



Acknowledgments

- National Institute for Mathematical and Biological Synthesis (NIMBioS)
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- bioRxiv
- Paul Bastide
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- Luke Harmon
- Anonymous reviewers of Systematic Biology

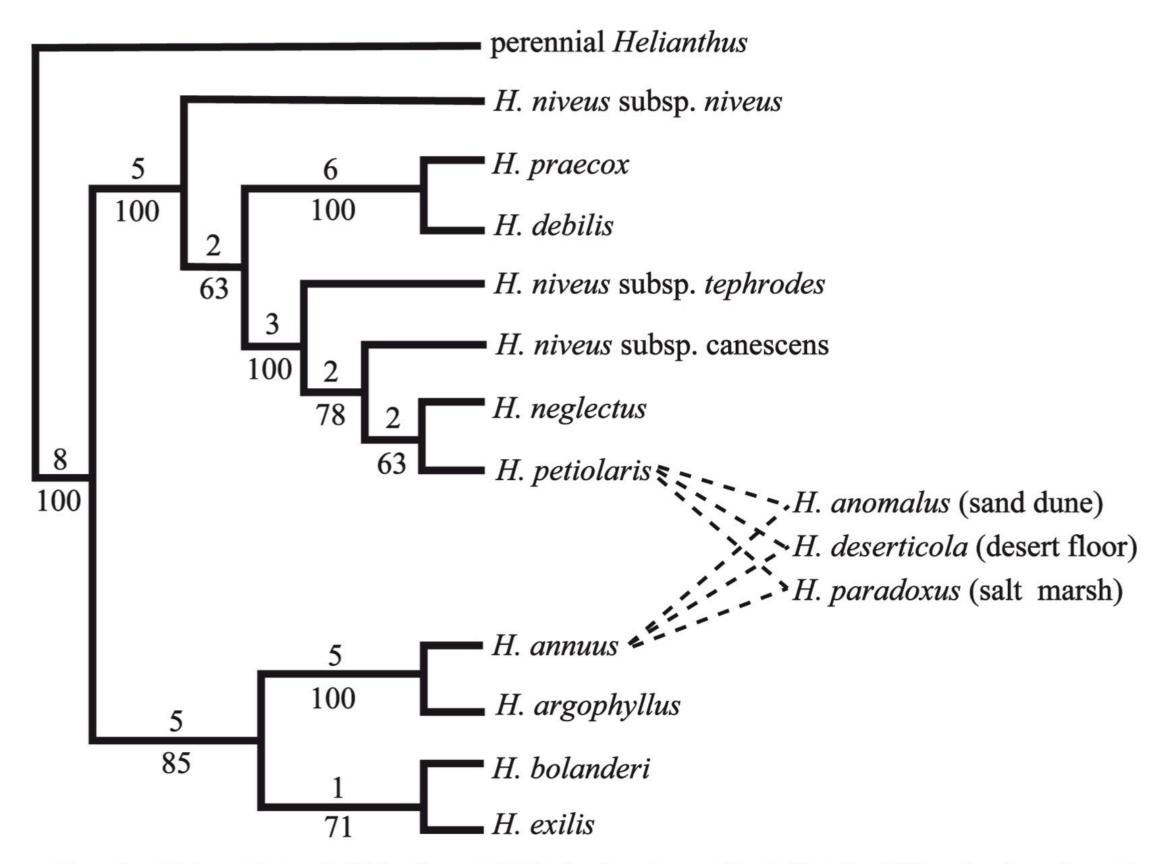


Figure 1. Phylogenetic tree for *Helianthus* sect. *Helianthus* based on combined chloroplast DNA and nuclear ribosomal DNA data (redrawn from Rieseberg, 1991, fig. 9). The number of mutations are given above and bootstrap percentages below each branch. Dashed lines indicate parentage of homoploid hybrid species.

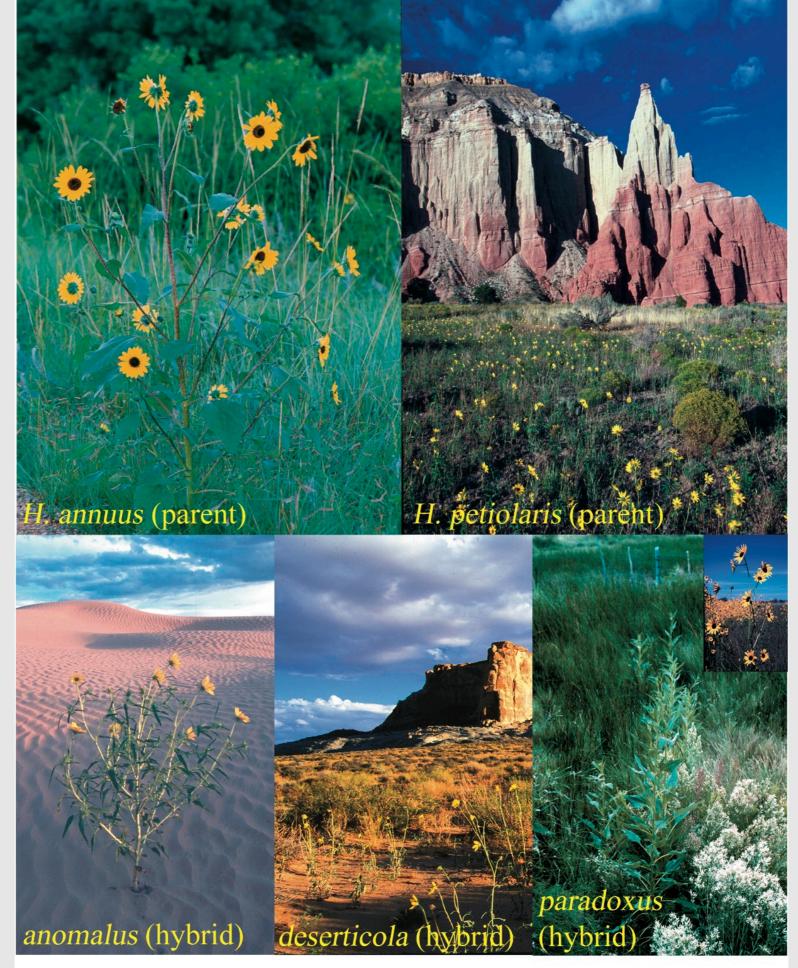
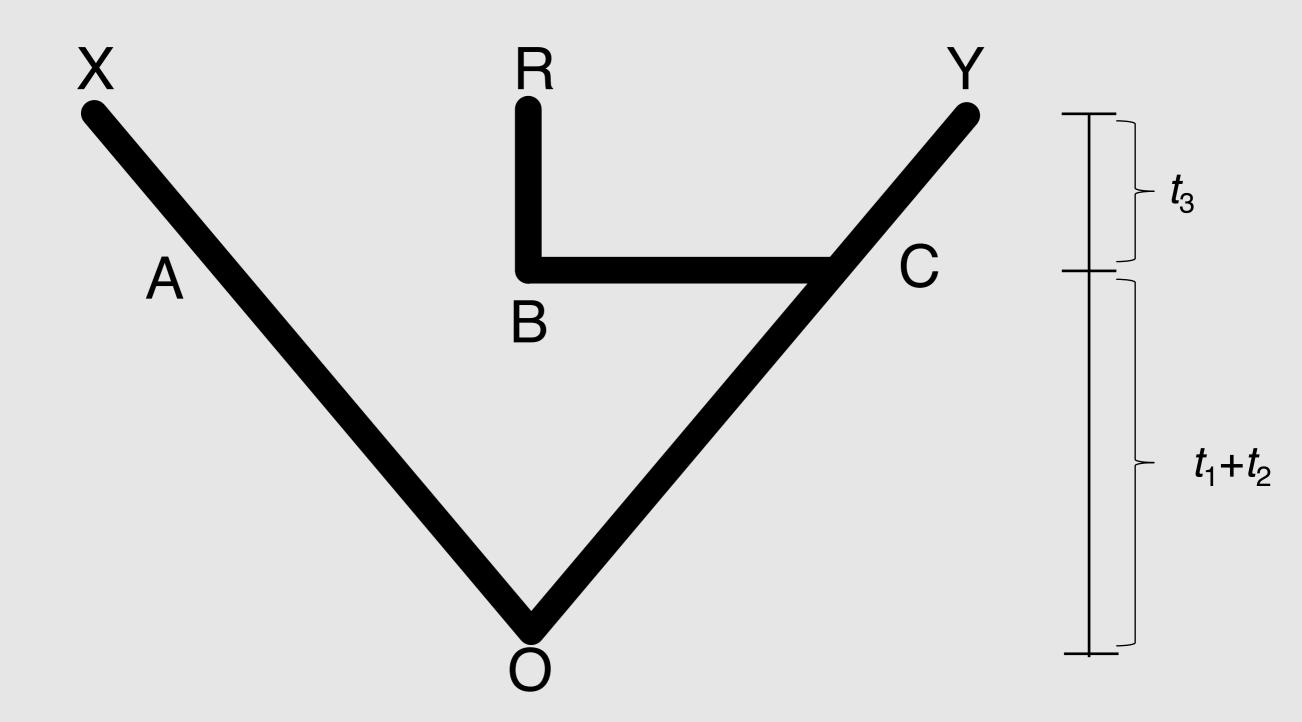


