## Methylation status of B cells of those afflicted with Transient Hypogammaglobulinemia

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Research Hypothesis and Experimental Proposal

## 1 B cell development

## 1.1 Stages

A B cell is a type of lymphocyte which contributes to the adaptive immune system by producing and secreting antibodies 1. Refer to later section which has

MOYC infO Production of B cells begins in foetal development and rises throughout gestation<sup>2</sup>. However, antibodies are predominantly provided for the foetus by the mother through active transfer across the placenta<sup>3,4</sup>. After birth, levels of serum antibody IgG are equal to or slightly greater than maternal levels. Subsequently, levels decrease rapidly reaching their lowest point at 3-6 months of age<sup>5</sup>. The low IgG is normal physiologic hypogammaglobulinemia. When serum IgG levels do not increase; when physiologic hypogammaglobulinemia is prolonged or accentuated, it can be described as Transient Hypogammaglobulinemia of infancy (THI)<sup>6</sup>.

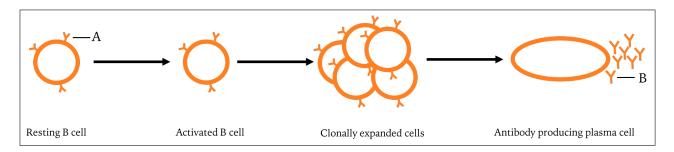


Figure 1: Schematic representation of B cell differentiation from resting cell to antibody producing plasma cell. Resting B cells remain inactive until encountering antigen corresponding to the B cell receptor (A). After clonal expansion, differentiation into plasma cells occurs. Plasma cells produce antibodies such as IgG (B).

## 2 Transient Hypogammaglobulinemia of infancy

THI is not consistently described across the literature. Antibodies are produced in many classes including IgG, IgM, IgA and IgE<sup>1</sup> Most THI definitions require that the concentration of at least one class of antibody be at a serum concentration more than two standard deviations (SD) below the mean for age matched controls<sup>4,5,7,8</sup>. Both IgG and IgA isotypes have been used for diagnosis.

## 2.1 Review of etiology and theories: Ovadia and Dalal 2014

- Most studies require that the concentration of at least one class of Ig be more than 2 standard deviations below the mean for age matched controls. (IgG makes sense, IgA is problematic because IgA levels reach only 20% of adult values at 1 year of age and there is high variability in the normal range of IgA levels anyway.
- In Dressler et al.<sup>5</sup>, they measured IgG levels at 2, 6, 12, 18, 24 and 40 months, and compared levels to age-matched controls
- The cause of THI remains unknown despite numerous pathogenic mechanisms being proposed.
  - 1. Fundenberg and Fundenberg demonstrated that gamma globulin antigenic determinants present in human fetal IgG molecules and not in maternal IgG can stimulate an immune response to fetal IgG during pregnancy. They concluded that alloantibodies might cross the placenta and cause transient suppression of fetal immunoglobulin production prospective study on the issue did not support the hypothesis.
  - 2. Soothill suggested that THI is a manifestation of genetic heterozygosity for some other ID diseases it remains a possibility but an example has not been found
  - 3. Siegel proposed a defect in TH cell maturation, following an observation of low T cell numbers, his observations have not been supported by many other studies
  - 4. More recent work has suggested a role for cytokines in the pathogenesis of THI. An enhanced production of  $\text{TNF}\alpha/\beta$  and IL-10 has been observed. Adding  $\text{TNF}\alpha/\beta$  was shown to inhibit IgG and IgA secretion. It was concluded that TNF may be involved in regulating IgG/IgA production and the balance between TNFs which suppress IgG production, and IL-10 which induces IgG production, may be important for the normal development of IgG secreting B cells.
  - 5. McGeady <sup>8</sup> mentioned that frequent administration of antibiotics could potentially diminish bacterial gut flora, however a number of patients in the study did not receive antibiotics frequently so it seems improbable.
- The defining factor of THI is that it has a gradual tendency to increase, unlike X linked or common variable agammaglobulinemia spontaneous recovery is said to occur between 2 and 4 years of age<sup>7</sup> "Most of the children did well after 2 years of age." <sup>8</sup>

