## **Group Meeting**

8/8/2007

#### Agenda

- Cell culture (C2C12 regular and 4R-CAT)
- Sonication
- ChIP preps and DNA concentration
- Solexa

### Large Cell Culture Effort

## Goal: collect chromatin to be used in a Solexa ChIP analysis

Setup: 36x 15 cm plate and 30x 6 cm plate

15 cm plates harvested at 0, ~12, 24 and 60 hrs

6 cm plates fixed at 0, 6, 12, 18, 24, 36, 48, 60, 72, 96 hrs

#### More Cell Culture

- As Biran and others have noted, cell density increases after the removal of serum and addition of differentiation medium.
- To compensate, 2x 15 cm plates were used per sample for the 0 hrs timepoint
- 1x 15 cm plate was used per sample for all post-differentiation timepoints (for ease of comparison)

#### Even More Cell Culture

Actual timepoints harvested:

C2C12 (WT): 0, 15, 24, 61

C2C12 (4R): 0, 12, 25, 60

Small plates fixed as planned (but there was always a fix accompanying the collection of chromatin)

#### Sonication

Which parameters are best?

Ali and Gordon get good results with 1 ml volume

Brian gets good shearing with ~200-300 μl volume

What is a "good result"?

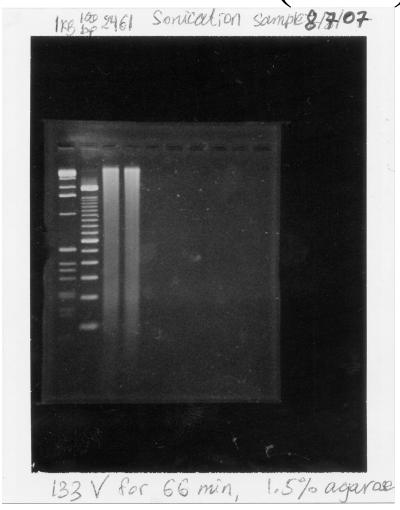
#### Sonication (5 months ago)



each lane = 1/2of a 3x 10 cm plate sample

Lanes 3-6 are sonicated using the number of cycles shown (2.0) Lanes 7-8 are the product from lane 6 (15 cycles of sonication) that was purified using the QIAquick purification kit / column.

Sonication (now)



Lanes 1-2 are the 1 KB and 100 bp ladders, respectively Lanes 3-4 are sonication products of the 24 and 61 hrs diff time points from the C2C12 (wt) t.c. (12 cycles @ 2.5, 300 µl)

#### Sonication

Switching back to using 1 ml now after having used 200-300 µl for the last several months

Ali and Gordon both use 1 ml

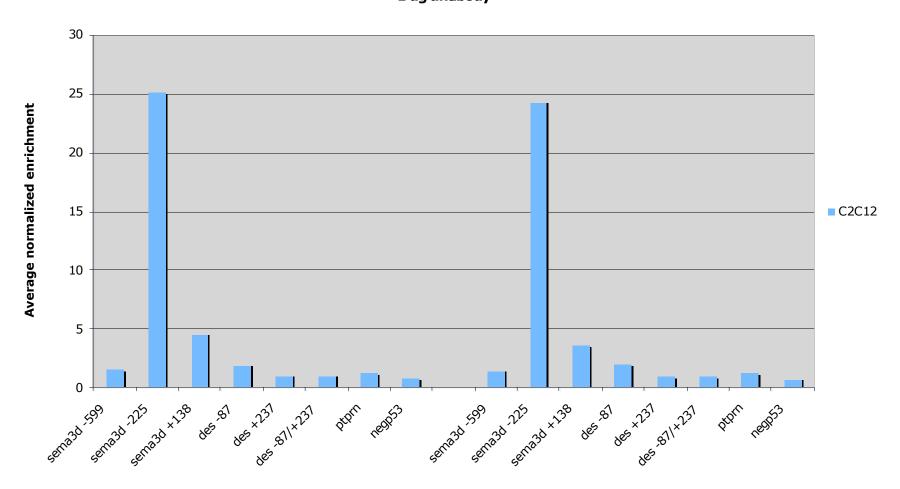
Did a batch of 8 sonications last night using Gordon's parameters

