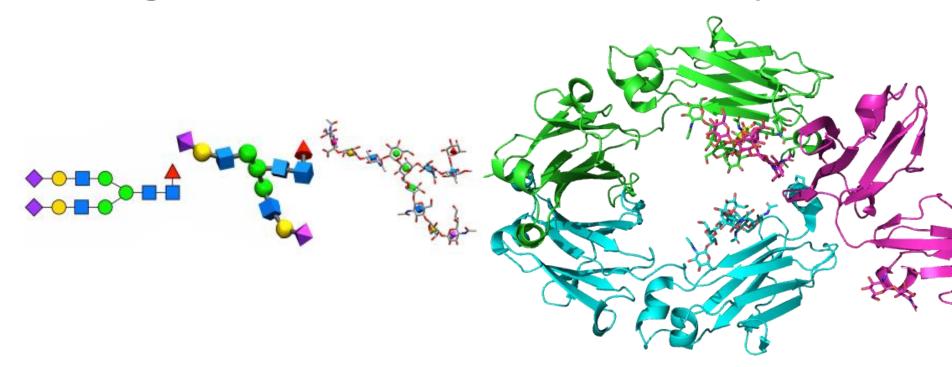
The Dynamics and Molecular Recognition of Complex Carbohydrates



Aoife Harbison Supervisor: Dr. Elisa Fadda



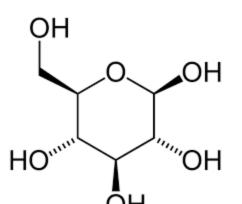




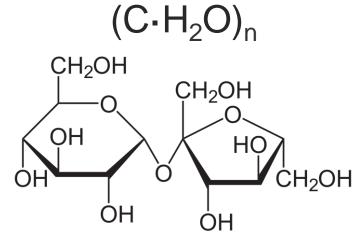
Overview

- Brief Introduction to carbohydrates and glycans
- Computational Chemistry
 - -Molecular Dynamics
 - -Methods for Enhanced Sampling
- Application to studying complex carbohydrates
 - -Free N-glycans
 - -N-glycans on IgG1 antibodies

