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### Chapter 4

## A data assimilating state-space model for algal growth under controlled conditions within a photo-bioreactor

### 4.1 Introduction

The key goal of biofuels production is the optimisation of biomass productivity in large-scale microalgal culturing systems such as open ponds or closed photobioreactors.

Carbon and light availability are two of the most common limiting factors of biomass productivity [?].

### 4.2 Methods

## 4.2.1 Data Model: Photo-bioreactor setup, experimental design and data collection methods

All data collection methods for this chapter were carried out by Peter Wood as part of a collection of PhD experiments (Peter Wood 2019 UTS PhD).

Microalgal culture Nannochloropsis oceanica (Droop) Green (strain CS-179) obtained from the Australian National Algae Culture Collection was cultured in 200 mL conical flasks; maintained in an incubator (Labec Pty Ltd) at 20°C, under an irradiance of  $50 \,\mu\text{mol}\,\text{m}^{-2}\,\text{s}^{-1}$  of cool-white fluorescent light at a 12 hour light/12 hour dark cycle. Stock cultures were grown in f/2 saltwater medium [5] and diluted 5 days prior to the start of experiments to ensure that N. oceanica was in the exponential growth phase and not nutrient deprived. f/2 was sparged prior to stock

culture dilutions to maximise carbon and oxygen content.

N. oceanica was cultured in four, 500 mL environmental photo-bioreactors (eP-BRs, Phenometrics Inc) with a 10% v/v inoculation of stock culture. Top-side illumination over a path length of 25 cm was provided by a cool-white light LED, whilst temperature was maintained at 27°C using a Peltier heater-cooler connected to a water jacket. In-built thermocouples, calibrated against external temperature sensors attached to the Firesting module (TeX4; PyroScience GmbH), measured every 5 minutes were used to control the Peltier heater-cooler jacket through a feedback loop to an accuracy of  $\pm$  0.2°C. pH was also measured in 5 min intervals by in-built pH electrodes (Van London Inc); controlled by periodic CO2 (5%) injections using valves in the ePBRs. pH was 3-point calibrated using pH buffer solutions at pH 4.00  $\pm$  0.02, pH 7.00  $\pm$  0.02 and pH 10.00  $\pm$  0.02. PBR mixing was controlled by magnetic stirring bars at 110 rpm. All four ePBRs were aerated with filtered/humidified air through a 1.2 mm needle valve (Terumo Co).

A period of 2 days was allowed for N. occulata to acclimate to the ePBRs at an irradiance of  $500 \,\mu\text{mol}\,\text{m}^{-2}\,\text{s}^{-1}$  and a temperature of  $27^{\circ}\text{C}$ . Following this acclimation period, the PBR was set to the experimental condition of  $2,000 \,\mu\text{mol}\,\text{photons}\,\text{m}^{-2}\,\text{s}^{-1}$  for another 2 days and a 12 hour light/12 hour dark cycle with a temperature of  $27^{\circ}\text{C}$ . ePBRs were maintained at an optical density (OD) of 0.4 using manual dilutions, creating a semi-batch culturing system. Dilutions occurred once per day (one hour before the light cycle), using aerated f/2 media. The experiment was conducted over a period of 4 days, samples were extracted post and prior dilution, as well as half way through the light cycle.  $50 \,\text{mL}$  was extracted to examine total alkalinity and dissolved inorganic carbon. Dissolved oxygen (DO) was measured using a  $3 \,\text{mm}$  robust optical probe (OXROB10-OI; PyroScience GmbH) attached to a FireStingO2 logger (PyroScience GmbH). DO measurements were taken every  $60 \,\text{seconds}$  and temperature-corrected using a temperature extension module

(TeX4; PyroScience GmbH). DO was two-point calibrated using air-saturated seawater (100% saturation) and sodium sulfate-saturated water (0% saturation). At 2 hour intervals, a solenoid valve (SMC Pneumatics Pty. Ltd.) was used to stop aeration for 10 minutes to allow for observations of net photosynthesis. Alkalinity was measured twice a day using TA titration; 0.1 M hydrochloric acid on 30 mL of N. oceanica media using an auto-titrator (800 Dosino; Metrohm AG).

[Chris: DIC measurement collection description]

[BM: confirm that reference with PW]

# 4.2.2 Data model: Data treatment, distributions and measurement error

Valve, temperature, light (normalised to 0/1) and dilution rates were used to force the model. Dissolved oxygen, pH, dissolved inorganic carbon and total alkalinity observations for 4 days post acclimation were assimilated. While pH observations were calibrated and corrected, it was visible that  $O_2$  observations were not completely calibrated and experienced some sensor drift during the experiment. An offset term  $(offset_{O_2})$  was added to the  $O_2$  ode to account for this. The offset was assigned a normally distributed prior distribution with mean 0 and standard deviation 2. [BM:check the offset is still there and you did it this way otherwise change/remove.]

The data model assigned log normally distributed observation errors for each instrument;  $O_{2obs} \sim \text{Log}\mathcal{N}(\log(O_2), \sigma_{O_2}), pH_{obs} \sim \text{Log}\mathcal{N}(\log(pH), \sigma_{pH}), DIC_{obs}$   $\sim \text{Log}\mathcal{N}(\log(DIC), \sigma_{DIC}), TA_{obs} \sim \text{Log}\mathcal{N}(\log(TA), \sigma_{DIC}),$  where the standard deviations  $(\sigma_{O_2}, \sigma_{pH}, \sigma_{DIC})$  were unknown parameters to be estimated as part of the assimilating model. Dissolved inorganic carbon and total alkalinity measurements were obtained from the same instrument thus the error is shared between these states. Initial observation error priors started at  $\sigma_{O_2} \sim \text{Log}\mathcal{N}(\log(0.1), 0.5)$  and

then were adjusted during the PMMH tuning phase.

[Chris/SW: should I talk about the thinning out of  $O_2$  and pH obs?]

### 4.2.3 Process model: Carbon chemistry

Ideally CO2SYS [6] would have been used to calculate the carbon chemistry of the photo-bioreactor. CO2SYS is a program developed for CO<sub>2</sub> system calculations (CO2SYS) that calculates and returns a detailed state of the carbonate system of oceanographic water samples in seawater and freshwater [6]. Using two of the four measurable carbonate system parameters (total alkalinity, total inorganic CO<sub>2</sub>, pH, and either fugacity fCO<sub>2</sub> or partial pressure of CO<sub>2</sub>) to calculate the other two parameters at a set of input conditions (temperature and pressure).

To incorporate CO2SYS into LiBbi for solving carbon chemistry on the timescale of the microalgae model, we explicitly defined 3 iterations (Eq. 4.4 - 4.12) of the Newton-Raphson method for finding approximations to roots of real valued functions. The Newton-Raphson method is an iterative process  $x_{n+1} = x_n - \frac{f(x_n)}{f'(x_n)}$  considering a function  $f(x_n)$ , its derivative  $f'(x_n)$  and an initial starting value  $x_0$ . The approximate root  $x_{n+1}$  converges to the exact solution very quickly if a close initial starting value is picked. To ensure the quick convergence of the Newton-Raphson method, an approximating equation for  $pH_0$  (Eq. 4.3) was obtained by fitting a stepwise regression with interactions to a range of simulated CO2SYS input parameters (temperature: 20-30, salinity: 30-40, DIC: 200-2500, and alkalinity: 1500-3000). A range of initial conditions and parameter values were tested, and each converged with RMSE < 0.01 across pH,  $HCO_3^-$ ,  $CO_2$ , and  $CO_3$ , DIC,  $O_2$ , TA by the 3rd iteration (Figure 4.1 and Table 4.1).

