

Engineering cytochromes P450 from ancestral predictions using the novel tool **GRASP**

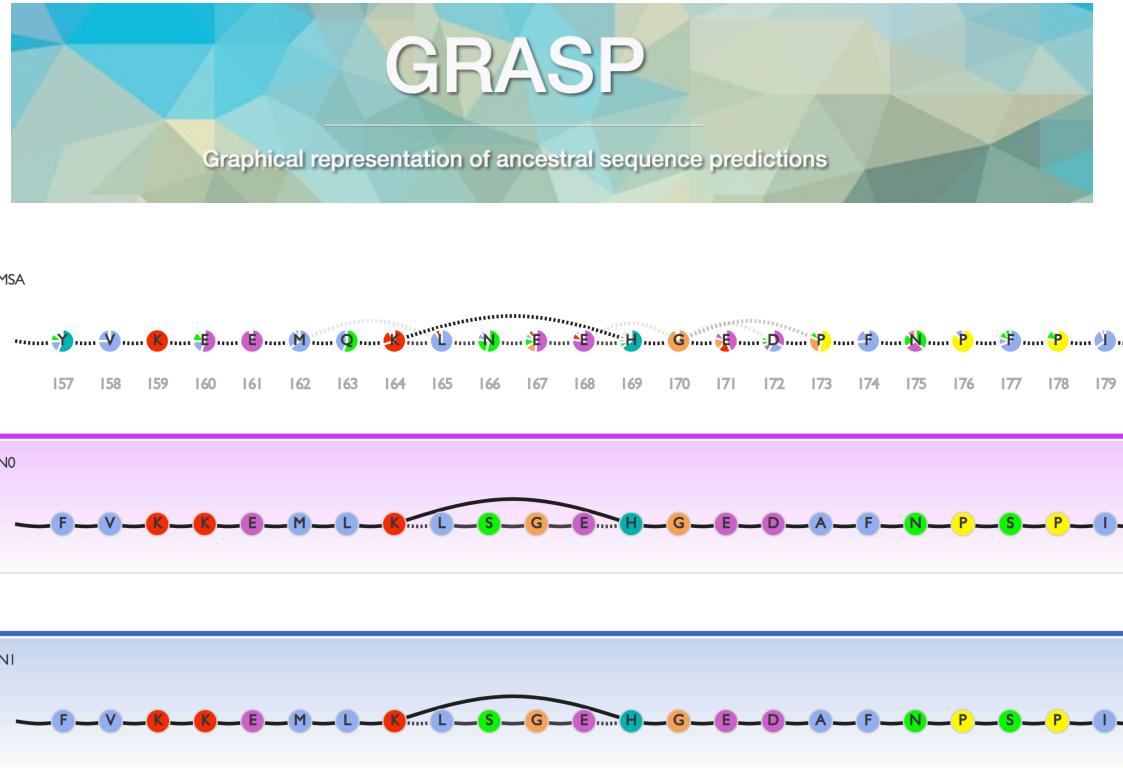
Gabe Foley

School of Chemistry and Molecular Biosciences

The University of Queensland

27/06/2019

Graphical representation of ancestral sequence predictions (GRASP)



- Ancestral sequence reconstruction (ASR) tool designed for large data sets
- Successful reconstruction of cytochromes P450 in collaboration with Liz Gillam at UQ

<http://grasp.scmb.uq.edu.au>

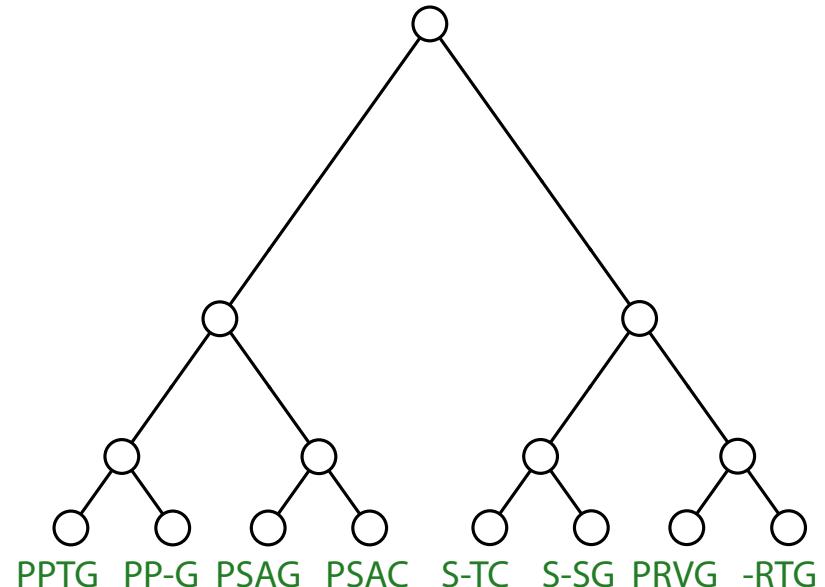
Overview

- *What* is ancestral sequence reconstruction (ASR) ?
- *Why* use it?
- ASR on big data
- How **GRASP** enables big data and extends the reach of ASR



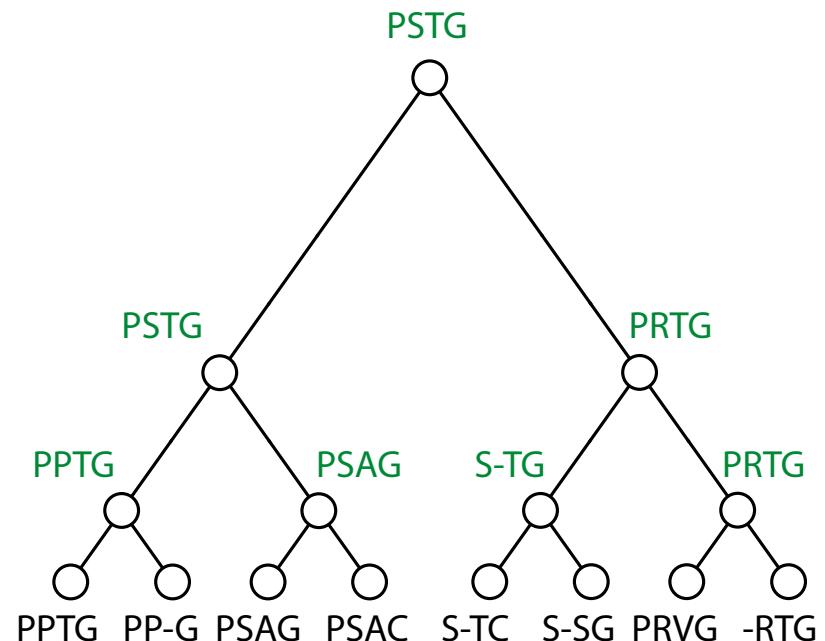
What is ancestral sequence reconstruction?

- Using the information in **modern day biological sequences** to infer what their ancestors looked like



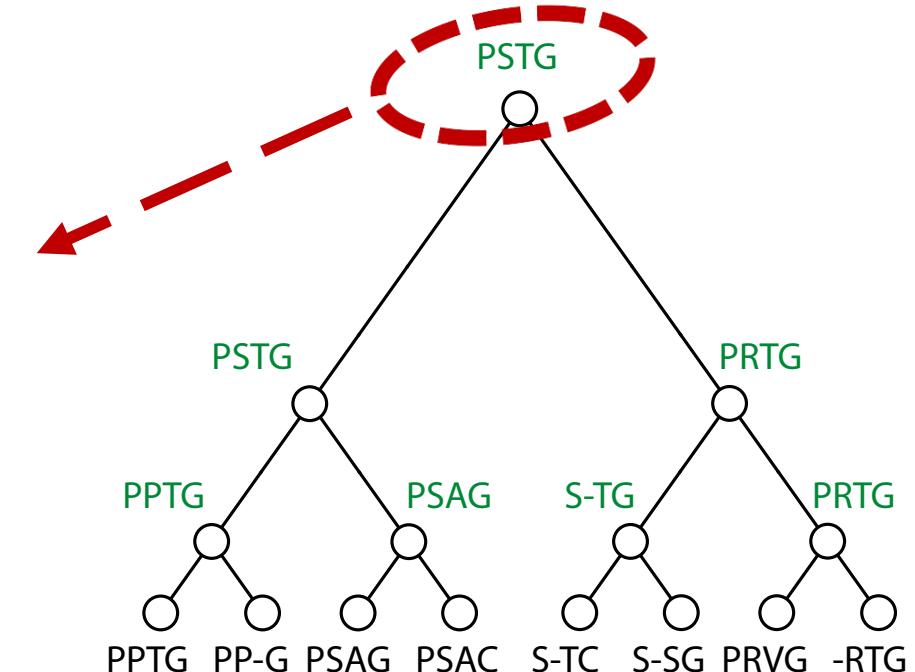
What is ancestral sequence reconstruction?

