) spectral library

**Usage**

```
createComplementaryIsotopeLibrary(sl)
```

**Arguments**

<code>sl</code>	tibble or data frame
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**Details**

No details

**Value**

data.frame

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

createLibrarySpectrum *Parse spectrum framgment match information listed in MaxQuant 'Identifications' results*

---

**Description**

Parse spectrum framgment match information listed in MaxQuant 'Identifications' results

**Usage**

```
createLibrarySpectrum(psm)
```

**Arguments**

psm                      a row of MaxQuant 'Identifications' data.frame

**Details**

No details

**Value**

list with attributes intensity, ionType, charge, mz, isNL

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

createSpectralLibrary *Create Spectral Library*

---

## Description

Create Spectral Library

## Usage

```
createSpectralLibrary(tb, minFragNb = 3, minNbTransitions = 5,  
  maxNbTransitions = 5, minBasePeakFraction = 0, ionTypeFilter = c("a",  
  "b", "x", "y"), includeNL = T)
```

## Arguments

tb	tibble maxQuant spectrum level search results
minFragNb	min frag number default 3 (i.e. b3 and y3 will be kept)
minNbTransitions	minimum number of fragments
minBasePeakFraction	minimum intensity fraction of base peak (most intense kept fragment)
ionTypeFilter	selected ion type default a,b,x,y
includeNL	include neutral loss peaks default TRUE

## Details

No details

## Value

data.frame

## Note

No note

## References

NA

## Examples

```
print("No examples")
```

---

digestProteome	<i>Get peptide candidates per protein</i>
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---

**Description**

Get peptide candidates per protein

**Usage**

```
digestProteome(proteins, proteaseRegExp = "[KR](?!P)", dispProgressBar = T,  
  peptideLengthRange = c(6, 21), nbMisleavages = 0,  
  exclusivePeptides = T, trimAC = T)
```

**Arguments**

proteins	list
proteaseRegExp	character '[KR](?!P)' - trypsin
dispProgressBar	T/F
peptideLengthRange	integer vector default c(6,21)
nbMisleavages	0
exclusivePeptides	TRUE/FALSE get exclusive peptides only i.e. peptides mapping to a single protein. default T
trimAC	TRUE/FALSE default TRUE splP62258 I433E_HUMAN -> P62258

**Value**

data.frame peptide,protein

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

getCMDLineOptions	<i>Define and User Specified Command Line Options</i>
-------------------	---

---

**Description**

Define and User Specified Command Line Options

**Usage**

```
getCMDLineOptions(version = version)
```

**Arguments**

version	SpectroX version number
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**Details**

No details

**Value**

list() cmd line options

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

getEmpiricalIRT	<i>Add iRT metric to data.frame</i>
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---

**Description**

Add iRT metric to data.frame

**Usage**

```
getEmpiricalIRT(tb, fit)
```

**Arguments**

tb	tibble containing colum labelled "Retention time"
fit	lm object

**Details**

No details

**Value**

vector of normalised rt (empirical irt)

**Note**

No note

**References**

Using iRT, a normalized retention time for more targeted measurement of peptides, Escher et al. (2012), <http://www.ncbi.nlm.nih.gov/pubmed/22577012>

**Examples**

```
print("No examples")
```

---

getFragmentSequence	<i>Get Fragment Sub Sequence</i>
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---

**Description**

Get Fragment Sub Sequence

**Usage**

```
getFragmentSequence(peptide = peptide, ionType = ionType,  
  fragmentNb = fragmentNb)
```

**Arguments**

peptide	character string
ionType	c("a","b","x","y")
fragmentNb	integer

**Details**

No details

**Value**

character string

**Note**

No note

## References

NA

## Examples

```
print("No examples")
```

---

getIRTModel	<i>Get linear model predicting iRT as a function retention time (column name "rt")</i>
-------------	--

---

## Description

Get linear model predicting iRT as a function retention time (column name "rt")

## Usage

```
getIRTModel(tb)
```

## Arguments

tb                      tibble including column "Retention time" and "Sequence"

## Details

Uses Robust Linear Regression. Retention times of at least 3 iRT unique peptides needs to be listed in the input tibble

## Value

list including fit rlm object, data.frame peptide,rt,irtRef

## Note

No note

## References

Using iRT, a normalized retention time for more targeted measurement of peptides, Escher et al. (2012), <http://www.ncbi.nlm.nih.gov/pubmed/22577012>

## Examples

```
print("No examples")
```



---

getPeptides	<i>Get Mz Shift of complementary peptide/fragment ion</i>
-------------	---

---

**Description**

Get Mz Shift of complementary peptide/fragment ion

**Usage**

```
getPeptides(proteinSeq, proteaseRegExp = "[KR](?!P)", nbMiscleavages = 0)
```

**Arguments**

proteinSeq	protein sequence
proteaseRegExp	protease Regular Expression
nbMiscleavages	default 0
aaSeq	character string
charge	default 1
isHeavy	c(T,F)
annotSpectrum	Annotated Spectrum object

**Details**

Calculate mass shift of complimentary spectrum.

