Temperature and moisture effects on greenhouse gas emissions from deep active-layer boreal soils

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#### Abstract

Rapid climatic changes, rising air temperatures, and increased fires are expected to drive permafrost degradation and alter soil carbon (C) cycling in many high-latitude ecosystems. How these soils will respond to changes in their temperature, moisture, and overlying vegetation is highly uncertain, but critical to understand given the large soil C stocks in these regions. We used a laboratory experiment to examine how temperature and moisture control CO2 and CH4 emissions from mineral soils sampled from the bottom of the annual active layer, i.e. directly above permafrost, in an Alaskan boreal forest. Gas emissions from thirty cores, subjected to two temperatures and either field moisture conditions or experimental drought, were tracked over a 100-day incubation; we also measured a variety of physical and chemical characteristics of the cores. Gravimetric water content was 0.31 ± 0.12 (unitless) at the beginning of the incubation; cores at field moisture were unchanged at the end, but drought cores had declined to 0.06 ± 0.04. Daily CO2 fluxes were positively correlated with incubation chamber temperature, core water content, and percent soil nitrogen, and had a temperature sensitivity (Q10) of 1.3 and 1.9 for the field moisture and drought treatments, respectively. Daily CH4 emissions were most strongly correlated with percent nitrogen, but neither temperature nor water content was a significant first-order predictor of CH4 fluxes. The cumulative production of C from CO2 was over six orders of magnitudes higher than that from CH4; cumulative CO2 was correlated with incubation temperature and moisture treatment, with drought cores producing 52% - 73% lower C. Cumulative CH4 production was unaffected by any treatment. These results suggest that deep active-layer soils may be quite sensitive to changes in soil moisture under aerobic conditions, a critical factor as discontinuous permafrost thaws in interior Alaska. Deep but unfrozen high-latitude soils have been shown to be strongly affected by long-term experimental warming, and these results provide insight into their future dynamics and feedback potential with future climate change.

#### 1 Introduction

High latitude ecosystems are being subjected to rapid changes in climate (IPCC, 2013) and increases in fire frequency and intensity (Kasischke et al., 2010), notably in northwestern North America and Alaska (Hinzman et al., 2005; Ju and Masek, 2016). This will have a wide variety of ecosystem effects (Alexander and Mack, 2016): in particular, rising temperatures and increasing fire will likely result in permafrost degradation (Pastick et al., 2015; Zhang et al., 2015; Genet et al., 2013; Helbig et al., 2016) and changes in soil temperature, with subsequent hydrology changes that will influence soil greenhouse gas (GHG) fluxes to the atmosphere (Schädel et al., 2014). Such fluxes are a large component of the global C cycle and, because of the high C stocks of northern soils (Tarnocai et al., 2009), could result in a significant and positive climate feedback (Treat et al., 2015; Koven et al., 2011; Schaefer et al., 2014).

The magnitude, timing, and form-in particular as methane (CH4) or carbon dioxide (CO2)-of such any such feedback remain highly uncertain (Schuur et al., 2015). While northern soils hold enormous quantities (Hugelius et al., 2014) of potentially mineralizable soil organic carbon (SOC), vegetation and succession dynamics (for example, thermal insulation by mosses) promote permafrost resilience to even large temperature changes (Jorgenson et al., 2010; Turetsky et al., 2012). Vegetation type also influences SOC quality and quantity, with microbial communities (Högberg et al., 2007), soil respiration (Raich and Tufekcioglu, 2000), and SOC all linked to aboveground factors such as woody versus nonwoody stems, deciduous versus evergreen canopies, and the presence of nitrogen-fixing plants.

A number of factors may however disrupt these these feedbacks between vegetation type, ground cover, permafrost, and SOC stability. Climate changes, in particular regional warming and drying, cause vegetation stress (Ju and Masek, 2016; Barber et al., 2000) and increased mortality. Conversely, increasing plant productivity in some regions can stimulate the decomposition of older SOC (Hartley et al., 2012). Climate also drives fire regime changes, and ecosystem disruption is particularly likely after more-intense fire events (Johnstone et al., 2010; Genet et al., 2013). Even absent disturbance, the stability of SOC is highly uncertain, as it depends on soil temperature and moisture, the ages of and ratio between the carbon (C) and nitrogen (N) pools (Weiss et al., 2015; Karhu et al., 2014), and its protection from competent microorganisms, enzymes, and resources (Bailey et al., 2012), whether by organomineral sorption, chemical lability, or physical location (Schmidt et al., 2011).