Foaming is a problem

Results pending

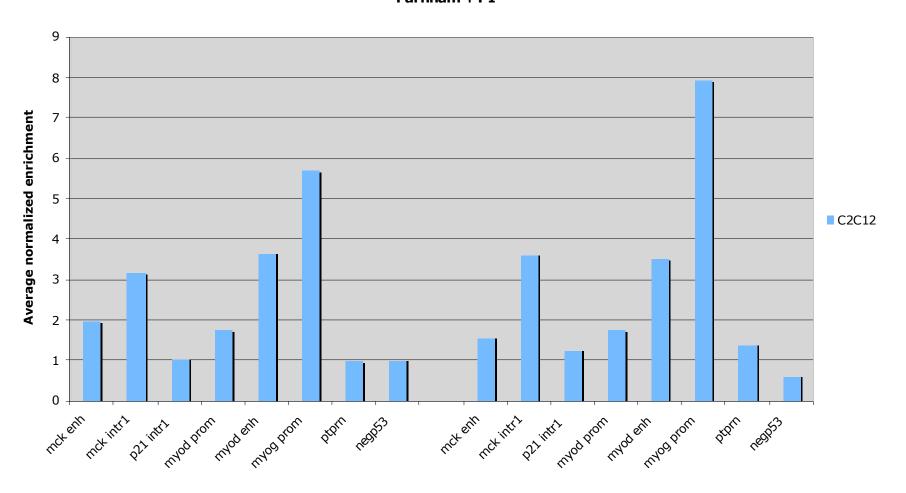
#### F5D ChIP (new primers)

F5D Santa Cruz ChIP 15 (3/23/2007)
100 hrs post differentiation C2C12s, 15 cycles of sonication, 100 ul beads,
1 ug antibody



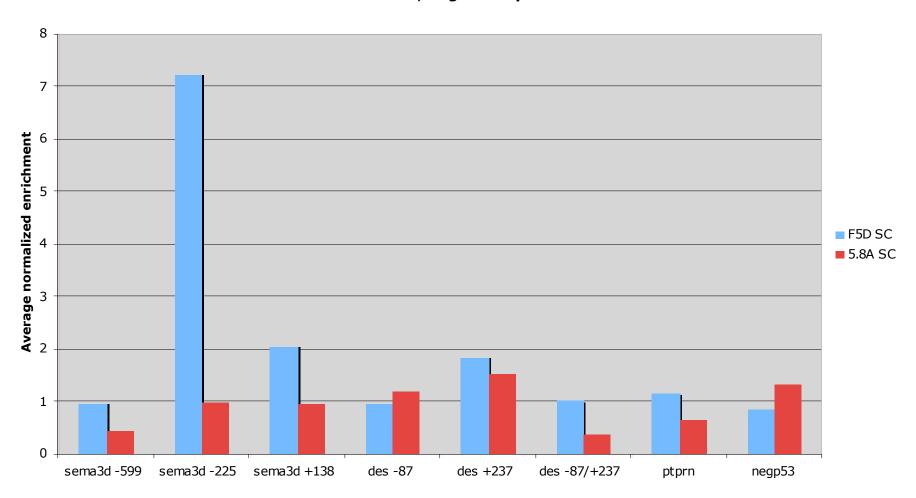
## F5D ChIP (old primers)

F5D Santa Cruz ChIP 16 (3/23/2007)
100 hrs post differentiation C2C12s, 100 ul beads, 1 ug antibody, 15 cycles sonication,
Farnham + PI



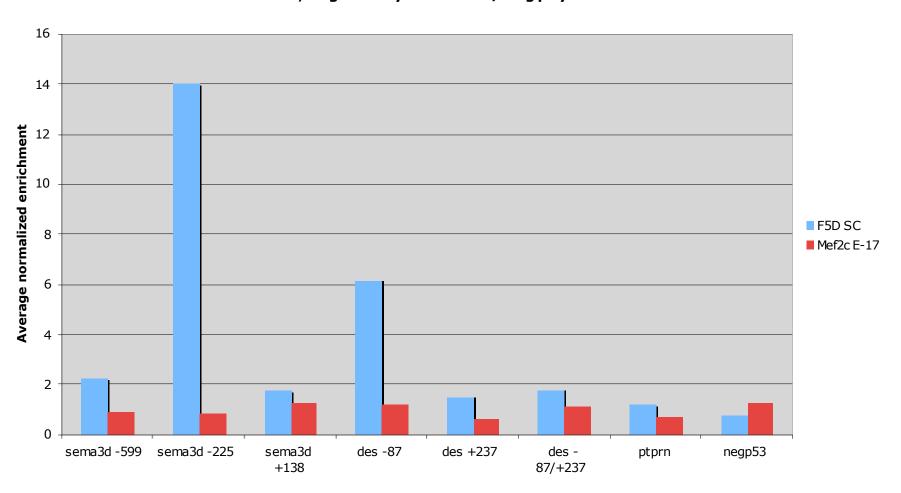
## F5D and 5.8A ChIP (new primers)

F5D SC and 5.8A ChIP 18 (4/23/2007)
63 hrs post differentiation C2C12s + ara-C, Farn + PI /20 cycles of sonication, 100 ul beads, 1 ug antibody



