

# Python - Application examples

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### **Availability of slides**

- All materials are freely available (CC BY) after the lectures:
  - StudIP: 'Python for Life Scientists'
  - GitHub: https://github.com/bpucker/teaching
- Questions: Feel free to ask at any time
- Feedback, comments, or questions: b.pucker[a]tu-bs.de

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#### Reverse complement of nucleotide sequence

What happens here?

```
Sequence of bases e.g. ATGACATGA
   pdef revcomp( seq ):
2
        seq = seq.lower() 			 Converts input to lower case: atgacatga
 3
4
 5
        #key:value (=dictionary)
        complement = { 'a':'t', 't':'a', 'c':'g', 'g':'c' }
6
8
        new seq = []
                                Get complement for each base
9
10
        for nt in seq:
11
            new seq.append( complement[ nt ] )
12
13
        #list[::-1] inverts list (last element becomes first)
14
        new seq = "".join(new seq[::-1])
15
16
        return new seq
                                             Inverts list (=reverse)
```

#### How to use dictionaries

- Values are accessible via keys
- Keys need to be unique
- Quick access to data based on key
- Higher memory occupation than lists/strings

```
my_dict = {"k1": "v1", "k2": {"x1": "y1"}, 5: ["one", "two", "three"], "hello world": "hello world" }
print(my_dict.keys()) #all keys of a dictionary
print(my_dict.values()) #all values of a dictionary
print(my_dict["k1"])
print(my_dict["k2"]["x1"])

dict_keys(['k1', 'k2', 5, 'hello world'])
dict_values(['v1', {'x1': 'y1'}, ['one', 'two', 'three'], 'hello world'])
v1
y1
```



#### **Exercises - Part 6a**

- 6.1) Write a function to get the reverse complement (upper case letters) of a DNA sequence given in upper case letters!
- 6.2) Write a function to convert a DNA sequence into a RNA sequence!
- 6.3) Write a function to translate a DNA sequence into amino acids (first frame only)!
- 6.4) Write a function to translate DNA sequences in all 6 frames into peptide sequences!
   The longest peptide sequence per DNA sequence should be returned!
- 6.5) Write a function to grep a sequence from a FASTA file based on the name of this sequence!



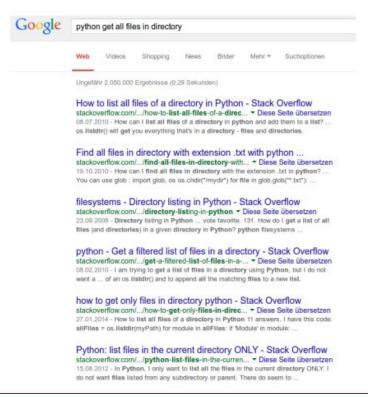
### How to approach a challenge in bioinformatics?

- Identify the problem that needs to be solved
- Split the problem into smallest possible parts
- Solutions for small parts of the problem might be available already
- Precise description of the problem is required for online search
- Best hit often leads to stackoverflow:
  - Problem is described by the asking person
  - Multiple solutions are suggested by the community
  - Community votes to identify the best solution
  - Green marking highlights the answer that solved the problem



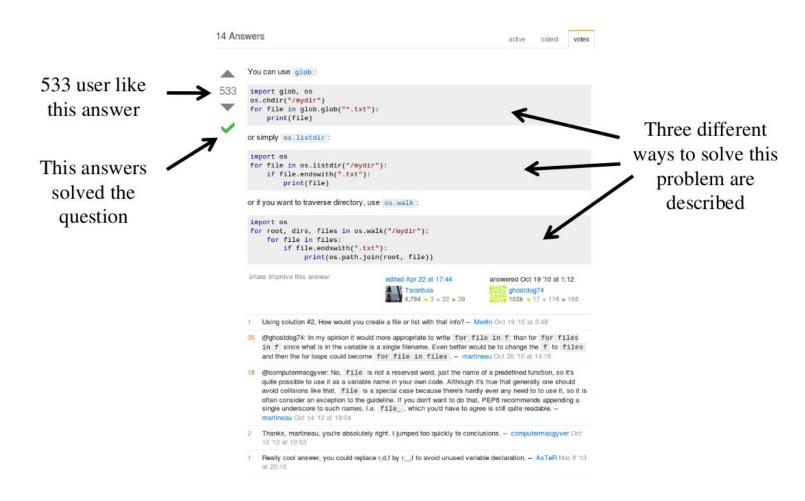
#### **Example**

- Problem: get paths of all files in a certain directory
- Search expression: 'python get all files in directory'