Temperature and moisture typically have strong and often interactive influences on soil GHG emissions. Laboratory incubations, field observations, and meta-analyses have documented changing greenhouse gas (GHG) fluxes with rising temperature (Olefeldt et al., 2013; Davidson and Janssens, 2006; Hashimoto et al., 2015; Treat et al., 2015). GHG responses to wetting and thawing dynamics exhibit substantial variability between studies, probably due to differences in soil type, antecedent conditions, phase changes, experimental protocols, etc. (Kim et al., 2012). The anaerobic conditions common following permafrost thaw are expected to lower CO2 emissions but increase those of CH4 (Treat et al., 2015; Treat et al., 2014), but emissions from aerobic soils will likely dominate the permafrost C feedback (Schädel et al., 2016). Decadal warming and drying trends in Alaska (Bieniek et al., 2014) thus seem likely to increase GHG emissions from soils, and incubation experiments are critical to understand these dynamics (Elberling et al., 2013).

Most previous studies have focused on surface soils or permafrost soils, neglecting deep active-layer soils that were identified as subject to strong effects from a two-decade warming experiment in the Alaskan Arctic (Sistla et al., 2013). Such deeper soils have particular characteristics distinguishing them from both shallow active layer soils and underlying permafrost: they are most affected by interannual variability in thaw depth, potentially flipping the C source/sink status of entire ecosystems (Goulden et al., 1998; Harden et al., 2012); they are subject to distinctive freeze-thaw, cryoturbation, and microbial dynamics, which are likely to change their sensitivity to climate change and feedback potential (Schuur et al., 2008); and they are known to pose particular problems for accurate modeling of high-latitude carbon dynamics (Nicolsky et al., 2007). These soils are likely to be a highly important contributor to future climate feedbacks, with modeling studies suggesting that one-third of 21st century climate-induced carbon loss may originate from seasonally frozen soil carbon (Koven et al., 2015).

The goal of this study was to examine how temperature and moisture control GHG (CO2 and CH4) emissions from soils sampled from the bottom of the annual active layer–i.e., directly above permafrost–in an Alaskan boreal forest. We also aimed to characterize the chemical and structural properties of these soils following a 100-day incubation at different temperatures, subjecting some cores to drying treatments. We hypothesized that (i) CO2 would be the dominant pathway for C loss in these largely aerobic soils; (ii) soils maintained at field moisture and high temperature would lose more C-CO2 than cores incubated at 4˚C, due to increased aerobic and anaerobic microbial activity; and (iii) core CH4 fluxes would be sensitive only to temperature, as no anaerobic conditions were imposed on the cores.

#### 2 Methods

**2.1 Field sampling**

The field component of this research took place in Caribou-Poker Creeks Research Watershed (CPCRW), part of the Bonanza Creek LTER (<http://www.lter.uaf.edu/research/study-sites-cpcrw>). CPCRW is located in the Yukon-Tanana Uplands northeast of Fairbanks, AK, a part of the boreal forest that has seen strong increases in air temperature and forest browning (Ju and Masek, 2016) over several decades. Annual average air temperature is -2.5 °C, and annual average precipitation 400 mm (Petrone et al., 2006). The watershed's lowlands and north-facing slopes are dominated by black spruce (*Picea mariana* (Mill.) BSP), feathermoss (*Pleurozium schreberi* and others), and *Sphagnum* spp.; the drier south slopes tend to be deciduous with a mixture of trembling aspen (*Populus tremuloides* Michx.), paper birch (*Betula neoalaskana*), and patches of alder (*Alnus crispa*).

We sampled soils from a southeast slope (65.1620 °N, 147.4874 °W) at CPCRW, in a 60 m transition zone between lowland *Picea mariana* and upland *Betula neoalaskana*, with significant white spruce (*Picea glauca*) presence as well. Stand density in this transition zone was 4060 ± 2310 trees ha-1, with basal area of 27.9 ± 7.0 m2 ha-1. The forest was at least 90 years old (cf. Morishita et al., 2014) according to tree rings taken at the stem base of several of the largest white spruce. The soil is characterized as a poorly-drained silt loam, and on average had ~20 cm of organic material over the mineral soil.

Thirty-nine soil cores, each 30 cm high by 7.5 cm wide, were taken using a soil recovery augur (AMS Inc., American Falls, ID) on 3-5 August 2015. We sampled from the bottom (within 0-2 cm of permafrost) of the active layer, which averaged 80 cm depth. Sample points were randomly located in the transition zone described above, and separated by 2-5 m. Cores were kept cool in the field before being packed in dry ice and shipped to Richland, WA within 48-72 hours of collection.