- McGeady <sup>8</sup> said that the children they investigated had fewer infections as they grew older and cited two papers with similar observations
- most studies have found that lymphocyte subpopulations and cellular immunity are intact, including the level of memory and class-switched B cells.
- Investigators have found reduced frequencies of both circulating IgM+ and "switched" memory B cells as well as an inability to produce IgG in vitro<sup>9</sup>.
- McGeady<sup>8</sup> said that the concept of delayed activation is consistent with most immunologists' current perception of THI

## 2.2 B cell features in transient hypogammaglobulinemia of infancy

#### 2.2.1 B cells in hypogammaglobulinemia - Fiorilli et al. 1986

- Immunoglobulin deficiency can be the result of (1) a failure of pre-B cells to differentiate into B cells, as in X-linked hypogammaglobulinemia, (2) a defect of isotype switch or (3) a failure of B cells to differentiate terminally into antibody-secreting plasma cells.
- (3) is commonly seen in patients with common variable immunodeficiency, which may present with low, normal or increased numbers of circulating B cells carrying IgM, IgG or IgA molecules on their surface.
- A number of studies have indicated that distinct mechanisms may prevent patients' B cells from differentiating properly, including the so-called "intrinsic B cell defects", deficiencies, deficiencies of helper T cells, and the presence of activated autosuppressive T cells
- B cells of some patients with CVI have been said to display patterns of membrane immunoglobulin isotypes resembling those of immature B lymphocytes
- Study investigated the presence of immature B cells, ie, cells carrying simultaneously IgG and IgM molecules on their surface, in 12 patients with primary IgG deficiencies.
- In 10 patients (except 2 with CVI) have the majority of the circulating IgG bearing B cells also expressing surface IgM.
- Found that patients with a profound deficiency of serum IgG usually have significant numbers of circulating IgG bearing B cells. In most cases they resembled immature B lymphocytes in that they express multiple surface immunoglobulin isotypes
- Platts-Mills et al. found that B cells from the majority of CVI patients behaved as functionally immature cells in the sense that they produced IgM but very little IgG or IgA in vitro. The other CVI patients had CVI which was uniformly associated with autoimmune disorders or thymomas as well as increased suppression, suggesting a secondary (acquired) hypogammaglobulinemia. Furthermore, B cells from patients of the latter group responded to polyclonal activation with Epstein-Barr virus in a manner qualitatively similar to that of mature B cells.

#### 2.2.2 Subsets of transient hypogammaglobulinemia of infancy Dalal et al. 1998

- Study aimed to characterise the subsets of THI, in an attempt to define the disorder.
- Used 35 patients, assigned to three categories:
  - 1. Patients eventually have normal total serum levels with normal IgG subclass division, and normal specific antibody production. The process may have transient phase whereby some IgG subclasses may become unbalanced
  - 2. Patients continue to have low serum IgG levels, and poor antibody titres
  - 3. Serum IgG becomes normal, but individuals remain incapable of mounting an adequate antibody response
- The groups are likely to represent a heterogenous group of different genotypes, and it is important to understand the phenotypes, before the genotypes are explored

# 3 Lineage commitment and methylation status in other cells

Epigenetic changes have been coined as the hallmark of cell differentiation. T cell activation and skewing, a certain type of cell differentiation, is governed in great parts by epigenetic changes which insure that the clone of a T cell will retain its phenotype (Th1/Th2 etc).

As said by Choukrallah and Matthias <sup>11</sup> the same TFs can be equally expressed in different cell types, and yet have different binding profiles. So, addition to the DNA sequence recognition, TF binding strongly depends on chromatin structure and epigenetic modification.