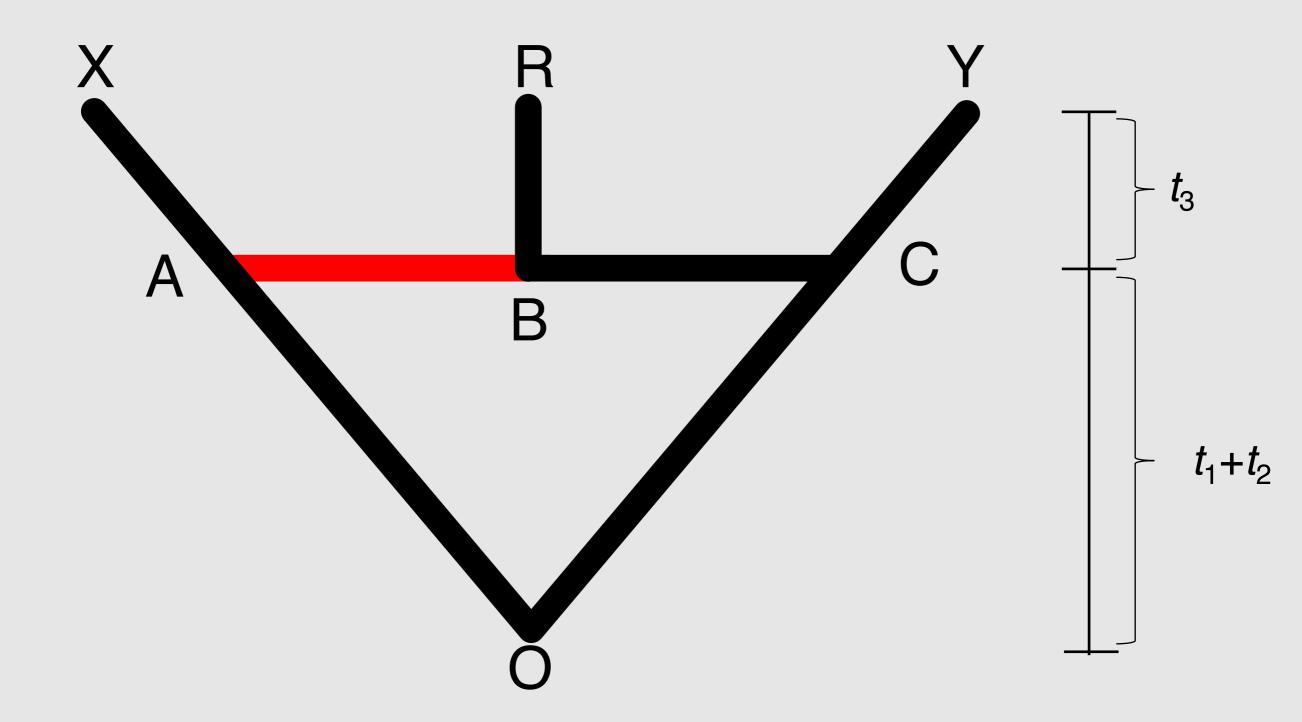
Figure 3. Photographs of the two parental species and their three hybrid derivative species in typical habitats. Photographs by Jason Rick.