**2.2 Laboratory incubation**

In the lab, the soil cores were stored at 4 °C for several days until they were weighed and prepared for incubation. At that point (11-12 August 2015), three fragmented or otherwise damaged cores were discarded, and the remaining cores were randomly assigned to one of six groups (N=6 in each group). These included two incubation temperatures of 4 and 20 °C, following the protocol of a number of previous boreal incubation studies (Treat et al., 2015). Within each temperature there were two moisture treatments: one in which soil moisture was maintained at field conditions (~28% moisture by volume), and a drought treatment in which no water was added and cores were allowed to dry down to ~5% moisture by volume. The fifth group was a 20 °C "controlled drought" one, in which water was added so that these cores' moisture status would close match those of the 4 °C "drought" cores, which we anticipated would dry more slowly than their 20 °C counterparts. The final 6-core group was used for destructive, pre-incubation measurements including moisture content, pH, soil carbon and N, and bulk density. Subsamples were collected and stored at -20 °C for dissolved organic carbon measurements or air-dried for soil C and N (see below).

On 18 August 2015 cores were placed into one of two growth chambers (Conviron Control Systems BDW80, Winnipeg, Canada) maintained at 4 and 20 °C temperatures and 70% relative humidity and allowed to equilibrate for two weeks. Starting on 31 August 2015 we measured the cores' mass and GHG (CH4 and CO2) emissions four times in the first week, then twice per week for the first month, and then once per week for the rest of the 100-day incubation. Throughout the incubation, cores had a 200 µm mesh screen fit to the base and were mounted on porous ceramic plates (Soil Moisture Equipment Corp., Santa Barbara, CA, USA) so that, when the plates were placed in contact with water, water would move up into the cores via capillary action. The "drought" cores were mounted on dry plates, but not allowed to drop below 5% water content. After each flux measurement, cores received additional wetting from the top to maintain their water status at the desired level.

For each measurement, a six-core treatment group was connected to a Picarro A0311 multiplexer that was in turn connected to a Picarro G2301 GHG Analyzer (Picarro Inc., Santa Clara, CA, USA). Dry CH4 and CO2 concentrations were monitored for 2 minutes, and this was repeated 2-3 times before moving on to a new treatment group. Cores were weighed immediately after gas measurements. Ambient air was measured between treatment groups, and before starting measurements in a chamber, as a check on ambient CO2 conditions and instrument stability.

The incubation experiment concluded on 9 December 2015, following the final CO2 and CH4 readings. Each soil core was maintained at the treatment-dependent temperature and moisture content (by mass) until removed for destructive sampling, December 14-18, 2015. Sub-samples were collected and composited throughout each soil core for dissolved organic carbon analysis (110 ± 24 g dry mass equivalent) and dry-mass calculations (~28 g each). The remaining core material was air-dried and separated into particles (>2 mm diameter) and soil (≤2 mm) using a U.S. Standard Test Sieve No. 10 (Fisherbrand, Pittsburg, PA, USA). The dry mass and volume of soil were used in calculations of gravimetric and volumetric soil moisture content, respectively (Gardner, 1986). Soil volume was calculated as the total core volume minus the volume of particles >2 mm diameter, with the latter determined by water displacement. Air-dried soil and sub-samples stored at -20 °C were sent to the Agricultural and Environmental Services Laboratory at the University of Georgia Extension in February 2016 for total C, N, and dissolved organic C. Samples were combusted in an oxygen atmosphere at 1350 °C, and measured for gaseous C and N using a Elementar Vario Max CNS. DOC was measured using a Shimadzu 5000 TOC Analyzer.

**2.3 Data and statistical analysis**

For each measurement of each sample throughout the 100-day incubation (i.e., each gas, core, and date/time), we used the rise in gas concentrations to calculate a flux rate in ppm s-1 (CO2) or ppb s-1 (CH4), a linear rate of change (δc/δt) based on the concentration rise from a minimum (up to 10 seconds after measurement began) to a maximum (at 10-45 seconds). Each core’s respiration flux (*F*) was then calculated as where *V* is the core-specific system volume, *M* the core dry mass as determined at the end of the incubation, *Pa* atmospheric pressure (101 kPa; the incubation chambers were ~120 m a.s.l.), *R* the universal gas constant (8.3 x 10-3 m3 kPa mol-1 K-1) and *T* the chamber air temperature (K) at time of measurement. The final respiration rate was expressed on a dry soil mass basis (µg C g soil-1 day-1).

Anomalous data were excluded based on their gas fluxes being more than 5 (for CO2) or 10 (for CH4) mean absolute deviations (Davies and Gather, 1993) from the treatment mean within a 10-day period, for a given treatment and temperature. We excluded 172 of 2686 (6.4%) measurements for this reason. If the coefficient of variability (CV) of fluxes from any core on a single day exceeded 140%, a value chosen based on the distribution of CVs across all cores, the entire core was excluded for that day (90 data points, 3.4%). Other data (4.8%) were removed because of known instrument problems, e.g. the analyzer was left running after leaving a chamber. The final number of valid flux samples from the 100-day incubation was 2198.