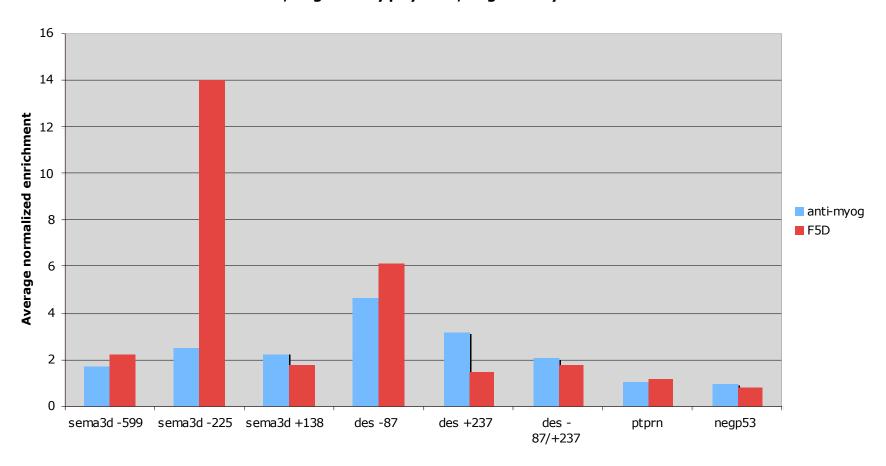
## F5D and Mef2c - E17 ChIP (new primers)

F5D Santa Cruz and Mef2c E-17 ChIP 19 (4/25/2007)
63 hrs post differentiation C2C12s + ara-C, Farn + PI /20 cycles of sonication, 100 ul beads, 1 ug antibody monoclonal / 4 ug polyclonal



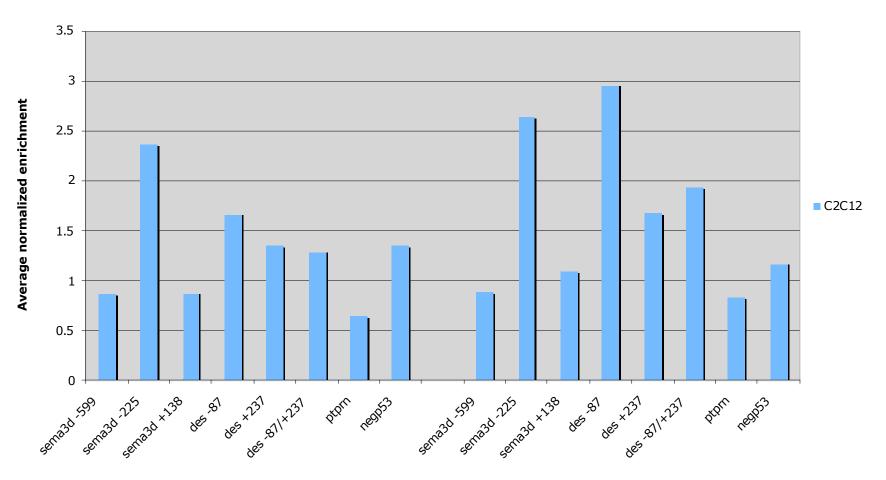
# F5D SC and anti-myog polyclonal ChIP (new primers)

Polyclonal Rabbit anti-myog and F5D
63 hrs post differentiation C2C12s + ara-C, Farn + PI
15/20 cycles of sonication (respectively),
100 ul beads, 2 ug antibody polyclonal, 1 ug antibody monoclonal



#### F5D Home ChIP (new primers)

F5D "Home" ChIP 21 (8/5/2007)
24 and 61 hrs post differentiation C2C12s, no ara-C, Farn + PI,
12 cycles of sonication, 100 ul beads, 1 ug antibody



#### Cell Density/Growth

Cells (aka cell body) grow during differentiation → larger pellet at later timepoints

However, amount of DNA appears to increase as well

Will test the effect of ara-C on postdifferentiation growth (results pending)

#### Cell Density/Growth

Density after 24 hrs diff: 197 ng/µl

Density after 61 hrs diff: 308 ng/µl

Extrapolated total chromatin:

- $24 \text{ hrs} = 325 \mu g$
- $61 \text{ hrs} = 508 \mu g$

## Factors Affecting Final DNA Conc.

- Time
- Scraping efficiency
- Transfer efficiency during washes
- Phenol/chloroform extraction efficiency
- ara-C

#### Solexa

Myogenin Solexa run pending completion of:

- Test ChIPs
- Sonication tests
- Immunostaining

#### The End

#### RNA Timecourse Data (from Brian)