- Using the information in modern day biological sequences to infer what **their ancestors** looked like



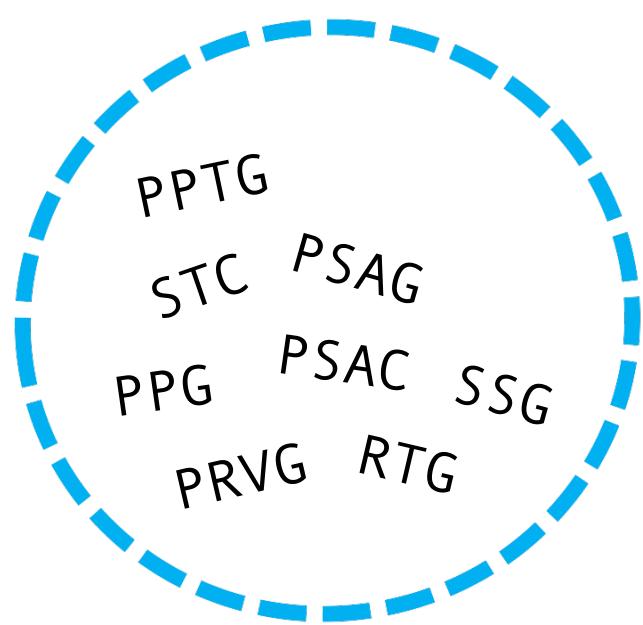
What is ancestral sequence reconstruction?

- Using the information in modern day biological sequences to infer what **their ancestors** looked like
- Ancestral sequences can be '**resurrected**' – synthesised and studied alongside modern day proteins