The effects of temperature, gravimetric water content, percent C, percent N, and DOC concentration on instantaneous gas fluxes were evaluated using a linear mixed-effects model fit by the R function *lme* in the R 'nlme' package, version 3.1.128. Because the dependent variable (CO2 or CH4 flux) was non-normally distributed, it was transformed using a natural-logarithm (+0.1 µg C g C-1 day-1 to ensure all positive fluxes, following Treat et al. 2015) transformation. Soil core was treated as a random effect in the model. We then performed stepwise model selection by Akaike's information criterion (AIC) using the *stepAIC* function in the R 'MASS' package, version 7.3.45. A linear mixed-effects model was also used to evaluate the effect of treatment on core water content.

Cumulative respiration for each core and gas was calculated by linearly interpolating flux rates between measurement dates and summing respired C over the entire incubation. The effect of temperature and treatment (drought, controlled drought, or field moisture conditions) on cumulative gas fluxes was evaluated with a post-hoc Tukey Honest Significant Differences test. Temperature sensitivity (Q10) was calculated for each gas and treatment as where *F1* and *F2* are the cumulative gas fluxes (mg C g C-1) at temperatures *T1* and *T2* (°C), respectively.

All data analysis and statistics were performed using R version 3.3.1 (2016-06-21) (R Development Core Team, 2016). This experiment was run as an 'open experiment' (Bond-Lamberty et al., 2016b) with all analysis code, data (from raw instrument data to final summaries), diagnostics, etc., available at <https://github.com/bpbond/cpcrw_incubation>. The summarized flux data backing the main results have been archived at [*DOI to be filled in*].

#### 3 Results

The 30 experimental cores had a bulk density of 1.00 ± 0.18 (mean ± sd) g cm-3. Large (>2 mm) particles, primarily schist, comprised 41% ± 11% of the cores' total mass. Soil (≤2 mm) dry mass was 886 ± 154 g. Sample DOC was 157.93 ± 55.74 mg kg-1. Carbon content was 1.20% ± 1.19%, while N content was 0.06% ± 0.06%. Mean C:N was 20.7. Neither temperature nor moisture treatment exerted any significant effect (P > 0.1 for all) on these highly variable properties (**Table 1**).

Gravimetric water content was 0.31 ± 0.12 (min 0.19, max 0.77) at the beginning of the incubation (**Figure 1**). "Field moisture" cores were on average unchanged (0.33 ± 0.13) at the end of the incubation, but both the drought treatments, which did not differ from each other in their effect on gravimetric water content (P = 0.880), had declined to 0.06 ± 0.04. Volumetric water content values ranged from 0.29 ± 0.05 (min 0.23, max 0.43) at the beginning of the experiment to 0.15 ± 0.11 (min 0.03, max 0.38) at the end across all cores. Water filled pore space, assuming a particle density of 2.65 g cm-3, was 22-65% over all cores, moisture treatments, and temperatures.

Carbon dioxide fluxes during the incubation ranged from 1.1 µg C g C-1 day-1 to a maximum of 5245.1 µg C g C-1 day-1, with a mean of 248.9 µg C g C-1 day-1 over the 100 days. CH4 rates ranged from 0.00 ng C g C-1 day-1 to a maximum of 1.31 ng C g C-1 day-1, with a mean of 0.06 ng C g C-1 day-1.

These means conceal considerable variability over the course of the incubation (**Figures 2 and 3**). In the linear mixed-effects model (AIC = 2992.6), instantaneous CO2 flux was positively correlated with incubation chamber temperature, core gravimetric water content, and percent soil N (all P < 0.05, and the latter two P < 0.001; **Table 2**). Temperature sensitivity decreased significantly (P < 0.001) over the course of the incubation, while moisture sensitivity was unaffected by time. Percent C and percent N were highly correlated (r = 0.99) for these cores. Because percent N was a slightly stronger predictor, it was retained in the model while percent C was excluded; cf. Colman and Schimel (2014). The interaction between water content and percent N was also highly significant (P < 0.001), although cores with N > 0.2% exhibited little relationship between water content and CO2 flux (data not shown). Instantaneous CH4 fluxes were positively correlated with percent N, while water content exhibited significant interactions with percent N and DOC as a predictor (**Table 3**). This model had little predictive power (AIC = -10879.2), however, and neither temperature nor water content was a significant first-order predictor of CH4 fluxes.

The cumulative production of C from CO2 (**Figure 4**) was over six order of magnitudes higher than that from CH4, with CO2:CH4 C ratios ranging from 1.4 million in the 4 °C "Field moisture" treatment, to 6.2 million in the 20 °C "Field moisture" treatment. Cumulative CO2 evolved was highly affected by temperature (P = 0.003), and "field moisture" cores emitted significantly more CO2 than the other two moisture treatments at both temperatures (P < 0.001 for both, with no significant interactive effect). There was no difference between fluxes from the 20 °C "drought" and "controlled drought" treatments (P = 0.377). "Drought" cores' cumulative production was 73% (4 °C) and 52% (20 °C) lower than the cores kept at field moisture. Neither temperature (P = 0.200) nor moisture treatment (mean P = 0.975) was a significant factor in predicting cumulative CH4 fluxes.