### CO2SYS constants and iterative solution for pH, $HCO_3^-$ , $CO_2$ , and $CO_3$

Total Sulfur:

$$\begin{split} TS &= \frac{0.14}{96.062} * \frac{S}{1.8065} \\ IS &= 19.924 \frac{S}{(1000 - 1.005S)} \\ KS_{int} &= -\frac{4276.1}{T_K} + 141.328 - 23.093log(T_K) + (-\frac{13856.0}{T_K} + 324.57 \\ &- 47.986log(T_K))\sqrt{IS} + (\frac{35474}{T_K} - 771.54 + 114.723log(T_K))IS \\ &- \frac{2698}{T_K}IS^{1.5} + \frac{1776}{T_K}IS^2 \\ KS &= (1 - 0.001005S)e^{(KS_{int})} \end{split}$$

Fluorine:

$$TF = \frac{\frac{0.000067S}{18.9984}}{1.80655}$$

$$KF = e^{\left(-\left(-\frac{874.0}{T_K} - 0.111\sqrt{S} + 9.68\right)\right)}$$

$$SWS_{2_T} = \frac{\left(1 + \frac{TS}{KS}\right)}{\left(1 + \frac{TS}{KS} + \frac{TF}{KF}\right)}$$

$$Free_{2_T} = 1 + \frac{TS}{KS}$$

H2O dissoc:

$$KW = e^{(148.9802 - \frac{13847.26}{T_K} - 23.6521log(T_K) + (\frac{118.67}{T_K} - 5.977 + 1.0495log(T_K))\sqrt{S} - 0.01615S)}$$

Boron:

$$KB = exp(\frac{(-8966.90 - 2890.53\sqrt{S} - 77.942S + 1.728S\sqrt{S} - 0.0996S^2)}{T_K} + 148.0248 + 137.1942\sqrt{S} + 1.62142S - (24.4344 + 25.085\sqrt{S} + 0.2474S)log(T_K) + 0.053105T_K\sqrt{S})$$

$$TB = 0.0004326\frac{S}{35}$$

Choice of carbonate dissociation constants  $K_1$  and  $K_2$  were Mehrbach [7] (refit by Dickson and Millero [2]) with  $1.23K_1$  and  $0.53K_2$  measured experiment specific adjustments:

$$K_1 = 10^{\left(-\left(\frac{3633.86}{T_K} - 61.2172 + 9.6777log(T_K) - 0.011555S + 0.0001152S^2\right)\right)} * 1.23$$
 (4.1)

$$K_2 = 10^{\left(-\left(\frac{471.8}{T_K} + 25.9290 - 3.16967log(T_K) - 0.01781S + 0.0001122S^2\right)\right)} * 0.53$$
(4.2)

Approximating equation for the starting value of pH:

$$pH_0 = 12.26 - 0.0030605DIC - 0.043752T - 0.013625S + 0.00011315TA$$
$$+ 1.3463e - 5DIC * T + 5.2215e - 7DIC * TA$$
(4.3)

Newton-Raphson iterations:

$$h_n = 10^{-pH_n} (4.4)$$

$$h_{n_{free}} = \frac{h_n}{Free_{2_T}} \tag{4.5}$$

$$f_n = (DIC * 1e - 6 * \frac{K_1 h_n + 2K_1 K_2}{h_n^2 + K_1 h_n + K_1 K_2} - h_{n_{free}} + \frac{KW}{h_n} - TA * 1e - 6 + \frac{TB}{1 + \frac{h_n}{KB}}) * 1e6$$

$$(4.6)$$

$$df_n = (DIC * 1e - 6 * \frac{K_1 + 2K_1K_2}{h_n^2 + K_1h_n + K_1K_2}$$
$$- DIC * 1e - 6 * \frac{(K_1h_n + 2K_1K_2)}{(h_n^2 + K_1h_n + K_1K_2)^2} (2h_n + K_1)$$

$$-TB \frac{1}{(1 + \frac{h_n}{KB})^2} / KB$$

$$-\frac{KW}{h_n^2} - \frac{1}{Free_{2T}}) * 1e6 * (-log(10) * 10^{-pH})$$
(4.7)

$$pH_{n+1} = pH_n - \frac{f_n}{df_n} \tag{4.8}$$

$$H_{n+1} = 10^{-pH_{n+1}} (4.9)$$

$$CO_{2_{n+1}} = \frac{H_{n+1}^2 DIC}{H_{n+1}^2 + K_1 H_{n+1} + K_1 K_2}$$
(4.10)

$$HCO_{3_{n+1}} = \frac{H_{n+1}K_1DIC}{H_{n+1}^2 + K_1H_{n+1} + K_1K_2}$$
(4.11)

$$CO_{3_{n+1}} = \frac{K_1 K_2 DIC}{H_{n+1}^2 + K_1 H_{n+1} + K_1 K_2}$$
(4.12)

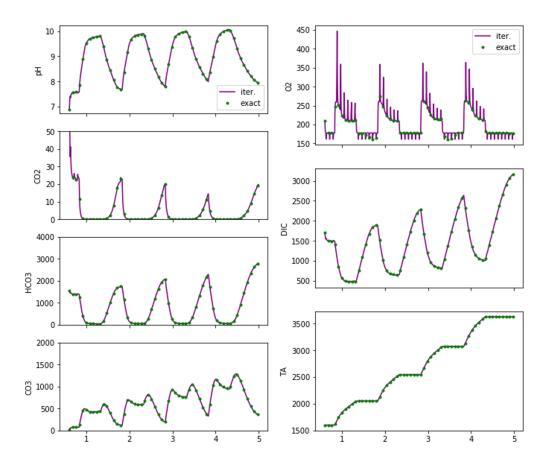


Figure 4.1 : Iterative (3rd iteration) vs exact solution for carbon chemistry  $CO_2$ ,  $HCO_3$ ,  $CO_3$ , pH and state variables  $O_2$ , DIC, and TA.

Variable	Iter. 1	Iter. 2	Iter. 3	Iter. 4	Iter. 5
pH	0.036092734	0.002355758	1.41E-05	6.93E-06	6.93E-06
$CO_2$	2.109401968	0.145719349	0.001222812	0.000866728	0.000866727
$HCO_3$	19.81869214	1.21021115	0.008016765	0.001025002	0.001025139
$CO_3$	20.89660704	1.307061652	0.00867642	0.001102278	0.001102434
DIC	16.78775711	0.958511825	0.005229318	0.002305054	0.002333411
$O_2$	0.308389964	0.016044284	4.18E-05	6.89E-05	7.59E-05
TA	2.607767674	0.160897272	0.000688102	0.001257725	0.001218981

Table 4.1: RMSE for 5 iterations of the Newton-raphson carbon chemistry iterative solution.

# 4.2.4 Process model: Gas transfer equilibrium concentrations for $O_2$ and $CO_2$

The equilibrium concentration for  $CO_2$  solubility in water  $CO_{2H}$  ( $\mu$ mol/L) is calculated using Henry's law,

$$CO_{2H} = K0_{CO2} * fCO2 * 1.0220 * 1e6$$
 (4.13)

where fCO2 (atm) is the fugacity or approximately the partial pressure of CO<sub>2</sub>, 1.0220 is the density of seawater (kg/L) at salinity 34 ppt and temperature 27°C [8] [3].  $KO_{CO2}$  (mol/kg<sub>soln</sub>/atm) is the solubility of gas in seawater [BM: ask Chris: solubility of gas? is this right] and is calculated from the fitted van't Hoff equation and the logarithmic Setchenow salinity dependence [9],

$$K0_{CO2} = e^{\left(-60.2409 + 93.4517 \frac{100}{T_K} + 23.3585 * log\left(\frac{T_K}{100}\right) + S\left(0.023517 - 0.023656 \frac{T_K}{100} + 0.0047036\left(\frac{T_K}{100}\right)^2\right)\right)} \tag{4.14}$$

where  $T_K$  is the temperature (K) and S is salinity (ppt). Similarly the equilibrium concentration for  $O_2$  solubility in water  $O_{2H}$  is calculated using Henry's law,

$$O_{2H} = K0_{O2} * fO2 * 1.0220 * 1e - 6 (4.15)$$

where fO2 (atm) is the fugacity or approximately the partial pressure of  $O_2$ , 1.0220 is the density of seawater (kg/L) at salinity 34 ppt and temperature 27°C [8] [3], and  $KO_{O2}$  (mol/kg<sub>soln</sub>/atm) is the solubility of oxygen in seawater with an adjusted salinity dependence [1],

