

Methylation status of B cells in Transient Hypogammaglobulinemia of Infancy

Brittany Howell

a1646948

Research Hypothesis and Experimental Proposal

Hypothesis

B cells from Transient Hypogammaglobulinemia of Infancy (THI) patients will exhibit more DNA methylation, indicating incomplete B cell differentiation, than age-matched controls.

1 Background

Immunoglobulins are a vital component of the adaptive immune system¹. The B cells which produce antibodies (active immunoglobulin) in adults are not fully mature in young infants, resulting in a decrease in serum immunoglobulin levels after birth²⁻⁴. Physiologic hypogammaglobulinemia refers to the point when serum immunoglobulin reaches its lowest point, commonly at 4-6 months of age⁵. THI is a disorder whereby affected persons have a prolongation or exacerbation of regular hypogammaglobulinemia, followed by spontaneous recovery⁴⁻¹³. The mechanism causing low serum immunoglobulin in THI patients has not yet been elucidated⁶.

Cause of THI

Immunoglobulin deficiency can result from B cell precursors failing to mature into B cells or fail to differentiate into antibody secreting plasma cells¹⁴. Studies investigating THI have found that levels of circulating B cells are normal and subpopulations of B cells are intact^{4,5,12–15}. With no obvious B cell deficiency, the cause of THI has been speculated extensively, but no proposed cause has been supported by replicated evidence^{9,10,12,14–20}.

In regards to genetic inheritance, THI was initially thought to be familial¹⁸. Soothill¹⁹ proposed that THI was a manifestation of genetic heterozygosity for other immunodeficiency diseases, noting the high number of patients who had immunodeficient relatives. While it remains a possibility as noted by McGeady¹⁰, no proceeding studies have shown supporting evidence^{12,14,20}.

2 Lineage commitment

Activation and differentiation of T cells is governed greatly by epigenetic changes which insure the phenotype of the T cell²¹. DNA methylation was the first epigenetic mechanism recognised, and the one that is most extensively studied²². In T regulatory cells (Treg), the methylation status of the Treg-specific demethylated region (TSDR) is imperative in Treg differentiation²³. In the thymus, where T cells mature, Tregs are induced by T cell receptor engagement. Subsequent demethylation occurs at the TSDR allowing FOXP3 to bind to its own gene to stabilise FOXP3 expression, stabilising differentiation to Treg. FOXP3 is also expressed during the activation of other T cell subsets, but due to the methylation of the TSDR, FOXP3 expression is transient²⁴. Therefore, demethylation permits FOXP3 binding and thence confirms Treg lineage.

3 Proposal

The most intriguing feature of THI is its self-limited nature; recurrent infections gradually subside and serum IgG levels increase with no obvious cause^{5,10–12,15,19,25}. Furthermore, the lack of evidence supporting a genetic basis suggests that the cause of THI is not within the genome^{12,14,20}

In common variable immunodeficiency, a disease related to THI, some B cells resemble immature B cells, producing very little IgG¹⁴. Incomplete maturation caused lack of IgG production, so it is possible that the delayed onset of IgG synthesis in THI is also due to incomplete B cell maturation. Activation and differentiation is greatly influenced by epigenetic changes; latent maturity could be caused by inappropriate methylation of B cell development or differentiation genes.

To study incomplete lineage commitment, B cells will be sampled from THI patients and age-matched controls and characterised using whole-genome bisulfite sequencing. As in Kulis et al.²⁶, DNA methylation maps will be generated for sorted human B cell populations: uncommitted haematopoietic progenitor cells (HPCs), pre-BII cells, naïve B cells from peripheral blood, germinal center B cells, memory B cells from peripheral blood and plasma cells from bone marrow. Global demethylation normally occurs as B cells mature²⁷. If methylation is a cause of delayed maturation, the methylome of THI patients should be distinct to the age-matched controls.

References

- [1] Simon, A. K., Hollander, G. A., and McMichael, A. (2015). Evolution of the immune system in humans from infancy to old age. *Proceedings. Biological sciences / The Royal Society*, 282(1821):20143085.
- [2] Martin, R., Nauta, A. J., Ben Amor, K., Knippels, L. M. J., Knol, J., and Garssen,

- J. (2010). Early life: gut microbiota and immune development in infancy. *Beneficial microbes*, 1(4):367–82.
- [3] Rechavi, E., Lev, A., Lee, Y. N., Simon, A. J., Yinon, Y., Lipitz, S., Amariglio, N., Weisz, B., Notarangelo, L. D., and Somech, R. (2015). Timely and spatially regulated maturation of B and T cell repertoire during human fetal development. *Science translational medicine*, 7(276):276ra25.
- [4] Stiehm, E. R. and Fulginiti, V. A. (1980). *Immunologic Disorders in Infants and Children*, chapter The immunodeficiencies of immaturity, pages 219–238. W.B Saunders Company, Philadelphia, second edition.
- [5] Dressler, F., Peter, H. H., Müller, W., and Rieger, C. H. (1989). Transient hypogammaglobulinemia of infancy: Five new cases, review of the literature and redefinition. *Acta paediatrica Scandinavica*, 78(5):767–74.
- [6] Al-Herz, W., Bousfiha, A., Casanova, J.-L., Chatila, T., Conley, M. E., Cunningham-Rundles, C., Etzioni, A., Franco, J. L., Gaspar, H. B., Holland, S. M., Klein, C., Nonoyama, S., Ochs, H. D., Oksenhendler, E., Picard, C., Puck, J. M., Sullivan, K., and Tang, M. L. K. (2014). Primary immunodeficiency diseases: an update on the classification from the international union of immunological societies expert committee for primary immunodeficiency. *Frontiers in immunology*, 5:162.
- [7] Gitlin, D. and Janeway, C. A. (1956). Agammaglobulinemia, congenital, acquired and transient forms. *Progress in hematology*, 1:318–29.
- [8] Al-Herz, W., Bousfiha, A., Casanova, J.-L., Chapel, H., Conley, M. E., Cunningham-Rundles, C., Etzioni, A., Fischer, A., Franco, J. L., Geha, R. S., Hammarström, L., Nonoyama, S., Notarangelo, L. D., Ochs, H. D., Puck, J. M., Roifman, C. M., Seger, R., and Tang, M. L. K. (2011). Primary immunodeficiency diseases: an update on the