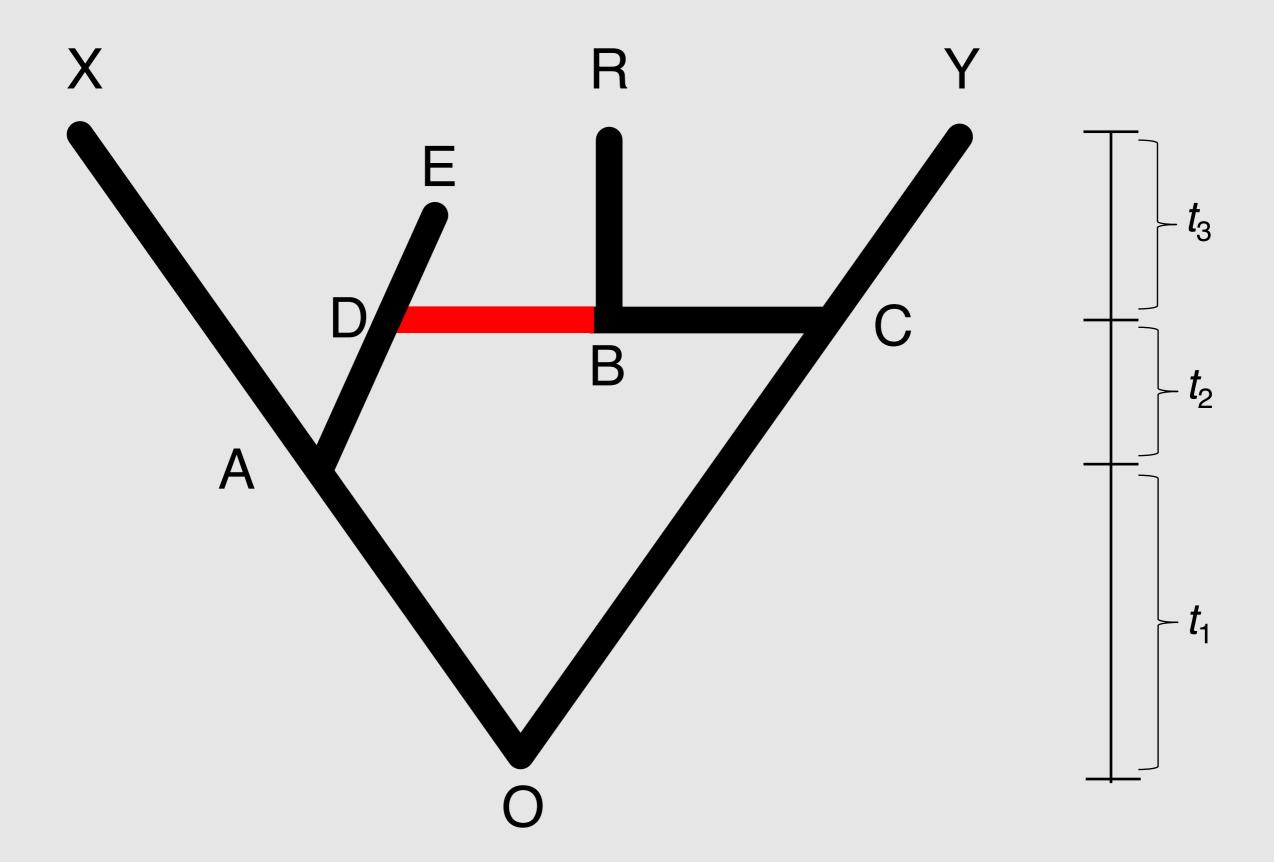
Rieseberg 2006 Photos by James Rick

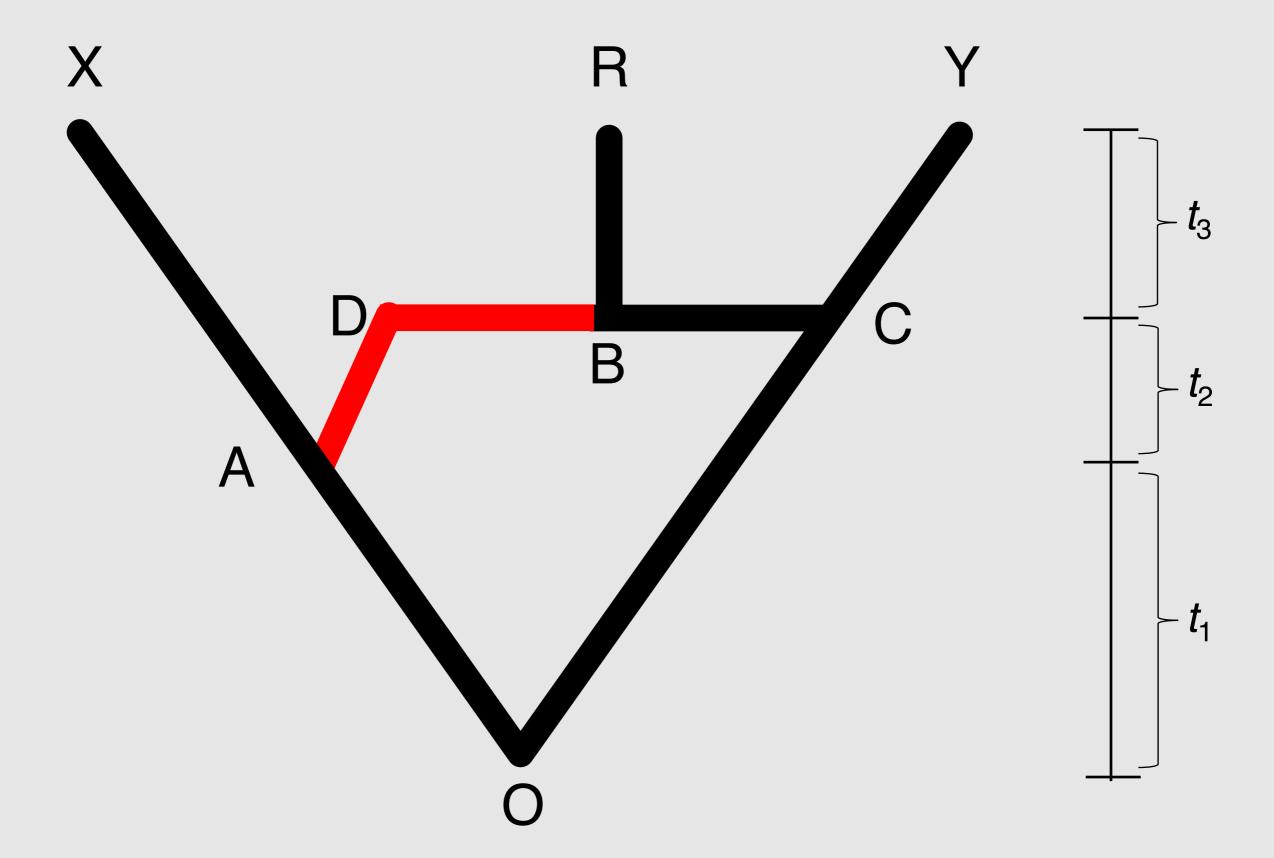
Model requirements

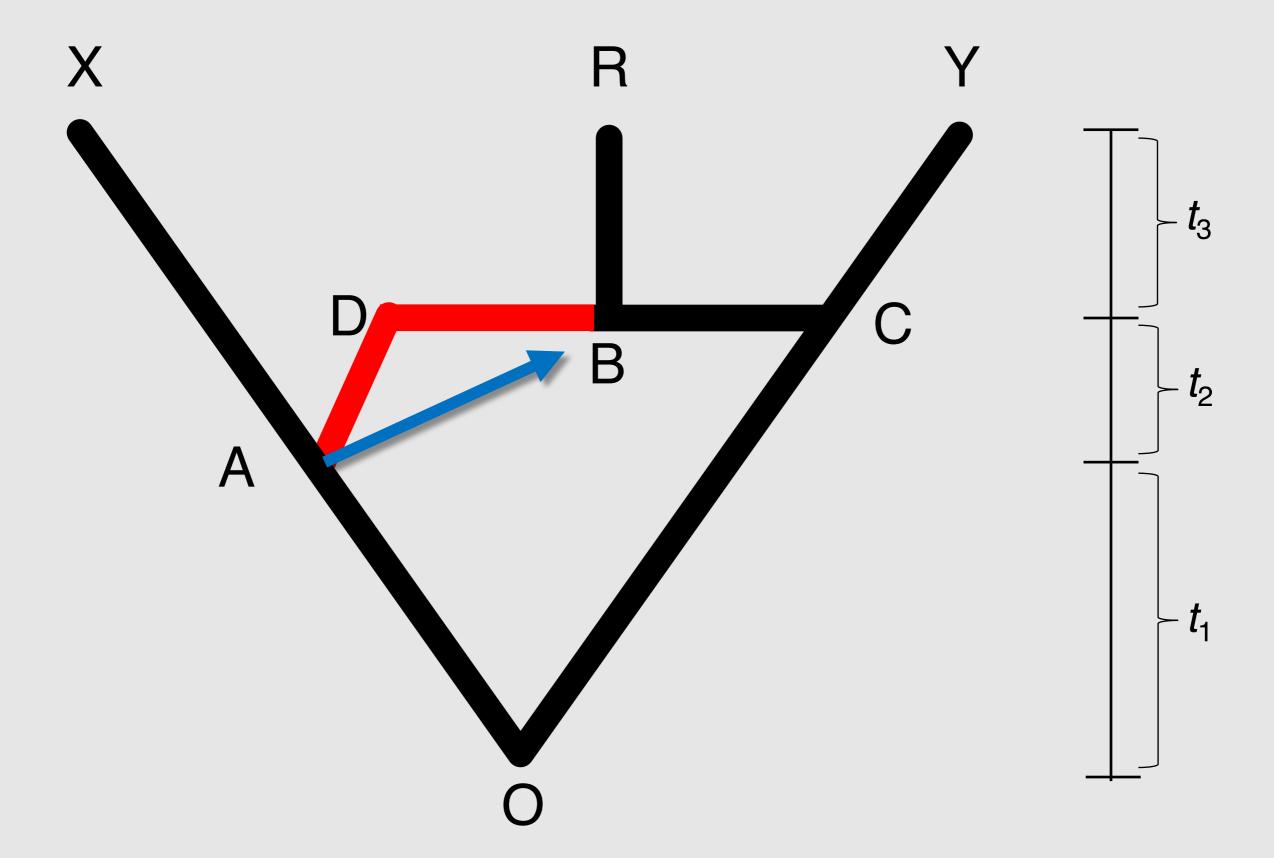
- Work on networks, not just trees
- Get parameter estimates
- Estimate mean shifts (i.e., hybrids bigger than parents)
- Estimate increased variation (hybrids weirder than parents)
- Incorporate measurement error (life happens)

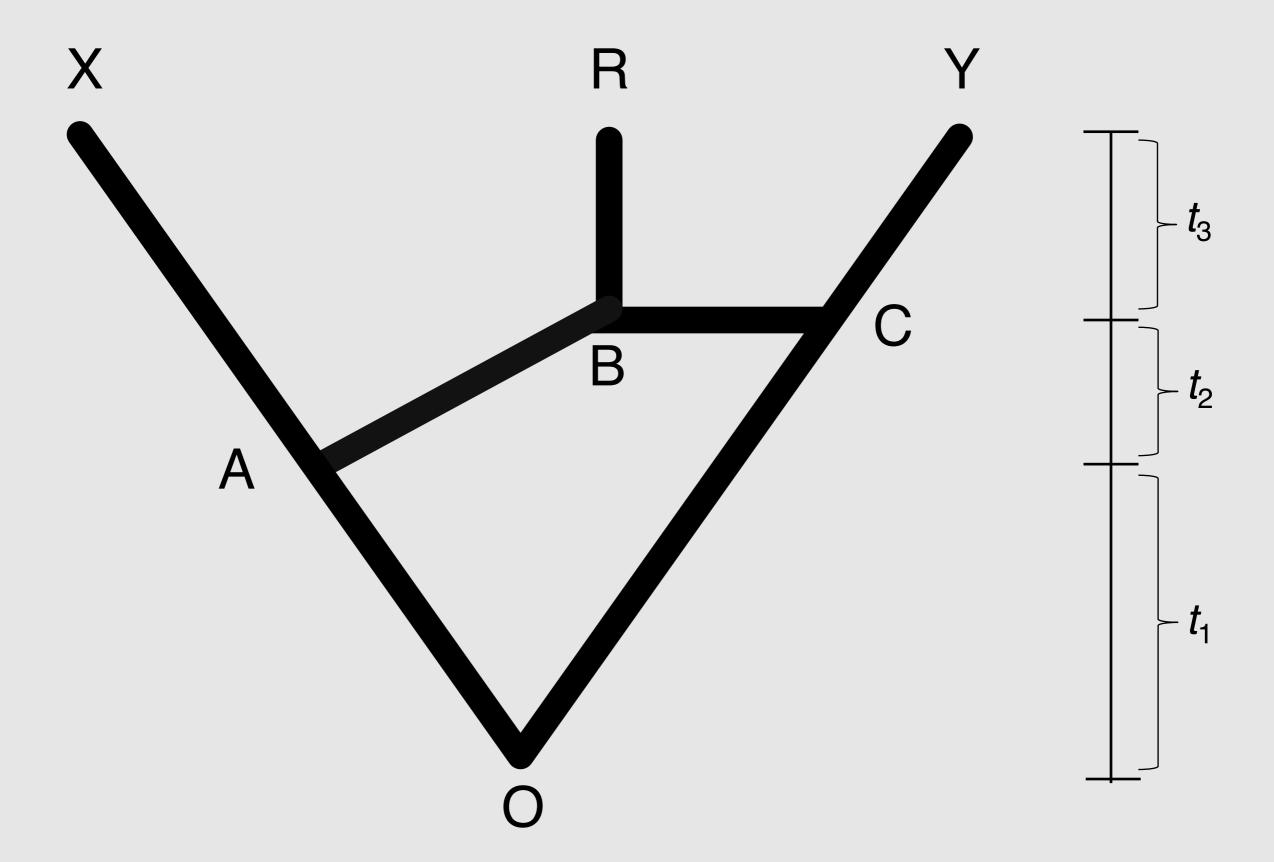












$$f(x) = \frac{1}{\sqrt{2\pi}\sqrt{|\Sigma|}} \exp\left(-\frac{1}{2}(x-\mu)'\Sigma^{-1}(x-\mu)\right)$$