$$K0_{O2} = \frac{e^{(-1282.8704 + \frac{36619.96}{T_K} + 223.1396log(T_K) - 0.354707T_K + S(5.957e - 3 - \frac{3.7353}{T_K}) + 3.68e - 6S^2)}{0.2094e - 6}$$

$$(4.16)$$

where  $T_K$  is the temperature (K) and S is salinity (ppt). The equilibrium concentrations for  $O_2$  and  $CO_2$  are modelled together with the gas turning on and off during the experiment, as

$$Q^{air}kLa_{O_2}^{air}(O_{2H} - O_2) (4.17)$$

$$Q^{air}kLa_{CO_2}^{air}(CO_{2H} - CO_2) (4.18)$$

where  $Q^{air}$  is the gas state (1= on, 0= off),  $kLa_{O_2}^{air}$  and  $kLa_{CO_2}^{air}$  are the mass transfer coefficients for air (d<sup>-1</sup>), and 0.893 is the ratio between measured  $O_2$  and  $CO_2$  mass transfer constants [4].

### 4.2.5 Process model: Photosynthesis and respiration

Net photosynthesis

$$\frac{\partial DIC}{dt} = -(P_1 I \frac{HCO_3^-}{K_m + HCO_3^-} - R_1) \tag{4.19}$$

$$\frac{\partial O_2}{\partial t} = \frac{1}{(RQ_d I + RQ_n(1-I))} (P_1 I \frac{HCO_3^-}{K_m + HCO_3^-} - R_1)$$
 (4.20)

$$\frac{\partial TA}{\partial t} = R_R(P_1 I \frac{HCO_3^-}{K_m + HCO_3^-}) \tag{4.21}$$

$$\frac{\partial C_p}{\partial t} = \qquad (P_1 I \frac{HCO_3^-}{K_m + HCO_3^-} - R_1) \tag{4.22}$$

Photosynthesis  $(P_1)$  and respiration  $(R_1)$  are both modelled as random walks, by taking P and R, previously constant parameters, and replacing them by  $P_1(t)$  and  $R_1(t)$ . Here, we take  $P_1(t)$  and  $R_1(t)$  to be such that

$$P_1(t + \Delta t) = P_1(t) + r_P$$

$$R_1(t + \Delta t) = R_1(t) + r_R$$

where  $r_P \sim N(0, \sigma_{r_P})$ ,  $r_R \sim N(0, \sigma_{r_R})$ , and  $\Delta t$  is the length of discrete time-step. For the purpose of the Bayesian analysis here,  $\sigma_{r_P}$  and  $\sigma_{r_R}$  are treated as parameters to be inferred.

 $RQ_d$  and  $RQ_n$  are the day and night respiratory quotients, the ratio of  $CO_2$  produced and  $O_2$  consumed by a cell.

The michaelis menton term  $\frac{HCO_3^-}{K_m + HCO_3^-}$  represents the photosynthetically active carbon that the microalgae use for photosynthesis.

#### 4.2.6 Process model: Dilution

$$\frac{Q^{M}}{V}(DIC^{M} - DIC)$$

$$\frac{Q^{M}}{V}(O_{2}^{M} - O_{2})$$

$$\frac{Q^{M}}{V}(TA^{M} - TA)$$

$$\frac{Q^{M}}{V}(C_{p}^{M} - C_{p})$$

### 4.2.7 Process model: Ordinary differential equations

A summary of the ode's that make up the process model:

Rate flux into cells gas transfer dilution 
$$\frac{\partial DIC}{\partial t} = -(P_{1}I\frac{HCO_{3}^{-}}{K_{m} + HCO_{3}^{-}} - R_{1}) + \hat{Q}^{air}kLa^{air}_{CO_{2}}(CO_{2}^{air} - CO_{2}) + \frac{Q^{M}}{V}(DIC^{M} - DIC)$$

$$\frac{\partial O_{2}}{\partial t} = \frac{(P_{1}I\frac{HCO_{3}^{-}}{K_{m} + HCO_{3}^{-}} - R_{1})}{(RQ_{d}I + RQ_{n}(1 - I))} + \hat{Q}^{air}kLa^{air}_{O_{2}}(O_{2}^{air} - O_{2}) + \frac{Q^{M}}{V}(O_{2}^{M} - O_{2})$$

$$\frac{\partial TA}{\partial t} = R_{R}(P_{1}I\frac{HCO_{3}^{-}}{K_{m} + HCO_{3}^{-}}) + \frac{Q^{M}}{V}(TA^{M} - TA)$$

$$\frac{\partial C_{p}}{\partial t} = (P_{1}I\frac{HCO_{3}^{-}}{K_{m} + HCO_{3}^{-}} - R_{1})$$

$$(4.23)$$

	Symbol	Description	Prior / Value	Unit
ions	$DIC^0$	Dissolved inorganic carbon	$Log \mathcal{N}(log(1300), 0.2)$	$\mu { m M~L^{-1}}$
conditions	$O_2^0$	Oxygen	$\log \mathcal{N}(\log(225), 0.2)$	$ m \mu M~L^{-1}$
ial cc	$TA^0$	Total alkalinity	$\log \mathcal{N}(\log(1750), 0.1)$	$ m \mu M~L^{-1}$
Initial	$C_p^0$		$\log \mathcal{N}(\log(300), 0.2)$	$ m \mu M~L^{-1}$
	$pH^0$	-	$Log \mathcal{N}(log(8.5), 0.2)$	$\log_{10}(-\text{mol/L H+})$
	$CO_2{}^0$	Carbon dioxide	$Log \mathcal{N}(log(5), 0.4)$	$ m \mu M~L^{-1}$
	$HCO_3^{-0}$	Bicarbonate	$\log \mathcal{N}(\log(1500), 0.3)$	$ m \mu M~L^{-1}$
	$CO_3^{2-0}$	Carbonate	$\log \mathcal{N}(\log(100), 0.4)$	$ m \mu M~L^{-1}$

_	ı			
lls	$P_1$	Maximum photosynthesis rate	*	$\mu$ M L <sup>-1</sup> hour <sup>-1</sup>
to ce	$R_1$	Respiration rate	*	$\mu$ M L <sup>-1</sup> hour <sup>-1</sup>
Flux into cells	$K_m$	Carbon restriction	*	$\mu M L^{-1}$
FI	$RQ_d$	Daytime respiratory quotient	*	-
	$RQ_n$	Night respiratory quotient	*	-
	$R_R$	Redfield ratio	*	-
	I	Light indicator	forcing $(0/1)$	-
rms	$\hat{Q}^{air}$	Indicator for flow in air line	forcing $(0/1)$	-
Gas transfer terms	$x_{CO_2}^{air}$	Mole fraction of CO <sub>2</sub> atmosphere	400	ppm
ransf	$CO_{2H}$	Equilibrium CO <sub>2</sub> concentration	Eq. 4.13	$\mu M L^{-1}$
Gas t	$CO_2^{air}$	Sat CO <sub>2</sub> conc with atmosphere	$x_{CO_2}^{air}CO_{2H}$	
	$kLa_{CO_2}^{air}$	Mass transfer coefficient for CO <sub>2</sub>	$0.893kLa^{air}_{O_2}$	$day^{-1}$
	$x_{O_2}^{air}$	Mole fraction of $O_2$ atmosphere	0.2094	atm
	$O_{2H}$	Equilibrium $O_2$ concentration	Eq. 4.15	$\mu M L^{-1}$
	$O_2^{air}$	Sat O <sub>2</sub> conc with atmosphere	$x_{O_2}^{air}O_{2H}$	
	au	half-life of $kLa_{O_2}^{air}$	range(2-20)	$\min^{-1}$
	$kLa^{air}_{O_2}$	Mass transfer coefficient for $O_2$	$\ln(2) * 24 * 60/\tau$	$day^{-1}$
SU	$Q^M$	Dilution rate	forcing	$\mathrm{ml}\ \mathrm{day}^{-1}$
Dilution terms	V	Volume of the reactor	500	ml
ution	$DIC^{M}$	Media dissolved inorganic carbon	1724.20	$\mu M L^{-1}$
Dil	$O_2^M$	Media oxygen concentration	226.65	$\mu { m M~L^{-1}}$
	$TA^{M}$	Media total alkalinity	1797.90	$\mu M L^{-1}$

Table 4.2: State variable, parameter and forcing definitions with units, and their assignments: either fixed values, priors on initial condition or priors on parameters (\*) defined later in Section 4.2.9.