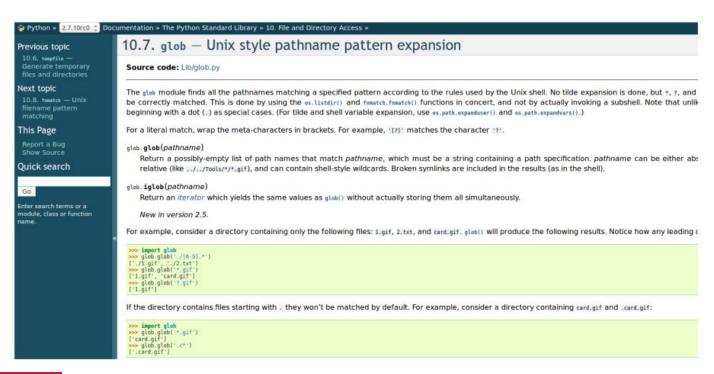
#### Best hit on stackoverflow





#### Official Python documentation

- Systematic documentation of all functions in a module with all possible arguments
- Sometimes examples are given





### Linux (Ubuntu)



- Ubuntu is an operating system with a graphical user interface
- Excellent environment to perform bioinformatics
- Offers a powerful terminal and comes with comprehensive support
- Processing of large data sets is more efficient via command line tools
- Dual boot system with Windows is possible
- Configuration of USB stick for Ubuntu is possible to explore opportunities



#### **How to run BLAST?**

- Running at the NCBI website does not give you full control over all parameters
- Python can be used to run a local search:
  - blastn \
  - -query <query\_file> \
  - -subject <subject file> \
  - -out <output\_file> \
  - -outfmt 6 \
  - -evalue 0.01 \
  - -word size 4

1.	qseqid	query (e.g., gene) sequence id
2.	sseqid	subject (e.g., reference genome) sequence id
3.	pident	percentage of identical matches
4.	length	alignment length
5.	mismatch	number of mismatches
6.	gapopen	number of gap openings
7.	qstart	start of alignment in query
8.	qend	end of alignment in query
9.	sstart	start of alignment in subject
10.	send	end of alignment in subject
11.	evalue	expect value
12.	bitscore	bit score



#### How to execute processes via shell?

Running shell commands through the subprocess module:

```
p = subprocess.Popen( arg='ls -lh', shell=True )
p.communicate()
```

- Can be used to run everything via Python
- Python waits until the command is completed
- Example:

```
import subprocess
p = subprocess.Popen( arg="mkdir test && cd test && ls -lh", shell=True )
p.communicate()
```



### **How to process BLAST results?**

```
□def load BLAST results( input file ):
      2
                  """! @brief load all BLAST results from file """
      3
      4
                  data = []
      5
                  with open( input file, "r" ) as f:
      6789
                       line = f.readline()
                       while line:
                            parts = line.strip().split('\t')
                            data.append( {
                                                 'query': parts[0],
     10
                                                  'subject': parts[1],
     11
                                                  'query start': int( parts[6] ),
     12
                                                  'query end': int( parts[7] ),
     13
                                                  'score': float( parts[-1] )
     14
     15
                            line = f.readline()
     16
                  return data
                                                          429
                                                                       429
                                                                             0.0
                                                                                    895
AT1G01010
            NdCChrl.ql.tl 100.00
                               429
                                                   1
                                                                1
AT1G01010
            NdCChr4.q18734.t1
                                32.84
                                      469
                                             247
                                                   15
                                                          1
                                                                428
                                                                       2
                                                                             443
                                                                                    4e-56
                                                                                           194
AT1G01010
            NdCChr1.q127.t1 34.23
                                336
                                      157
                                             10
                                                   1
                                                          330
                                                                1
                                                                       278
                                                                             1e-41
                                                                                    152
AT1G01010
            NdCChr1.g128.t1 32.38
                                349
                                      157
                                             14
                                                   1
                                                          331
                                                                1
                                                                       288
                                                                             4e-39
                                                                                    146
AT1G01010
            NdCChr4.g18730.t1
                                39.33
                                     178
                                             95
                                                          1
                                                                175
                                                                       2
                                                                             169
                                                                                    1e-32
                                                                                           126
                                             89
AT1G01010
            NdCChr3.g12773.t1
                                39.63 164
                                                          1
                                                                162
                                                                       1
                                                                             156
                                                                                    1e-28
                                                                                           115
                                40.74 162
                                             79
                                                          5
                                                                159
                                                                      11
                                                                                    3e-28
AT1G01010
            NdCChr4.g22969.t1
                                                                             162
                                                                                           117
            NdCChr4.g18733.t1
                                40.00
                                     165
                                                                162
                                                                       2
                                                                             160
                                                                                    4e-28
AT1G01010
                                                                                           115
            NdCChr3.g17122.t1
                                42.31 156
                                             74
                                                                153
                                                                       15
                                                                             161
                                                                                    4e-27
                                                                                           112
AT1G01010
```



