

Methylation status of B cells of those afflicted with Transient Hypogammaglobulinemia

Brittany Howell

a1646948

Research Hypothesis and Experimental Proposal

1 Background

Immunoglobulins are a vital component of the adaptive immune system¹. The B cells which produce antibodies (active immunoglobulin) in adults are not fully mature in young infants, resulting in a decrease in serum immunoglobulin levels after birth²⁻⁴. Physiologic hypogammaglobulinemia refers to the point when serum immunoglobulin reaches its lowest point, commonly at 4-6 months of age⁵. Transient hypogammaglobulinemia (THI) is a disorder whereby affected persons have a prolongation or exacerbation of regular hypogammaglobulinemia, followed by spontaneous recovery⁴⁻¹³. The mechanism causing low serum immunoglobulin in THI patients has not yet been elucidated⁶.

Proposed mechanism

Immunoglobulin deficiency can result from B cell precursors failing to either mature into B cells or further fail to differentiate into antibody secreting plasma cells¹⁴. Studies investigating THI have found that levels of circulating B cells are normal and subpopulations of B cells are intact^{4,5,12-15}. With no obvious B cell deficiency, the cause of THI remains unknown⁶.

Fudenberg and Fudenberg¹⁶ first proposed that the fetal IgG molecules produced during pregnancy were enough to stimulate an immune response from the mother. The immune response was thought to produce maternal antibodies which caused transient suppression of fetal immunoglobulin production. Rosen and Janeway⁹ and Nathenson et al.¹⁷ were unable to find agglutinators in four mothers of infants with THI. Willenbockel¹⁸ observed THI as familial, Soothill¹⁹ further proposed THI as a manifestation of genetic heterozygosity for other immunodeficiency diseases, noting the high number of patients who had immunodeficient relatives. While it remains a possibility as noted by McGeady¹⁰, no studies have shown supporting evidence^{12,14,20}. Siegel et al.¹⁵ hypothesised that a T cell deficiency was the cause after the observation of low T cell numbers in THI patients, but T cell numbers have been normal in proceeding studies. McGeady¹⁰ suggested that frequent antibiotic treatment could induce hypogammaglobulinemia by diminishing bacteria gut flora. The explanation was not described in any other THI literature, and was indeed described by McGeady¹⁰ to be improbable due to the mostly brief courses of antibiotic treatments given to THI patients. In summary, a wealth of theories have been proposed for mechanism of THI but none have been supported by replicated evidence.

2 Lineage commitment

Activation and differentiation of T cells is governed greatly by epigenetic changes which insure the phenotype of the T cell²¹. DNA methylation was the first epigenetic mechanism recognised, and the one that is most extensively studied²². In T regulatory cells (Treg), the methylation status of the Treg-specific demethylated region (TSDR) is imperative in Treg differentiation²⁴. In the thymus, where T cells mature, Tregs are induced by T cell receptor engagement. Subsequent demethylation occurs at the TSDR allowing FOXP3 to bind to its own gene to stabilise FOXP3 expression, stabilising differentiation to Treg. FOXP3 is also expressed during the activation of other T cell subsets, but due to the methylation of the

TSDR, FOXP3 expression is transient²⁵. Therefore, demethylation permits FOXP3 binding and thence confirms Treg lineage.

3 Proposal

The most intriguing feature of THI is its self-limited nature; that gradually recurrent infections subside and serum IgG levels increase with no obvious cause^{5,10–12,15,19,26}. An investigation into common variable immunodeficiency, a disease related to THI, B cells resembling immature B cells were observed¹⁴. The observed cells were found to produce little IgG. It is possible that the delayed onset of IgG synthesis in THI is due to incomplete lineage commitment, caused by inappropriate methylation of differentiation genes. In normal B cell maturation, global methylation loss is observed²⁷. Tagoh et al.²⁸ explained that epigenetic programs are engraved into the chromatin of lineage-specific genes before cell lineage specification and the onset of detectable gene expression.