# 4.2.8 Design and setup of data assimilation model with both simulated and experimental data

A simulated dataset was created by running the process model described in Section 4.2.7 with a fixed set of parameters ( $P_1 = 200$ ,  $R_1 = 30$ ,  $kLA_{O_2}^{air} = 200$ ,  $K_m = 150$ ,  $RQ_d = 0.85$ ,  $RQ_n = 0.95$ ,  $R_R = 0.075$ ) and initial conditions ( $O_2^0 = 225$ ,  $DIC^0 = 1250$ ,  $TA^0 = 1750$ ).

The first set of results will assimilate the simulated observations and attempt to recover the true value parameters.

[Chris: noise on simulated dataset? sigmas? can you double check what I've written here is right?]

#### 4.2.9 Parameter Model: Priors

\*\* include simulated data priors, and real data priors.

Decide whether the parameters vary in time or not.

Parameter	Prior	Proposal
S	34	*
fCO2	397e-6	*
fO2	0.21	*
$kLA_{O2}$	LogNormal( $log(200.0), 0.5$ )	$LogNormal(log(kLA_{O2}), 0.5prop_{std})$
$K_m$	LogNormal(log(200.0), 0.8)	$LogNormal(log(K_m), 0.8prop_{std})$
$R_R$	Uniform(0.0001, 0.2)	$\operatorname{TrunNormal}(\mathbf{R}_{R},\ 0.2\operatorname{prop}_{std},\ 0.0001,\ 0.2)$
$R_Q$	Uniform(0.66, 1)	$TrunNormal(R_Q, 0.2prop_{std}, 0.66, 1.0)$
$\sigma_P$	Normal(0.05, 0.01)	$Normal(\sigma_P, 0.01prop_{std})$
$\sigma_R$	Normal(0.01, 0.001)	$Normal(\sigma_R, 0.001prop_{std})$

Table 4.3: Table of Parameters, their priors and proposal distributions. \* indicates the parameter was held fixed. (prop<sub>std</sub> =0.1)

### 4.3 Results and Discussion

### 4.3.1 Posteriors with simulated data

Parameter	Quantiles (25%, 75%)	Quantiles (5%, 95%)	True value
$P_1$	(237.2167, 302.0565)	(200.8971, 365.1751)	200
$R_1$	(17.5370, 32.3501)	(10.6537, 45.3096)	30
$kLA_{O_2}^{air}$	(177.1383, 222.9641)	(148.2838, 267.5934)	200
$K_m$	(182.6643, 362.6714)	(118.4125, 643.8518)	150
$R_R$	(0.0504, 0.1041)	(0.0196, 0.1458)	0.075
$RQ_d$	(0.7529, 0.8718)	(0.6894, 0.9666)	0.85
$RQ_n$	(0.7469, 0.9338)	(0.6400, 0.9920)	0.95

Table 4.4: True values for parameters used to create the simulated observations, and posterior (25%, 75%), (5%, 95%) quantiles for parameters after assimilating observations.

After 50,000 samples the true values of parameters  $R_1$ ,  $kLA_{O_2}^{air}$ ,  $R_R$ , and  $RQ_d$  were recovered to the 25th and 75th percentile (Table 4.4). While the true values of  $K_m$  and  $RQ_n$  did not lie within the 25th and 75th quantile, they were captured by the 5th and 95th percentiles (Table 4.4).  $P_1$  was the only parameter whose true value lied on the cusp of the 5th percentile. Parameters  $P_1$ ,  $R_1$ ,  $kLA_{O_2}^{air}$ , and  $R_R$  mixed well, while parameters  $K_m$ ,  $RQ_d$ , and  $RQ_n$  did not mix as well, some autocorrelation between samples was present (Figure A.3).

Log-likelihood stopped rising (Figure A.2). Acceptance rate was XX.

Observed state variables  $O_2$ , DIC, TA, and observed pH posteriors were in excellent agreement with observations, with all observations fitting within the 95% credible interval posteriors (Figure 4.2, Figure 4.3).

Unobserved state variable  $C_p$  .. (Figure 4.3).

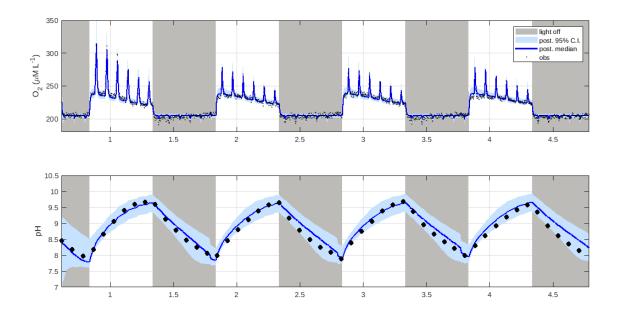


Figure 4.2 : Posterior medians (solid blue line), 95% credible intervals (shaded blue), and simulated observations (black) for  $O_2$  and pH across 4 days.

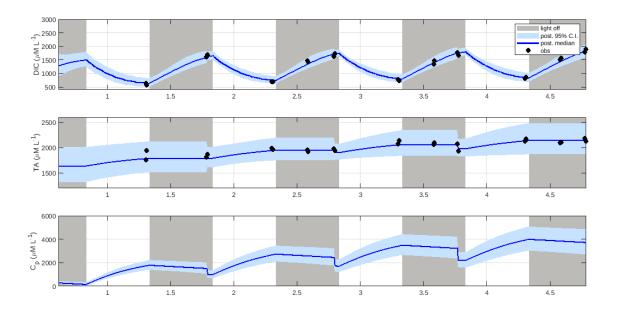


Figure 4.3: Posterior medians (solid blue line), 95% credible intervals (shaded blue), and simulated observations (black) for DIC, TA and  $C_p$  across 4 days.

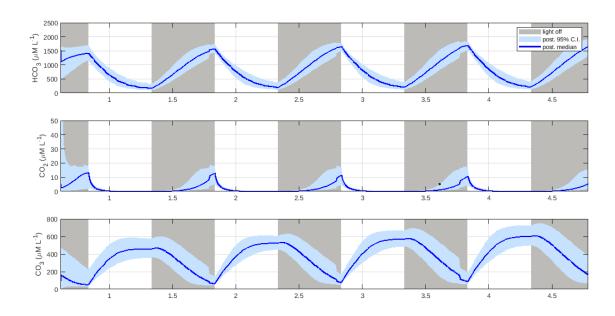


Figure 4.4 : Posterior medians (solid blue line) and 95% credible intervals (shaded blue) for  $HCO_3$ ,  $CO_2$  and  $CO_3$  across 4 days.

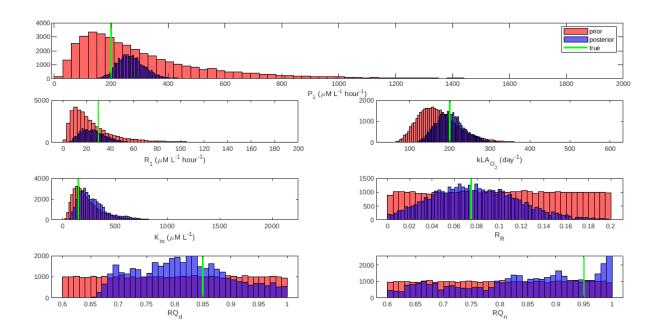


Figure 4.5: Priors (pink), posteriors (purple) and true values (green) for model parameters.