The cumulative flux numbers above result in CO2 temperature sensitivity (Q10) values of 1.3 and 1.9 for the field moisture and drought treatments, respectively; the corresponding Q10 values based on cumulative CH4 were 1.2 and 1.3. Computing Q10 values based on fluxes normalized by water filled pore space changed these values only slightly: to 1.2 and 1.7 for CO2, respectively, and 1.1 and 1.2 for CH4.

#### 4 Discussion

Rises in boreal air temperatures, and unpredictable precipitation changes, are changing fire disturbance regimes, warming and dry many soils, increasing vegetation stress, degrade permafrost and deepen the active layer (Schuur et al., 2015), all with uncertain consequences for soil dynamics and GHG fluxes. In this laboratory experiment we found that CO2, but not CH4, fluxes from these oxic active-layer mineral soils were sensitive to temperature and, in particular, moisture.

A number of studies have measured microbial respiration and GHG fluxes very close to our study site. Morishita et al. (2014) quantified GHG fluxes at CPCRW and nearby forests, and found CO2 production to be correlated with both temperature and moisture in upland Cryosols, consistent with our results. They found however that CH4 uptake was driven by temperature only. Waldrop et al. (2010) incubated active-layer and permafrost soils from *Picea mariana* sites near Fairbanks, AK, observing aerobic Q10 values of 9.0 (active layer) and 2.3 (permafrost) from -5 to 5 °C, and flux rates of 0.001-0.10 µmol CH4 day-1 g-1 (~0.001-0.133 ng C g C-1 day-1), and ~1-5 µg C-CO2 hr-1 g-1 (~2000-10000 µg C g C-1 day-1), considerably higher than the CO2 rates observed here. During the first 100 days of an incubation of Fairbanks-area 0-10 cm mineral soils, Neff and Hooper (2002) observed fluxes of ~55-409 µg C-CO2 g C-1 day-1, in line with the results here, while Wickland and Neff (2008) reported that temperature and moisture exhibited interactive and similar-magnitude effects on decomposition in *P. mariana* soils. Finally, Sierra et al. (2016) incubated homogenized organic soils from CPCRW and observed interactions between high temperatures, moisture, and oxygen concentration.

A number of synthesis studies have produced robust results documenting the dynamics and C feedback potential of Arctic and boreal soils; comparing to these results is useful because although the response of soil biota to stresses such as drought tends to differ between soil types, organisms, and vegetation, it is often broadly similar across biomes and climatic conditions (Manzoni et al., 2012). Using two meta-analyses of aerobic and anaerobic permafrost soil incubations, Schädel et al. (2016) showed that C release was highly sensitive to temperature, and that soils released far more (220-520%) C under aerobic conditions. Our incubation was fully aerobic, but its results are consistent with the conclusion that respiration in the form of CO2 is likely to dominate the high latitude C feedback, and that aerobic soils, and the conditions under which currently waterlogged soils may drain, deserve particular attention. In terms of absolute flux rates, in a pan-Arctic synthesis of anaerobic soil incubations, Treat et al. (2015) reported mean CO2 rates of 47 (all mineral soils) and 101 (for 20-100 cm soils) µg C-CO2 g C-1 day-1, somewhat lower than our aerobic incubation results. Treat et al. (2014) also found CO2 and CH4 emissions to be strongly correlated with temperature and moisture based on an incubation of Alaskan peats. Whether climate change makes northern regions wetter or driers is thus a critical factor affecting the quantity and form of C release.

**4.1 Temperature versus moisture sensitivity for cumulative emissions**

Warming usually increases soil GHG fluxes, for example at depth in a long-term Arctic tundra experiment (Sistla et al., 2013), as increased temperatures enhance the production of extracellular enzymes, increase enzyme activities, and enhance desorption rates of organic matter from minerals. A key question, for both experimentalists and modelers (Falloon et al., 2011), is to what degree such soils' emissions could by constrained by their moisture status, that is itself driven by changes in factors such as high-latitude temperatures, vapor pressure deficit, and precipitation.

In contrast to the controls on instantaneous fluxes, the controls on the cumulative fluxes (**Figure 4**) integrate the entire incubation time and are important to understand. Our results suggest that moisture limitation could exert a large effect on CO2 production for deep active-layer soils: "drought" cores' cumulative production was 73% (4 °C) and 52% (20 °C) lower than the cores kept at field moisture. This effect was highly significant, and suggests that moisture limitations could exert a significant constraint on deep active-layer soils as they slowly warm. Such moisture constraints are thought to be already exerting effects on vegetation and soil fluxes at large scales (Ju and Masek, 2016; Bond-Lamberty et al., 2012), but our understanding of the interactive effects involved is poor.