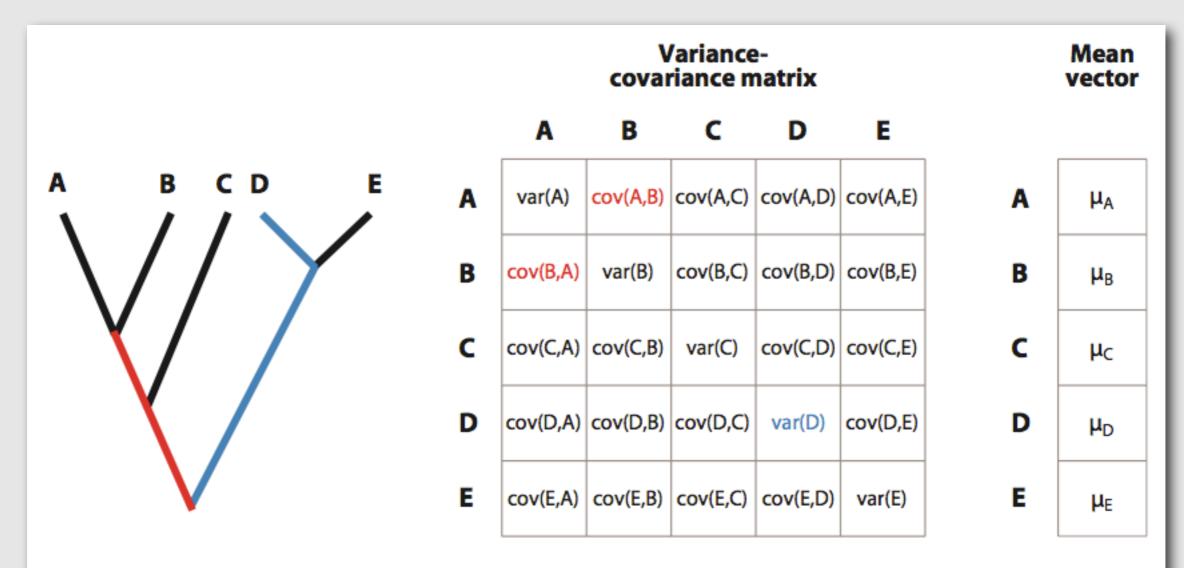
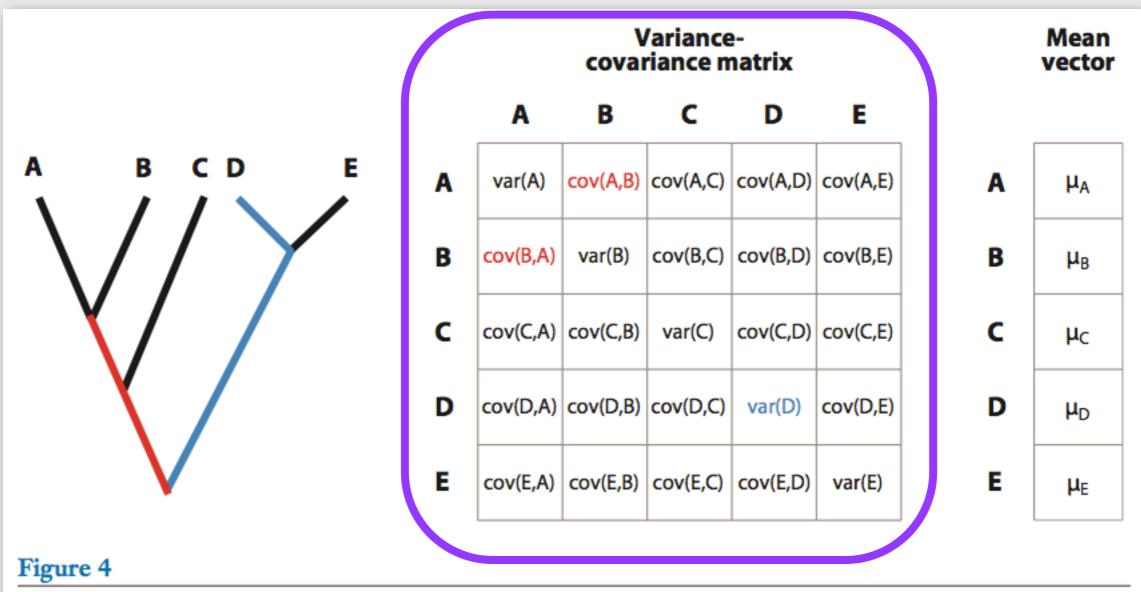


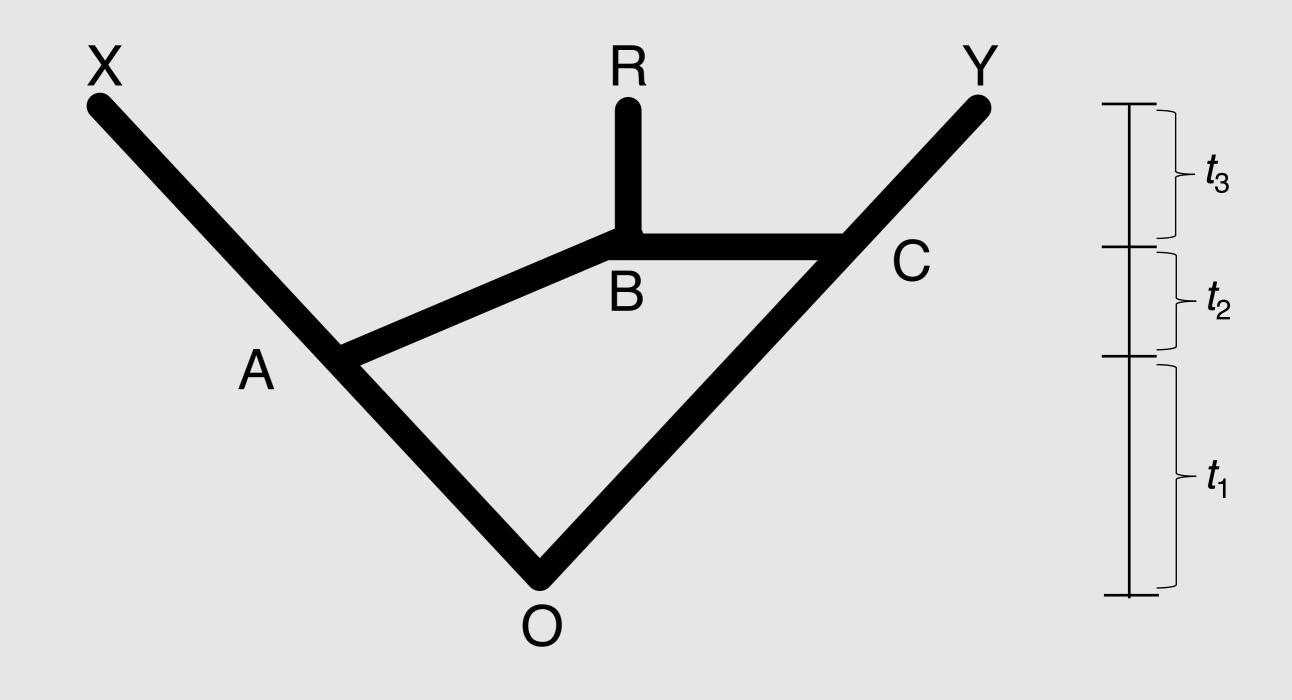
Figure 4

Multivariate normal distribution. The figure shows a tree, the tree's variance-covariance matrix, and the vector of means (which, under Brownian motion, would equal the root state). Highlighted are the branches leading to covariance between taxa A and B (red) and the branches leading to variance in D (blue).

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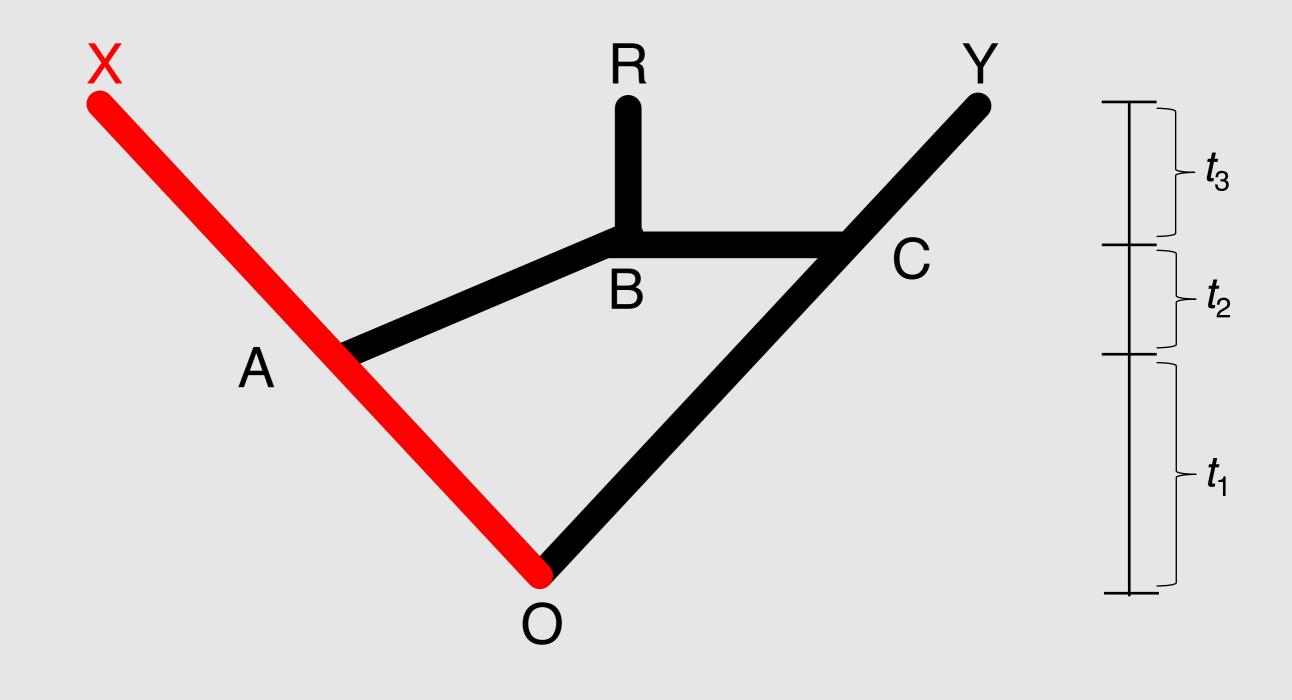