Carbon chemistry variables  $HCO_3,\ CO_2,\ {\rm and}\ CO_3$  ... (Figure 4.4).

### 4.3.2 Posteriors with simulated data version 2

Parameter	Quantiles (25%, 75%)	Quantiles (5%, 95%)	True value
$P_1$	()	()	200
$R_1$	()	()	30
$kLA_{O_2}^{air}$	(170.9854, 216.0578)	(145.6899, 253.4652)	200
$K_m$	(187.3586, 386.4103)	(93.9670, 641.1432)	150
$R_R$	(0.0521, 0.1075)	(0.0192, 0.1544)	0.075
$RQ_d$	(0.7906, 0.8965)	(0.6833, 0.9711)	0.85
$RQ_n$	(0.6654, 0.8334)	(0.6164, 0.9739)	0.95

Table 4.5: True values for parameters used to create the simulated observations, and posterior (25%, 75%), (5%, 95%) quantiles for parameters after assimilating observations.

Ran for 50,000 samples, discarded first 10,000 as burn-in.  $P_1$  and  $R_1$  as random walks with  $\sigma_{rP}$  and  $\sigma_{rR}$  as standard deviations of the random walks and parameters to be estimated.

After 50,000 samples the true values of parameters  $kLA_{O_2}^{air}$ ,  $R_R$ , and  $RQ_d$  were recovered to the 25th and 75th percentile (Table 4.5). While the true values of  $K_m$  and  $RQ_n$  did not lie within the 25th and 75th quantile, they were captured by the 5th and 95th percentiles (Table 4.5). Parameters  $\sigma_{rP}$ ,  $\sigma_{rR}$ ,  $kLA_{O_2}^{air}$ , and  $R_R$  mixed well, while parameters  $K_m$ ,  $RQ_d$ , and  $RQ_n$  did not mix as well, some autocorrelation between samples was present (Figure 4.11).

\*\*\*\*This results section came out the same as the first model with the simulated observations.

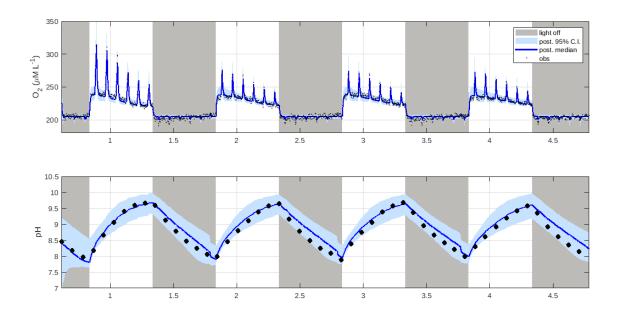


Figure 4.6 : Posterior medians (solid blue line), 95% credible intervals (shaded blue), and simulated observations (black) for  $O_2$  and pH across 4 days.

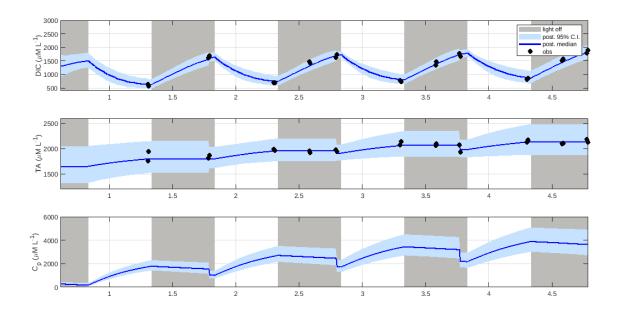


Figure 4.7: Posterior medians (solid blue line), 95% credible intervals (shaded blue), and simulated observations (black) for DIC, TA and  $C_p$  across 4 days.

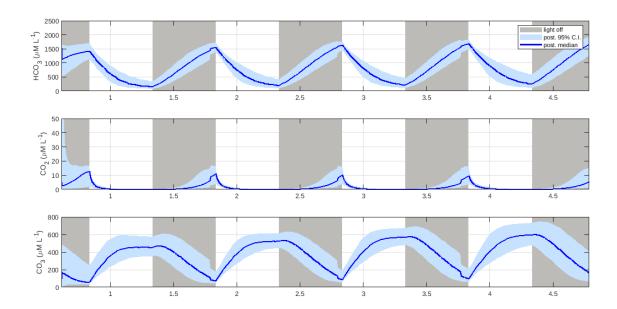


Figure 4.8 : Posterior medians (solid blue line) and 95% credible intervals (shaded blue) for  $HCO_3$ ,  $CO_2$  and  $CO_3$  across 4 days.

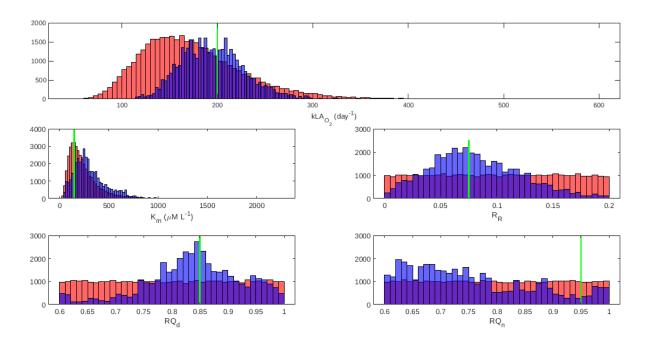


Figure 4.9: Priors (pink), posteriors (purple) and true values (green) for model parameters.

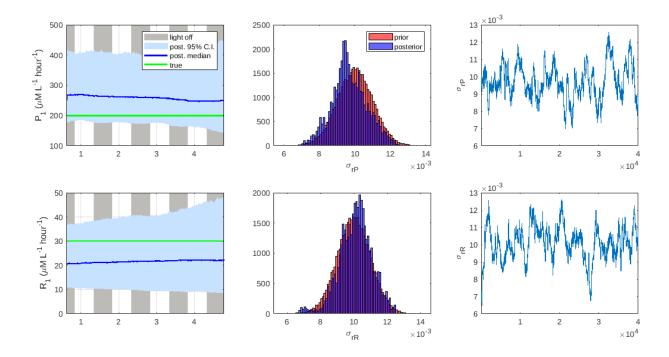


Figure 4.10: Random walk posteriors  $P_1$  and  $R_1$  medians (solid blue), 95% credible intervals (shaded blue), and true values (green).  $\sigma_{rP}$  and  $\sigma_{rR}$  priors (pink), posteriors (purple), true values (green) and traces.

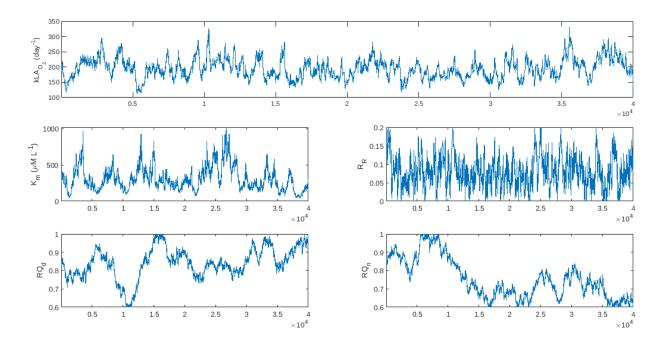


Figure 4.11 : Traces for model parameters.

### 4.3.3 Posteriors with experimental data

40,000 samples run with 1024 particles (40,000 was all that could complete in the maximum time allocation on the hpc cluster). 10,000 discarded as burn-in, acceptance rate was XX.

 $P_1$  and  $R_1$  are random walks.  $RQ_d$  and  $RQ_n$  are random walks. State posteriors are visualised by plotting the median and shading 95% credible intervals, while parameter priors and posteriors are displayed by histograms.