#### **Exercises - Part6b**

- 6.6) Collect the best CHS BLAST result per contig from the CHS\_vs\_Digitalis.txt file.
- 6.7) Count the number of BLAST hits that show a similarity >80%, an alignment length >200, and an e-value<10<sup>-10</sup>.



### How to organize a Python script

- Make a script recognize that it needs to run with Python:
   #!/usr/bin/env python3
- Other information to include:
  - Author
  - Version
  - Usage
  - Imports

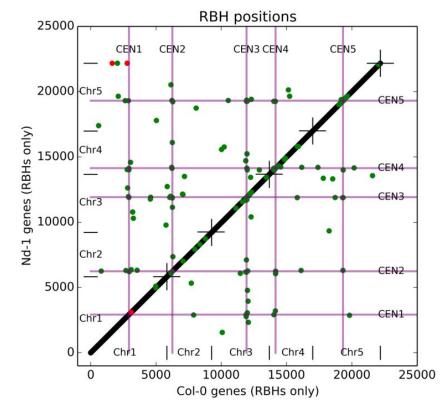
```
### Boas Pucker ###
     ### bpucker@cebitec.uni-bielefeld.de ###
     ### v0.2 ###
      usage = """
                 python construct RNA seq coverage file.py\n
                 --in <BAM FILE>
                 --out <OUTPUT FILE>
 8
10
                 --bam is sorted <PREVENTS EXTRA SORTING OF BAM FILE>
11
12
                 feature requests and bug reports: bpucker@cebitec.uni-bielefeld.de
13
14
15
     cite = """ Pucker & Brockington, 2018: https://doi.org/10.1186/s12864-018-5360-z """
16
17
18
     import os, sys
19
20
     # --- end of imports --- #
22 pdef main( arguments ):
```

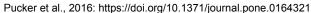
#### How to pass arguments to a Python script?

```
usage = """ how to run the script and list of arguments """
 3
 4
   □def main( arguments ):
         """! @brief run everything """
 5
 6
 7
         fasta file = arguments[ arguments.index( '--fasta' )+1 ]
         gff3 file = arguments[ arguments.index( '--gff3' )+1 ]
 8
         species = arguments[ arguments.index( '--species' )+1 ]
 9
         output dir = arguments[ arguments.index( '--tmp' )+1 ]
10
11
         hints file = arguments[ arguments.index( '--hints' )+1 ]
12
         if '--cutoff' in arguments:
13
             cutoff = int( arguments[ arguments.index( '--cutoff' )+1 ] )
14
15
         else:
16
             cutoff = 1
17
18
         #everything happens here
19
20 ☐ if '--fasta' in sys.argv and '--gff3' in sys.argv and '--species' in sys.argv and '--tmp' in sys.argv and '--hints' in sys.argv:
21
         main( sys.argv )
22 Felse:
23
         sys.exit( usage )
```

### matplotlib

- Importing matplotlib: import matplotlib.pyplot as plt
- Visualization of complex data
- Automatic generation of plots
- Unlimited customization options







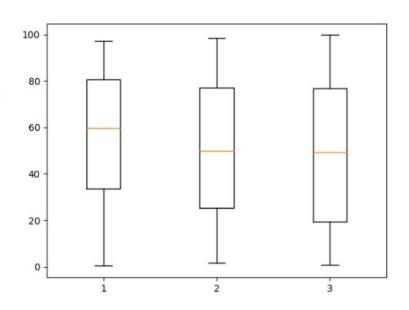
### **Box plot**

```
import matplotlib.pyplot as plt
import numpy as np

dl = np.random.rand(50) * 100  #generate random numbers
d2 = np.random.rand(50) * 100
d3 = np.random.rand(50) * 100

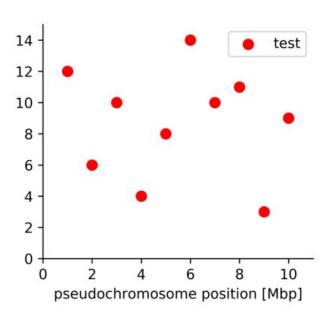
data = [d1, d2, d3] # multiple box plots on one figure

plt.figure()
plt.boxplot(data)
plt.show()
```



