```
library(sf)
library(spdep)
library(tidyverse)
library(tmap)
NC <- read sf("data/NC REGION.shp")</pre>
NC_UTM <- st_transform(</pre>
 x = NC
  crs = st_crs("EPSG:26917")
queen nb <- NC UTM %>%
  tibble::column to rownames("NAME") %>%
  st_as_sf() %>%
  poly2nb(queen = TRUE)
queen_nb_w <- nb2listw(</pre>
  neighbours = queen nb,
  style = "W",
  zero.policy = TRUE
```

Question 1

Calculate Moran's I for the following variables in the dataset. List the Moran's I values and p-values for each variable in a single summary table:

- MNEM2000
- MNEM1990
- TOTJOB2000
- TOTJOB1990

Provide your R code for the Moran's I tests (not the results themselves – that's what the table is for).

```
# It's totally OK if students do this part manually; I just was curious to
find
# a way to specify a set of variables to input automatically and then get a
# table as an output.

vars <- c("MNEM2000", "MNEM1990", "TOTJOB2000", "TOTJOB1990", "POP2000",
"POP1990")

moran_results <-
NC_UTM %>%
```

```
st drop geometry() %>%
  dplyr::select(any_of(x = vars)) %>%
  map(function(x) {
    moran.test(
      X = X
      listw = queen_nb_w,
      zero.policy = TRUE,
      alternative = "two.sided"
   )[c("estimate", "p.value")]
  })
moran_table <- moran_results %>%
  transpose() %>%
  as tibble() %>%
  unnest wider(estimate) %>%
  mutate(Variable = vars, .before = `Moran I statistic`)
knitr::kable(moran table)
```

Variable	Moran I statistic	Expectation	Variance	p.value
MNEM2000	0.3884704	-0.010101	0.0040192	3.238771e-10
MNEM1990	0.4283699	-0.010101	0.0040993	7.468829e-12
TOTJOB2000	0.1631958	-0.010101	0.0035409	0.003587933
TOTJOB1990	0.1642235	-0.010101	0.0037208	0.004264945
POP2000	0.1647821	-0.010101	0.0035306	0.003248218
POP1990	0.1642235	-0.010101	0.0037208	0.004264945

Question 2:

Calculate Moran's I for the MNEM2000 variable using four different versions of W_{ij} :

- Queen's Case
- Rook's Case
- k = 4 nearest neighbors
- Maximum Distance (100% threshold)

Make sure your weights matrices are row standardized. Provide a table that summarized the calculated I, p-values, and average number of connections per county. Discuss any systematic changes you observe in I, p-values, and average links.

```
NC_centroids <- st_centroid(NC_UTM)

q_w <- NC_UTM %>%
   poly2nb(queen = TRUE)
r_w <- NC_UTM %>%
   poly2nb(queen = FALSE)
```

```
k_w <- knn2nb(knearneigh(NC_centroids,</pre>
k = 4
))
max_dist <- knn2nb(</pre>
  knearneigh(
    x = NC_centroids,
   k = 1
  )
) %>%
  nbdists(coords = NC_centroids$geometry) %>%
  unlist() %>%
  max()
d_w <- dnearneigh(</pre>
  NC_centroids,
 d1 = 0,
 d2 = max dist
)
list_w <- list(
  "Queen's Case" = q_w,
  "Rook's Case" = r_w,
  "k = 4" = k_w,
  "Max Distance" = d_w
)
list w weighed <-
  map(list_w, ~ nb2listw(.x, style = "W", zero.policy = TRUE))
moran_w <- list_w_weighed %>%
  map(function(w) {
    moran.test(
      x = NC_UTM$MNEM2000,
      listw = w,
      zero.policy = TRUE,
      alternative = "two.sided"
    )[c("estimate", "p.value")]
  })
neighbors <- list_w_weighed %>%
  map(function(w) {
    w[["neighbours"]] %>%
      unclass() %>%
      map(length) %>%
      unlist() %>%
      mean()
  }) %>%
  enframe() %>%
```

```
rename(Links = value)

moran_table_w <- moran_w %>%
    transpose() %>%
    as_tibble() %>%
    unnest_wider(estimate) %>%
    mutate(Method = names(list_w_weighed), .before = `Moran I statistic`) %>%
    bind_cols(neighbors["Links"])

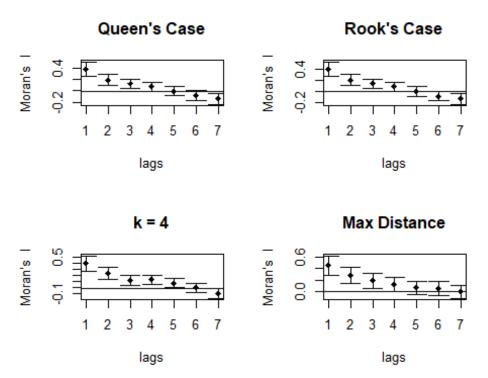
knitr::kable(moran_table_w)
```