To investigate incomplete lineage commitment, B cells will be sampled from THI patients and age-matched controls and tested for methylation of key development and differentiation genes. Transcription factor families which show hypomethylation in B cell development include are AP-1, EBF, RUNX, OCT, IFF and NF κ B Genes crucial for B cell identity (*Pax5*, *Spib*, *Ebf1*) maintain an active epigenetic state^{29,30}. Using bisulfite sequencing, the methylation status of loci imperative for B cell development can be established.

Hypothesis

At key B cell development loci, THI patients will exhibit more DNA methylation than age matched controls.

References

- [1] Simon, A. K., Hollander, G. A., and McMichael, A. (2015). Evolution of the immune system in humans from infancy to old age. *Proceedings. Biological sciences / The Royal Society*, 282(1821):20143085.
- [2] Martin, R., Nauta, A. J., Ben Amor, K., Knippels, L. M. J., Knol, J., and Garssen, J. (2010). Early life: gut microbiota and immune development in infancy. *Beneficial microbes*, 1(4):367–82.
- [3] Rechavi, E., Lev, A., Lee, Y. N., Simon, A. J., Yinon, Y., Lipitz, S., Amariglio, N., Weisz, B., Notarangelo, L. D., and Somech, R. (2015). Timely and spatially regulated maturation of B and T cell repertoire during human fetal development. *Science translational medicine*, 7(276):276ra25.
- [4] Stiehm, E. R. and Fulginiti, V. A. (1980). *Immunologic Disorders in Infants and Children*, chapter The immunodeficiencies of immaturity, pages 219–238. W.B Saunders Company, Philadelphia, second edition.
- [5] Dressler, F., Peter, H. H., Müller, W., and Rieger, C. H. (1989). Transient hypogammaglobulinemia of infancy: Five new cases, review of the literature and redefinition. *Acta paediatrica Scandinavica*, 78(5):767–74.
- [6] Al-Herz, W., Bousfiha, A., Casanova, J.-L., Chatila, T., Conley, M. E., Cunningham-Rundles, C., Etzioni, A., Franco, J. L., Gaspar, H. B., Holland, S. M., Klein, C., Nonoyama, S., Ochs, H. D., Oksenhendler, E., Picard, C., Puck, J. M., Sullivan, K., and Tang, M. L. K. (2014). Primary immunodeficiency diseases: an update on the classification from the international union of immunological societies expert committee for primary immunodeficiency. *Frontiers in immunology*, 5:162.
- [7] Gitlin, D. and Janeway, C. A. (1956). Agammaglobulinemia, congenital, acquired and transient forms. *Progress in hematology*, 1:318–29.

- [8] Al-Herz, W., Bousfiha, A., Casanova, J.-L., Chapel, H., Conley, M. E., Cunningham-Rundles, C., Etzioni, A., Fischer, A., Franco, J. L., Geha, R. S., Hammarström, L., Nonoyama, S., Notarangelo, L. D., Ochs, H. D., Puck, J. M., Roifman, C. M., Seger, R., and Tang, M. L. K. (2011). Primary immunodeficiency diseases: an update on the classification from the international union of immunological societies expert committee for primary immunodeficiency. *Frontiers in immunology*, 2:54.
- [9] Rosen, F. S. and Janeway, C. A. (1966). The gamma globulins: the antibody deficiency syndromes. *New England Journal of Medicine*, 275(13):709–715.
- [10] McGeady, S. J. (1987). Transient hypogammaglobulinemia of infancy: need to reconsider name and definition. *The Journal of pediatrics*, 110(1):47–50.
- [11] Dalal, I., Reid, B., Nisbet-Brown, E., and Roifman, C. M. (1998). The outcome of patients with hypogammaglobulinemia in infancy and early childhood. *The Journal of pediatrics*, 133(1):144–6.
- [12] Tiller, Jr, T. L. and Buckley, R. H. (1978). Transient hypogammaglobulinemia of infancy: review of the literature, clinical and immunologic features of 11 new cases, and long-term follow-up. *The Journal of pediatrics*, 92(3):347–53.
- [13] Buckley, R. H. (1983). Immunodeficiency. *The Journal of allergy and clinical immunology*, 72(6):627–41.
- [14] Fiorilli, M., Crescenzi, M., Carbonari, M., Tedesco, L., Russo, G., Gaetano, C., and Aiuti, F. (1986). Phenotypically immature igg-bearing b cells in patients with hypogammaglobulinemia. *Journal of clinical immunology*, 6(1):21–5.
- [15] Siegel, R. L., Issekutz, T., Schwaber, J., Rosen, F. S., and Geha, R. S. (1981). Deficiency of t helper cells in transient hypogammaglobulinemia of infancy. *The New England journal of medicine*, 305(22):1307–13.