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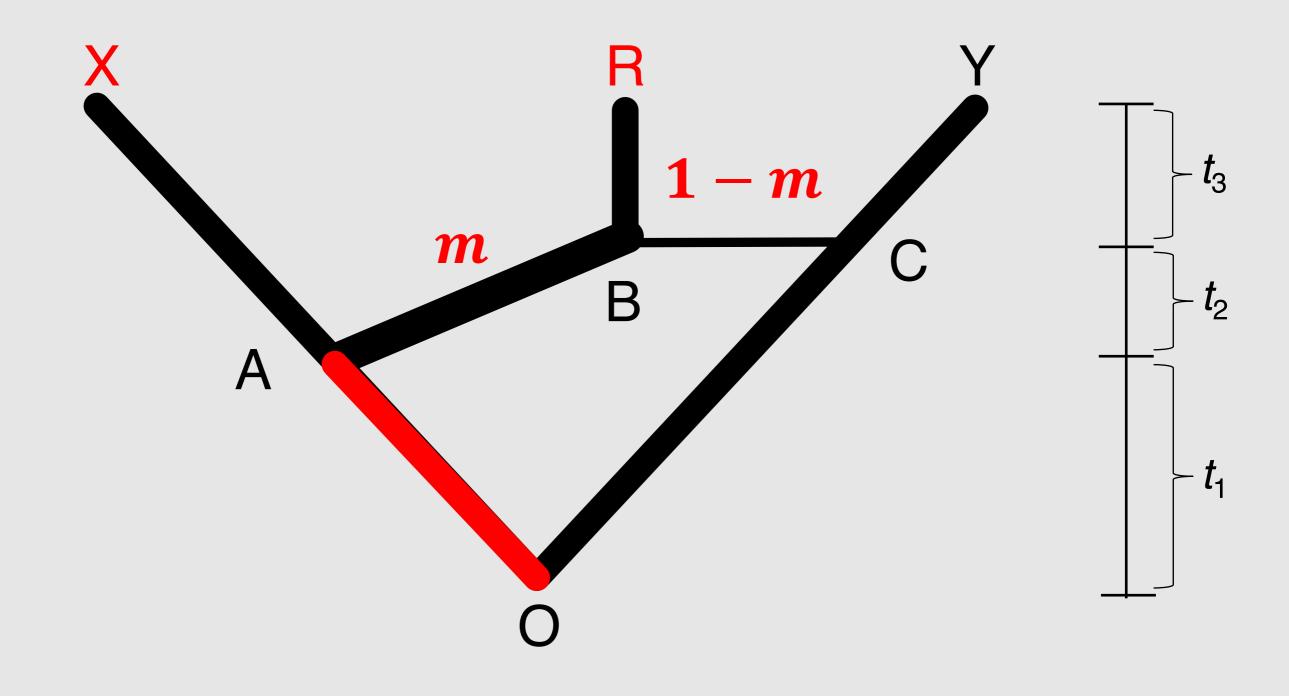
X R Y

$$\begin{bmatrix} \sigma^{2}(t_{1}+t_{2}+t_{3}) & \sigma^{2}mt_{1} & 0 \\ \sigma^{2}mt_{1} & \sigma^{2}(m^{2}+(1-m)^{2})(t_{1}+t_{2})+\sigma^{2}t_{3}+v_{H} & \sigma^{2}(1-m)(t_{1}+t_{2}) \\ 0 & \sigma^{2}(1-m)(t_{1}+t_{2}) & \sigma^{2}(t_{1}+t_{2}+t_{3}) \end{bmatrix}$$



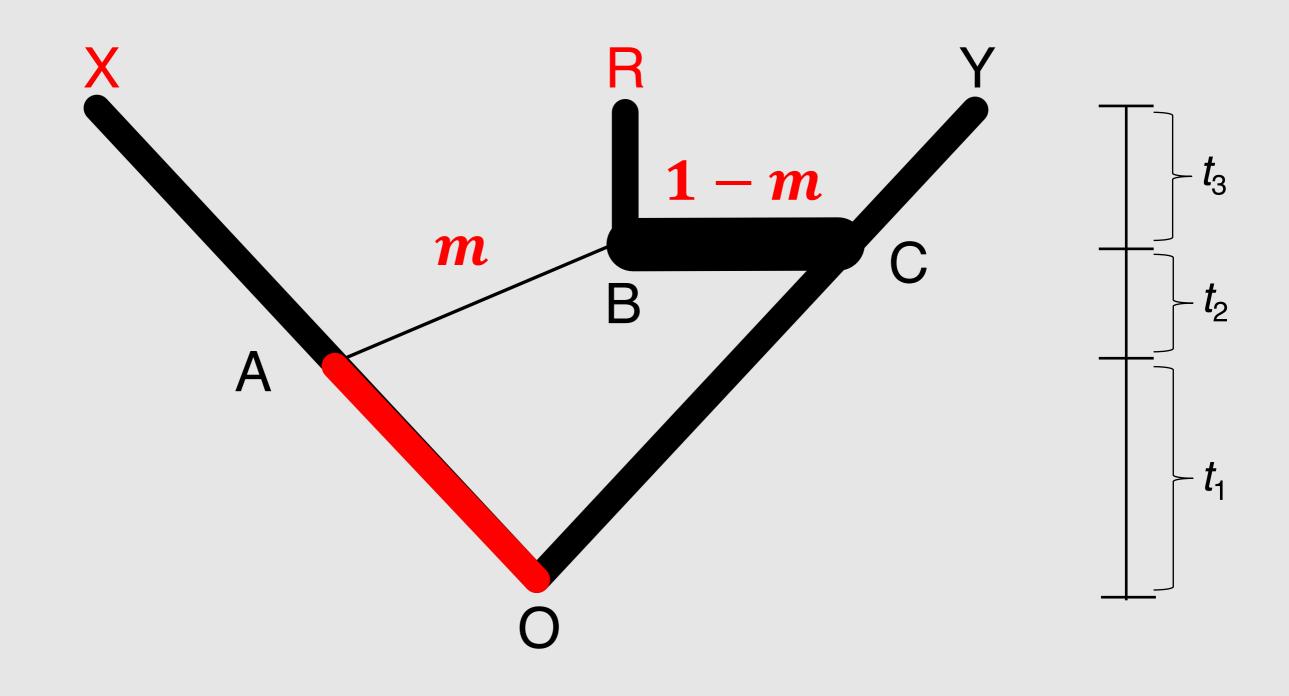
X Y

$$\begin{bmatrix}
\sigma^{2}(t_{1}+t_{2}+t_{3}) & \sigma^{2}mt_{1} & 0 \\
\sigma^{2}mt_{1} & \sigma^{2}(m^{2}+(1-m)^{2})(t_{1}+t_{2})+\sigma^{2}t_{3}+v_{H} & \sigma^{2}(1-m)(t_{1}+t_{2}) \\
0 & \sigma^{2}(1-m)(t_{1}+t_{2}) & \sigma^{2}(t_{1}+t_{2}+t_{3})
\end{bmatrix}$$



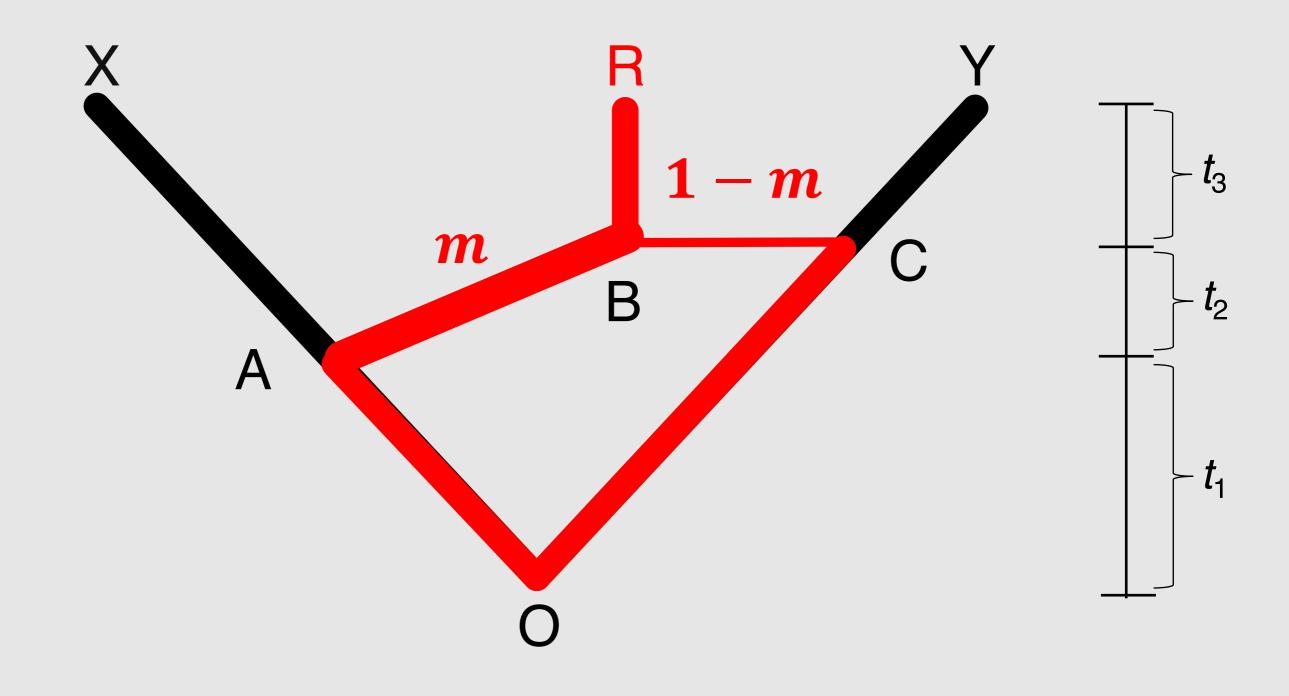
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\end{bmatrix}$$