- PETIDEK (light) -> 8.014199
- PETIDEK (heavy) -> -8.014199
- PETIDER (light) -> 10.008269
- PETIDER (heavy) -> -10.008269

No details

No details

**Value**

numeric

annotSpectrum Annotated Spectrum object

vector of peptides

**Note**

No note

No note

No note

**References**

NA

NA

**Examples**

```
print("No examples")
Created complimentary Heavy/Light annotated spectrum with updates precursor and fragment m/z values.
print("No examples")
Digest protein
print("No examples")
```

---

getSearchedModifications

*Get list of variable modifications considered by MaxQuant*


---

**Description**

Get list of variable modifications considered by MaxQuant

**Usage**

```
getSearchedModifications(tb, ignoreArgLysIsoLabel = T)
```

**Arguments**

ignoreArgLysIsoLabel	default T Do not consider Arg Lys C,N heavy isotope label
df	tibble or data.frame

**Details**

No details

**Value**

character vector of modification names (does not return "Arg10", "Lys8" labels )

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

getUserOptions	<i>check and return User Specified Command Line Options</i>
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---

### Description

check and return User Specified Command Line Options

### Usage

```
getUserOptions(cmdlineOptions)
```

### Arguments

list()	command line options
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### Details

No details

### Value

list() user options

### Note

No note

### References

NA

### Examples

```
print("No examples")
```

---

parseMaxQuantMSMS	<i>Parse MaxQuant msms.txt</i>
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---

### Description

Parse MaxQuant msms.txt

### Usage

```
parseMaxQuantMSMS(file, pepCutoff = 0.05, targetPeptides = NA,
  targetProteins = NA, filterContaminants = T,
  contaminantRegExp = "^CON_", selectedPTMRegExp = NA,
  filterNonExclusivePeptides = T, minPepLength = 0, chargeState = c(1,
  Inf), label = NA, maxMissedCleavages = 0, keepBestSpectrumOnly = T,
  requiredPepSeqRegExp = ".", ...)
```

**Arguments**

file	path
pepCutoff	numeric default 0.05
targetPeptides	character default NA
targetProteins	character default NA
filterContaminants	TRUE
contaminantRegExp	'^CON_'
selectedPTMRegExp	NA
filterNonExclusivePeptides	default TRUE
minPepLength	default 0 minimum peptide length
chargeState	default [1,Inf)
label	(Arg10 and Lys8 only filter) options NA (deafault,no filter), 'light','heavy'
keepBestSpectrumOnly	keep top-scoring spectrum only, per peptidoform, default TRUE
requiredSequenceRegExp	dafault '.' (no filter)

**Details**

No details

**Value**

data.frame of maxQuant psm level results

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

parseRange	<i>Parse range in format 2:4 and return integer vector. In this case 2,3,4</i>
------------	--

---

**Description**

Parse range in format 2:4 and return integer vector. In this case 2,3,4

**Usage**

```
parseRange(rangeStr)
```

**Arguments**

rangeStr	e.g. '2:4'
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**Details**

No details

**Value**

vector of integers

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

plotIRTCalibration	<i>Plot IRT calibration curve</i>
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---

**Description**

Plot IRT calibration curve

**Usage**

```
plotIRTCalibration(irtModel, cex = 1.5, col = "blue", pch = 19, ...)
```

**Arguments**

irtModel	list
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**Details**

No details

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

```
proteotypicPeptideExport
# Export top X peptide per protein (ordered by 'adjustedIntensitySum')
# and add theoretical peptides ranked by length if fewer than X peptides
# were identified by MaxQuant
```

---

**Description**

# Export top X peptide per protein (ordered by 'adjustedIntensitySum') and add theoretical peptides ranked by length if fewer than X peptides were identified by MaxQuant

**Usage**

```
proteotypicPeptideExport(spectralLibrary, theoPeptides = NA,
  targetProteins = unique(spectralLibrary$protein),
  nbPeptidesPerProtein = 5, outFile = paste0(tempdir(), "/tmp.xls"))
```

**Arguments**

```
spectralLibrary      tibble
theoPeptides        data.frame 'peptide' 'protein' 'length'
targetProteins      list of protein accession numbers e.g. P62258. default all proteins in spectralLi-
                    brary
nbPeptidesPerProtein number of peptides per protein to be exported
outFile              path to output file default tmp.xls in tmp dir
```

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

skylineExport	<i>Write skyline compatible xls file</i>
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---

**Description**

Write skyline compatible xls file

**Usage**

```
skylineExport(sl, file)
```

**Arguments**

sl	tibble or data frame
----	----------------------

**Details**

No details

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

spectroDiveExport	<i>Write SpectroDive compatible xls file</i>
-------------------	--

---

**Description**

Write SpectroDive compatible xls file

**Usage**

```
spectroDiveExport(sl, file)
```

**Arguments**

sl	tibble or data frame
----	----------------------

**Details**

No details

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```

---

spectronautExport	<i>Write Spectronaut compatible xls file</i>
-------------------	--

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**Description**

Write Spectronaut compatible xls file

**Usage**

```
spectronautExport(sl, file)
```

**Arguments**

sl	tibble or data frame
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**Details**

No details

**Note**

No note

**References**

NA

**Examples**

```
print("No examples")
```



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