In contrast, the temperature sensitivities observed in this experiment were low (all less than 2.0, even when controlling for changes in soil moisture), but not unprecedented in comparison to a wide range of other laboratory soil incubations (Hamdi et al., 2013). It is more typical to observe Q10 values of around 2 (Dutta et al., 2006; Schädel et al., 2016), although the temperature sensitivity of C release can change over time of incubation (Dutta et al., 2006) and vary between soil fractions cycling over different time horizons (Karhu et al., 2010; Schädel et al., 2014). We observed Q10 change over the course of this incubation, emphasizing the need for longer-term studies (Lee et al., 2012; Elberling et al., 2013).

Observed surface CO2 fluxes at this CPCRW site exhibited a Q10 of 5.1 ± 1.4 over a temperature range of 3.5-15 °C (personal communication, C. Anderson); these surface fluxes were measured over multiple months and include root respiration, however, confounding any direct comparison. It is also important to note that while increased temperature does not always drive C mineralization rates in forest mineral soils (Giardina and Ryan, 2000), it is linked with increases in soil moisture content can lead to changes in microbial community structure and GHG fluxes (Xue et al., 2016).

More surprisingly, Q10 values were lower in the drought treatment cores, a mathematical consequence of the fact that drought restricted CO2 respiration more at 4 °C than at 20 °C. There is evidence that climate warming changes the microbial decay dynamics of soil organic C compounds generally considered to be stable(Frey et al., 2013; Bond-Lamberty et al., 2016a). Unlike surface soils, deep active layer soils, which store large quantities of soil C (Mueller et al., 2015), are not subject to abundant inputs of fresh C from vegetation, so the starting quality of the native soil C in active layer soils may be older, more microbially processed, and dominated by more stable "heavy" organic C (Karlsson et al., 2011). Thus, it is not surprising that these more stable C compounds would be metabolized by processes that have been reported to be less temperature-sensitive.

We observed very low but positive CH4 production from these upland mineral soils. This is contrast to many field studies that have observed CH4 uptake (oxidation) in dry boreal sites (Matson et al., 2009; Schaufler et al., 2010). Anoxic microsites in soil can however provide enough CH4 production to balance low-level consumption in otherwise aerobic soils (Kammann et al., 2009). In addition, our results are broadly consistent with data from 65 studies summarized by Olefeldt et al. (2013), who found that CH4 emissions were more sensitive to soil temperature in wetter ecosystems; it would have been a surprise if the little methanogenic activity in our upland, well-drained soils was temperature-sensitive at all. Methane was also a far smaller C flux than CO2 from these soils, in particular at higher temperatures (as CO2 was responsive to temperature, but CH4 was not). This is true more generally: for example, Treat et al. found a median CO2:CH4 production ratio of 387 for anaerobic incubations of boreal soils. This is naturally far lower than our observed aerobic (and thus high-CO2) ratios, but nonetheless consistent with them. Thus we see little opportunity for CH4 to be a significant contributor to these upland soils' C fluxes and climate feedback risk, even accounting for the 25x stronger radiative forcing of this gas over a 100-year time horizon (Lee et al., 2012).

**4.2 Soil nitrogen**

Somewhat unexpectedly, percent soil N was very significantly and positively correlated with both CO2 and CH4 fluxes (**Tables 2 and 3**). N interacts with microbial respiration via a number of complex, interactive, and still unclear mechanisms (Luo and Zhou, 2006), including reductions in belowground plant allocation, shifts in energy source or population of the saprotrophic community (Saiya-Cork et al., 2002) that leave it less capable of decomposing recalcitrant compounds, and perhaps abiotic stabilization mechanisms (Janssens et al., 2010). Meta-analyses have generally shown negative to neutral effects of N deposition on microbial biomass (Treseder, 2008) and respiration (Ramirez et al., 2012), and total soil respiration across ecosystems and biomes (Janssens et al., 2010; Zhou et al., 2014). These effect are likely due to several one or more mechanisms involving soil pH, ligninase enzymes, and phenol oxidase activity (Luo and Zhou, 2006).

discuss - C/N was not a good predictor (they're highly correlated here) ref Schadel 2014

These conclusions have generally come from examining the effect of anthropogenic N deposition or experimental N amendments. Individual studies, including this one, have however observed positive correlations between ambient soil N and microbial respiration. For example, Weiss et al. (2015) found CO2 production from Siberian Yedoma permafrost samples to be correlated with both percent C and N, consistent with our active-layer results (**Table 2**). *In situ* respiration rates have also been shown to be negatively correlated with C:N at large spatial scales (Allaire et al., 2012). Percent C and N both varied widely in our soil cores, even though they were collected within tens of meters of each other, suggesting that active-layer SOC response to temperature and moisture may also be highly spatially variable, even in a visually uniform boreal forest. Spatially explicit analyses of soil properties, temperatures (Bond-Lamberty et al., 2005), and respiration (Allaire et al., 2012) are likely necessary to accurately constrain and predict soil fluxes in this ecosystem.