### Scatter plot

```
import matplotlib.pyplot as plt
 1
 2
       fig, ax = plt.subplots(figsize=(10, 4)) #defining size of plot
 3
 4
 5
       x values = [1, 2, 3, 4, 5, 6, 7, 8, 9, 10]
 6
       y values = [ 12, 6, 10, 4, 8, 15, 10, 11, 3, 9 ]
 7
       ax.scatter( x values, y values, color="red", s=10, marker="0", label="test" )
 8
       #setting color, marker size, marker shape and label of this group
 9
10
11
       ax.legend( numpoints=1)
12
       #each group is represented by only one marker in the legend (default=3)
13
14
       ax.set xlim(0, 11) #set range of x-axis
15
       ax.set ylim(0, 15) #set range of y-axis
16
17
       ax.set xlabel( "pseudochromosome position [Mbp]" )
18
19
       ax.spines["top"].set visible(False)
                                              #remove lines and ticks
20
       ax.spines["right"].set visible(False) #remove lines and ticks
21
22
       plt.subplots adjust(left=0.05, right=0.99, top=0.97, bottom=0.12)
23
       #adjust size of plot within figure
24
25
       plt.show()
26
       fig.savefig( "my_plot.png", dpi=600 ) #write figure into output file
27
       plt.close( "all" ) #destroy created figures (cleaning up)
```

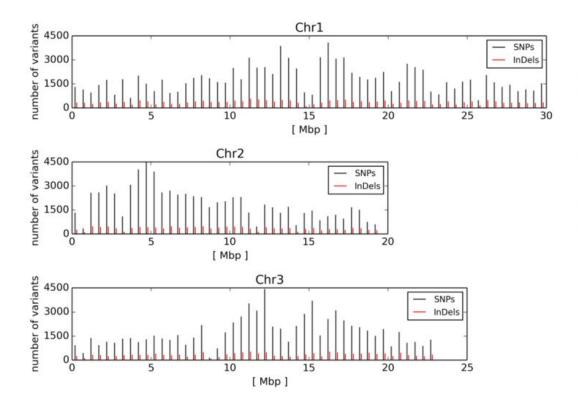


#### **Histogram**

```
import matplotlib.pyplot as plt
 2
 3
      # --- end of imports --- #
 4
 5
     Egene_space = [ 3, 3, 6, 6, 9, 9, 12, 3, 3, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,
 6
                      11, 12, 13, 14, 15, 16, 17, 18, 19, 20,
                                                                                                           CDS
                                                                                                                                  not CDS
                                                                                                1000
                                                                                                                        50000
                      12, 15, 18, 21, 24, 27, 30 ]
 8
     □intergenic = [ 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 1, 2, 3, 4, 5, 6, 7, 8, 9,
 9
                       1, 2, 3, 4, 5, 6, 7, 1, 2, 3, 4, 1, 2, 1]
                                                                                                 800
                                                                                                                        40000
10
11
                                                                                                                        30000
12
      fig, (ax1, ax2) = plt.subplots(1, 2, sharey=False)
13
      counts, bins, patches = axl.hist( gene space, bins=max( gene space ), align="left"
14
      ax1.set title( "CDS" )
                                                                                                                        20000
15
      ax1.set xlim( 0, 30 )
16
      ax1.set xlabel( "InDel size [bp]" )
                                                                                                                        10000
                                                                                                 200
17
      ax1.set ylabel( "number of InDels" )
18
19
      counts, bins, patches = ax2.hist( intergenic, bins=max( intergenic ), align="left" )
                                                                                                                                      20
                                                                                                         10
                                                                                                           15
                                                                                                               20
                                                                                                                  25
                                                                                                                     30
                                                                                                                                 10
                                                                                                                                    15
20
                                                                                                         InDel size [bp]
      ax2.set title( "not CDS" )
                                                                                                                                 InDel size [bp]
21
      ax2.set xlim(0, 30)
22
      ax2.set xlabel( "InDel size [bp]" )
23
      plt.subplots adjust( wspace=0.3 ) #increase space between figures
24
25
      plt.show()
26
      fig.savefig( prefix + "InDel size distribution.png", dpi=300 )
27
      plt.close('all')
```



### **Barplot figure**



barplots.py generates barplots at specific positions by drawing a normal line

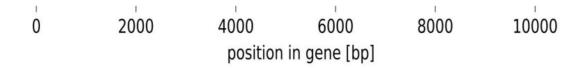
(script is available in course repository)



### **Gene structure plot**

 gene\_structure\_plot.py generates visualizations of gene/transcript structures based on GFF annotations







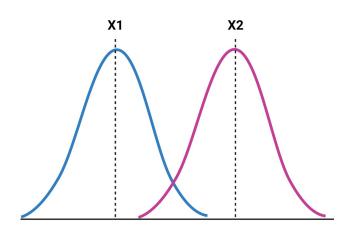
#### **Exercises - Part6c**

- 6.8) Construct a figure to illustrate the order and orientation of genes in the gum gene cluster of *Xanthomonas campestris* pv. campestris!
- 6.9) Save this figure in different file formats (png, jpg, pdf, svg)!