Carbohydrates



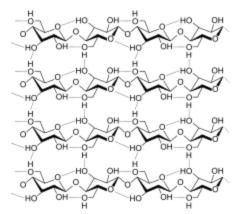
Monosaccharide



Disaccharide



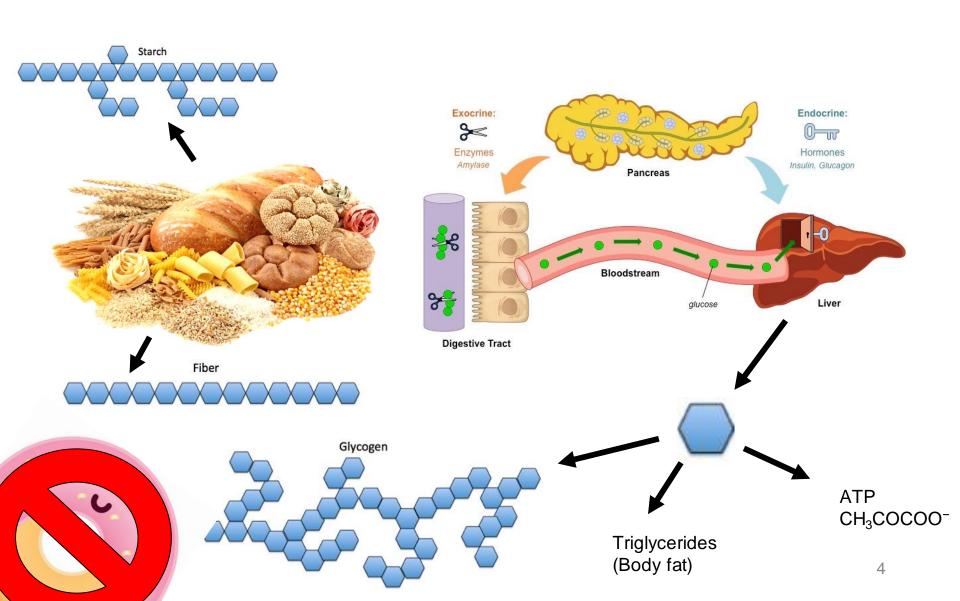




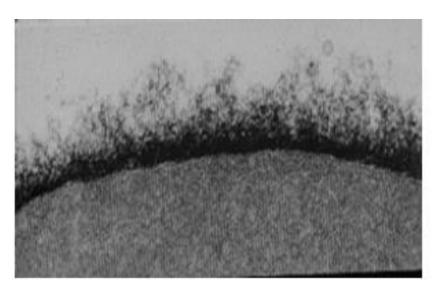
Polysaccharide



Carbohydrates



Complex Carbohydrates: Glycans

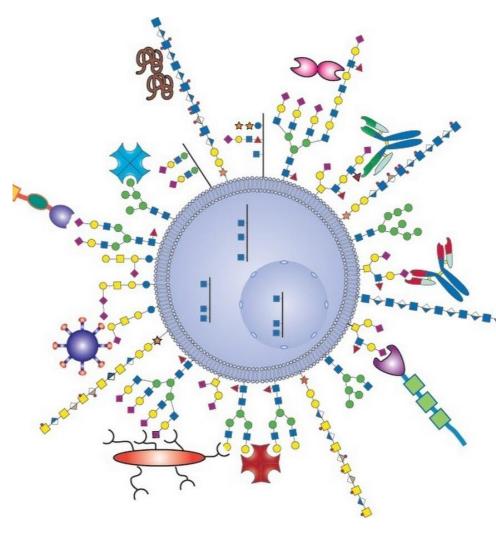


Allon Weiner, The Weizmann Institute of Science, Rehovot, Israel. 2006.

Glycans are post translational modifications of proteins and lipids

They cover the cell membrane constituting what is called glycocalyx

Glycans modulate protein or other biomolecules' activity and function



Richard Cummings, NCFG (2013)

Complex Carbohydrates: Glycans

White (Generic)	Blue	Green	Yellow	Orange	Pink	Purple	Light Blue	Brown	Red
Hexose	Gle	Man	Ga1	Gul	A1t	A11	Tai	Ido	
HæxNAc □	GleNAe	ManNAe	GalNAc	GulNAc	AltNAc	A11NAe	TalNAc	IdoNAc	
Hexos ami ne	G1dN	ManN	GaiN	GulN	AltN	AliN	TaiN	IdoN	
Hexuronate	GlcA	ManA	GalA	GulA ♦	Alt A	A11A	Ta1A ♦	IdoA	
Deoxyhexose	Qui	Rha _			6dAlt ▲		6dTal △		Fue _
DeoxyhexNAc △	QuiNAc //a	RhaNAe 🕰							FueNAc
Di-deoxyhexose	Oli	Tyv		Abe	Par	Dig	Col		
Pentose ☆		Ara ★	Lyx ☆	Xyl ★	Rib ☆				
Nonulos onate		Kdn				Neu5Ac ◆	Neu5Gc	Neu	
Unknown	Bae —	LDManHep	Kdo —	Dha —	DDManHep	MurN Ac	MurNGe	Mur	
Assigned	Api	Fru	Tag	S or	Psi				

Complex Carbohydrates: Glycans

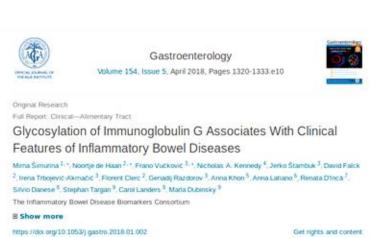


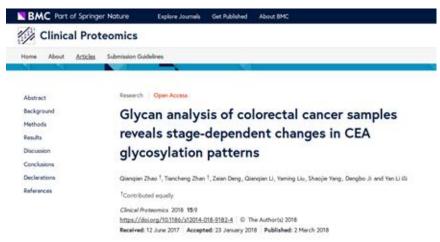
University of Utrecht, "Illuminating the Secret World of your Glycans".

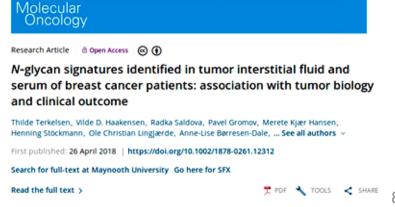
Interest In Glycobiology

By modifying the structure and content of a glycan, the functional properties of the glycoprotein can change, and so enable or disable particular desired/undesired interactions with the protein and other biomolecules, making them attractive as biomarkers and for therapeutic targeting.

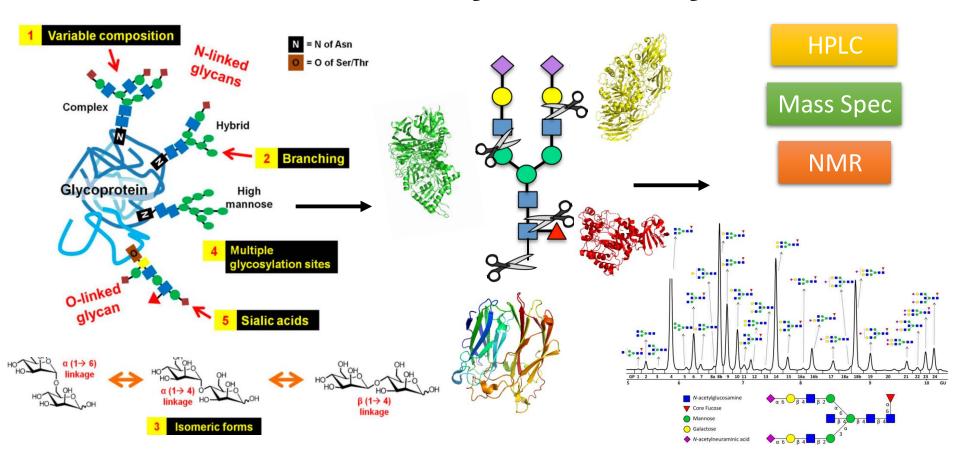








Structural Analysis of Glycans



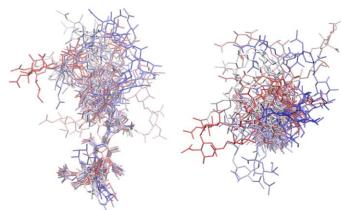
Limitations: not able to account for 3D structure, difficult to differentiate isomers, expensive and time consuming