## 3.1 Methylation

- DNA methylation was the first epigenetic mechanism recognised, and the one that is most extensively studied. 12
- De novo methylation occurs in response to various cellular stressors, and results in the addition of a methyl group to position 5 of a cytosine residue <sup>12</sup>.
- CpG islands have clusters at promoters and enhancers. Up to 90% of genome CpG sites are methylated, with most unmethylated CpG islands being in active genes <sup>12</sup>
- Schmidl et al. <sup>13</sup>: Methylation of cytosine residues in genomic DNA is an important epigenetic mark that is essential for normal embryonic development in mammals, imprinting, X inactivation and silencing of potential hazardous genetic elements such as transposons

• DNA methylation provides an additional mechanism for gene regulation, it can occur at the fifth position of cytosine and can repress via the following mechanisms: (1) Inhibit protein binding (like TFs). (2) Recruit proteins containing domains which interfere with transcription by recruiting repressors <sup>11,12</sup>.

#### 3.2 T cells

### 3.2.1 Bégin and Nadeau2014: T cell methylation in development

- The Th1/Th2 balance of T cells can be affected by environmental exposures which change epigenetic controls.
- In resting CD4<sup>+</sup> cells, the IL-4 and IFN- $\gamma$  genes are methylated
- Upon allergic sensitisation, the IL-4 promoter in allergen-specific T cells is demethylated, correlating with IL-4 expression.
- The IL-4 locus of Th2 cells is also marked with permissive histone modifications (H3K4me3) which are absent in Th1 or naïve T cells.
- At the main Th2 gene locus on chromosome 5, a chromatin hub that interacts with GATA-3 is formed.
- GATA-3 interacts with HAT enzyme p300 and Chd to induce permissive histone and chromatin changes.
- GATA-3/Chd complex binds HDAC to repress the locus encoding TBET, the master regulator which activates Th1 and suppresses Th2 genes
- Further suppression of Th1 cytokines is achieved by the increase of their DNA methylation from naïve state.
- Th2 genes are demethylated mechanism is still incompletely understood.
- Epigenetics is complicated: the GATA-3 promoter has been shown to keep its repressive histone modification despite Th2 activation. It presents a bivalent state with repressive and activating histone modifications positive feedback is important to insure stable expression.
- In the thymus, iTregs are induced by TCR engagement, subsequent NF- $\kappa$ B signalling induces permissive histone modifications and potentially initiates chromatin remodelling in the Foxp3 locus.
- DNA demethylation of the CNS2, also called the Treg-specific demethylated region (TSDR) is a major event in tTreg differentiation, and carries an important function in Foxp3 stabilising Foxp3 expression. CNS2 is the site at which Foxp3 binds to its own gene to maintain expression in a positive feedback mechanism allowing for a persistent phenotype and suppressive function

- Activated, Non Treg T cells also express Foxp3 upon TCR engagement, however the expression is transient owing to the continued methylation of the CNS2.
- Comparing tTregs to Foxp3<sup>+</sup> activated effector T cells, there are hundreds of loci throughout the genome which show demethylation and correspond to binding sites for Foxp3. The methylation changes are not induced by Foxp3, but rather, allow Foxp3 to access its targets and exert its function.
- In activated T cells, the loci are methylated, which could explain the difference in function despite the expression of Foxp3 in activated T cells.

## 3.2.2 Schmidl et al. 2009: Differentially methylated regions that may be enhancers

Study used conventional CD4<sup>+</sup> T cells to compare to CD4<sup>+</sup> CD25<sup>+</sup> T regs. Differentially methylated regions were found and thought to indicate methylation sensitive enhancers

- It has been shown that the functional program of Treg cells is at least partially controlled by the miRNA pathways
- For continuous expression of the lineage-directing transcription factor Foxp3, the methylation status of a methylation-sensitive, Treg cell-specific enhancer in intron I is important.
- The restriction of cell type-specific enhancers is a key function of DNA methylation.
- STAT5:
  - Regions specifically demethylated in Treg cells were enriched for STAT5 consensus sites.
  - TF STAT5 is activated by IL-2
  - Treg cell survival requires IL-2
  - STAT5 also has an essential role in Treg homeostasis and is known to regulate Foxp3 through an intronic, methylation-sensitive enhancer.
  - It would make sense that STAT5 bind at this region, but it has not been confirmed in vivo
- In T-reg cells, A functionally important intronic enhancer of the Foxp3 gene was shown to be methylation sensitive properties.
- In the study, conventional CD4<sup>+</sup> T cells were compared to CD4<sup>+</sup> CD25<sup>+</sup> T regs
- Half of the tested DMRs (differentially methylated regions) significantly enhanced the activity of a heterologous promoter in transient reporter gene assays performed in a T cell leukaemia line.
- All regions lost enhancer activity upon CpG methylation.

