# COMP 204 - Assignment #2

**Due date: October 11, 23:59** 

# NO LATE ASSIGNMENTS WILL BE ACCEPTED, TO ALLOW US TO DISTRIBUTE SOLUTIONS BEFORE THE MIDTERM EXAM.

- Submit *one* Python program on MyCourses, which should contain all your functions.
- Write your name and student ID at the top of the program
- For each question, complete the relevant function. Do not change its name or arguments.
- Your program should not print anything. Each function should just **return** the appropriate object.
- Do not use any modules except the math module, or any pieces of code you did not write yourself.
- For each question, your function will be automatically tested on a variety of test cases, which will account for 75% of the mark.
- For each question, 25% of the mark will be assigned by the TA based on (i) your appropriate naming of variables; (ii) commenting of your program; (iii) simplicity of your program (simpler = better)

# **Background information:**

Transcription factors (TF) are proteins that recognize and bind to DNA, and in doing so regulate the expression of a nearby gene. Each type of TF has an affinity for a particular short DNA pattern. For example, the E-box TF likes to bind the sequence CAGCTG, whereas the Runx TF likes to bind CCACA. However, every transcription factor has a certain flexibility in the DNA sequences it can bind to. For example, the E-box TF will sometimes bind CAGGTG, or CAGCAG. The Runx TF may also bind CCAGA or ACACA. Biologists can identify DNA sequences bound by a given transcription factor using experiments such as chromatin immunoprecipitation followed by sequencing (ChIP-seq). The outcome of such an experiment is a set of DNA sequences from the genome that are bound by the TF. For example, a ChIP-seq experiment on transcription factor E2F1 may produce the following set of 10 sequences:

ACGATG ACAATG

ACGATC

ACGATC

TCGATC

TCGAGC

TAGATC

TAAATC

AAAATC

**ACGATA** 

These DNA sequences are probably only a small subset of all the sites bound by E2F1 in the genome. We would like to be able to learn a model of what type of pattern E2F1 likes to bind, in order to predict, in the human genome, sites that may have been missed by the experiment. In order to do so, bioinformaticians have developed a model called Position Weight Matrix (PWM). Here, you will learn how this model works, and you will have to implement it in Python.

To build a PWM, one first needs to build a Position Frequency Matrix (PFM). A PFM is matrix (table) of 4 rows and L columns, where L is the length of the binding site sequences. In the E2F1 example, L=6. The entry in row i and column j of the PFM, which we denote PFM[i][j], is the frequency of nucleotide i at position j of the binding sites. For E2F1, based on the 10 sites listed above, the PFM is

The PFM can be converted to a PWM using the following formula:

$$PWM[i][j] = \log_{10} \left( \frac{PFM[i][j] + p}{PFM[0][j] + PFM[1][j] + PFM[2][j] + PFM[3][j] + 4p} \right) - \log_{10} (0.25)$$

The variable p, called the pseudocount is generally set to a small value such as 0.1. For our E2F1 example, the PWM is:

	0	1	2	3	4	5
A	0.37	0.07	0.07	0.59	-1.41	-0.37
C	-1.41	0.43	-1.41	-1.41	-1.41	0.43
G	-1.41	-1.41	0.43	-1.41	-0.37	-0.09
${f T}$	0.19	-1.41	-1.41	-1.41	0.54	-1.41

You will notice that large positive values in the PWM (e.g. 0.59) correspond to positions where the TF has a strong preference for a particular nucleotide (e.g. an A at position 3).

We can now use a PWM to calculate the score of a match between a candidate sequence of *L* nucleotides and the PWM. For example, the score of candidate sequence TCGATC is obtained by summing up the appropriate values from the PWM:

Score(TCGATG) = 
$$PWM[T][0] + PWM[C][1] + PWM[G][2] + PWM[A][3] + PWM[T][4] + PWM[G][5]$$
  
=  $0.19 + 0.43 + 0.43 + 0.59 + 0.54 + (-0.09) = 2.11$ 

whereas for sequence ACATAG, we get Score(ACATAG) = PWM[A][0] + PWM[C][1] + PWM[A][2] + PWM[T][3] + PWM[A][4] + PWM[G][5] = 0.37 + 0.43 + 0.07 + (-1.41) + (-0.09) = -2.03 The larger the score, the higher the affinity of the transcription factor for the sequence.

Finally, we can use a PWM to scan a longer sequence to identify potential binding sites for the TF. This is done by calculating the PWM score for every possible portion of L nucleotides of the sequence. For example:

```
Sequence = ATCGATGAGACTGA

Score(0) = Score(ATCGAT) = -3.87...

Score(1) = Score(TCGATG) = 1.65...

Score(2) = Score(CGATGA) = -4.16...

Score(3) = Score(GATGAG) = -4.16...

Score(4) = Score(ATGAGA) = -0.01...

Score(5) = Score(TGAGAC) = -2.55...
```

Positions whose score exceeds a user-chosen threshold are reported as a putative binding sites for the TF. For example, if we choose a threshold of -0.2, then positions 1 and 4 would be reported as putative sites.

# Download TFBSscanning.py from MyCourses.

Your work will consist in completing the each of the functions given in this program. *You must not change the function names or their arguments.* 

For each question, it is your responsibility to test your program to make sure it works on all possible cases, not just those provided as examples.

# Question 1 (10 points). Encoding of DNA nucleotides into integers.

It is often convenient to represent a nucleotide ("A","C","G", or "T") as a number between 0 and 3, where "A" is represented as 0, "C" as 1, "G" as 2, and "T" as 3.