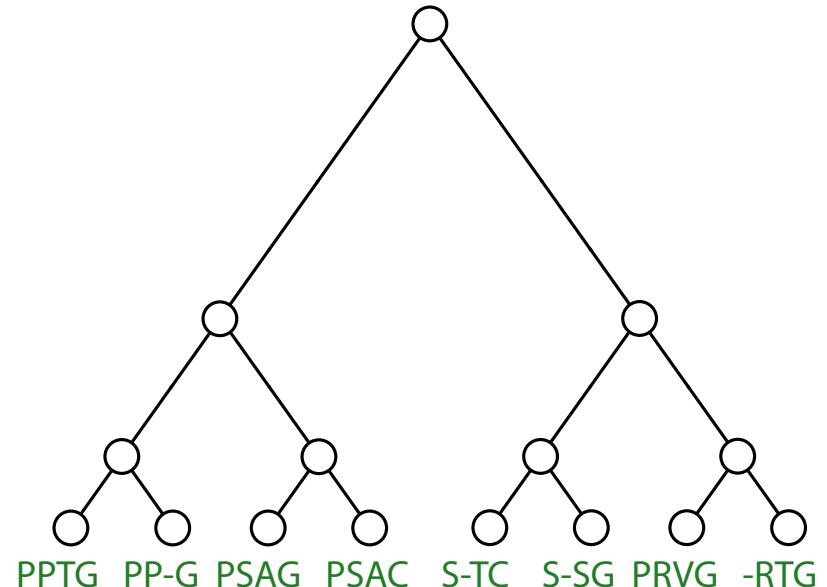


Ancestral sequence reconstruction steps

1. Collect sequences

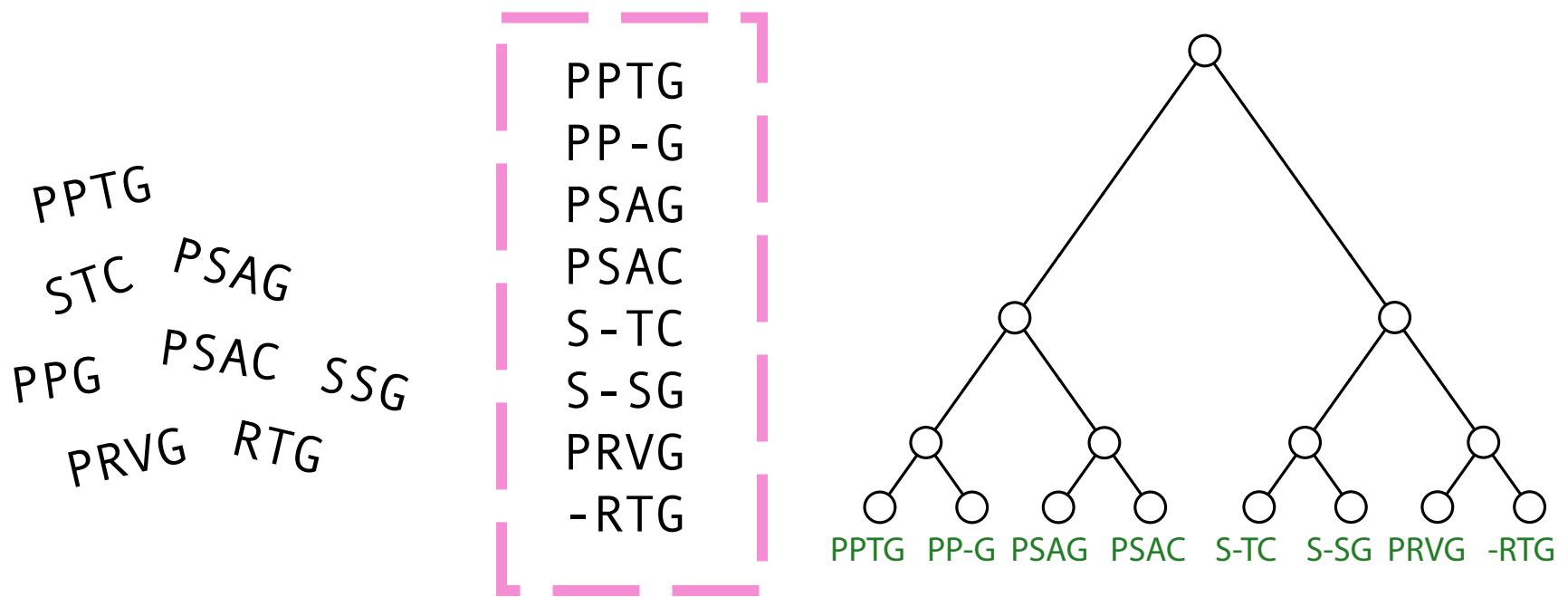


PPTG
PP -G
PSAG
PSAC
S -TC
S -SG
PRVG
-RTG



Ancestral sequence reconstruction steps

2. Align sequences

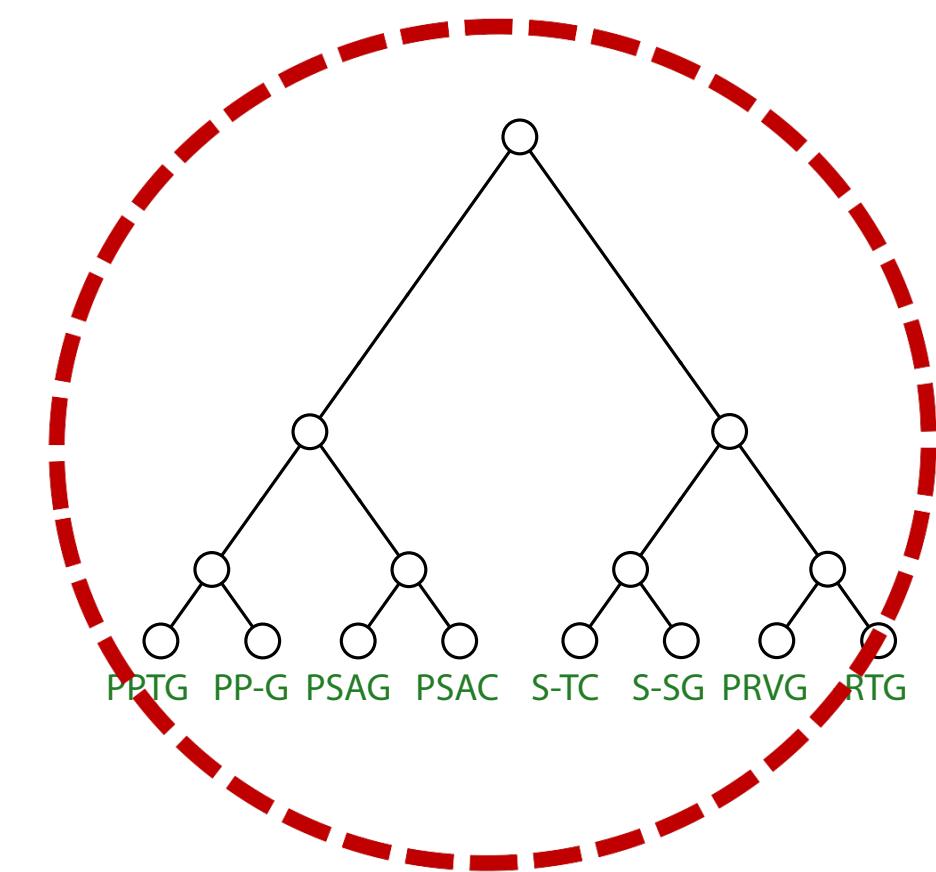


Ancestral sequence reconstruction steps

3. Infer phylogenetic tree

PPTG
STC PSAG
PPG PSAC SSG
PRVG RTG

PPTG
PP-G
PSAG
PSAC
S-TC
S-SG
PRVG
-RTG

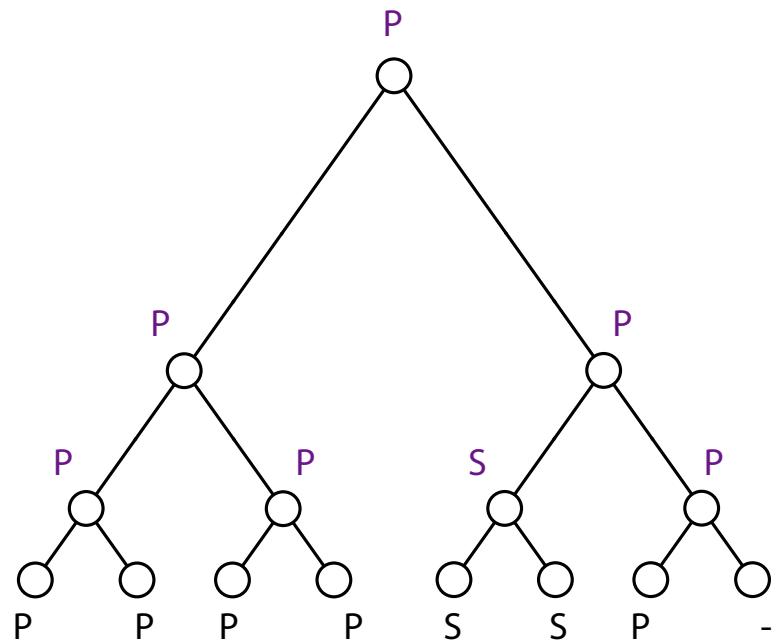


Ancestral sequence reconstruction steps

4. Infer ancestors for individual columns

PPTG
STC PSAG
PPG PSAC SSG
PRVG RTG

PPTG
PP-G
PSAG
PSAC
S-TC
S-SG
PRVG
-RTG

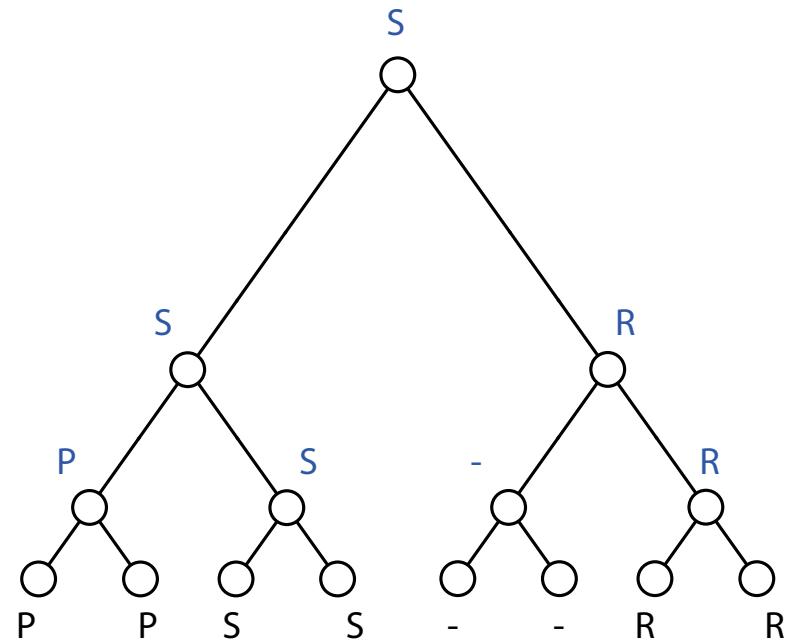


Ancestral sequence reconstruction steps

4. Infer ancestors for individual columns

PPTG
STC PSAG
PPG PSAC SSG
PRVG RTG

PPTG
PP-G
PSAG
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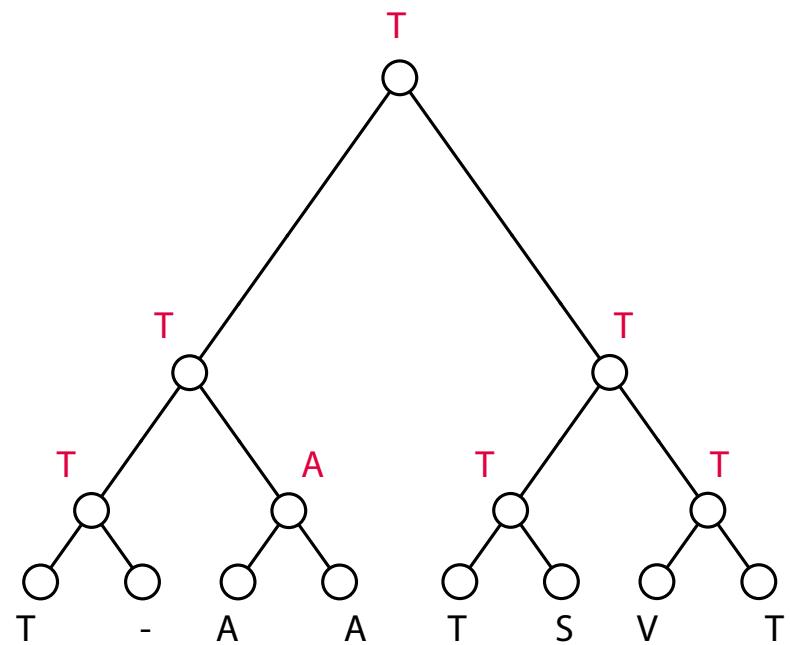


Ancestral sequence reconstruction steps

4. Infer ancestors for individual columns

PPTG
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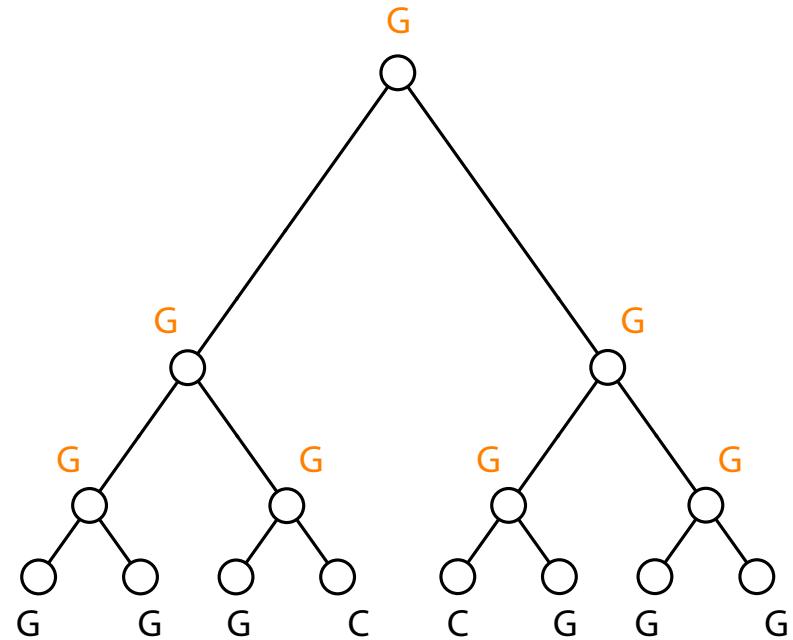


Ancestral sequence reconstruction steps

4. Infer ancestors for individual columns

PPTG
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PPTG
PP -G
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S -TC
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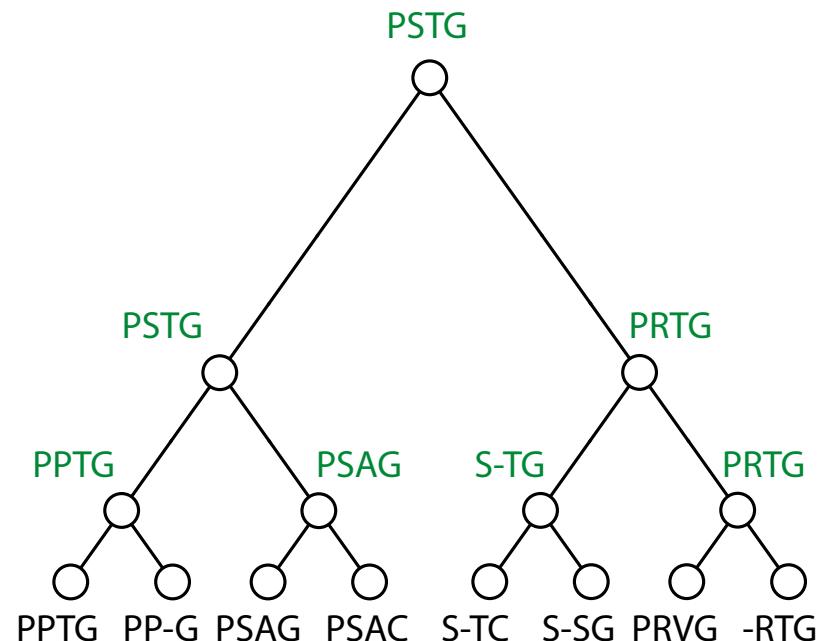