Eg. Found methylation sensitive enhancer in intron 4 of CD40LG in Tconv (CD4<sup>+</sup> cells). CD40L is important in regulating B cell function through interaction with CD40 on B cells and Dendritic cells.

## 4 B cell development and differentiation

## 4.1 Stages and markers of B cell differentiation:

- Pro-B cells express B220, which coincides with entry into B cell lineage
- pre-BI cells express CD19 and complete recombination of heavy chain IgH D to J segments.
- Next stage sees generation of IgH V(D)J alleles, allowing heavy chain expression which assembles with the surrogate light chain to form the pre-B cell receptor.
- Cells need to pass functional tests here
- small pre-BII cells rearrange the light chain allowing formation and exposure of a functional Ig molecule (BCR)
- immature cells can leave the bone marrow and enter the periphery <sup>11</sup>]

## 4.2 Transcription factors in B cell development

#### 4.2.1 Pu.1

- Very upstream TF, essential for the development of lymphoid cells as well as macrophages and neutrophils.
- Disruption of PU.1 in mouse was shown to prevent commitment of MPPs towards lymphoid lineage

#### **4.2.2 IKAROS**

- Also upstream
- Mutational disruption of Ikaros DNA-binding domain leads to an early block in lymphopoiesis before commitment to lymphoid restrictd stages.
- Also involved in later stages of B cell development, where it promotes heavy chain gene rearrangement by inducing expression of RAG1/2 genes.
- Also required for differentiation of large pre-B cells to small pre-B cells and for transcription and rearrangement of the IgL locus.

#### 4.2.3 E2A

- Required for Ebf1 and FoxO1 expression at the Common Lymphoid Progenitor stage
- E2A mutant mice lack B cells

#### 4.2.4 EBF1

- Essential for B cell specification and commitmnt.
- Regulates expression of genes required for B cell development including FoxO1 and Pax5

#### 4.2.5 Pax5

- Essential for B cell commitment and maintenance of B cell identity through activation of B cell specific genes and repression of lineage inappropriate genes
- Deletion of Pax5 in mature B cells leads to de-differentiation to lymphoid progenitors, which can differentiate into functional T cells

#### 4.2.6 FoxO1\*\*

- Early deletion of FoxO1 causes substantial block at pro-B cell stage due to failure to express IL-7 receptor alpha chain
- Inactivation of FoxO1 in late pro-B cells results in arrest at pre-B cell stage due to impaired expression of RAG1/2 (direct targets of FoxO1)
- Deletion in peripheral B cells leads to reduced number of LN B cells, due to down regulation of L-selectin and defect in class-switch recombination

### 4.2.7 c-Myb and Runx

- Deletion of c-Myb in mice leads to a block at the pre-pro B cell stage which is accompanied with impaired expression of the  $\alpha$  chain receptor and Ebf1
- Deletion of Runx1 also causes a developmental block at the pro-B cell stage accompanied by reduced expression of E2A, Ebf1 and Pax5.
- Runx1-deficient pro-B cells were shown to harbour excessive amounds of the repressive histone mark H3K23me3 in the Ebf1 proximal promoter
- Retroviral transduction of Ebf1, not Pax5, into Runx1-deficient progenitors restores B cell development.