**4.3 Limitations and weaknesses**

There were weaknesses in our approach and experimental design that should be considered. Laboratory experiments offer precise control, but lack the *in situ* nature of field manipulations (Sistla et al., 2013), raising uncertainties to what degree their results can be extrapolated. Soils isolated during incubation may, for example, underestimate temperature sensitivity of respiration (Podrebarac et al., 2016) or exhibit lag effects (Treat et al., 2015). Nonetheless, the controlled environments of incubations provide an important way to elucidate the key mechanisms controlling GHG from high-latitude soils (Schuur et al., 2015). It should also be noted that our 100-day incubation was not long enough to observe slowly-cycling soil fractions, which may vary in their response to experimental manipulation (Karhu et al., 2010).

The soils studied here were from an upland, mixed conifer-deciduous boreal forest, and care needs to be taken before drawing regional inferences, or about other ecosystem types. We focused on an experimental drought, rather than flooding, because of the well-drained nature of the field site: it is unlikely that the mid-slope forest we sampled in will ever suffer from thermokarst or excessive soil moisture, but too-dry conditions are a serious possibility in this low-precipitation ecosystem (Barber et al., 2000).

Finally, the soils here are not surface layer soils (where the majority of microbial activity and C mineralization of labile C takes place); removing them from *in situ* conditions (where they are less exposed to O2, for example) may significantly change the abiotic conditions to which the microbial community is adapted. However, focusing on the active layer provides crucial information about the potential loss of C from these soils, a risk that needs to be well understood as permafrost degradation leads to expansions in the depth of the active layer across the Arctic.

#### 5 Conclusions

In this laboratory experiment, we found that CO2 fluxes were strongly influenced by temperature and water content, and correlated with soil C and N, while CH4 fluxes were much smaller and not sensitive to temperature or water content in these well-drained mineral soils. These results add to a growing body of Arctic permafrost and active layer incubation literature, and underscore the importance of understanding moisture effects on CO2 fluxes in particular. How soil moisture might change with spatially variable permafrost degradation, how soil biota will respond to these changes, and how models should treat soil organic matter decomposition with respect to multiple and interacting drivers are all critical areas of research going forward. Further controlled field and laboratory studies, ideally tightly integrated with modeling experiments, are important to understand GHG emission dynamics from high-latitude soils.

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#### Author contributions

B.B.-L., A.P.S., and V.L.B. designed this experiment. B.B.-L. and A.P.S. performed field sampling, and A.P.S. led the laboratory incubation and analyses. B.B.-L. wrote the manuscript, with contributions from all authors.

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**Table 1.** Summary of dissolved organic carbon (DOC), percent C, percent N, and CO2 and CH4 fluxes by treatment. Each column summarizes (mean ± s.d.) 12 cores, combining two groups of N=6 at each incubation temperature, except for the pre-incubation values shown in the right-hand column (N=6).

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Variable | Controlled drought | Drought | Field moisture | Pre-incubation |
| DOC (mg/kg) | 165.68 ± 66.46 | 154.60 ± 57.15 | 173.62 ± 46.67 | 125.43 ± 49.07 |
| C (%) | 0.87 ± 0.50 | 0.76 ± 0.60 | 1.67 ± 1.60 | 1.44 ± 1.32 |
| N (%) | 0.04 ± 0.03 | 0.03 ± 0.03 | 0.08 ± 0.08 | 0.07 ± 0.06 |
| BD (g/cm3) | 1.06 ± 0.17 | 1.08 ± 0.14 | 0.89 ± 0.18 | - |
| CO2 (µg C/g C/day) | 159.77 ± 116.41 | 97.03 ± 96.38 | 456.40 ± 543.91 | - |
| CH4 (µg C/g C/day) | 0.10 ± 0.00 | 0.10 ± 0.00 | 0.10 ± 0.00 | - |