X Y

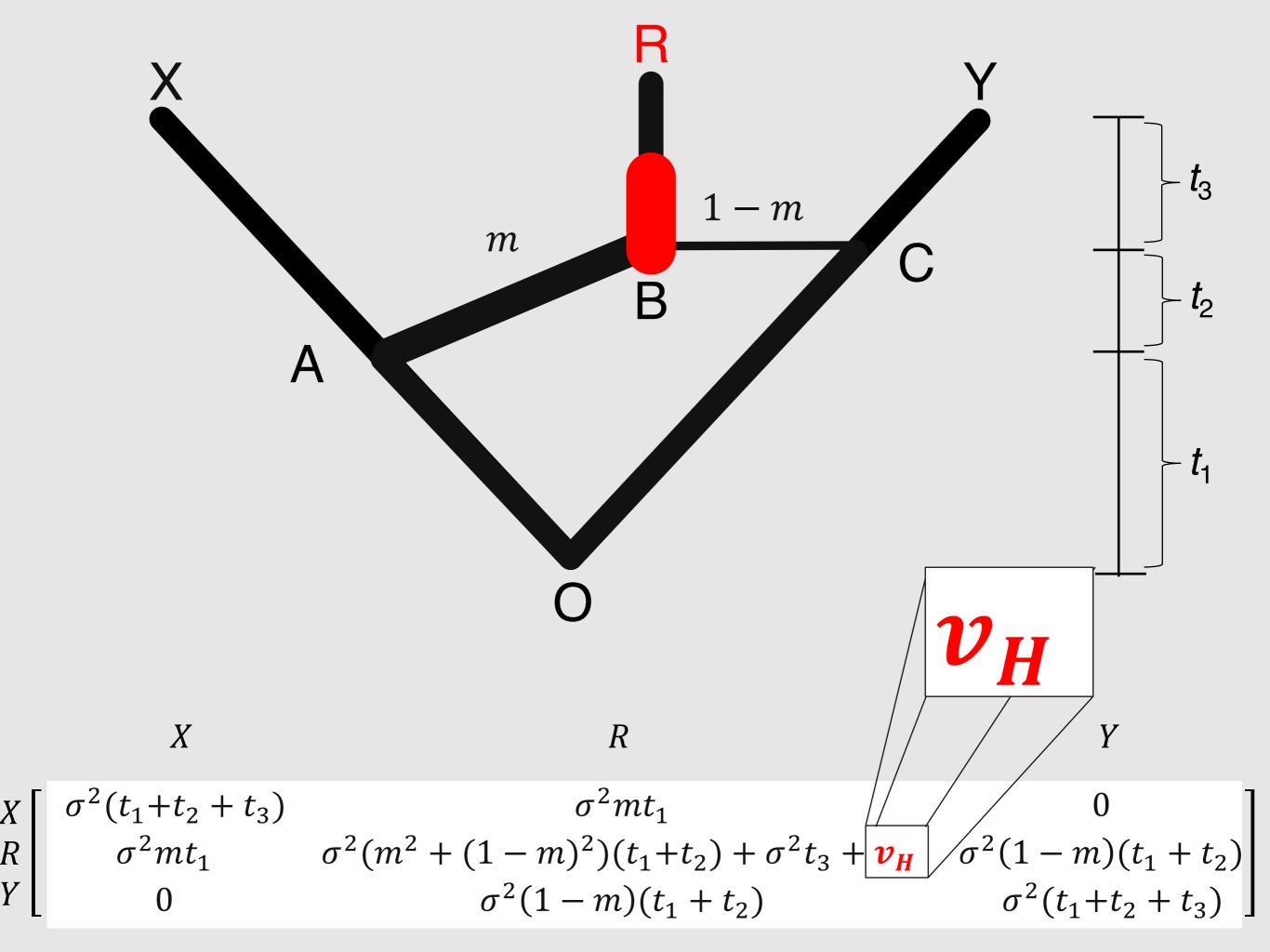
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\end{bmatrix}$$



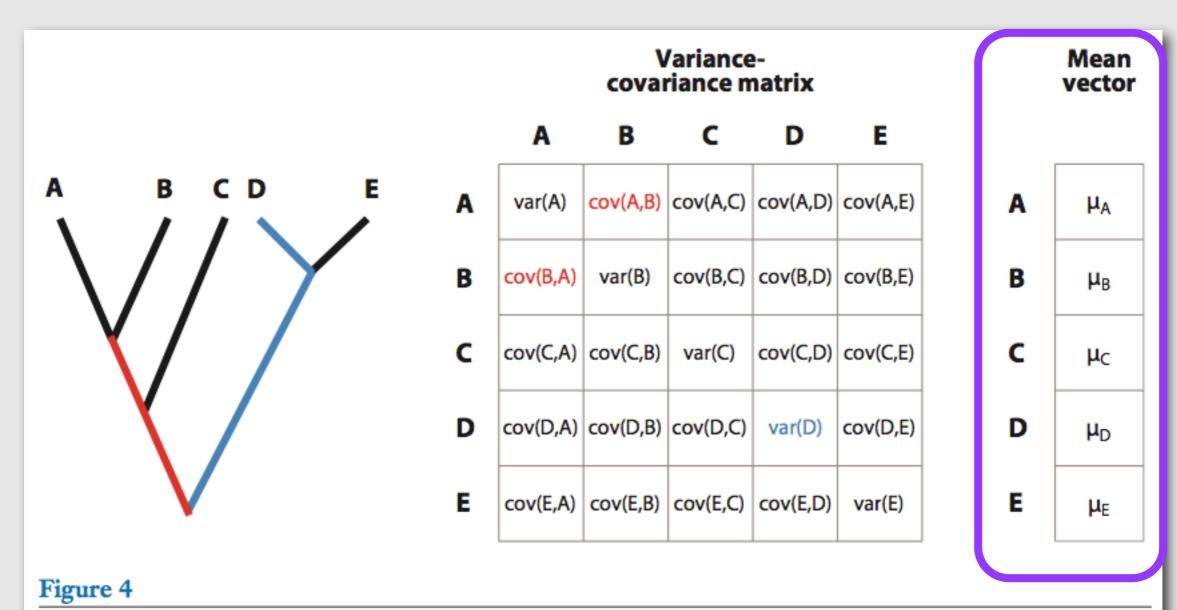
$$\sigma^{2}(t_{1}+t_{2}+t_{3}) \qquad \sigma^{2}mt_{1} \qquad 0$$

$$\sigma^{2}mt_{1} \qquad \sigma^{2}(m^{2}+(1-m)^{2})(t_{1}+t_{2})+\sigma^{2}t_{3}+v_{H} \qquad \sigma^{2}(1-m)(t_{1}+t_{2})$$

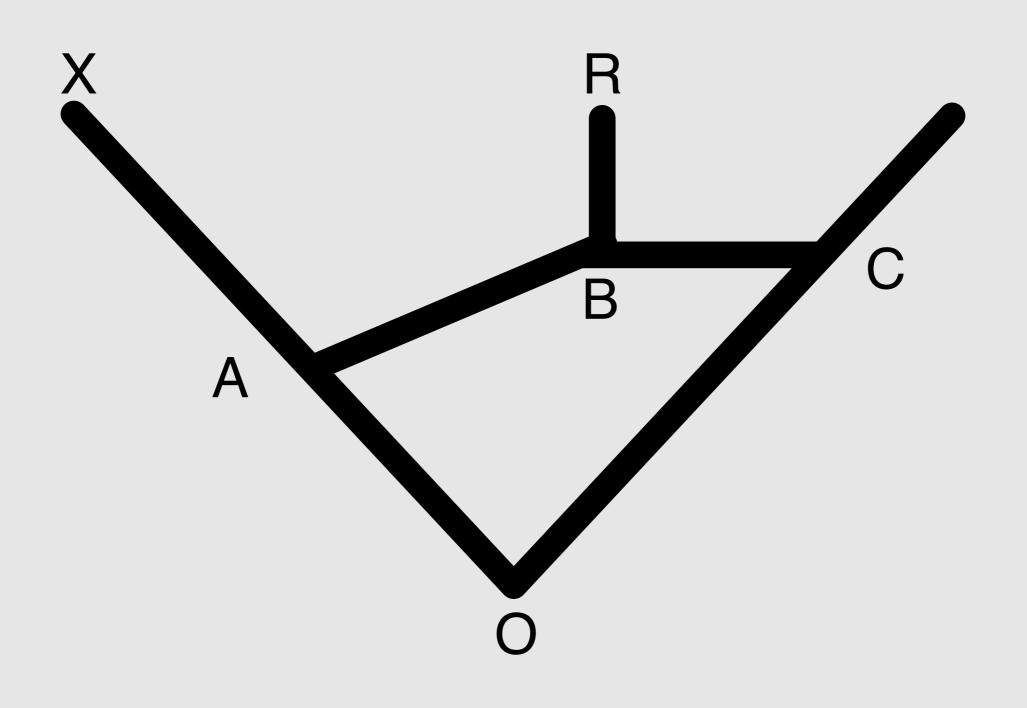
$$\sigma^{2}(1-m)(t_{1}+t_{2}) \qquad \sigma^{2}(t_{1}+t_{2}+t_{3})$$