- classification from the international union of immunological societies expert committee for primary immunodeficiency. *Frontiers in immunology*, 2:54.
- [9] Rosen, F. S. and Janeway, C. A. (1966). The gamma globulins: the antibody deficiency syndromes. *New England Journal of Medicine*, 275(13):709–715.
 - [10] McGeady, S. J. (1987). Transient hypogammaglobulinemia of infancy: need to reconsider name and definition. *The Journal of pediatrics*, 110(1):47–50.
 - [11] Dalal, I., Reid, B., Nisbet-Brown, E., and Roifman, C. M. (1998). The outcome of patients with hypogammaglobulinemia in infancy and early childhood. *The Journal of pediatrics*, 133(1):144–6.
 - [12] Tiller, Jr, T. L. and Buckley, R. H. (1978). Transient hypogammaglobulinemia of infancy: review of the literature, clinical and immunologic features of 11 new cases, and long-term follow-up. *The Journal of pediatrics*, 92(3):347–53.
 - [13] Buckley, R. H. (1983). Immunodeficiency. *The Journal of allergy and clinical immunology*, 72(6):627–41.
 - [14] Fiorilli, M., Crescenzi, M., Carbonari, M., Tedesco, L., Russo, G., Gaetano, C., and Aiuti, F. (1986). Phenotypically immature igg-bearing b cells in patients with hypogammaglobulinemia. *Journal of clinical immunology*, 6(1):21–5.
 - [15] Siegel, R. L., Issekutz, T., Schwaber, J., Rosen, F. S., and Geha, R. S. (1981). Deficiency of t helper cells in transient hypogammaglobulinemia of infancy. *The New England journal of medicine*, 305(22):1307–13.
 - [16] Fudenberg, H. H. and Fudenberg, B. R. (1964). Antibody to hereditary human gamma-globulin (GM) factor resulting from maternal-fetal incompatibility. *Science*, 145(3628):170–1.

- [17] Nathenson, G., Schorr, J. B., and Litwin, S. D. (1971). Gm factor fetomaternal gamma globulin incompatibility. *Pediatric Research*, 5(1):2–9.
- [18] Willenbockel, U. (1960). Transitorisch-protrahiertes Antikörpermangelsyndrom bei zweieiigen Zwillingen. *Zeitschrift für Kinderheilkunde*, 84(5):477–83.
- [19] Soothill, J. F. (1968). Immunoglobulins in first-degree relatives of patients with hypogammaglobulinaemia. transient hypogammaglobulinaemia: a possible manifestation of heterozygosity. *Lancet*, 1(7550):1001–3.
- [20] Ovadia, A. and Dalal, I. (2014). Transient hypogammaglobulinemia of infancy. *LymphoSign Journal*, 1(1):1–9.
- [21] Zeng, W.-p. (2013). ‘all things considered’: transcriptional regulation of T helper type 2 cell differentiation from precursor to effector activation. *Immunology*, 140(1):31–8.
- [22] Bégin, P. and Nadeau, K. C. (2014). Epigenetic regulation of asthma and allergic disease. *Allergy Asthma Clinical Immunology*, 10(1):27.
- [23] Polansky, J. K., Kretschmer, K., Freyer, J., Floess, S., Garbe, A., Baron, U., Olek, S., Hamann, A., von Boehmer, H., and Huehn, J. (2008). Dna methylation controls Foxp3 gene expression. *European journal of immunology*, 38(6):1654–63.
- [24] Ohkura, N., Kitagawa, Y., and Sakaguchi, S. (2013). Development and maintenance of regulatory T cells. *Immunity*, 38(3):414–423.
- [25] Kowalczyk, D., Mytar, B., and Zembala, M. (1997). Cytokine production in transient hypogammaglobulinemia and isolated IgA deficiency. *The Journal of allergy and clinical immunology*, 100(4):556–62.
- [26] Kulis, M., Merkel, A., Heath, S., Queirós, A. C., Schuyler, R. P., Castellano, G., Beekman, R., Raineri, E., Esteve, A., Clot, G., et al. (2015). Whole-genome fingerprint of the DNA methylome during human B cell differentiation. *Nature genetics*, 47(7):746–756.

- [27] Oakes, C. C., Seifert, M., Assenov, Y., Gu, L., Przekopowicz, M., Ruppert, A. S., Wang, Q., Imbusch, C. D., Serva, A., Koser, S. D., Brocks, D., Lipka, D. B., Bogatyrova, O., Weichenhan, D., Brors, B., Rassenti, L., Kipps, T. J., Mertens, D., Zapatka, M., Lichter, P., Döhner, H., Küppers, R., Zenz, T., Stilgenbauer, S., Byrd, J. C., and Plass, C. (2016). DNA methylation dynamics during B cell maturation underlie a continuum of disease phenotypes in chronic lymphocytic leukemia. *Nature genetics*, 48(3):253–64.