SPATIALLY VARIABLE DECONVOLUTION FOR LIGHTSHEET MICROSCOPY

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LIGHTSHEET MICROSCOPY

Light sheet microscopy is a type of fluorescence microscopy used in cell biology due to its fast acquisition times and low photo-damage to the sample. This is achieved by selectively illuminating a slice of the sample using a sheet of light and detecting the emitted fluorescence signal using a dedicated objective orthogonal to the plane of the sheet.



Figure credit: Jörg Ritter, PhD thesis (2011)

PROBLEM

the lightsheet beam and the

detection/objective point spread

function (PSF), the effective PSF of

the system is spatially varying, and

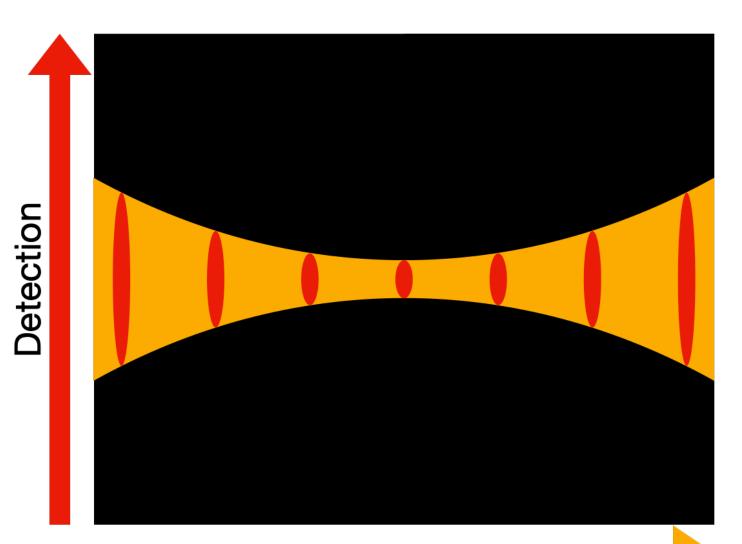
therefore standard deconvolution

approaches are not applicable.

the

interaction

SPATIALLY VARYING PSF



Illumination/lightsheet beam GOAL

In this work, we propose a model for image formation that describes the interaction between the illumination and the detection PSF which replicates the physics of the microscope while leading to a tractable inverse problem.

PSF MODEL

The detection PSF h is modelled as the Fourier transform of the pupil function multiplied by defocus [3]:

$$h(x,y,z) = \left| \iint g_{\sigma} * p(\kappa_x, \kappa_y) e^{2i\pi z} \sqrt{(n/\lambda)^2 - \kappa_x^2 - \kappa_y^2} e^{2i\pi(\kappa_x x + \kappa_y y)} \, d\kappa_x \, d\kappa_y \right|^2$$
 (1)

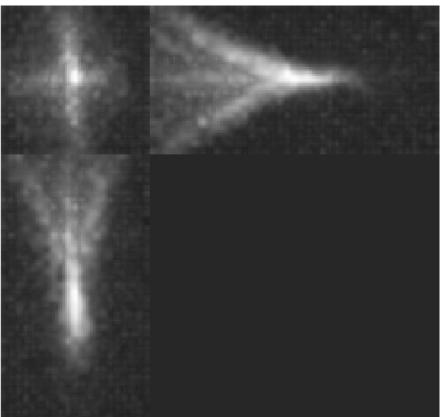
where p is the pupil function, defined as

$$p(\kappa_x, \kappa_y) = \begin{cases} e^{2i\pi \sum_{j=1}^{15} c_i Z_j(\kappa_x, \kappa_y)} & \text{for } \rho = \sqrt{\kappa_x^2 + \kappa_y^2} \le NA/\lambda, \\ 0, & \text{otherwise.} \end{cases}$$
(2)