## 4.3 Transcription factors in B cell differentiation - Li et al. 2013

- Resting B cells display genome wide DNA hypomethylation
- Genes crucial for the maintenance of B cell identity (*Pax5*, *Spib*, *Ebf1*) and B cell marker genes (*CD19*), display active epigenetic state
- Chromatin in transcribed  $Igh \ V_H DJ_H$  regions, the intronic  $\mu$  enhancer and the  $Igh \ 3$ ' locus control region contains hypomethylated DNA and activating histone modifications
- The epigenetic marks were likely introduced during B cell development, because the open chromatin state of these regions is required for V(D)J recombination
- Active epigenetic marks in the *Igh* locus and in the *Pax5*, *Spib*, *Ebf1* and *CD19* loci persist during naïve B cell activation
- Upon activation by antigens, B cells undergo DNA demethylation and histone modifications, and express a specific set of miRNAs.
- Repression of the *Aicda* gene in naïve B cells is mediated by promoter hypermethylation, during B cell activation, *Aicda* DNA is demethylated and the locus becomes enriched in active histone modifications
- DNA hypomethylation seems to be important, as B cells carrying identical pre-rearranged  $Ig\kappa$  alleles, only the hypomethylated allele is hypermutated despite comparable transcription of both alleles.
- S regions: the genes which contain IgG, IgA etc genes. They are acted upon when undergoing Class Switch Recombination
- Active epigenetic state is found in even naïve B cells, indicating that  $S\mu$  is in a constitutively open state, poised for switching.
- For plasma cell differentiation: Blimp-1 (encoded by *Prdm1*. Epigenetic induction of Blimp-1 causes events which drives plasma cell differentiation and possibly maintains plasma cell identity
- Differentiating into memory cells not likely a problem
- Overall DNA hypomethylation has been associated with systemic autoimmune diseases

Region/gene	Epigenetic mark	function of epigenetic mark
V(D)J	DNA hypomethylation	Increases region accessibility
Igh 3' LCR	DNA hypomethylation	Mediates germline VDJ and $I_H$ -S- $C_H$ transcription

## 4.4 DNA methylation in B cells during maturation and differentiation

### 4.4.1 Why epigenetics?

- Lara-Astiaso et al. <sup>15</sup> noted that mutations in loss of chromatin factors lead to haematopoiesis defects and disease
- Tagoh et al. <sup>16</sup> said that even before the onset of gene expression and stable TF binding, specific chromatin alterations are observed (including methylation changes). Hence the idea that epigenetic programs guiding blood cell differentiation are engraved into the chromatin of lineage-specific genes, and such chromatin changes are implemented **before cell lineage specification**; Epigenetic programs are engraved into the chromatin of lineage-specific genes before cell lineage specification and the onset of detectable gene expression
- Differentiation and lineage commitment are associated with specific methylation or demethylation events <sup>13</sup>

#### 4.4.2 Methylation status in B cells

- Methylation loss is observed as B cells mature <sup>17</sup>.
- Hypomethylation is enriched in enhancer/promoter regions
- The TF families which show hypomethylation are AP-1, EBF, RUNX, OCT, IFF and NF $\kappa$ B
- The cell subtypes which show the most pronounced methylation changes in comparison to the preceding stage are germinal centre B cells, memory B cells and BM plasma cells <sup>18</sup>
- It is possible to accurately classify B cells into their maturation stage by the methylation state of 5 CpGs in genes important to B cell differentiation <sup>18</sup>.
- Transition from HPCs to pre-B1 cells has an inverse corelation between the expression of TFs and the methylation of their binding sites; High methylation status occurs with low expression of TFs<sup>18</sup>.

## 5 Proposal

- Bisulfite sequencing of enhancers or transcription factors that are important to B cell development
- Take B cells from infants with CVID and THI, as well as controls. B cells should be taken at different stages, up until age 5