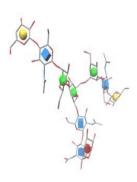
Computational Chemistry

(Computational Biophysics)



- Allows us to understand the significance of the 3D structure of molecules
- Able to examine and explore the physical properties of a molecular system
- Easy (enough) to implement and inexpensive relative to experimental analysis







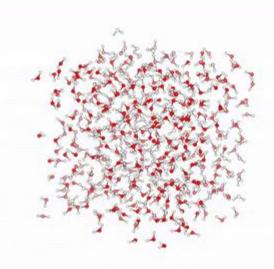
Molecular Dynamics

Representing the **motion** of the atoms and molecules within a specified system **over time**, using classical Newtonian equations of motions

The initial state of the molecule/system is taken from crystal structure, homology model, built using modelling software

Atoms are represented as single point masses inside Van der Waals potentials → behave as hard spheres

Bond and angle restraints are treated as simple harmonic oscillators



Molecular Dynamics

Then interaction potentials are applied to the atoms and molecules using **force fields**, which are composed of:

a functional form

&

parameter sets

Potential energy of the system can be calculated by terms:

Etotal = Ebonded + Enonbonded

Ebonded = Ebond + Edihedral

Enonbonded = Eelectrostatics + Evan der Waals

Empirical set of data for each atom type and molecule type

Atom: atomic mass, Van der Waals radius, partial charge

Molecule: equilibrium values for bond lengths, bond angles, dihedrals angles

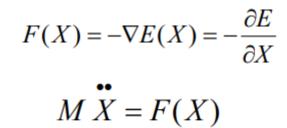
Selecting the right force field is imperative, have to choose wisely!

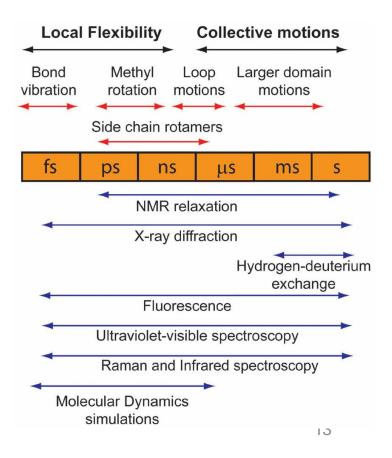
Molecular Dynamics

Starting the simulation, velocities from the Boltzmann distribution @ 300K randomly allocated Then for each time step:

- Compute forces on each particle
- Solve Newton's 2nd Law of motion for each atom to calculate the new coordinates and new velocity for each atom position
- Repeat for a specified number of steps

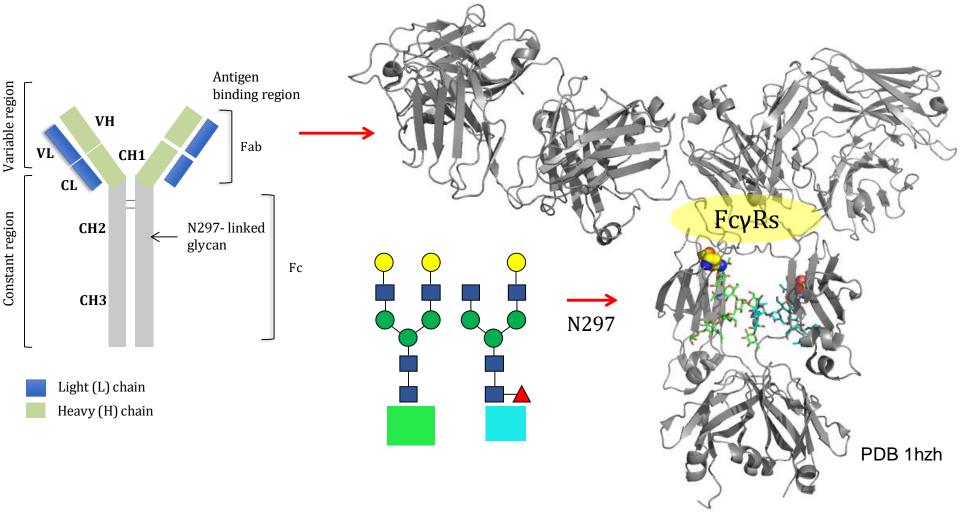
A stepwise numerical integration method is used to solve the equation Usually a time step of 1 or 2fs is used, which gives a reasonable approximation to the solution