#### **Statistics**

- Compare observed sample against the expected distribution
- Check if two samples are derived from same distribution
- H0 = samples were taken from same distribution
- H0 can only be rejected or kept due to insufficient evidence against it
- H0 can NEVER be confirmed





### **Shapiro-Wilk test**

- Testing data set for normal distribution
- Important for decision about potential tests

```
from scipy import stats
x = [1, 2, 3, 3, 3, 2, 1]
stats.shapiro(x)
```



#### Correlation

- Pearson correlation coefficient is suitable for data following a normal distribution
- Spearman correlation coefficient is better if data distribution is unknown

```
from scipy import stats
x = [1,2,3,4,5]
y = [2,4,6,8,10]
r,p = stats.pearsonr(x,y)
r,p = stats.spearmanr(x,y)
```



#### t-test

- Samples need to show normal distribution
- Comparison of one sample against a reference value
- Comparison of two samples:
  - Paired samples (ttest\_rel)
  - Unpaired samples (ttest\_ind)

```
from scipy import stats
x = [1,2,3,4,5]
y = [4,6,8,10,11]
t,p = stats.ttest_ind(x,y) #independent samples
t,p = stats.ttest_rel(x,y) #paired samples
```



#### W-test

- Wilcoxon (W) test compares two paired samples
- Normal distribution is not required

```
from scipy import stats
x = [1,2,3,4,5]
y = [4,6,8,10,11]
w,p = stats.wilcoxon(x,y)
```

#### **U-test**

- Comparison of unpaired samples
- Normal distribution is not required

```
from scipy import stats
x = [1,2,3,4,5]
y = [4,6,8,10,11]
w,p = stats.mannwhitneyu(x,y)
```

### Chi square test

- Comparison of an observation against an expectation
- Comparisons of two observations

```
from scipy import stats
obs = [1,2,3,4,5]
exp = [4,6,8,10,11]
x, p = stats.chisquare(obs,exp)
```

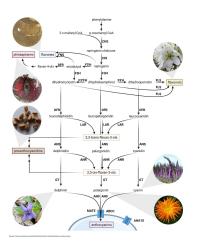
### Exercises - Part6c (UNKNOWN\_DATA.ods)

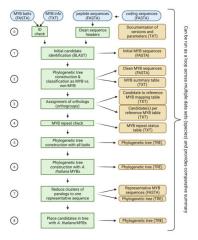
- 6.10) Construct a suitable visualization!
- 6.11) Analyze distribution and trends!
- 6.12) Apply statistical test to investigate difference!



### Looking for more opportunities to apply Python?

- Plant Biotechnology and Bioinformatics group is working on:
  - Plant genomics
  - Plant transcriptomics (RNA-seq)
  - Specialized plant metabolites
  - Synthetic biology
  - Big data comparative studies
  - Tool development
  - Data reuse
- Details: <a href="https://www.tu-braunschweig.de/en/ifp/pbb">https://www.tu-braunschweig.de/en/ifp/pbb</a>
- Web server: <a href="https://pbb-tools.de/">https://pbb-tools.de/</a>





https://doi.org/10.1186/s12864-022-08452-5



## Time for questions!



### References (biological background of examples)

- Nd-1 genome assembly (Pucker et al., 2016)
  - https://doi.org/10.1371/journal.pone.0164321
- Non-canonical splice sites (Pucker et al., 2017)
  - https://doi.org/10.1186/s13104-017-2985-y
- Croton tiglium transcriptome assembly (Haak et al., 2018)
  - https://doi.org/10.3389/fmolb.2018.00062
- Genome-wide non-canonical splice sites in plants (Pucker & Brockington, 2018)
  - https://doi.org/10.1186/s12864-018-5360-z
- Chromosome-level Nd-1 genome assembly (Pucker et al., 2019)
  - https://doi.org/10.1371/journal.pone.0216233
- NAVIP (Baasner et al., 2019)
  - https://doi.org/10.1101/596718