Parameter	Quantiles (25%, 75%)	Quantiles (5%, 95%)
$kLA_{O_2}^{air}$	(139.5979, 170.8102)	(120.9171, 204.5474)
$K_m$	(168.1931, 378.8001)	(104.2598, 599.6881)
$R_R$	(0.0815, 0.1512)	(0.0284, 0.1844)
$\sigma_{r_P}$	(0.0178, 0.0202)	(0.0159, 0.0222)
$\sigma_{r_R}$	(0.0094, 0.0113)	(0.0081, 0.0121)

Table 4.6 : Posterior (25%, 75%), (5%, 95%) quantiles for parameters after assimilating observations.

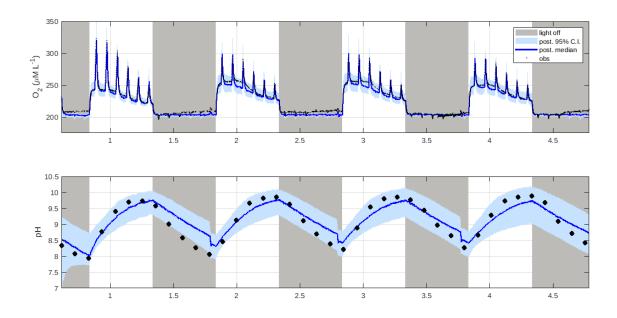


Figure 4.12 : Posterior medians (solid blue line), 95% credible intervals (shaded blue), and simulated observations (black) for  $O_2$  and pH across 4 days.

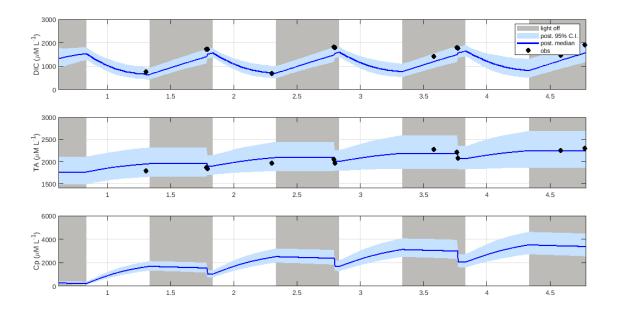


Figure 4.13: Posterior medians (solid blue line), 95% credible intervals (shaded blue), and simulated observations (black) for DIC, TA and  $C_p$  across 4 days.

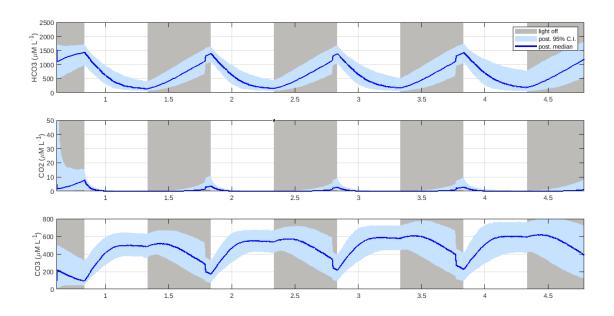


Figure 4.14: Posterior medians (solid blue line) and 95% credible intervals (shaded blue) for  $HCO_3$ ,  $CO_2$  and  $CO_3$  across 4 days.

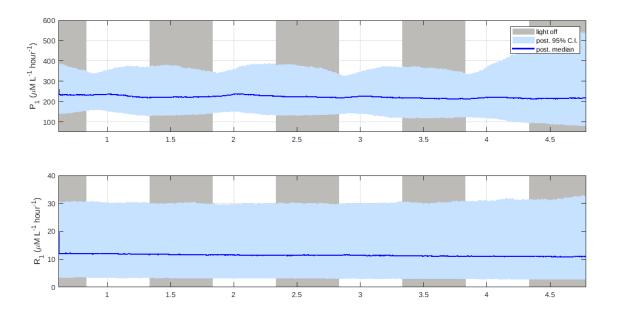


Figure 4.15: Posterior medians (solid blue line) and 95% credible intervals (shaded blue) for photosynthesis  $P_1$  and respiration  $R_1$ .

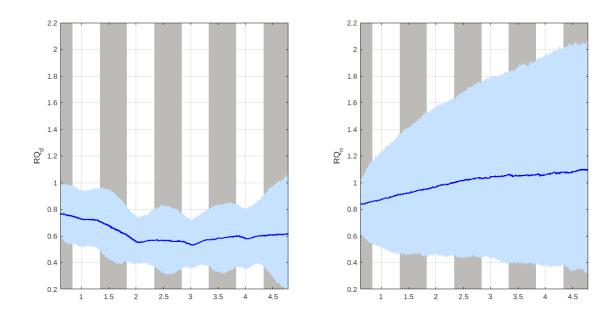


Figure 4.16 : Posterior medians (solid blue line) and 95% credible intervals (shaded blue) for  $RQ_d$  and  $RQ_n$ .

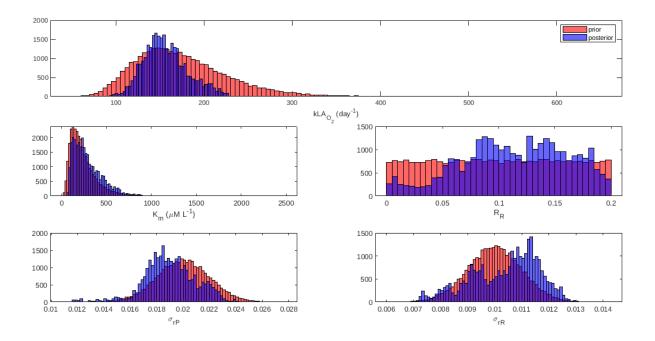


Figure 4.17: Priors (pink) and posteriors (purple) for model parameters.

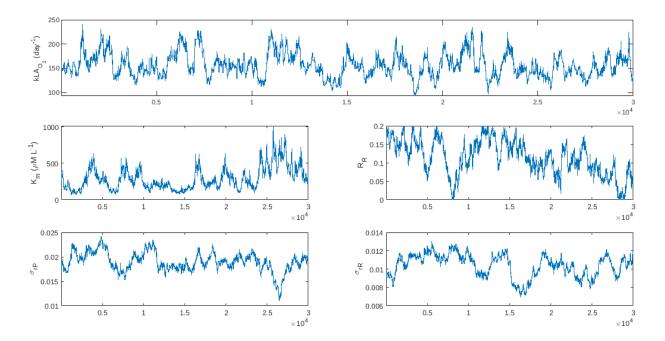


Figure 4.18: Traces for model parameters.