IgG1 Fc N-glycosylation





N-Glycosylation of IgG1 @N297

94% of the glycoforms in Fc are fucosylated

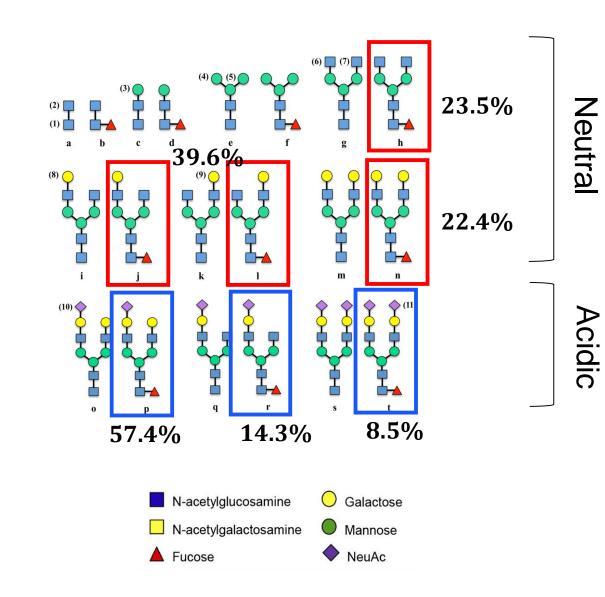
Raju *et al. Glycobiology* (2000), **10**, 477-486

Gal and Sia levels are lower in RA patients

Pucic et al, Mol. Cell Proteomics (2011)

Core-Fuc reduces ADCC

Parekh *et al*, Nature (1985), **316**: 452-457



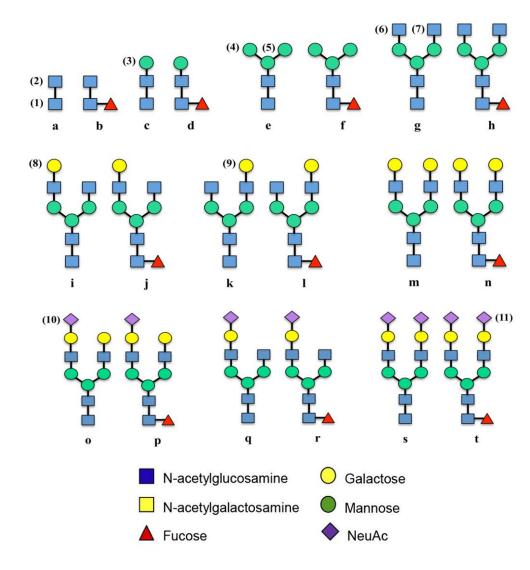
MD analysis of N-glycan series

Structures built with glycam-WEB (glycam.org)

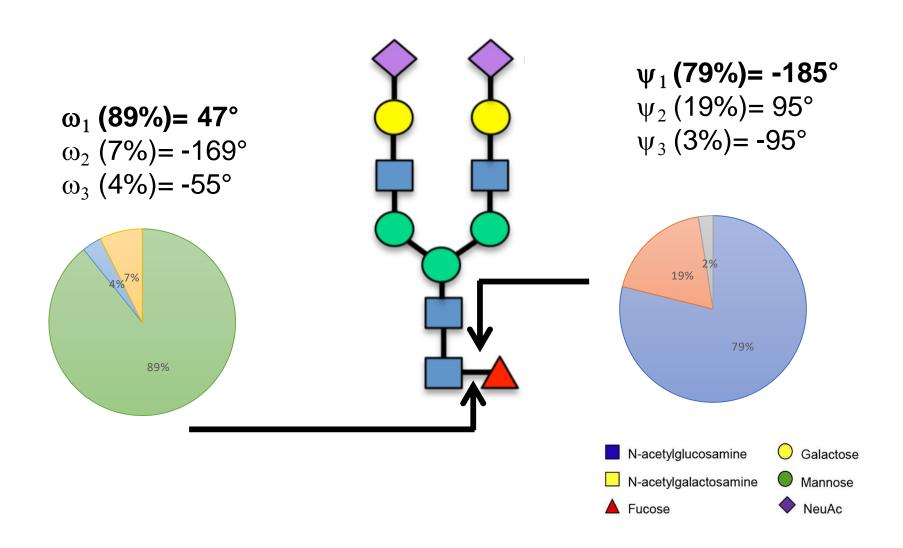
Forcefields:

Glycam06h-12SB/TIP3P AMBER v.12/16

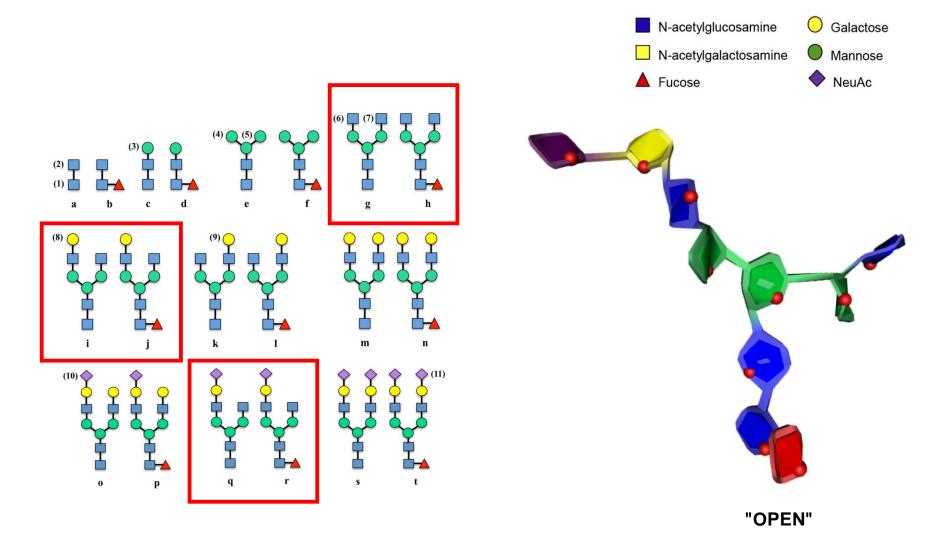
62 μs of cumulative simulation time 1,785,600 CPU hours on Fionn (2 nodes, 24ppn, 60 hrs per 100ns)



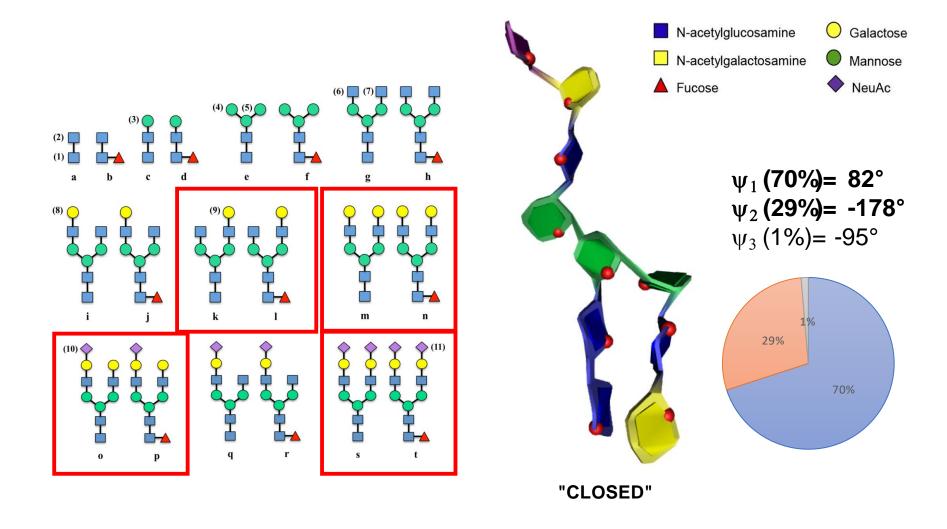
Conformational Propensities: Fucose



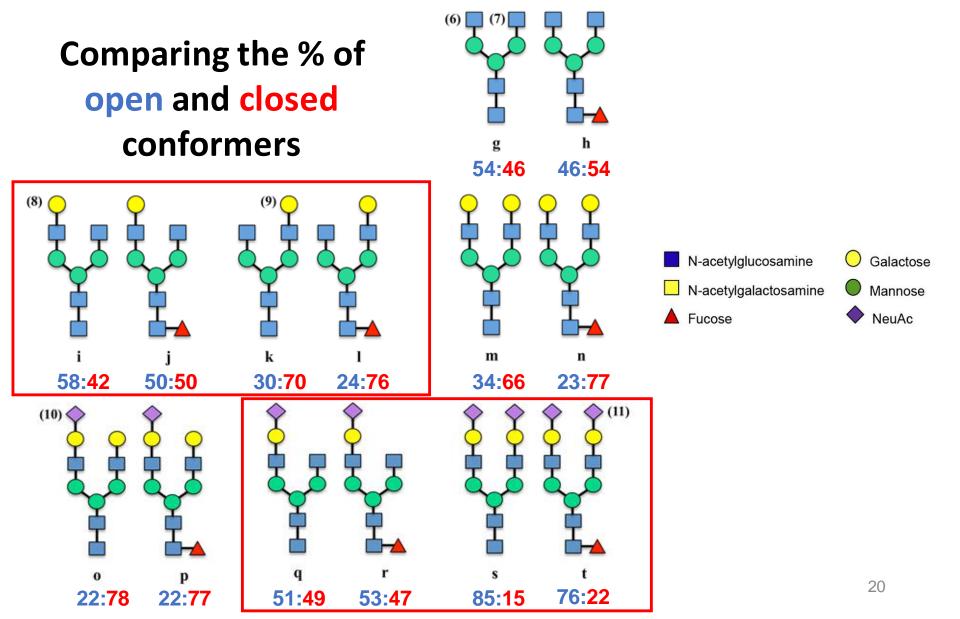
Conformational Propensities: (1-6) Arm



Conformational Propensities: (1-6) Arm



Conformational Propensities: (1-6) Arm



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Sequence-to-structure dependence of isolated IgG Fc complex biantennary N-glycans: a molecular dynamics study