Complete the **encode** function that takes as argument a string of one character, and **returns** an integer between 0 and 3. If the string provided as argument is not one of "A","C","G", or "T", the function should return -1.

```
Example of execution:

code = encode("G")

print(code) > 2

code = encode("A")

print(code) > 0

code = encode("X")

print(code) > -1
```

# Question 2 (20 points). Building a PFM.

Write a function called <code>build\_PFM</code> that takes as input a list of DNA sequences identified as binding sites for a given transcription factor, and **returns** a Position Frequency Matrix (PFM), represented as a list of lists of integers, where PFM[nuc][pos] = number of sequences that have nucleotide <code>nuc</code> at position <code>pos</code>. The function can assume that the input sequences all have the same length, but that length is not necessarily 6 (like in our example).

Example of execution:

#### **Hints:**

- **1.** Use the encode function you wrote in question 1.
- **2.** In your code, you may find it useful to be able to create a PFM with all entries initialized to zero. If L is the length of the desired PFM, this can be done as follows:

PFM = [[0 for i in range(L)] for j in range(4)]

### Question 3 (10 points). Building a PWM from a PFM

Write the function *get\_PWM\_from\_PFM*, which takes as argument a PFM and a real number called the pseudocount and **returns** a PWM, according to the formula given above.

Example of execution:

```
PFM = [[6, 3, 3, 10, 0, 1], [0, 7, 0, 0, 0, 7], [0, 0, 7, 0, 1, 2], [4, 0, 0, 0, 9, 0]]

PWM = get_PWM_from_PFM(PFM, 0.1)

print(PWM)

Output:

[[0, 370356487039949, 0,07638834586345467, 0,07638834586345467, 0,5893480]
```

 $\begin{bmatrix} [0.370356487039949, 0.07638834586345467, 0.07638834586345467, 0.5893480258118245, -1.4149733479708178, -0.37358066281259295], [-1.4149733479708178, 0.43628500074825727, -1.4149733479708178, -1.4149733479708178, 0.43628500074825727], [-1.4149733479708178, -$ 

# Question 4 (20 points). Scoring a sequence of length L

Write a function *score* that takes as argument a sequence and a PWM (both of the same length L), and calculates the score of the sequence with that PWM.

# Example of execution:

```
\begin{aligned} & \text{PWM} = [[0.370356487039949, 0.07638834586345467, 0.07638834586345467, \\ & 0.5893480258118245, -1.4149733479708178, -0.37358066281259295], [-1.4149733479708178, \\ & 0.43628500074825727, -1.4149733479708178, -1.4149733479708178, -1.4149733479708178, \\ & 0.43628500074825727], [-1.4149733479708178, -1.4149733479708178, 0.43628500074825727, \\ & -1.4149733479708178, -0.37358066281259295, -0.09275405323689867], \\ & [0.19781050874891748, -1.4149733479708178, -1.4149733479708178, -1.4149733479708178, \\ & 0.5440680443502756, -1.4149733479708178]] \end{aligned}
```

```
s= score("TCGATG",PWM)
print(s) → 2.1110425271706337
s=score("ACATAG",PWM)
print(s) → -2.039670915526873
```

# Question 5 (20 points). Identifying matches in longer sequences

Write a function predict\_sites that takes as input a DNA sequence and a PWM as positional arguments, as well a floating point number called threshold as keyword argument, and **returns** a list of the starting positions of substrings whose score with the given PWM is larger or equal to the threshold.

#### Example of execution:

```
\begin{aligned} \text{PWM} = & [[0.370356487039949, 0.07638834586345467, 0.07638834586345467, \\ 0.5893480258118245, -1.4149733479708178, -0.37358066281259295], [-1.4149733479708178, \\ 0.43628500074825727, -1.4149733479708178, -1.4149733479708178, -1.4149733479708178, \\ 0.43628500074825727], [-1.4149733479708178, -1.4149733479708178, 0.43628500074825727, \\ -1.4149733479708178, -0.37358066281259295, -0.09275405323689867], \\ [0.19781050874891748, -1.4149733479708178, -1.4149733479708178, -1.4149733479708178, \\ 0.5440680443502756, -1.4149733479708178]] \end{aligned}
```

```
hits = predictSites(sequence, PWM) print(hits) \rightarrow [3, 38, 43, 47]
```

# Question 6 (20 points): Identification of putative target genes

Genes are portions of DNA sequences. The transcription start site of a gene is the position of the start of the gene in the sequence. Suppose that you have a dictionary of gene names and associated transcription start site position. For example genes = {"BRCA1":3, "MYC":23, "RUNX": 45}

Our goal is to count the number of predicted binding sites located in the neighborhood of the transcription start site of each gene. For example, if we allow a maximum distance of 10 and the list of PWM hits is [12, 38, 43, 46], then the BRCA1 gene has 1 hit, the MYC gene has 0, and the RUNX gene has 3.

Your task is to write the function count\_hits\_per\_gene, which takes as positional arguments the genes dictionary and the list of hits, as well as keyword argument max\_dist, and **returns** a new dictionary with genes as keys and hit count as values.

```
Example of execution:
genes = {"BRCA1":3, "MYC":23, "RUNX": 45}
hits = [3, 38, 43, 47]
gene_hits = count_hits_per_gene(genes, hits, max_dist = 10)
print(gene_hits) → {'BRCA1': 1, 'MYC': 0, 'RUNX': 3}
gene_hits = count_hits_per_gene (genes, hits, max_dist = 20)
print(gene_hits) → {'BRCA1': 1, 'MYC': 3, 'RUNX': 3}
```