## References

- [1] Cooper, M. D. (2015). The early history of B cells. *Nature reviews. Immunology*, 15(3): 191–7.
- [2] Rechavi, E., Lev, A., Lee, Y. N., Simon, A. J., Yinon, Y., Lipitz, S., Amariglio, N., Weisz, B., Notarangelo, L. D., and Somech, R. (2015). Timely and spatially regulated maturation of B and T cell repertoire during human fetal development. *Science translational medicine*, 7(276):276ra25.
- [3] Kohler, P. F. and Farr, R. S. (1966). Elevation of cord over maternal IgG immunoglobulin: evidence for an active placental IgG transport. *Nature*, 210(5040):1070–1.
- [4] Dalal, I., Reid, B., Nisbet-Brown, E., and Roifman, C. M. (1998). The outcome of patients with hypogammaglobulinemia in infancy and early childhood. *The Journal of pediatrics*, 133(1):144–6.
- [5] Dressler, F., Peter, H. H., Müller, W., and Rieger, C. H. (1989). Transient hypogamma-globulinemia of infancy: Five new cases, review of the literature and redefinition. *Acta paediatrica Scandinavica*, 78(5):767–74.
- [6] GITLIN, D. and JANEWAY, C. A. (1956). Agammaglobulinemia, congenital, acquired and transient forms. *Progress in hematology*, 1:318–29.
- [7] Tiller, Jr, T. L. and Buckley, R. H. (1978). Transient hypogammaglobulinemia of infancy: review of the literature, clinical and immunologic features of 11 new cases, and long-term follow-up. *The Journal of pediatrics*, 92(3):347–53.
- [8] McGeady, S. J. (1987). Transient hypogammaglobulinemia of infancy: need to reconsider name and definition. *The Journal of pediatrics*, 110(1):47–50.
- [9] Ovadia, A. and Dalal, I. (2014). Transient hypogammaglobulinemia of infancy. *LymphoSign Journal*, 1(1):1–9.
- [10] Fiorilli, M., Crescenzi, M., Carbonari, M., Tedesco, L., Russo, G., Gaetano, C., and Aiuti, F. (1986). Phenotypically immature igg-bearing b cells in patients with hypogammaglobulinemia. *Journal of clinical immunology*, 6(1):21–5.
- [11] Choukrallah, M. A. and Matthias, P. (2014). The interplay between chromatin and transcription factor networks during B cell development: Who pulls the trigger first? *Frontiers in immunology*, 5:156.
- [12] Bégin, P. and Nadeau, K. C. (2014). Epigenetic regulation of asthma and allergic disease. Allergy Asthma Clinical Immunology, 10(1):27.
- [13] Schmidl, C., Klug, M., Boeld, T. J., Andreesen, R., Hoffmann, P., Edinger, M., and Rehli, M. (2009). Lineage-specific DNA methylation in T cells correlates with histone methylation and enhancer activity. *Genome research*, 19(7):1165–74.

- [14] Li, G., Zan, H., Xu, Z., and Casali, P. (2013). Epigenetics of the antibody response. Trends in immunology, 34(9):460–70.
- [15] Lara-Astiaso, D., Weiner, A., Lorenzo-Vivas, E., Zaretsky, I., Jaitin, D. A., David, E., Keren-Shaul, H., Mildner, A., Winter, D., Jung, S., et al. (2014). Chromatin state dynamics during blood formation. *Science*, 345(6199):943–949.
- [16] Tagoh, H., Melnik, S., Lefevre, P., Chong, S., Riggs, A. D., and Bonifer, C. (2004). Dynamic reorganization of chromatin structure and selective DNA demethylation prior to stable enhancer complex formation during differentiation of primary hematopoietic cells *in vitro*. *Blood*, 103(8):2950–5.
- [17] Oakes, C. C., Seifert, M., Assenov, Y., Gu, L., Przekopowitz, M., Ruppert, A. S., Wang, Q., Imbusch, C. D., Serva, A., Koser, S. D., Brocks, D., Lipka, D. B., Bogatyrova, O., Weichenhan, D., Brors, B., Rassenti, L., Kipps, T. J., Mertens, D., Zapatka, M., Lichter, P., Döhner, H., Küppers, R., Zenz, T., Stilgenbauer, S., Byrd, J. C., and Plass, C. (2016). DNA methylation dynamics during B cell maturation underlie a continuum of disease phenotypes in chronic lymphocytic leukemia. Nature genetics, 48(3):253-64.
- [18] Kulis, M., Merkel, A., Heath, S., Queirós, A. C., Schuyler, R. P., Castellano, G., Beekman, R., Raineri, E., Esteve, A., Clot, G., et al. (2015). Whole-genome fingerprint of the DNA methylome during human B cell differentiation. *Nature genetics*, 47(7):746–756.