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Abstract

Fc glycosylation of human immunoglobulins G (IgGs) is essential for their structural integrity and activity. Interestingly, the specific nature of the Fc glycoforms is known to modulate the IgG effector function and inflammatory properties. Indeed, while core-fucosylation of IgG Fc-glycans greatly affects the antibody-dependent cell-mediated cytotoxicity function, with obvious repercussions in the design of therapeutic antibodies, sialylation can reverse the antibody inflammatory response, and galactosylation levels have been linked to aging, to the onset of inflammation, and to the predisposition to rheumatoid arthritis. Within the framework of a structure-to-function relationship, we have studied the role of the N-glycan sequence on its intrinsic conformational propensity. Here we report the results of a systematic study, based on extensive molecular dynamics simulations in excess of 62 us of cumulative simulation time, on the effect of sequence on the structure and dynamics of increasingly larger, complex biantennary N-glycoforms isolated from the protein, i.e. from chitobiose to the larger N-glycan species commonly found in the Fc region of human IgGs. Our results show that while core fucosylation and sialylation do not affect the intrinsic dynamics of the unlinked N-glycans, galactosylation of the $\alpha(1-6)$ arm shifts dramatically its conformational equilibrium from an outstretched to a folded conformation. These findings are in agreement with and can help rationalize recent experimental evidence showing a differential recognition of positional isomers in glycan array data and also the preference of sialyltransferase for the more accessible, outstretched a(1-3) arm in both isolated, and Fc-bound N-glycans,

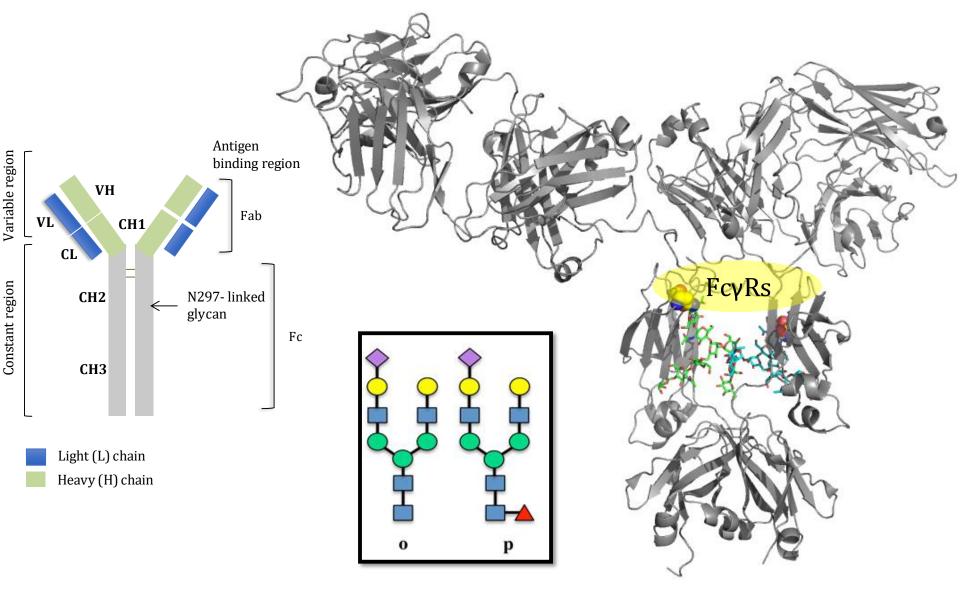
Key words: Fc-gly cosylation, gly coinformatics, IgG, molecular dynamics, N-glycans