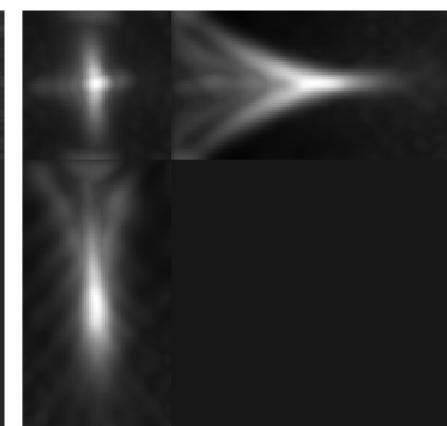
where the phase of the pupil function is computed by least-squares fitting of the coefficients of the first 15 Zernike polynomials using an image of a bead.

The parameters of the model are given by the experimental setup used to acquire the image:

- *n* refractive index
- λ wave length
- NA numerical aperture
- g_{σ} Gaussian blur to take into account other properties not accounted for in our model



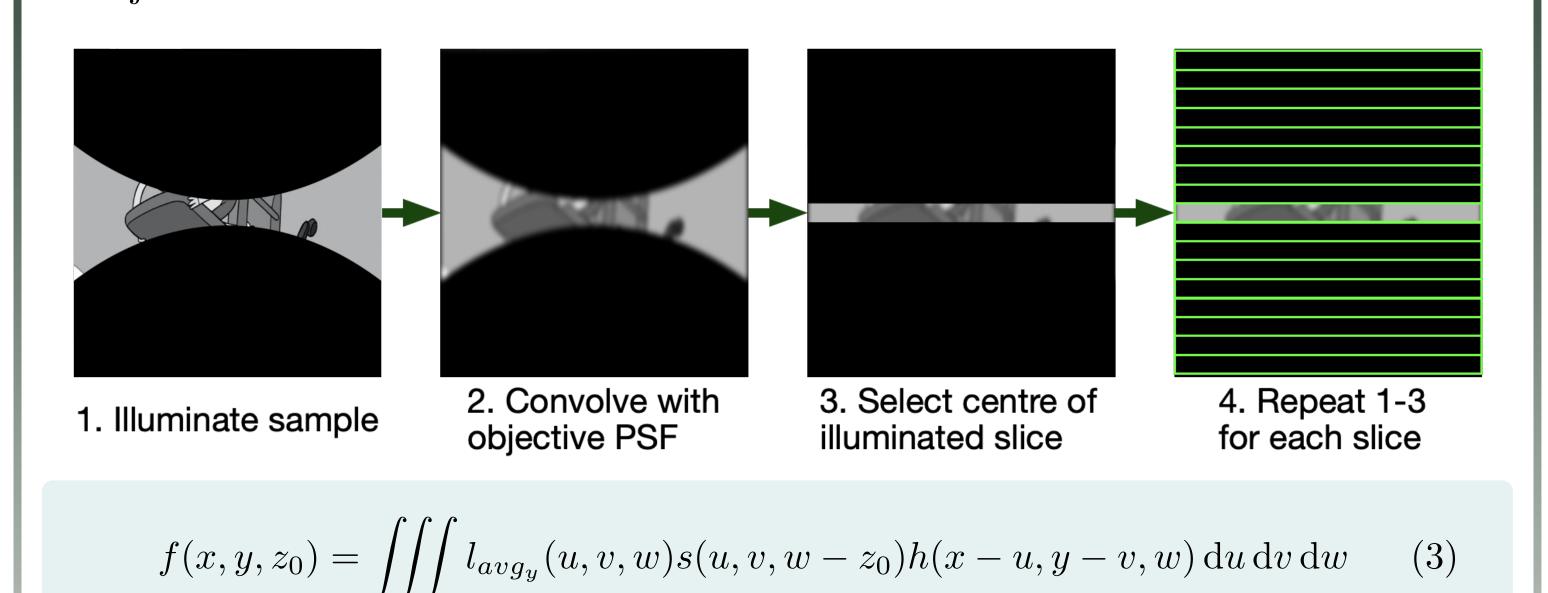
Bead image (MIP)