Ancestral sequence reconstruction steps

5. Concatenate predictions into a complete sequence

PPTG
STC PSAG
PPG PSAC SSG
PRVG RTG

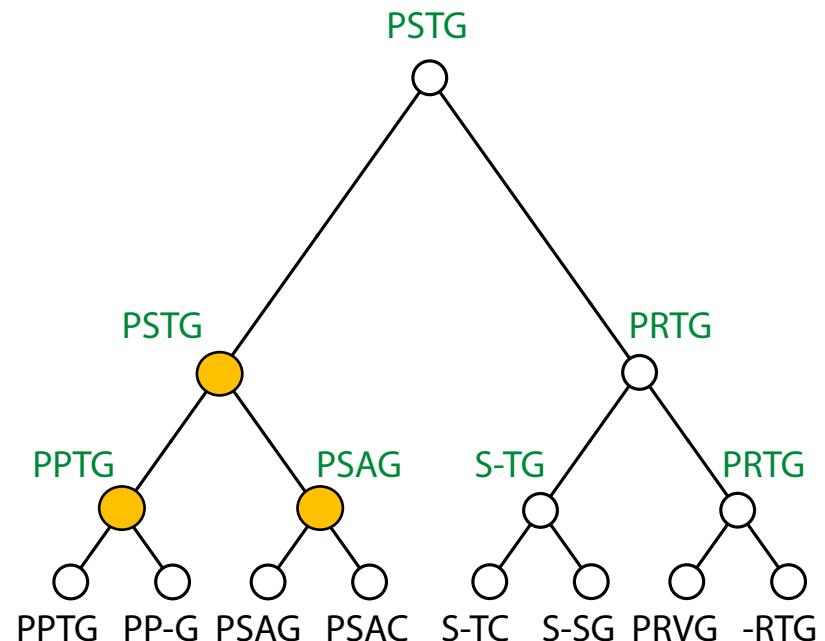
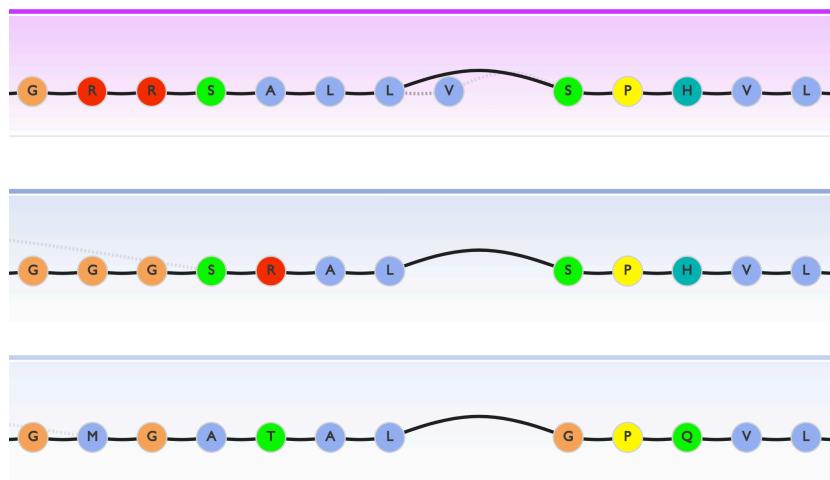
PPTG
PP -G
PSAG
PSAC
S - TC
S - SG
PRVG
- RTG



Ancestral sequence reconstruction steps

Joint reconstruction

Infer predictions for all ancestors

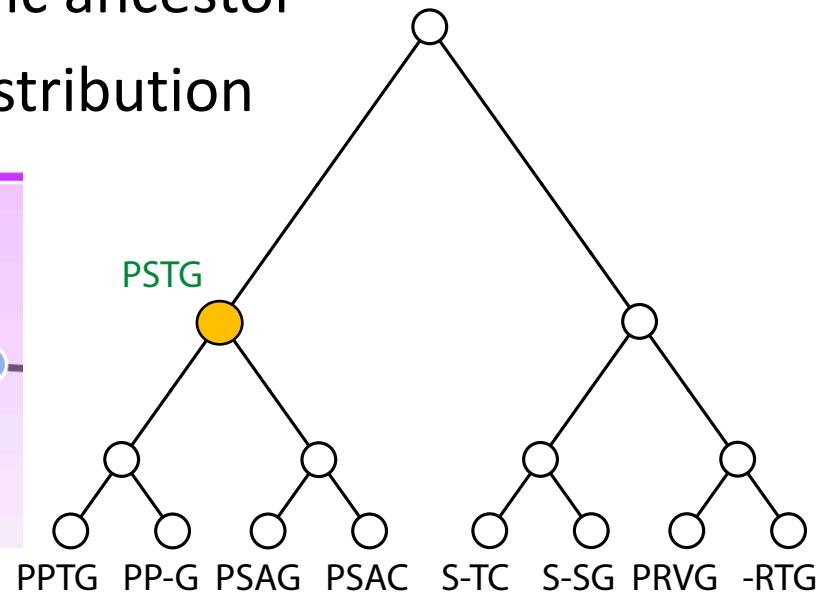
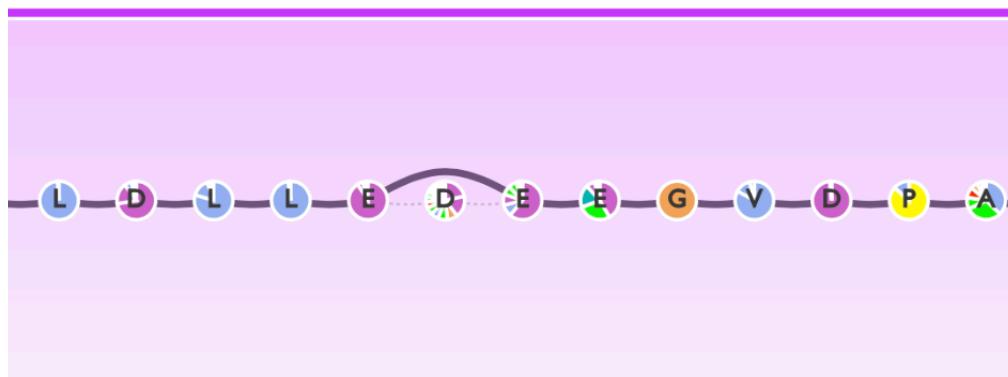


Ancestral sequence reconstruction steps

Marginal reconstruction

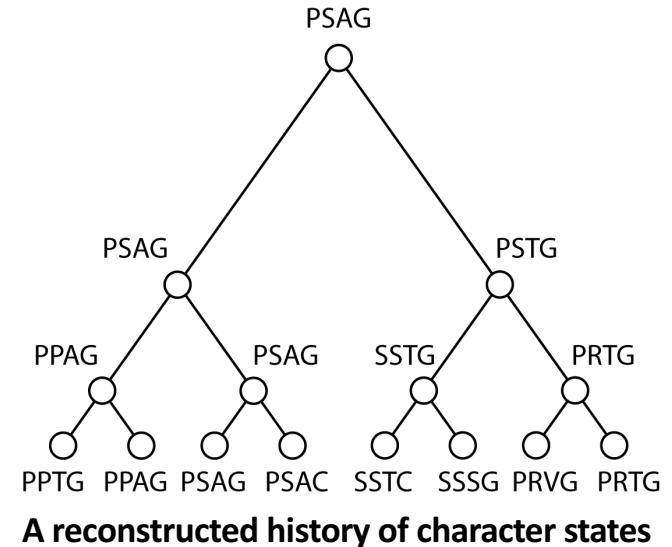
Center prediction around a specific ancestor

Each position has a probability distribution



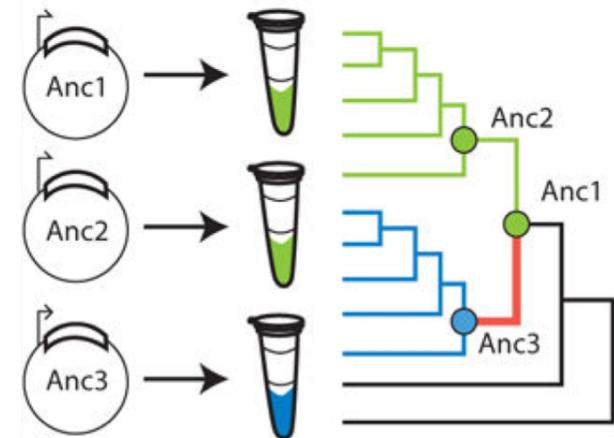
Why use ancestral sequence reconstruction?

- **Studying evolutionary histories**
- Determining important functional residues
- Engineering ancestors from templates
- Constructing novel sequences



Why use ancestral sequence reconstruction?

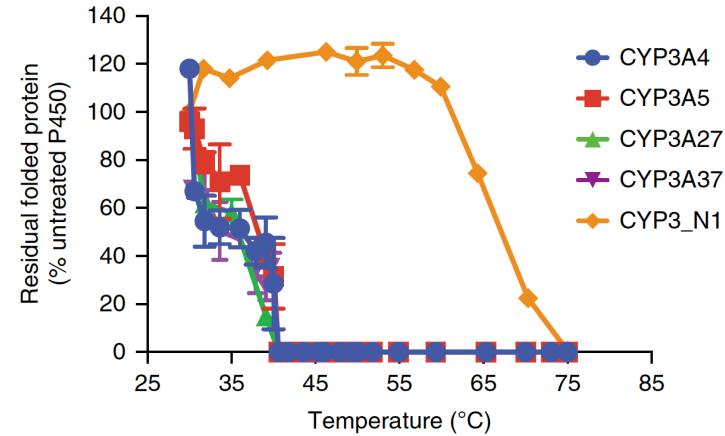
- Studying evolutionary histories
- **Determining important functional residues**
- Engineering ancestors from templates
- Constructing novel sequences



Adapted from Hochberg & Thornton, *Annu Rev Biophys* **46**, 247–269 (2017)

Why use ancestral sequence reconstruction?

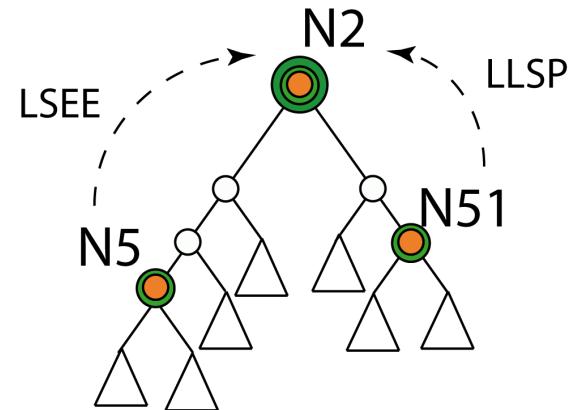
- Studying evolutionary histories
- Determining important functional residues
- **Engineering ancestors from templates**
- Constructing novel sequences



Adapted from Gumulya et al., *Nature Catalysis* 1, 878 (2018).