Introduction

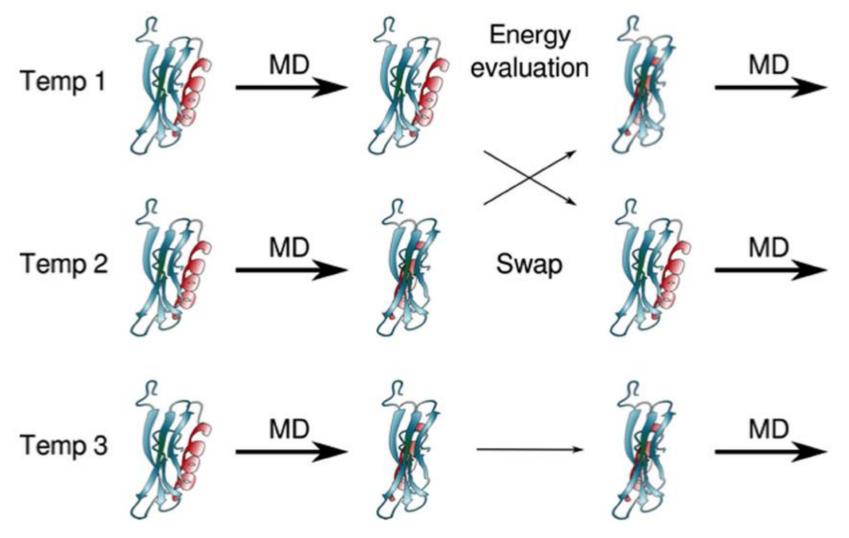
N-glycosylation of the immunoglobulin G (IgG) fragment crystallizable (Fc) region is essential for its structural stability and function (Krapp, Mimura, et al. 2003; Arnold, Wormald, et al. 2007; Kobata 2008; Fang, Richardson, et al. 2016). The sequence and branching of the Fc N-glycoforms, bound at the highly conserved Asn 297 in both CH3 domains of the Fc region, strongly affect the antibodymediated effector function (Raju 2008; Subedi and Barb 2016; Hayes, Frostell, et al. 2017) by modulating the binding to the immune cells' Fc receptors, thus the antibody-mediated immune response (Tao and Morrison 1989; Shields, Namenuk, et al. 2001). In this context the effects of core-fucosylation, sialylation and galactosylation are particularly interesting. Between 81 and 98.7% of the Fc N-glycans in human IgGs are core-fucosylated (Pucic, Knezevic, et al. 2011). Core fucosylation, where fucose is $\alpha(1-6)$ linked to the chitobiose core, greatly affects the IgG antibody-dependent cell-mediated cytotoxicity (ADCC) function. More specifically, a strongly enhanced ADCC corresponds to nonfucosylated Fc N-glycan species (Satoh, Iida, et al. 2006; Kanda, Yamada, et al. 2007; Matsumiya, Yamaguchi, et al. 2007; Strohl 2009; Ratner

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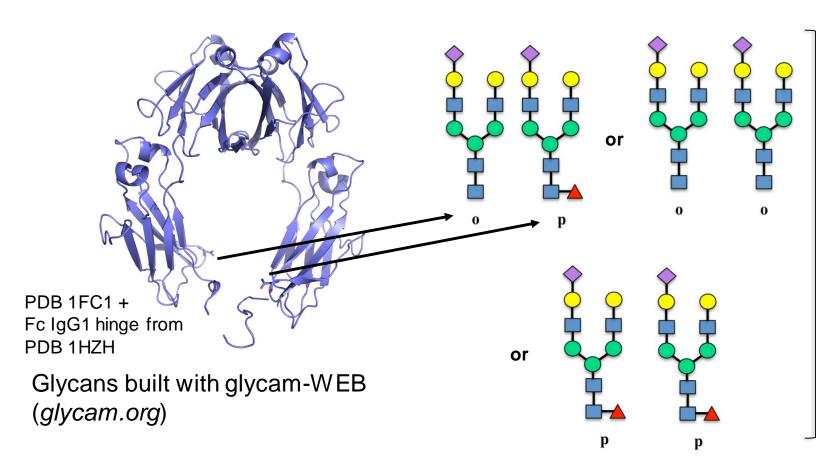
IgG1 Fc N-glycosylation