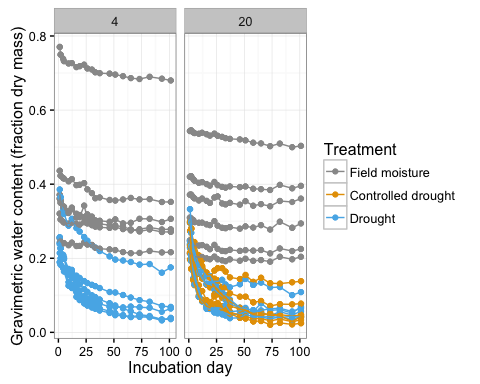
**Table 2.** Linear mixed-effects model parameters, testing effects of temperature (°C), gravimetric water content (unitless), soil C (%), soil N (%), and dissolved organic carbon (mg kg-1) on individual core CO2 fluxes (+0.1 µg C g C-1 day-1); a colon (":") indicates an interaction. Dependent variable has units of log(µg C g C-1 day-1). Columns include parameter value; standard error (SE); degrees of freedom (DF); T statistic; and P value.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  | Value | SE | DF | T | P |
| (Intercept) | 1.713 | 0.354 | 1153 | 4.839 | < 0.001 |
| Temperature | 0.046 | 0.020 | 26 | 2.336 | 0.027 |
| WC\_gravimetric | 3.496 | 1.052 | 1153 | 3.322 | 0.001 |
| N\_percent | 37.976 | 6.810 | 26 | 5.576 | < 0.001 |
| Temperature:WC\_gravimetric | 0.116 | 0.061 | 1153 | 1.905 | 0.057 |
| Temperature:N\_percent | -0.507 | 0.300 | 26 | -1.690 | 0.103 |
| WC\_gravimetric:N\_percent | -37.347 | 8.425 | 1153 | -4.433 | < 0.001 |

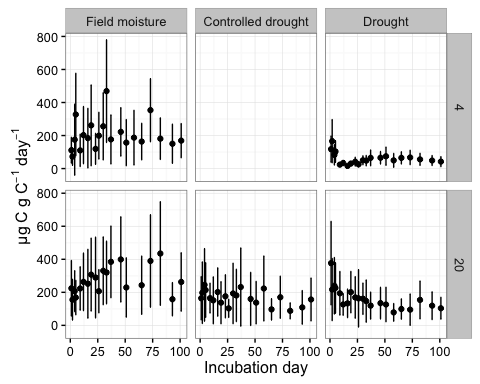
**Table 3.** Linear mixed-effects model parameters, testing effects of temperature (°C), gravimetric water content (unitless), soil N (%),and dissolved organic carbon (DOC, mg kg-1) on log-transformed, individual core CH4 fluxes (+0.1 µg C g C-1 day-1); a colon (":") indicates an interaction. Dependent variable has units of log(µg C g C-1 day-1). Columns include parameter value; standard error (SE); degrees of freedom (DF); T statistic; and P value.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  | Value | SE | DF | T | P |
| (Intercept) | 1.713 | 0.354 | 1153 | 4.839 | < 0.001 |
| Temperature | 0.046 | 0.020 | 26 | 2.336 | 0.027 |
| WC\_gravimetric | 3.496 | 1.052 | 1153 | 3.322 | 0.001 |
| N\_percent | 37.976 | 6.810 | 26 | 5.576 | < 0.001 |
| Temperature:WC\_gravimetric | 0.116 | 0.061 | 1153 | 1.905 | 0.057 |
| Temperature:N\_percent | -0.507 | 0.300 | 26 | -1.690 | 0.103 |
| WC\_gravimetric:N\_percent | -37.347 | 8.425 | 1153 | -4.433 | < 0.001 |

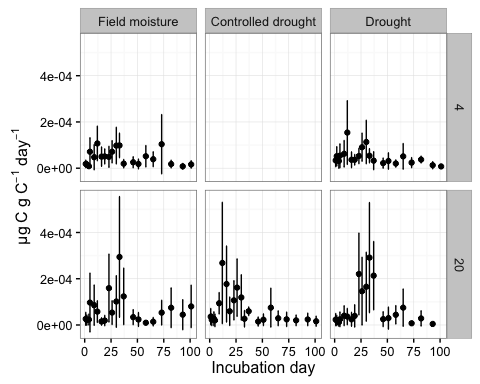
**Figure 1.** Core water content across the course of the incubation experiment by temperature (left panel 4 °C, right panel 20 °C) and treatment.



**Figure 2.** Mass-normalized CO2 fluxes over the 100-day incubation, by temperature (4 and 20 °C, rows) and treatment (field moisture, drought, and controlled drought; columns). Error bars show core-to-core standard deviation. The "controlled drought" treatment, for 20 °C only, was meant to dry cores at roughly the same rate as the drought cores at 4 °C.



**Figure 3.** Mass-normalized CH4 fluxes over the 100-day incubation, by temperature (4 and 20 °C, rows) and treatment (field moisture, drought, and controlled drought; columns). Error bars show core-to-core standard deviation. The "controlled drought" treatment, for 20 °C only, was meant to dry cores at roughly the same rate as the drought cores at 4 °C.



**Figure 4.** Cumulative C fluxes (mg g C-1) over the incubation, by gas (CO2 and CH4, top and bottom panels respectively) and treatment (columns). Letters within a panel indicate significant differences based on Tukey's HSD.