Why use ancestral sequence reconstruction?

- Studying evolutionary histories
- Determining important functional residues
- Engineering ancestors from templates
- **Constructing novel sequences**

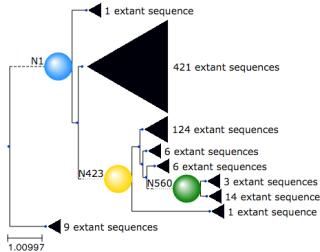


Successfully reconstructed CYP2U1 variants

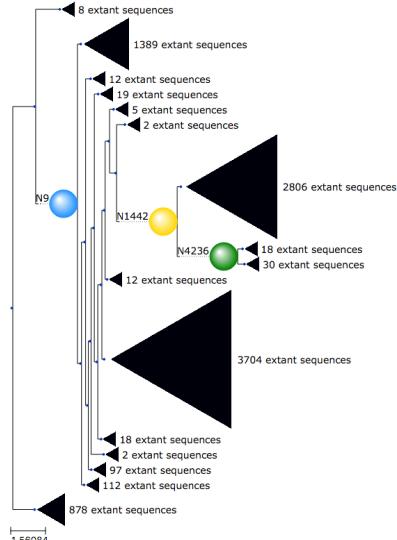
ASR in the era of big data

- Better coverage increases robustness of predictions
 - Enables us to classify allowable variation
- Incorporation of distant homologs can allow us to infer further back in evolutionary time
- Ancestral data sets become rich sources of information which can be mined and studied

Large data sets approach a canonical form of ancestor

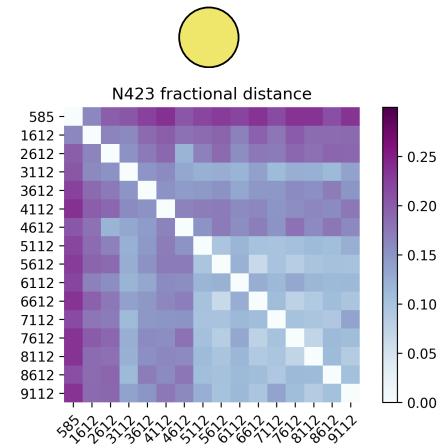
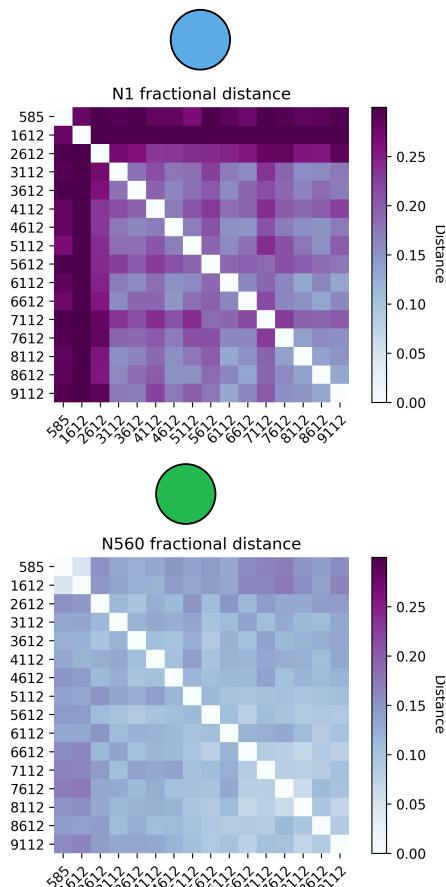


585 sequences



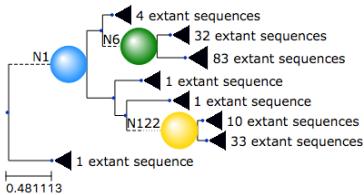
9112 sequences

Dihydroxy-acid dehydratase data set
Fractional distances between different ancestors

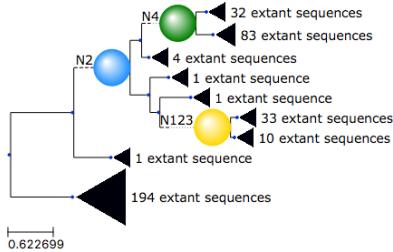


- Increasing sequence count mean predictions approach canonical forms
- Ancestors closer to extants are less affected

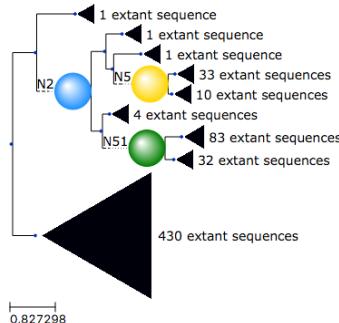
Large data sets approach a canonical form of ancestor



CYP2U1: 165 sequences

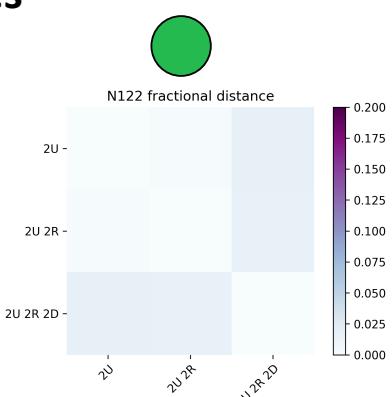
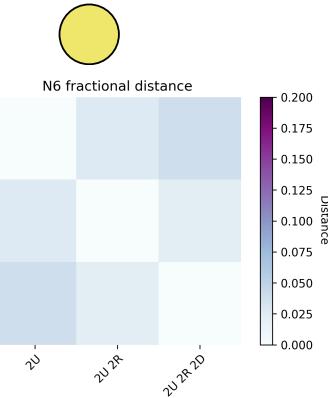
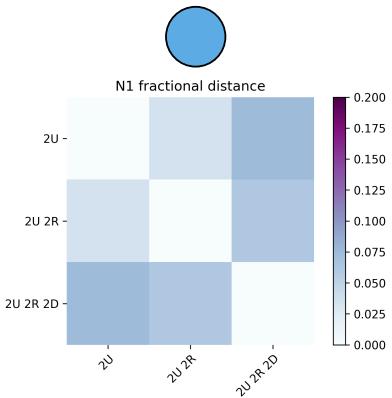


CYP2U1 / CYP2R1 : 359 sequences



CYP2U1 / CYP2R1 / CYP2D: 595 sequences

CYP2U1 / CYP2R1 / CYP2D data set
Fractional distances between different ancestors



- Increasing sequence count mean predictions approach canonical forms
- Ancestors closer to extants are less affected

ASR – challenges with big data

- Processing large data sets takes a long time or is impossible
 - Current tools, FastML and PAML capable of ~500 - 600 sequences
- Increased presence of insertions and deletions
 - Increases alignment length and must be dealt with in order to predict sensible ancestors
- Extracting information is a much harder tasker
 - More alternatives exist, scale of data is harder to examine

GRASP – solutions for big data

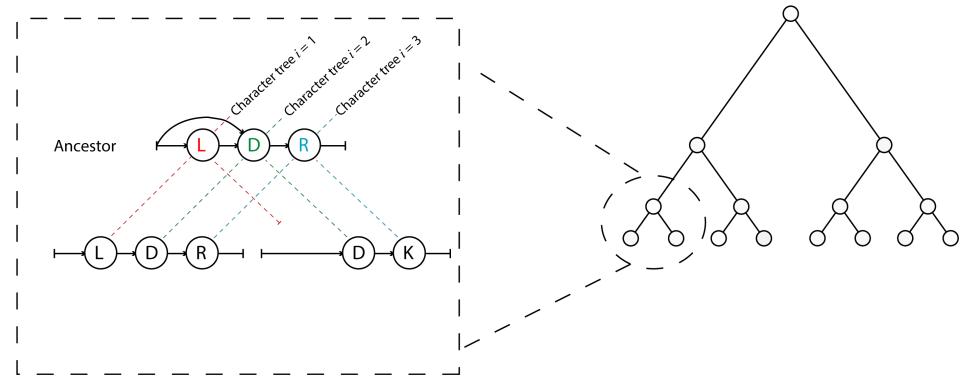
- Processing large data sets takes a long time or is impossible
 - GRASP is capable of inferring data set sizes of ~9000
- Increased presence of insertions and deletions
 - GRASP uses partial order graphs to discretely model insertion and deletion events
- Extracting information is a much harder tasker
 - GRASP is an interactive tool built for exploration, with annotations, mutant suggestions, and motif searching

Processing large data sets

- **Data structure** is a Bayesian network and we use variable elimination for efficient inference
- **Inference algorithm** is equivalent to FastML or PAML

INFERENCE STEPS

1. Calculate all possible state
 2. Calculate a consensus path
-
- Importantly, we can dynamically process these on demand



359 sequences

Tool	Run time (full output)	Run time (selected output)
GRASP	3 min	1 min 30 seconds
FastML	8 hours	Not possible
PAML	13 hours	Not possible