Replica Exchange MD



IgG1 Fc + N-glycans simulations

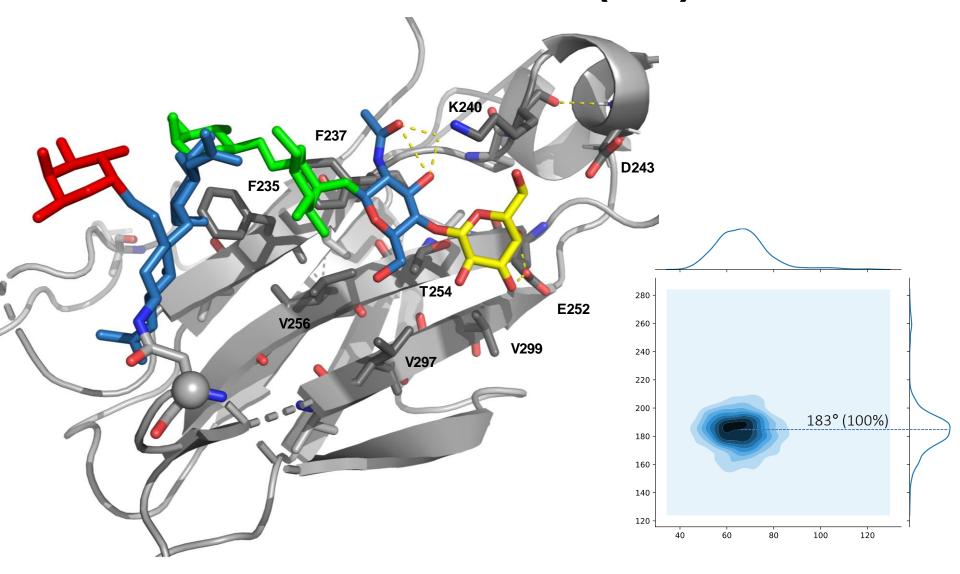


"OPEN" &
"CLOSED"
Starting
conformation

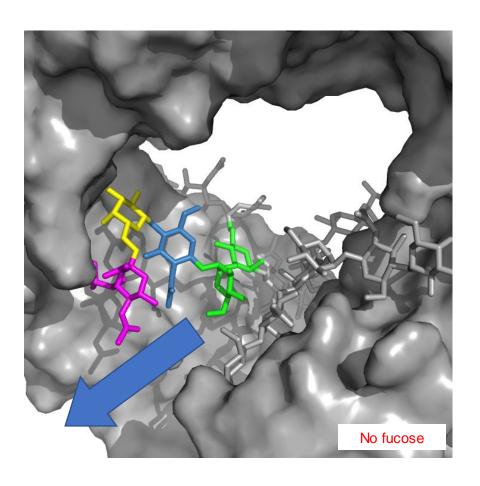
MD simulation

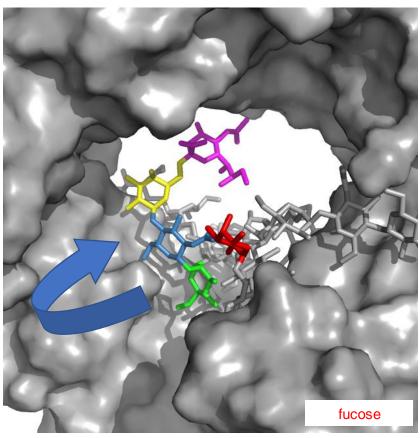
Glycam06j-1/TIP3P AMBER AMBER12SB with NAMD 2.31b1 Generated REMD: 17ns * 90 replicas for each starting structure

Restriction of the $\alpha(1-6)$ arm

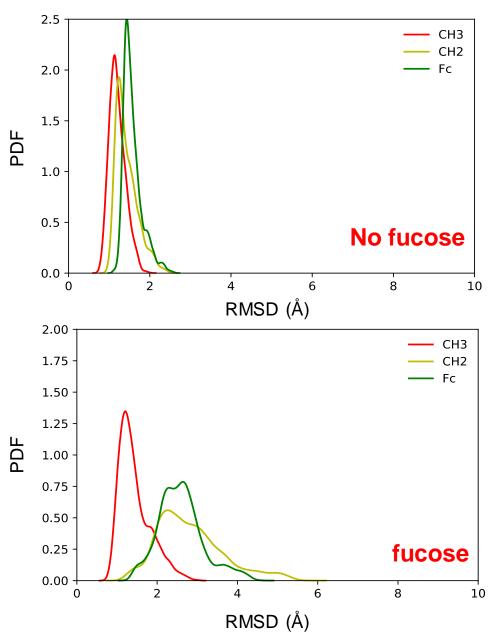


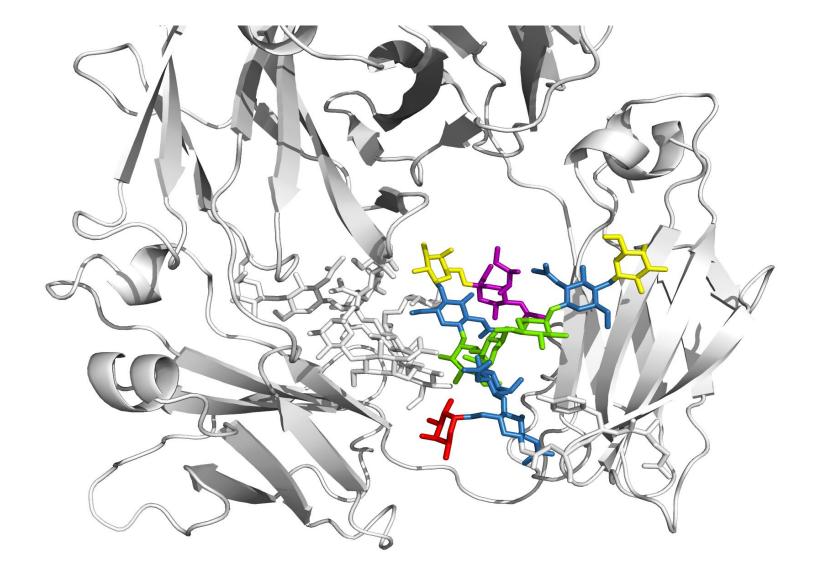
Core fucose regulates the $\alpha(1-3)$ arm





Dynamics of the CH2 domain (protein)









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The immunoglobulin type G (IgG) Fc N-glycans are known to modulate the interaction with membrane-bound Fc y receptors (FcyRs), fine-tuning the antibody's effector function in a sequence-dependent manner. Particularly interesting in this respect are the roles of galactosylation, which levels are linked to autoimmune conditions and aging, of core fucosylation, which is known to reduce significantly the antibody-dependent cellular cytotoxicity (ADCC), and of sialylation, which also reduces ADCC but only in the context of core-fucosylation. In this work we provide an atomistic level perspective through enhanced sampling computer simulations, based on replica

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Thank you for your attention!