- [16] Fudenberg, H. H. and Fudenberg, B. R. (1964). Antibody to hereditary human gamma-globulin (GM) factor resulting from maternal-fetal incompatibility. *Science*, 145(3628): 170–1.
- [17] Nathenson, G., Schorr, J. B., and Litwin, S. D. (1971). Gm factor fetomaternal gamma globulin incompatibility. *Pediatric Research*, 5(1):2–9.
- [18] Willenbockel, U. (1960). Transitorisch-protrahiertes Antikörpermangelsyndrom bei zweieiigen Zwillingen. *Zeitschrift für Kinderheilkunde*, 84(5):477–83.
- [19] Soothill, J. F. (1968). Immunoglobulins in first-degree relatives of patients with hypogammaglobulinaemia. transient hypogammaglobulinaemia: a possible manifestation of heterozygosity. *Lancet*, 1(7550):1001–3.
- [20] Ovadia, A. and Dalal, I. (2014). Transient hypogammaglobulinemia of infancy. *LymphoSign Journal*, 1(1):1–9.
- [21] Zeng, W.-p. (2013). 'all things considered': transcriptional regulation of T helper type 2 cell differentiation from precursor to effector activation. *Immunology*, 140(1):31–8.
- [22] Bégin, P. and Nadeau, K. C. (2014). Epigenetic regulation of asthma and allergic disease. *Allergy Asthma Clinical Immunology*, 10(1):27.
- [23] Li, E. and Zhang, Y. (2014). DNA methylation in mammals. *Cold Spring Harbor perspectives in biology*, 6(5):a019133.
- [24] Polansky, J. K., Kretschmer, K., Freyer, J., Floess, S., Garbe, A., Baron, U., Olek, S., Hamann, A., von Boehmer, H., and Huehn, J. (2008). Dna methylation controls Foxp3 gene expression. *European journal of immunology*, 38(6):1654–63.
- [25] Ohkura, N., Kitagawa, Y., and Sakaguchi, S. (2013). Development and maintenance of regulatory T cells. *Immunity*, 38(3):414–423.

- [26] Kowalczyk, D., Mytar, B., and Zembala, M. (1997). Cytokine production in transient hypogammaglobulinemia and isolated IgA deficiency. *The Journal of allergy and clinical immunology*, 100(4):556–62.
- [27] Oakes, C. C., Seifert, M., Assenov, Y., Gu, L., Przekopowicz, M., Ruppert, A. S., Wang, Q., Imbusch, C. D., Serva, A., Koser, S. D., Brocks, D., Lipka, D. B., Bogatyrova, O., Weichenhan, D., Brors, B., Rassenti, L., Kipps, T. J., Mertens, D., Zapatka, M., Lichter, P., Döhner, H., Küppers, R., Zenz, T., Stilgenbauer, S., Byrd, J. C., and Plass, C. (2016). DNA methylation dynamics during B cell maturation underlie a continuum of disease phenotypes in chronic lymphocytic leukemia. *Nature genetics*, 48(3):253–64.
- [28] Tagoh, H., Melnik, S., Lefevre, P., Chong, S., Riggs, A. D., and Bonifer, C. (2004). Dynamic reorganization of chromatin structure and selective DNA demethylation prior to stable enhancer complex formation during differentiation of primary hematopoietic cells *in vitro*. *Blood*, 103(8):2950–5.
- [29] Li, G., Zan, H., Xu, Z., and Casali, P. (2013). Epigenetics of the antibody response. *Trends in immunology*, 34(9):460–70.
- [30] Choukrallah, M. A. and Matthias, P. (2014). The interplay between chromatin and transcription factor networks during B cell development: Who pulls the trigger first? *Frontiers in immunology*, 5:156.