1529 sequences

Tool	Run time (full output)	Run time (selected output)
GRASP	1 hour 5 mins	9 min

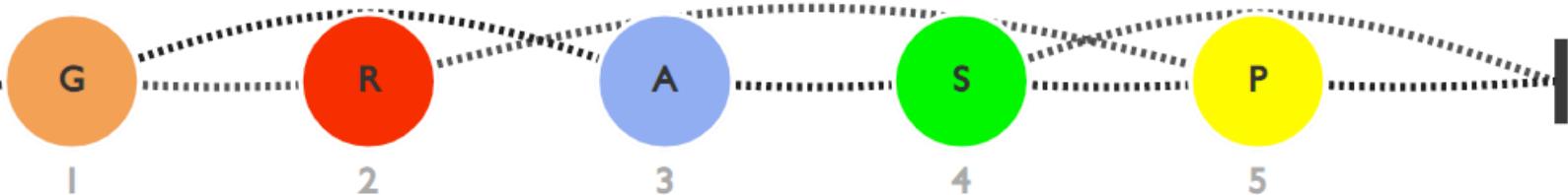
9112 sequences

Tool	Run time (full output)	Run time (selected output)
GRASP	~ 7 days	~ 1 day

Modelling indels with partial order graphs

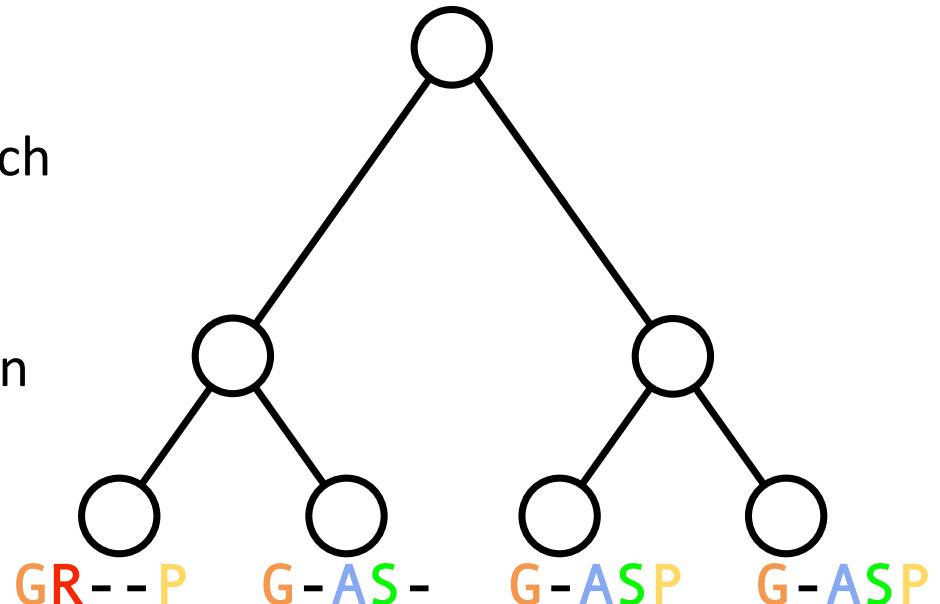
- Represents ambiguity
- Summarises indel events as edges on a graph

GR - - P
G - AS -
G - ASP
G - ASP

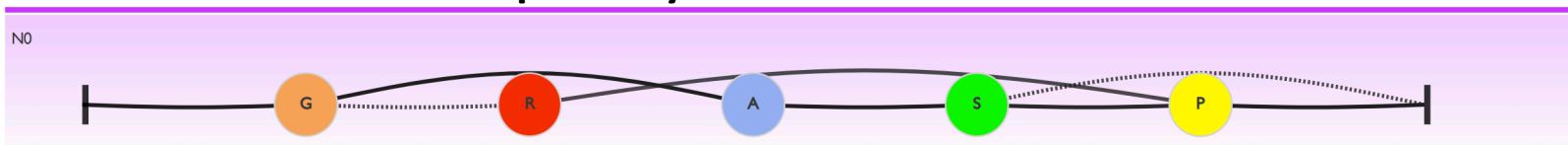


Inferring a consensus path

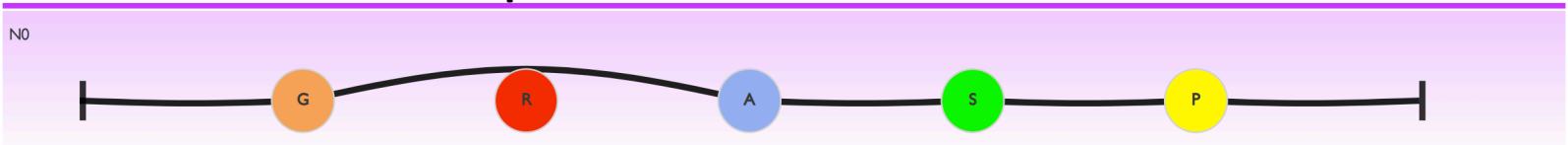
- Parsimony is used to score each out edge *and* each in edge
- Edges that are parsimonious in both directions are preferred



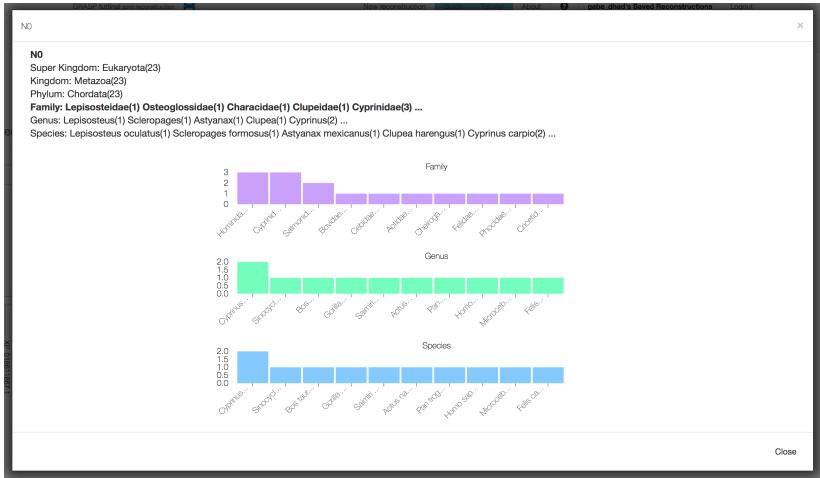
Ancestor with alternative pathways



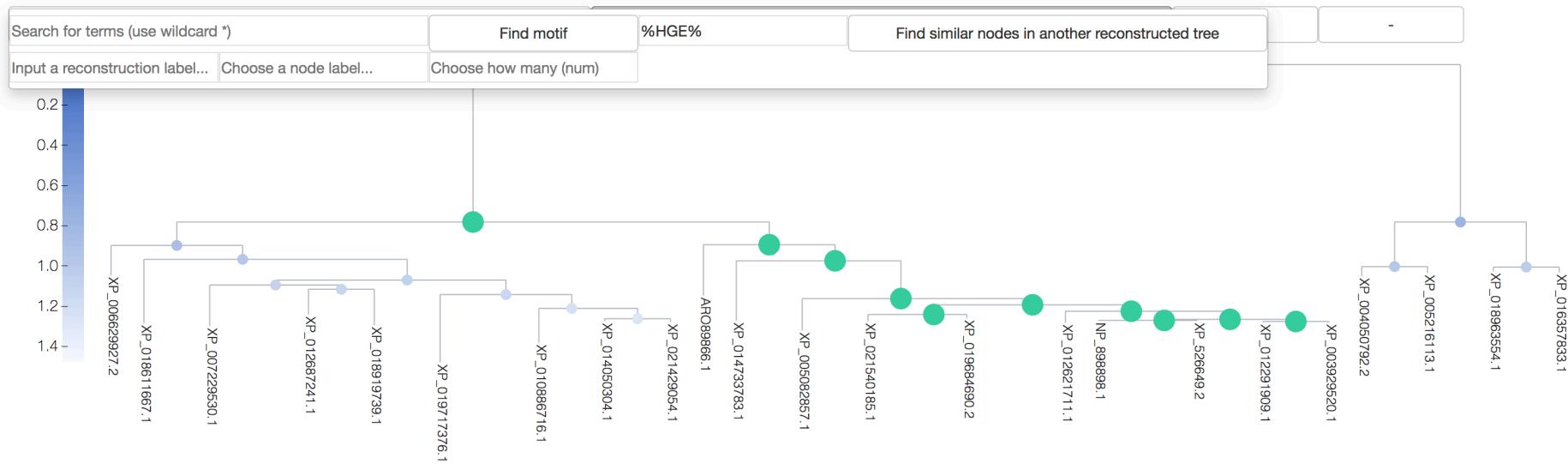
Ancestor with consensus path



GRASP annotations and searching



- Taxonomic annotation from UniProt / NCBI
- Searching ancestors for -
 - Annotations
 - Sequence motifs



SeqScrub for curation

- Annotates
- Cleans
- Checks for obsolete sequences
- Checks for given characters

SeqScrub

Upload a FASTA file to edit all headers into the specified cleaned format.