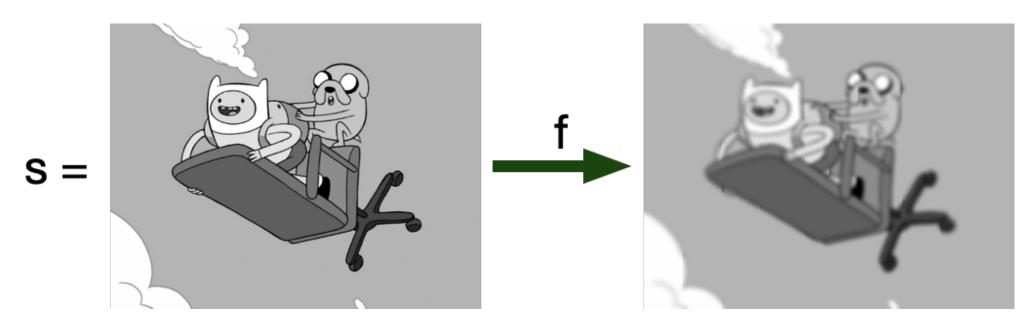
Estimated detection PSF (MIP)

IMAGE FORMATION MODEL

The sample s illuminated at $z = z_0$ by the light-sheet l and the photons are collected by an objective with PSF h:



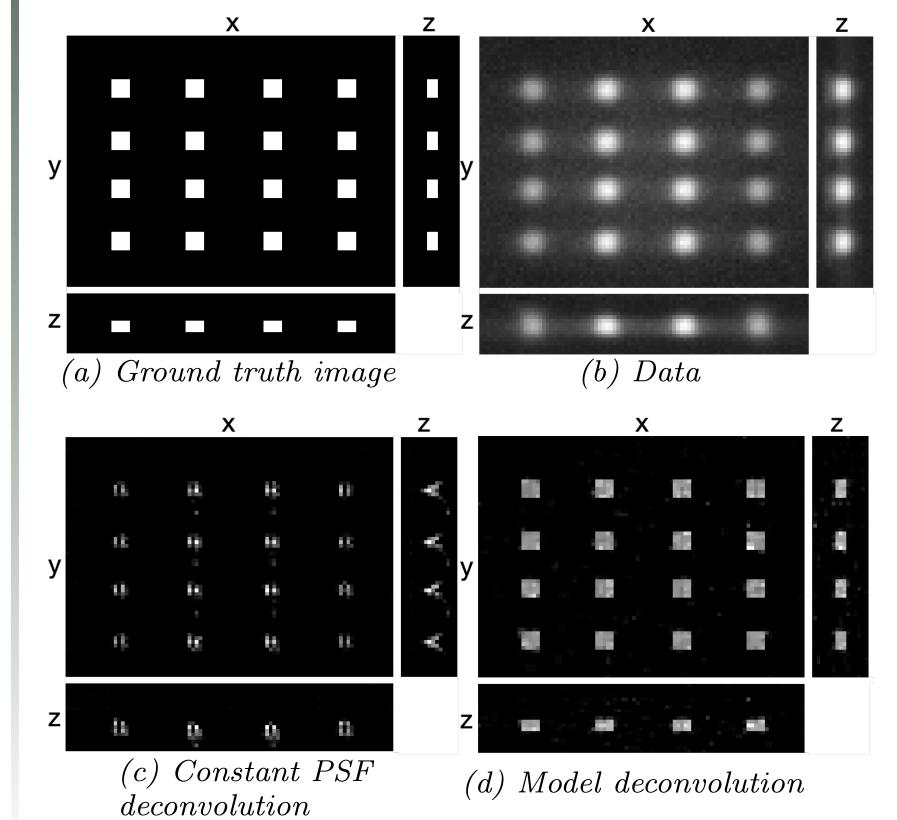
where h is the detection PSF, calculated using (1) and l_{avg_y} is the light-sheet, calculated by averaging the beam PSF obtained in a similar way to h, without Zernike polynomials.