### Appendix A

### LiBbi model code

### LiBbi model file: micro\_iterative.bi

```
1 model micro_iterative {
3 const FO2
                 = 0.2094
4 const FCO2
                 = 397e-6
5 const S
                  = 34.0
6 const V
                 = 500.0
                                          // volume of the
     reactor
                 = 1724.20
7 const DIC_M
                                          // calculated with
     CO2SYS[DIC_M = 1724.20, Alk = 1797.90, T = 27, S = 34]
                = 226.65
8 const O_2_M
                 = 1797.90
9 const alk_M
10 const tau
                 = 6.0
11 const kLA02_m = log(2.0)*24.0*60.0/tau
12
13 param kLAO2
14 param Km
15 param RR
 param RQ_d
17 param RQ_n
 param sigma_0_2
19 param sigma_pH
20 param sigma_DIC
```

```
_{21} param offset_0_2
22
23 input I
                           // light intensity
                           // temperature (C)
24 input T
  input gas
                           // gas on/off
                           // dilution rate
26 input dil
27
28 state DIC // state variables
29 state 0_2
30 state pH
31 state Cp
32 state mich_ment
33 state O2H_pr
34 state CO2H_pr
35 state R
36 state R1
37 state P
38 state P1
39 state alk
40 state CO2
41 state HCO3
42 state CO3
43 state O_2H
44 state CO2H
45 state h_3
46 state h_free_3
47
48 noise r_R
49 noise r_P
```

```
50
  /* random walk parameter */
51
  param sigma_r_R
  param sigma_r_P
54
  obs 02_obs
55
  obs pH_obs
  obs DIC_obs
  obs alk_obs
59
  sub parameter {/* prior distribution over parameters */
           ~ log_normal(log(100.0), 0.5)
  Km
61
  kLA02
          ~ log_normal(log(kLAO2_m), 0.3)
           ~ uniform(0.0001, 0.2)
  RR
  RQ_d
          " uniform(0.66, 1.0)
           ~ uniform(0.66, 1.0)
  RQ_n
65
66
   sigma_0_2 ~ log_normal(log(0.03), 0.5)
   sigma_pH ~ log_normal(log(0.03), 0.5)
68
   sigma_DIC \sim log_normal(log(0.03), 0.5)
69
70
   offset_0_2 ~ normal(0, 2.0)
71
72
                   ~ normal(0.01, 0.001)
  sigma_r_R
73
                   ~ normal(0.05, 0.01)
  sigma_r_P
75
76
  const prop_std = 0.1;
78 sub proposal_parameter {
```

```
~ log_normal(log(Km), 0.5*prop_std)
79 Km
           ~ log_normal(log(kLAO2), 0.3*prop_std)
  kLAO2
80
           ~ truncated_normal(RR, 0.2*prop_std, lower = 0.0001,
81 RR
      upper = 0.2)
         ~ truncated_normal(RQ_d, 0.2*prop_std, lower = 0.66,
      upper = 1.0)
83 RQ_n ~ truncated_normal(RQ_n, 0.2*prop_std, lower = 0.66,
      upper = 1.0)
84
85
   sigma_0_2
                    ~ log_normal(log(sigma_0_2), 0.5*prop_std)
86
                    ~ log_normal(log(sigma_pH), 0.5*prop_std)
   sigma_pH
87
   sigma_DIC
                    ~ log_normal(log(sigma_DIC), 0.5*prop_std)
89
   offset_0_2
                    ~ normal(offset_0_2, 2.0*prop_std)
90
91
                    ~ normal(sigma_r_R, 0.001*prop_std)
   sigma_r_R
92
                    ~ normal(sigma_r_P, 0.01*prop_std)
   sigma_r_P
  }
94
95
   sub initial {/* prior distribution over initial conditions,
      given parameters */
   // specify the initial condition model
           ~ normal(log(20.0), 0.4)
98
           ~ log_normal(log(20.0), 0.4)
99
           ~ normal(log(200.0), 0.4)
100
           ~ log_normal(log(200.0), 0.4)
  Ρ1
101
102
           ~ log_normal(log(300.0), 0.2)
103 Cp
```

```
~ log_normal(log(1750.0), 0.1)
  alk
104
            ~ log_normal(log(1300.0), 0.2)
105 DIC
            ~ log_normal(log(225.0), 0.2)
106 0_2
            ~ log_normal(log(8.5), 0.2)
107
  рΗ
           ~ log_normal(log(3.0), 0.4)
108 CO2
           ~ log_normal(log(1000.0), 0.3)
109
   HC03
110 CO3
           ~ log_normal(log(300.0), 0.4)
           ~ log_normal(log(200.0), 0.2)
111 O_2H
            ~ log_normal(log(10.0), 0.2)
  CO2H
113
  }
114
115
116 //sub transition(delta = 0.0023) { // obs are in days ie
      delta=1.0 for daily solving. delta=0.00069 for solving
      every minute, 0.0014 for every 2 mins, 0.0021 for 3 mins,
      0.0028 for 4mins, delta=0.000011574 for solving every
      second
   sub transition(delta = 0.0021) {
118
   /* processes */
119
120
121 inline TK
                 = T + 273.15
                                             // temp in kelvin
  inline KO_CO2 = exp(-60.2409 + 93.4517*(100.0/TK) + 23.3585*
      log(TK/100.0) + S*(0.023517 - 0.023656*(TK/100) +
      0.0047036*(TK/100.0)*(TK/100.0)))
   CO2H
                  <- K0_C02*FC02*1.0220*1e6
123
124
  inline KO_02 = (exp(-1282.8704 + 36619.96/TK + 223.1396*log)
125
      (TK) -0.354707*TK + S*(5.957e-3 -3.7353/TK) + 3.68e-6*S*S)
```

```
)/(0.2094e-06)
126 O_2H
               <- K0_02*F02*1.0220*1e-6
127
128 inline PAC = HCO3
                                   //PAC=photosynthetically
      active carbon. if the phyto are just using CO2 to
      photosynthesise then PAC=CO2
  inline mm = PAC/(Km + PAC)
129
130
131 // CO2SYS iterative solution
  // set up all the constants
133
134 inline logTK = log(TK)
135 inline S2 = S*S
  inline sqrtS = sqrt(S)
137
138 // total sulphur
139
inline TS = (0.14/96.062)*(S/1.80655)
             = 19.924*S/(1000.0 - 1.005*S)
141 inline IS
142
inline KS_{int} = -4276.1/TK + 141.328 - 23.093*logTK +
      (-13856.0/TK + 324.57 - 47.986*logTK)*sqrt(IS) + (
      35474.0/TK - 771.54 + 114.723*logTK)*IS - 2698.0/TK*IS
      **1.5 + 1776.0/TK*IS**2
  inline KS = \exp(KS_{int})*(1 - 0.001005*S)
144
145
146 // Fluorine
147
148 inline TF
                  = 0.000067*S/18.9984/1.80655
```

```
= \exp(-(-874.0) \text{TK} - 0.111 * \text{sqrtS} + 9.68))
149 inline KF
  inline SWS_2T = (1.0 + TS/KS)/(1.0 + TS/KS + TF/KF)
  inline Free_2_T = 1.0 + TS/KS
151
152
153 // H2O dissoc
154
   inline KW = \exp(148.9802 - 13847.26/TK - 23.6521*logTK +
      (118.67/TK - 5.977 + 1.0495*logTK)*sqrtS - 0.01615*S)
156
  // Boron
158
  inline KB = exp((-8966.90 - 2890.53*sqrtS - 77.942*S + 1.728*
      S*sqrtS - 0.0996*S2)/TK + 148.0248 + 137.1942*sqrtS +
      1.62142*S - (24.4344 + 25.085*sqrtS + 0.2474*S)*logTK +
      0.053105*sqrtS*TK)
  inline TB = 0.0004326*S/35.0
161
162 // Carbon eq constants
163
  inline K1 = 10**(-(3633.86/TK - 61.2172 + 9.6777 *logTK -
164
      0.011555*S + 0.0001152*S**2))*1.23 //1.23 experiment
      specific and measured
inline K2 = 10**(-(471.8/TK + 25.9290 - 3.16967*logTK -
      0.01781*S + 0.0001122*S**2))*0.53
                                         //0.53 experiment
      specific and measured
166
   // end all the constants
167
168
169 // intial guess at the pH (use the approximating equation)
```

```
170
inline pH_init = 12.26 -0.0030605*DIC -0.043752*T -0.013625*S
      + 0.00011315*alk + 1.3463e-05*DIC*T + 5.2215e-07*DIC*alk
172
173 // iteration 1
174
inline h_1 = 10.0**(-pH_init)
inline h_free_1 = h_1/Free_2_T
inline f0_1 = (DIC*1e-6*(K1*h_1 + 2.0*K1*K2)/(h_1*h_1 +
      K1*h_1 + K1*K2) - h_free_1 + KW/h_1 - alk*1e-6 + TB/(1.0 +
      h_1/KB) *1e6
inline df0_1 = (DIC*1e-6*(K1 + 2.0*K1*K2))/(h_1**2.0 + K1*)
      h_1 + K1*K2) - DIC*1e-6*(K1*h_1 + 2.0*K1*K2)/(h_1**2.0 + 6.0*K1*K2)
      K1*h_1 + K1*K2)**2.0*(2.0*h_1 + K1) - TB*1.0/(1.0 + h_1/KB)
      )**2.0/KB - KW/h_1**2.0 - 1.0/Free_2_T)*1e6*(-log(10.0)
      *10.0**(-pH_init))
inline pH_1 = pH_init - f0_1/df0_1
180
181 // iteration 2
182
inline h_2 = 10.0**(-pH_1)
inline h_free_2 = h_2/Free_2_T
185 inline f0_2
                = (DIC*1e-6*(K1*h_2 + 2.0*K1*K2)/(h_2*h_2 +
      K1*h_2 + K1*K2) - h_free_2 + KW/h_2 - alk*1e-6 + TB/(1.0 + FR)
       h_2/KB))*1e6
inline df0_2 = (DIC*1e-6*(K1 + 2.0*K1*K2)/(h_2**2.0 + K1*)
      h_2 + K1*K2) - DIC*1e-6*(K1*h_2 + 2.0*K1*K2)/(h_2**2.0 +
      K1*h_2 + K1*K2)**2.0*(2.0*h_2 + K1) - TB*1.0/(1.0 + h_2/KB)
      )**2.0/KB - KW/h_2**2.0 - 1.0/Free_2_T)*1e6*(-log(10.0)
```

```
*10.0**(-pH_1))
inline pH_2 = pH_1 - f0_2/df0_2
188
189 // iteration 3
190
191 h_3
                   <-10.0**(-pH_2)
192 h_free_3
                  <- h_3/Free_2_T
inline f0_3 = (DIC*1e-6*(K1*h_3 + 2.0*K1*K2)/(h_3*h_3 +
      K1*h_3 + K1*K2) - h_free_3 + KW/h_3 - alk*1e-6 + TB/(1.0 + B)
      h_3/KB))*1e6
inline df0_3 = (DIC*1e-6*(K1 + 2.0*K1*K2))/(h_3**2.0 + K1*)
      h_3 + K1*K2) - DIC*1e-6*(K1*h_3 + 2.0*K1*K2)/(h_3**2.0 + 2.0*K1*K2)
      K1*h_3 + K1*K2)**2.0*(2.0*h_3 + K1) - TB*1.0/(1.0 + h_3/KB)
      )**2.0/KB - KW/h_3**2.0 - 1.0/Free_2_T)*1e6*(-log(10.0)
      *10.0**(-pH_2))
                  <- pH_2 - f0_3/df0_3
195 pH
196
197 // iteration 4
198
199 //
          inline h_4 = 10.0**(-pH_3)
          inline h_free_4 = h_4/Free_2_T
200 //
201 //
          inline f0_4 = (DIC*1e-6*(K1*h_4 + 2.0*K1*K2)/(h_4)
      *h_4 + K1*h_4 + K1*K2) - h_free_4 + KW/h_4 - alk*1e-6 + TB
      /(1.0 + h_4/KB))*1e6
202 //
          inline df0_4 = (DIC*1e-6*(K1 + 2.0*K1*K2)/(h_4)
      **2.0 + K1*h_4 + K1*K2) - DIC*1e-6*(K1*h_4 + 2.0*K1*K2)/(
      h_4**2.0 + K1*h_4 + K1*K2)**2.0*(2.0*h_4 + K1) - TB
      *1.0/(1.0 + h_4/KB)**2.0/KB - KW/h_4**2.0 - 1.0/Free_2_T)
      *1e6*(-log(10.0)*10.0**(-pH_3))
```

```
203 //
         inline pH_4 = pH_3 - f0_4/df0_4
204
205 //
          рΗ
                            <- pH_4
206
  // calculate the final concentrations
208
209 inline H
                   = 10.0**(-pH)
210 inline H2
                   = H * H
211 inline denom
                   = (H2 + K1*H + K1*K2)
                   <- DIC*H2/denom
212 CO2
213 HCO3
                   <- DIC*H*K1/denom
214 CO3
                   <- DIC*K1*K2/denom
215
_{216} // end CO2SYS iterative solution
217
218
219 /* R and P as random walks */
220
221 r_R
          ~ normal(0.0, sigma_r_R)
222 R
           <-R+r_R
223 R1
          <- exp(R)
224
r_P \sim normal(0.0, sigma_r_P)
          <- P + r_P
226 P
           <- exp(P)
227 P1
228
229 ode(h = 0.1, atoler = 1.0e-6, rtoler = 1.0e-6, alg = 'RK4(3)
      '){
```

```
230 dDIC/dt = -P1*24.0*I*mm + R1*24.0
             + gas*0.893*kLA02*(CO2H - CO2) + dil/V*(
      DIC_M - DIC)
d0_2/dt = (P1*24.0*I*mm - R1*24.0)/(RQ_d*I + RQ_n*(1.0-I))
         + gas*kLAO2*(0_2H - 0_2)
                                                 + dil/V*(0_2_M
      - 0_2) + offset_0_2
232 dalk/dt = RR*P1*24.0*I*mm
                                                      + dil/V*(
      alk_M - alk)
233 dCp/dt = (P1*24.0*I*mm - R1*24.0)
                                                      + dil/V*(
      Cp)
234
235 }
236
237 mich_ment <- mm
238 O2H_pr <- O_2H
239 CO2H_pr <- CO2H
240
241 }
242
243
244 sub observation {
245
246 02_{obs} ~ log_normal(log(0_2), sigma_0_2)
247 pH_obs ~ log_normal(log(pH), sigma_pH)
248 DIC_obs ~ log_normal(log(DIC), sigma_DIC)
249 alk_obs ~ log_normal(log(alk), sigma_DIC)
250 }
```

```
251 }
```