Method	Moran I statistic	Expectation	Variance	p.value	Links
Queen's Case	0.3884704	-0.010101	0.0040192	3.238771e-10	4.9
Rook's Case	0.4044841	-0.010101	0.0042276	1.814863e-10	4.62
k = 4	0.4024844	-0.010101	0.0041371	1.412727e-10	4
Max Distance	0.4536758	-0.010101	0.0074840	8.278861e-08	2.8

Question 3:

```
corr_list <- list_w %>%
  map(~ sp.correlogram(
    neighbours = .x,
    var = NC_UTM$MNEM2000,
    order = 7,
    method = "I",
    zero.policy = TRUE
))

par(mfrow = c(2, 2))
corr_plots <- corr_list %>%
  imap(~ {
    plot(.x, main = .y)
  })
```



The general relationship is that the I statistic decreases with the number of lags. However, in certain graphs, e.g. k=4, there are small "bumps" where the value increases. This likely indicates that the "centers" of clusters are approximately $3\sim4$ units (counties) away from one another.

Question 4:

4. Adopt the code above to provide LISA maps for the MNEM1990 variable at an alpha of 0.1, 0.05, and 0.01 using the Queen's case W. Provide a brief explanation of your findings about any local spatial autocorrelation at these more restrictive values of alpha.

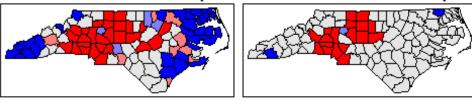
The map in this one is slightly different than the example in the "old" R Module 9. I'm using the quadrants generated by the function itself, which is still based on the mean. I'm not quite sure why the results seem to be different.

```
local_moran_queen <- localmoran(
    x = NC_UTM$MNEM2000,
    listw = queen_nb_w,
    # zero.policy = TRUE,
    alternative = "two.sided"
)

lm_pal <-
    c(
    "High-High" = "red",
    "High-Low" = "#FF000080",</pre>
```

```
"Insignificant" = "gray90",
    "Low-High" = "#0000FF80",
    "Low-Low" = "blue"
  )
quadr <- attr(local moran queen, "quadr")[, 2]</pre>
NC localmoran <- NC UTM
NC localmoran$Quadrant <- quadr
NC localmoran$p.value <- local moran queen[, 5]
sig_level <- 0.5
plot_LISA_sig <- function(sig) {</pre>
  NC localmoran <- NC localmoran %>%
    mutate(
      Quadrant_Sig = if_else(p.value <= sig,</pre>
        true = as.character(Quadrant),
        false = "Insignificant"
      ) %>%
        as.factor()
    )
  t <- tm_shape(NC_localmoran) +
    tm_polygons(
      col = "Quadrant_Sig",
      palette = lm_pal,
      legend.is.portrait = F,
      border.col = "black"
    ) +
    tm_layout(
      main.title = paste0("LISA Cluster at Alpha = ", sig),
      legend.outside = T,
      legend.outside.position = "bottom"
  return(t)
LISA_list \leftarrow map(c(0.5, 0.1, 0.05, 0.01), plot_LISA_sig)
tmap_arrange(LISA_list)
## Legend labels were too wide. Therefore, legend.text.size has been set to
0.47. Increase legend.width (argument of tm layout) to make the legend wider
and therefore the labels larger.
```

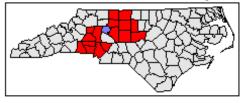
LISA Cluster at Alpha LISA Cluster at Alpha

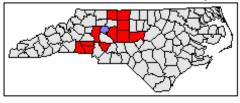




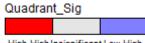


LISA Cluster at Alpha LISA Cluster at Alpha









High-High Insignificant Low-High