RECONSTRUCTION

Let f be the image data with Gaussian noise and f(s) the result of applying the forward model (3) to the sample s. To recover s, we solve:

Find
$$\hat{s} \in \operatorname{argmin}_{s} \left\{ \|\hat{f} - f(s)\|_{L_{2}} + \lambda TV(s) \right\}$$

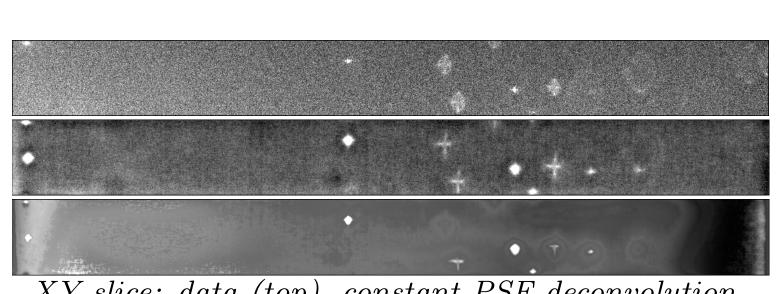


- variation totalregulariser TV(s) and ℓ_2 fidelity term due to the Gaussian noise in the data.
- To solve the optimisation problem, we apply a version of the Primal Dual Hybrid Gradient (PDHG) algorithm from [1, 2].
- In the figure on the left, we show an example of a simulated sample (a) to which we apply the forward model with Gaussian noise (b) and the result of deconvolving using only a spatially varying PSF (c) and the full forward model (d).
- The images are shown using maximum intensity projection.

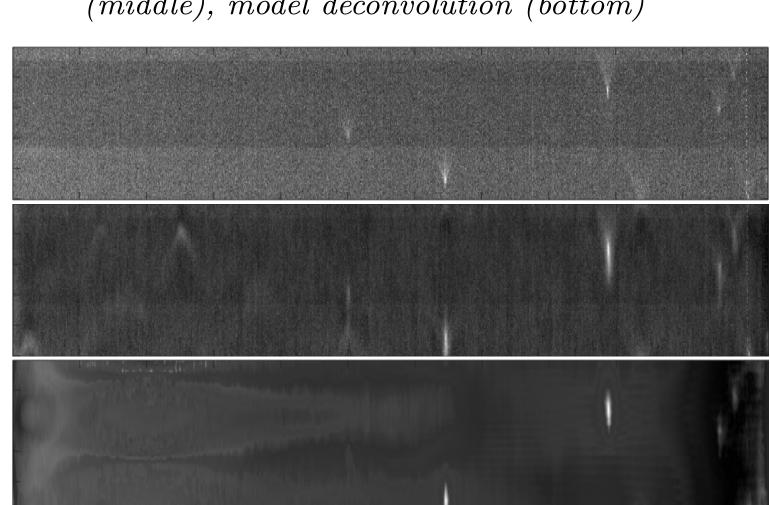
RESULTS

BEADS

- We apply the proposed method to a sample of beads in agarose.
- The dimensions of the sample are 1127 x 111 x 100 pixels.



 $XY \ slice: \ data \ (top), \ constant \ PSF \ deconvolution$ (middle), model deconvolution (bottom)



XY slice: data (top), constant PSF deconvolution (middle), model deconvolution (bottom)

MARCHANTIA

- We apply the proposed method to a sample of Marchantia plant.
- The dimensions of the sample are 1127 x 155 x 100 pixels.

XY slice: data (top), constant PSF deconvolution

(middle), model deconvolution (bottom)



XZ slice: data (top), constant PSF deconvolution (middle), model deconvolution (bottom)

ACKNOWLEDGMENTS

This work is funded by Isaac Newton Trust/Wellcome Trust ISSF/University of Cambridge Joint Research Grants Scheme, RG89305.

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