### LiBbi prior sampling file: prior.conf

```
1 --target prior
2 --model-file micro_iterative.bi
3 --nsamples 500
4 --start-time 0.61304
5 --end-time 4.7866
6 --noutputs 6049
7 --input-file data/input_all_2018_normalised.nc
8 --output-file results/prior_micro_iterative.nc
```

### LiBbi posterior sampling file: posterior.conf

```
--target posterior

--model-file micro_iterative.bi

--input-file data/input_all_2018_normalised.nc

--obs-file data/obs_all_2018.nc

--nsamples 500

--nparticles 1024

--start-time 0.61304

--end-time 4.7866

--noutputs 6049

--output-file results/posterior_micro_iterative.nc

--with-transform-initial-to-param
```



Figure A.1 : Directed Acyclic Graph of the LiBbi model file micro\_iterative.bi

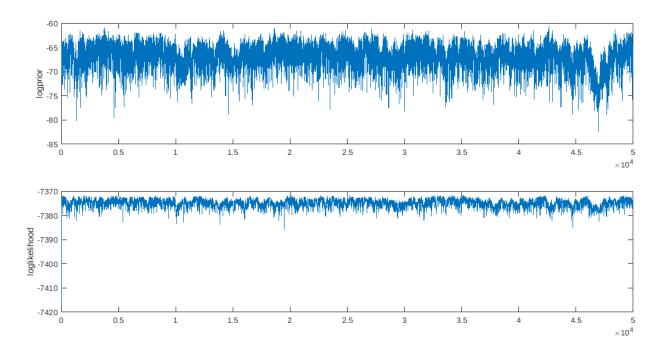


Figure A.2 : Log-prior and log-likelihood for the simulated data experiment.

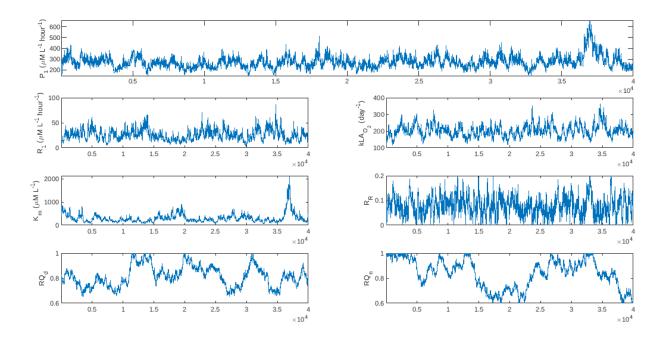


Figure A.3: Parameter posterior traces for the simulated data experiment.

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