Choose a file

Choose a tree (optional)

Type of sequence content: Amino acids

Submit

Select header output format: (Click field to add and drag items to rearrange)

Gene information Species

Curation options

Remove obsolete sequences

Remove un-mappable sequences

Remove sequences containing: B J O U X Z

Remove these characters from header: .,:;()

Keep original headers - just remove characters from headers: .,:;()

Don't check databases - just remove characters from headers: .,:;()

Retain only the first ID from headers with multiple IDs

Formatting options

Format UniProt IDs like this: |>|A0A1ABUQD|TA0A1ABUQI7_NOTFU\$

For PDB sequences - Keep the original header information and don't add annotations

Add this character after ID

Use this character to split gene information:

Use this character to split species name information:

Use this character to split taxonomic / common name:

Change spaces to underscores in header

Add square brackets around species name

Remove internal brackets in species name

If cleaning a tree and the new label contains whitespace, add quotation marks

Cleaned sequences:

Sequences with illegal characters:

Obsolete sequences:

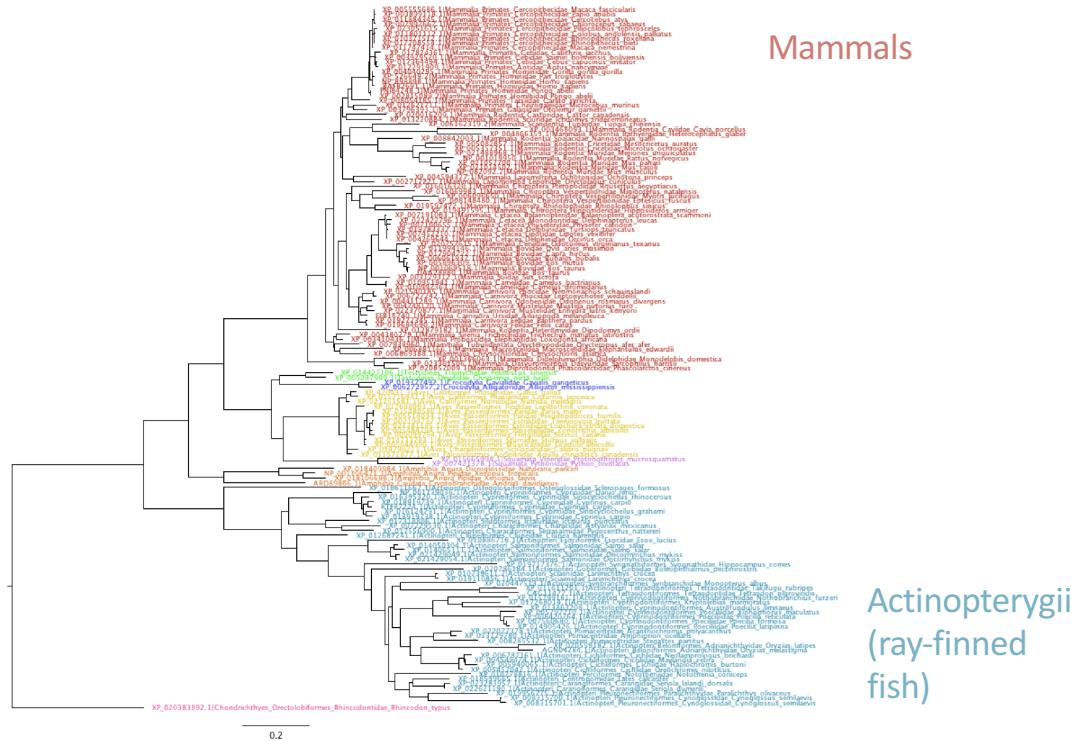
Un-mappable sequences:

Read the documentation and FAQ

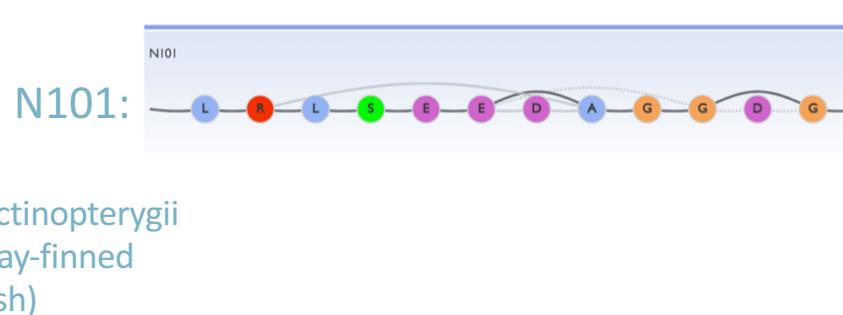
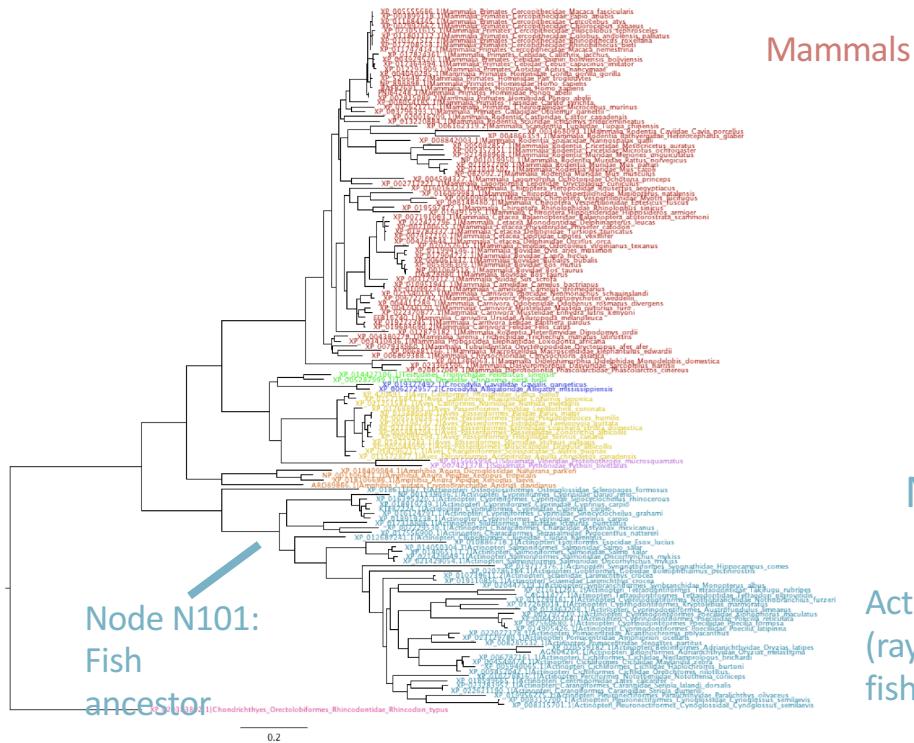
- Communicates with NCBI / UniProt
- Completely in-browser application

Foley, Sützl, D'Cunha, Gillam, Bodén, *BioTechniques* (2019) doi:[10.2144/btn-2018-0188](https://doi.org/10.2144/btn-2018-0188)

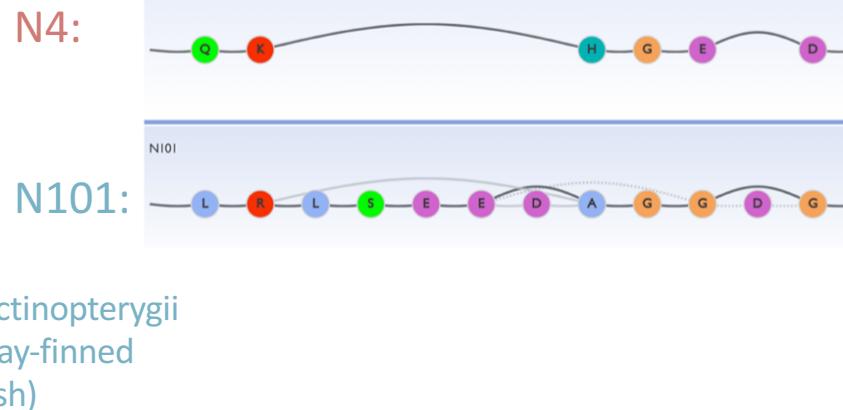
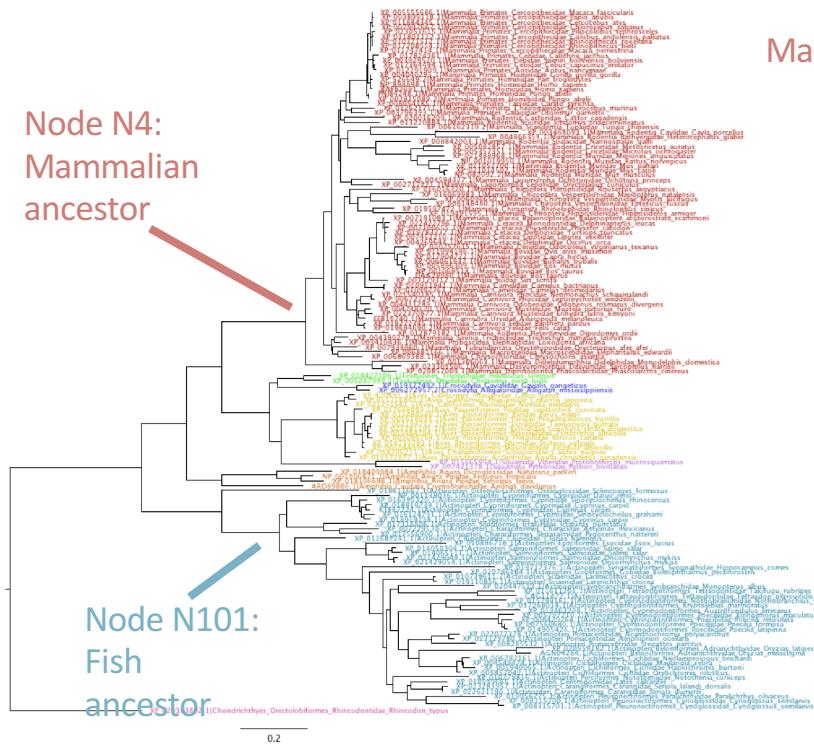
GRASP enables inspection of indel histories