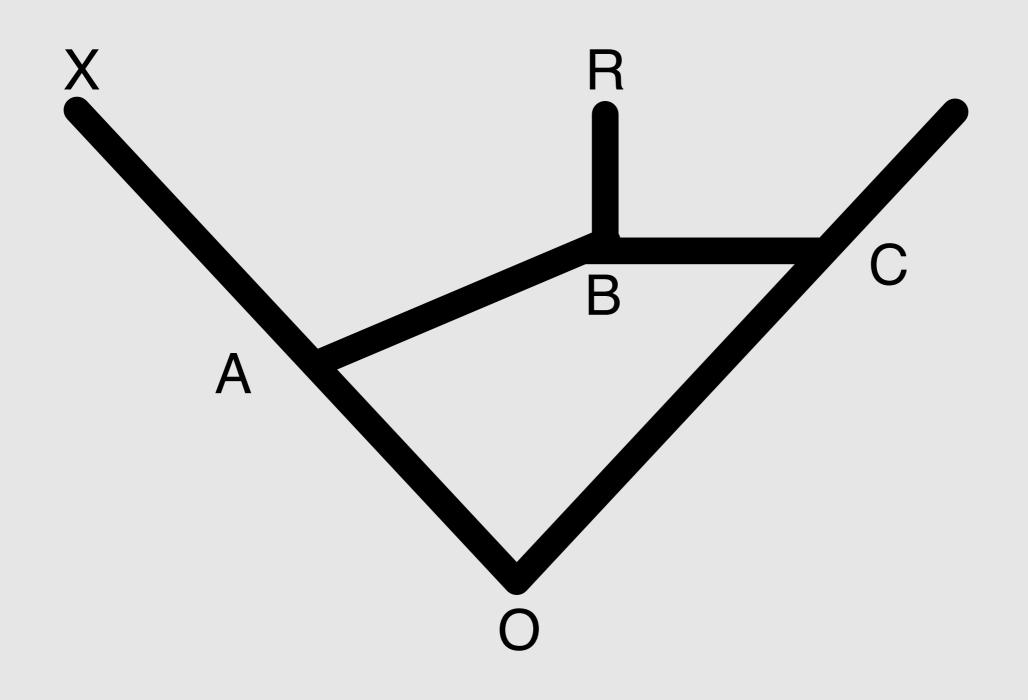
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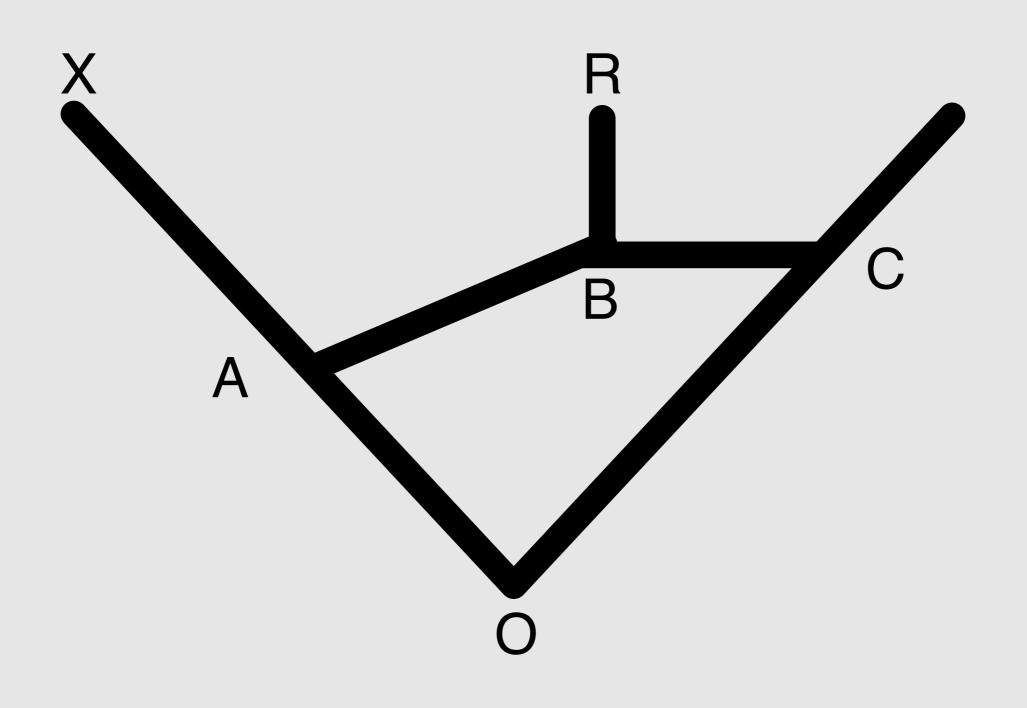
Multivariate normal distribution. The figure shows a tree, the tree's variance-covariance matrix, and the vector of means (which, under Brownian motion, would equal the root state). Highlighted are the branches leading to covariance between taxa A and B (red) and the branches leading to variance in D (blue).