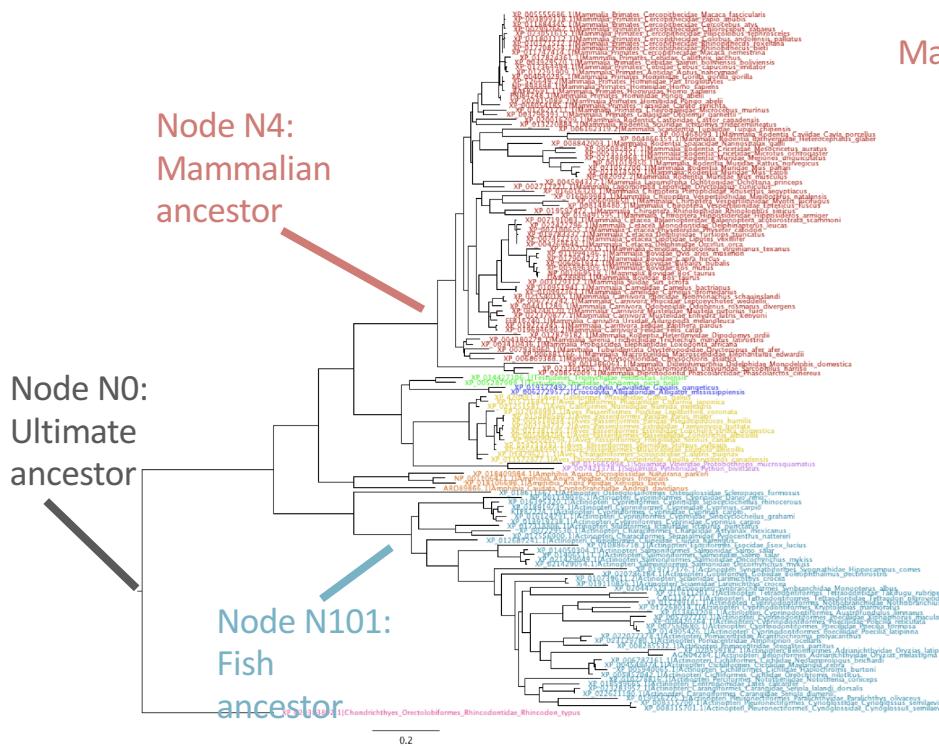
GRASP enables inspection of indel histories



GRASP enables inspection of indel histories



GRASP enables inspection of indel histories



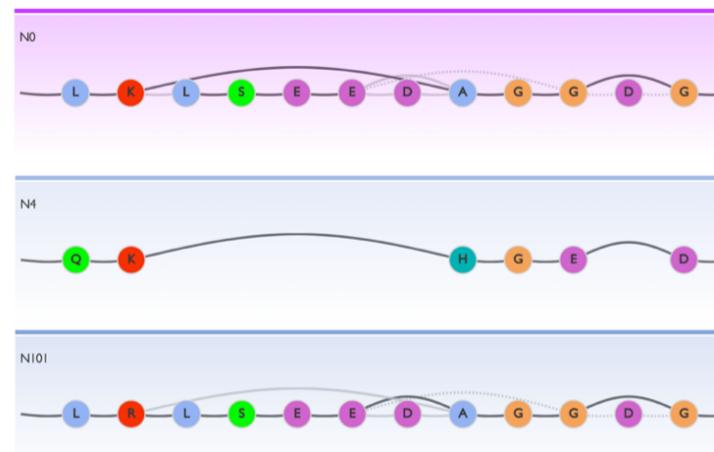
Mammals

N0:

N4:

N101:

Actinopterygii
(ray-finned fish)



Conclusion

- Ancestral sequence reconstruction is a valuable resource to understand, explore, and utilise evolution
- Large data sets allow us to extend the reach of ASR
- GRASP enables novel experiments on previously unobtainable data set sizes

Acknowledgements

PhD supervisors

Elizabeth Gillam

Mikael Bodén

Ross Barnard

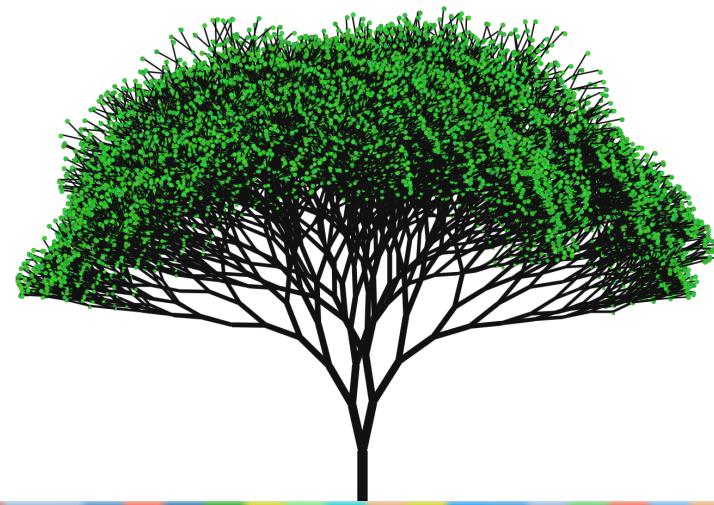


<http://grasp.scmb.uq.edu.au>

The Gillam and Bodén groups

Groups of Volker Sieber and Dietmar Haltrich

Connie Ross, Ariane Mora, Marnie Lamprecht,
Raine Thomson, Yosephine Gumulya,
Kurt Harris, Stephina D'Cunha

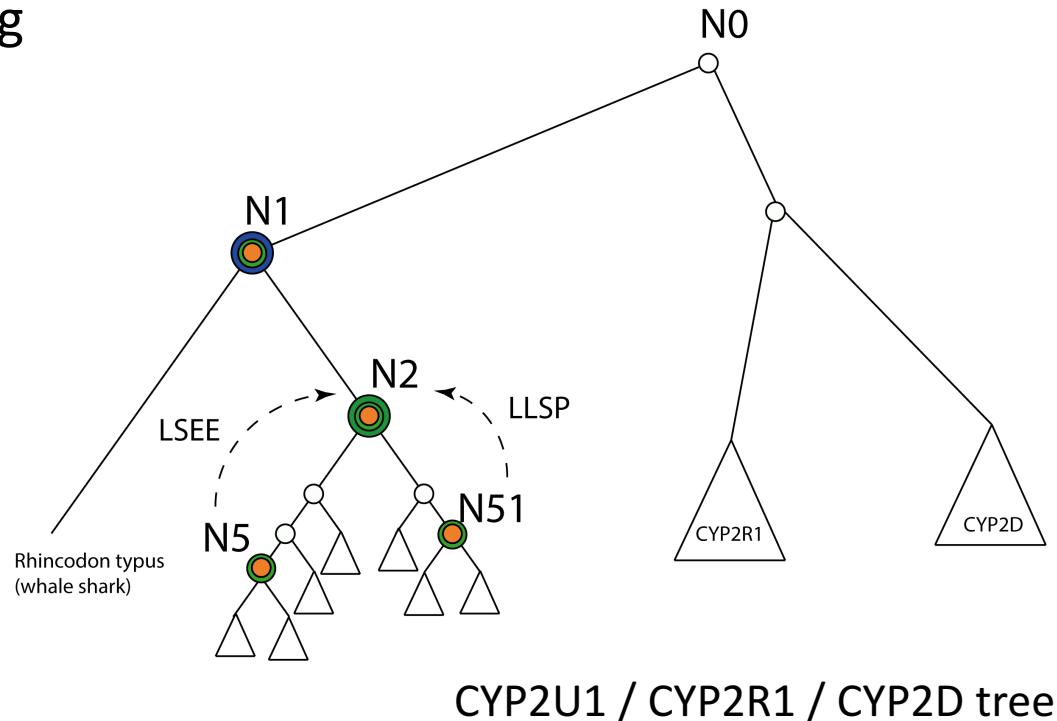


Additional slides

Constructing novel indel variants

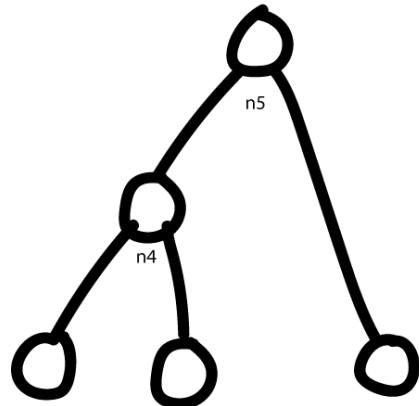
From this tree we **reconstructed**
10 CYP2U1 ancestors, including
six ancestors that either
reverted or pre-empted
insertions and deletions.

All ancestors were able to
express and show a
characteristic P450 spectrum.



Experimental work performed by Connie Ross

Marginal & joint differences



$$\begin{aligned} P(n4 = A, n5 = A) &= 0.4 \\ P(n4 = A, n5 = C) &= 0.3 \\ P(n4 = C, n5 = A) &= 0.05 \\ P(n4 = C, n5 = C) &= 0.25 \end{aligned}$$

Joint reconstruction of node n4 and node n5

Find the highest probability

$$P(n4 = A, n5 = A) = 0.4$$

Character at n5 is assigned A

Marginal reconstruction of node n5

Sum up all the ways we could get n5=A

$$\begin{aligned} P(n4 = A, n5 = A) + P(n4 = C, n5 = A) \\ = 0.4 + 0.05 \\ = 0.45 \end{aligned}$$

Sum up all the ways we could get n5=C

$$\begin{aligned} P(n4 = A, n5 = C) + P(n4 = C, n5 = C) \\ = 0.3 + 0.25 \\ = 0.55 \end{aligned}$$

Character at n5 is assigned C

Marginal & joint differences

Posterior probability distributions from the CYP2U1 CYP2R1 Realigned marginal reconstruction at positions where the marginal and joint reconstructions differ