$$\mu_B = [m\mu_D + (1 - m)\mu_C] + \log \beta$$

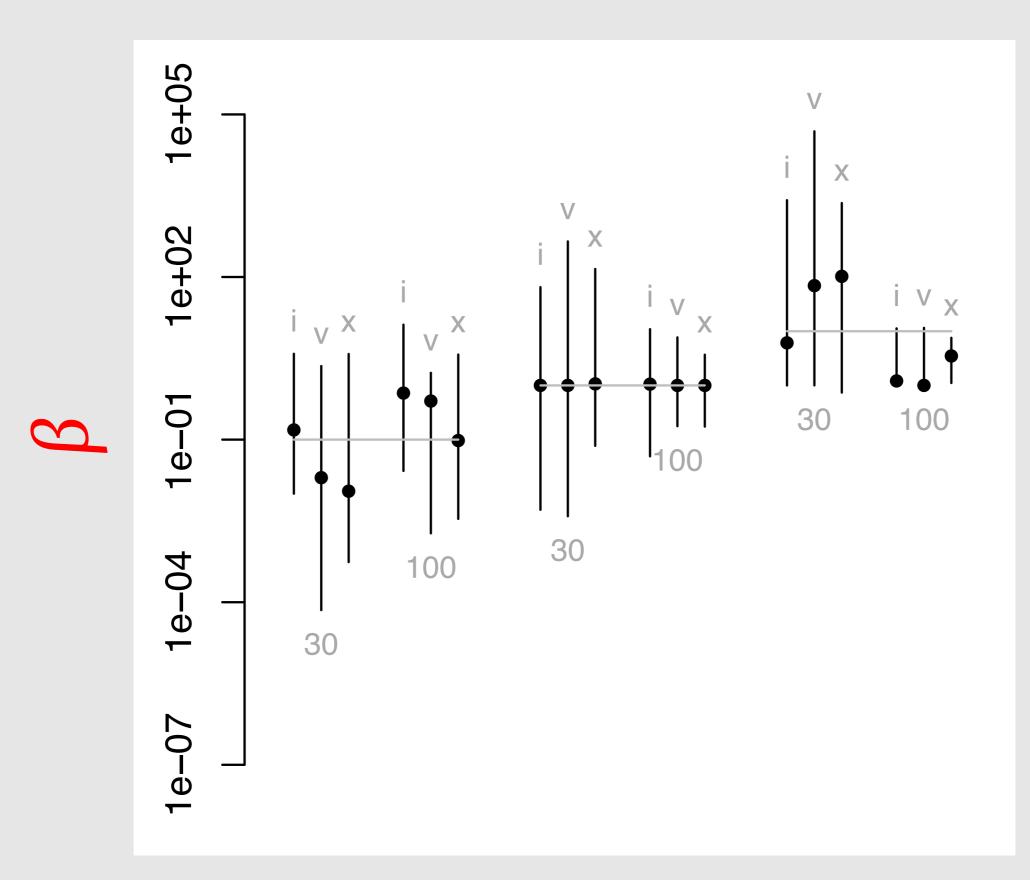


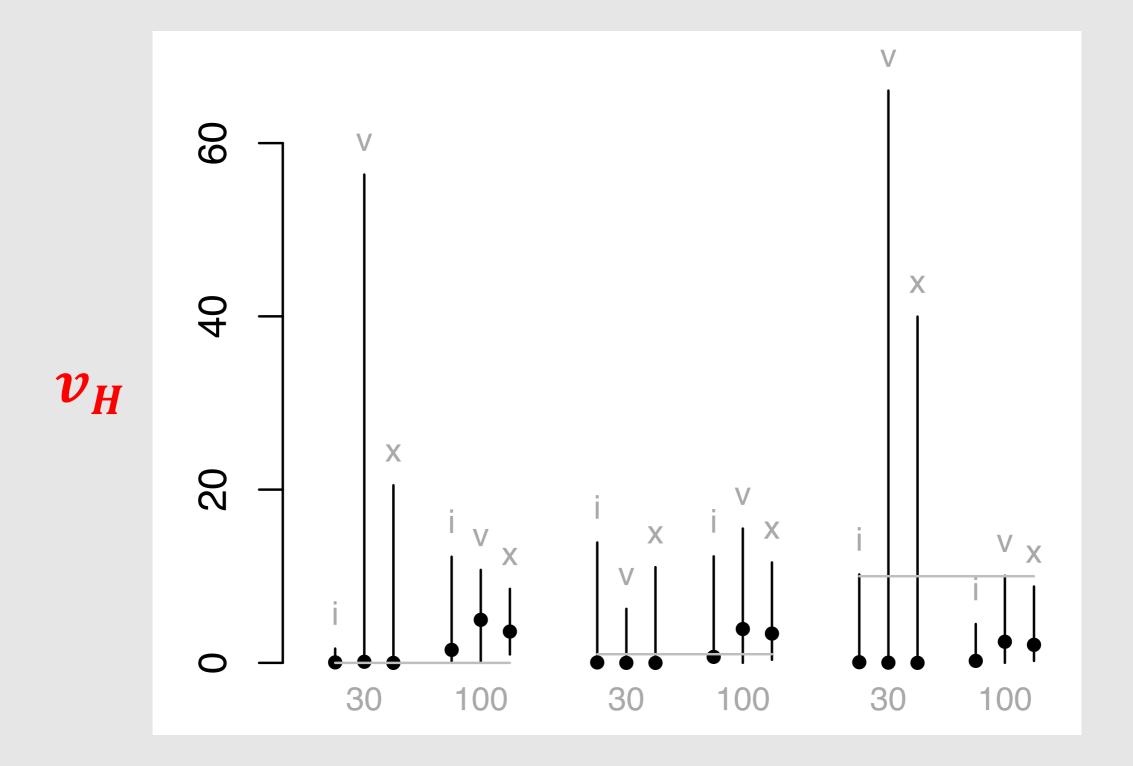
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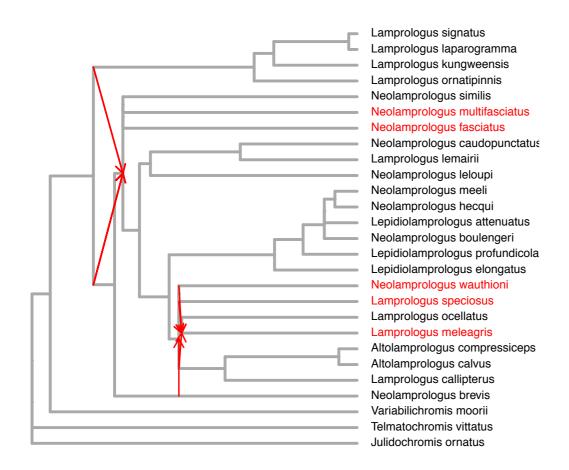
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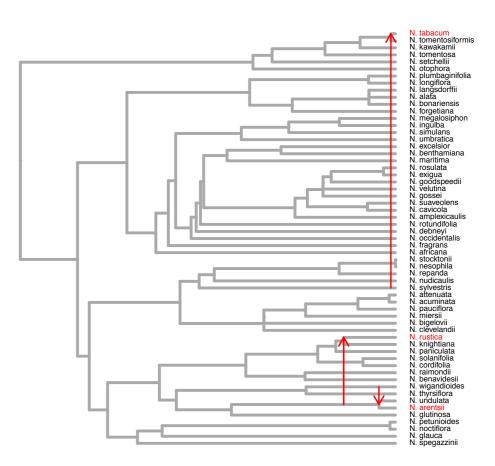


Cichlid



Network: Kobmüller et al. (2007) **Traits**: standard length from fishbase (Froese and Pauly, 2010) using rfishbase (Boettiger et al. 2012)

Nicotiana



Network: Chase et al. (2003)

Traits: seedling growth under mannitol

treatment (Komori et al. 2000)

Cichlids

Akaike weight	σ^2	μ	β	νH	SE	variance over tree
	(log(cm)) ² /MY	log(cm) ²	unitless scalar	log(cm) ²	log(cm)	log(cm) ²
0.525	0 (0, 0.093)	2.152 (1.737, 2.454)	1 (fixed)	0 (fixed)	0.324 (0.091, 0.558)	0 (0, 0.524)
0.129	0 (0, 0.08)	2.155 (1.751, 2.44)	1 (fixed)	0 (0, 1.052)	0.31 (0.104, 0.536)	0 (0, 0.448)
0.284	0 (0, 0.079)	2.216 (1.82, 2.509)	0.708 (0.419, 1.196)	0 (fixed)	0.306 (0.117, 0.527)	0 (0, 0.446)
0.062	0 (0, 0.068)	2.216 (1.848, 2.497)	0.707 (0.434, 1.192)	0 (0, 0.524)	0.305 (0.128, 0.501)	0 (0, 0.384)
Model average	0 (0, 0.086)	2.175 (1.77, 2.47)	0.899 (0.8, 1.068)	0 (0, 0.169)	0.316 (0.102, 0.543)	0 (0, 0.483)

Nicotiana

Akaike weight	σ^2	μ	β	νH	SE	variance over tree
	log(rel. growth) ² /MY	log(rel.growth) ²	unitless scalar	log(rel.growth) ²	log(rel.growth)	log(rel.growth) ²
0.084	0.04 (0.008, 0.083)	2.601 (2.02, 3.191)	1 (fixed)	0 (fixed)	0.472 (0.318, 0.801)	0.347 (0.071, 0.73)
0.036	0.053 (0.019, 0.132)	2.582 (1.945, 3.234)	1 (fixed)	0.412 (0.105, 5.568)	0.393 (0.2, 0.723)	0.462 (0.171, 1.156)
0.716	0.076 (0.037, 0.091)	2.601 (2.119, 3.188)	2.358 (1.522, 2.748)	0 (fixed)	0.329 (0.279, 0.479)	0.664 (0.324, 0.803)
0.165	0.08 (0.036, 0.119)	2.511 (2.045, 2.91)	2.42 (1.448, 2.982)	0.025 (0.007, 0.495)	0.332 (0.243, 0.502)	0.701 (0.314, 1.044)
Model average	0.072 (0.034, 0.097)	2.586 (2.093, 3.144)	2.206 (1.447, 2.578)	0.019 (0.005, 0.279)	0.344 (0.274, 0.519)	0.636 (0.296, 0.849)

Package: BMhyb Version: 1.3.5 Date: 2016-08-22

Type: Package

Title: PCM for Hybridization

Author: Dwueng-Chwuan Jhwueng <djhwueng@umail.iu.edu> and Brian O'Meara

<omeara.brian@gmail.com>

Maintainer: Dwueng-Chwuan Jhwueng <djhwueng@umail.iu.edu>

Description: The BMhyb package analyzes the phenotypic evolution of species of hybrid origin on a phylogenetic network. This package can detect the hybrid vigor effect, a burst of variation at formation, and the relative portion of heritability from its parents. Parameters are estimated by maximum likelihood. Users need to enter a comparative data set, a phylogeny, and information on gene flow leading to hybrids.

License: GPL (>=2)
RoxygenNote: 5.0.1



Package: BMhyb Version: 1.3.5 Date: 2016-08-22

Type: Package

Title: PCM for Hybridization

Author: Dwueng-Chwuan Jhwueng <djh

<omeara.brian@gmail.com>

Maintainer: Dwueng-Chwuan Jhwueng < Imports: corpcor, numDeriv, geiger

mvtnorm, grDevices, graphic

Description: The BMhyb package anal New Results

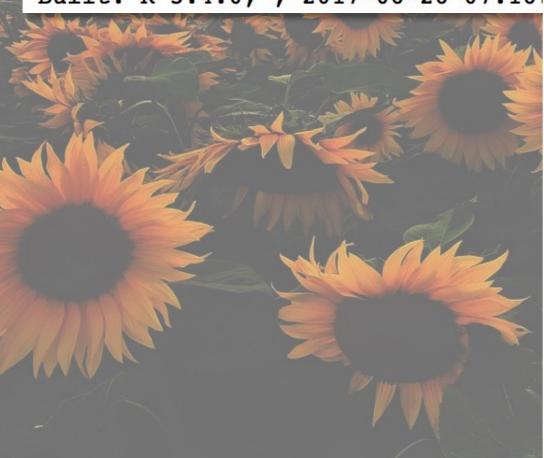
of hybrid origin on a phylogene vigor effect, a burst of variat

heritability from its parents. Users need to enter a comparat:

flow leading to hybrids.

License: GPL (>=2) RoxygenNote: 5.0.1

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Trait Evolution on Phylogenetic Networks

Dwueng-Chwuan Jhwueng, D Brian O'Meara doi: https://doi.org/10.1101/023986

This article is a preprint and has not been peer-reviewed [what does this mean?].

Abstract

Info/History

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Abstract

Species may evolve on a reticulate network due to hybridization or other gene flow rather than on a strictly bifurcating tree, but comparative methods to deal with trait evolution on a network are lacking. We create such a method, which uses a Brownian motion model. Our method seeks to separately or jointly detect a bias in trait value coming from hybridization (β) and a burst of variation at the time of hybridization (v_H) associated with the hybridization event, as well as traditional Brownian motion parameters of ancestral state (μ) and rate of evolution (σ^2) of Brownian motion, as well as measurement error of the tips (SE). We test the method with extensive simulations. We also apply the model to two empirical examples, cichlid body size and Nicotiana drought tolerance, and find substantial measurement error and a hint that hybrids have greater drought tolerance in the latter case. The new methods are available in CRAN R package BMhyd.

