Brighton Skeel and Myron Liu Lab Section 113 17 March 2016

# Ligand/Solvent Pair Efficacy in Aqueous Heavy Metal Complex Removal from Mixed Matrices

### Goal

This experiment aims to evaluate the efficacy of various organic solvent/phase transfer agent pairs in removing heavy metals from various aqueous matrices. To maintain a real world applicability, this experiment will use matrices representing the following matrices: acidified, seawater, distilled water, and hard water.

### Introduction

Since the dawn of the industrial age, the human race has developed a particular dependence upon metals-both rare and common. As these metal resources deplete, we turn to sources of lessening quality, many of which contain unwanted and toxic by-products such as chromium, cadmium, nickel, lead, cobalt, copper, and zinc[1]. Though care is taken to prevent these substances from infiltrating the environment, accidents do happen (think of the recent flint water crisis or recent mining accidents in South America). In absence of adequate preventative methods, we are forced to consider retroactive methods. One such method, explored in this experiment, is the use of phase transfer agents and organic solvents to complex heavy metals and physically extract them from contaminated water. Central to this process is the tendency of transition metals to preferentially form bonds with certain molecules (ligands) over water. These metal-ligand complexes have, in certain cases, properties analogous to those of micell structures—that is, they are much more soluble in non-polar solvents, and thus easily extracted from water. Exploiting these properties carefully, we plan to complex heavy metals under various conditions, with the expectation that more complex solutions will degrade heavy metal percentage elimination due to competitive effects (the heavy metals will, in many cases, have to compete with other, substantially less toxic cations such as sodium and magnesium).

Skeel, Liu 2

**Experimental Procedure** 

Week One:

Matrix: Distilled Water

Solvent Assignments: Brighton-Tetrachloroethylene; Myron-Hexanes

Both members will begin by going to ISF to obtain all glassware, equipment, and chemicals to be used throughout the day. Both will then proceed by acid washing the glassware they plan to use throughout the course of the day. For the first day, this also includes the 500m volumetric flask to be used for the preparation of the master heavy metal matrix solution. An outline of the procedure for acid washing is outlined below, as

taken from the Chemistry 4B lab manual [2].

1. Rinse with a small ( $\approx 3mL$ ) amount of 50%  $HNO_3$ 

2. Rinse three times with tap water

3. Rinse with a small ( $\approx 3mL$ ) amount of 10% HCl

4. Rinse three times with tap water

5. Rinse two times with distilled water

Following an acid wash of all glassware, the master solution will be prepared. This experiment is based on the premise of ppm metal concentrations, so metal salts will be added as such. To achieve appropriate metal concentrations (400-500ppm) for eventual dilution, we will add  $0.2250(\pm 0.0250)g$  of each of the following salts to the flask, mixing thoroughly:

 $Cadmium \ Chloride : CdCl_2$ 

Cobalt (II) Chloride :  $CoCl_2$ 

Chromium (III) Chloride:  $CrCl_3$ 

Copper (II) Chloride :  $CuCl_2$ 

 $Lead (II) Chloride : PbCl_2$ 

 $Zinc (II) Chloride : ZnCl_2$ 

This solution will be sampled according to what, from here onward, we will refer to as the AES sample preparation procedure. The procedure is outlined below, and has been optimized to reduce fluid waste:

- 1. Sample 1mL of the matrix with a volumetric pipet
- 2. Dilute this sample in a 10mL volumetric flask to volume with 0.2% trace metal grade  $HNO_3$
- 3. Dilute 5mL of this solution to 50mL in a 50mL volumetric flask using 0.2% trace metal grade  $HNO_3$ . (Note that this results in a net dilution factor of  $100\times$ , which is an acceptable level for MP-AES analysis[2])
- 4. Filter an aliquot of this dilute sample through a regenerated cellulose membrane and into a 50mL falcon tube for processing with MP-AES
- 5. Label the falcon tube with the sample name, date, and any other relevant information.

After sampling the master heavy metal matrix, pH and temperature data for the solution will be recorded. Following these measurements a  $\approx 100mL$  aliquot will be drawn off and split into two separate beakers. These beakers will be appropriately labeled, and will be used by each group member (I.e., each individual will receive one of these beakers). Remaining matrix solution will be stored in two 250mL plastic bottles (note the use of plastic bottles for long term storage—this mitigates error inherent from ion adsorption onto glass surfaces).

Following matrix partitioning so as to supply solution to each member, the complexion portion of the experiment will be carried out. Each member begins by placing exactly 5mL of their organic solvent into a 15mL plastic falcon tube (centrifuge style). Safety Note: As the fumes from organic solvents, particularly the tetrachloroethylene used in this experiment, are harmful, all solvents, when not in use, will be stored in sealed glass bottles. Falcon tubes containing solvent must be closed after delivery of solvent, so as to prevent excessive fuming. After addition of solvent, the complexing agent will be dissolved in the organic phase. So as to obtain meaningful data, the complexing agent will be added in sub-stoichiometric quantities (so as to elucidate phase transfer agent affinity to certain

metals). Regarding this step, we recognize that the three complexion agents used in this experiment tend to form differing numbers of bonds to metal atoms, as tabulated below.

Dithizone[4] (dithiocarbazone): Bidentate, 2 mol per complex (tetrahedral complexion)

Bipy (2,2'-bipyridine) : Bidentate, 3 mol per complex (octahedral complexion)

Dibenzo 24-crown-8[5] : Polydentate[3], 1-2 mol per complex (cation-size dependent)

These complexion agents have been chosen based off of empirically determined ability to form metal complexes (as is the case with dithizone and the crown ether) or based off of reasonable suspicion that they may form non-polar complexes (as is the case with bipy). Citations adjacent the complexion agents refer to literature examples of their use as effective agents.

As we know the concentration of heavy metal ions in solution (assuming the use of 0.2250g/500mL for a total metal concentration of  $2.7 \times 10^{-3} \frac{g}{mL}$ ), we compute that the total cation molarity must be  $1.7_0 \times 10^{-5} \frac{mol}{mL}$ . Using these data (assuming the crown ether will coordinate with, on average,  $\frac{2}{3}$  of a cation)[3], as well as the (arbitrarily assigned) sub-stoichiometric quantities of complexion agent to be 25%, 50%, and 75%, we arrive at the following mass values for complexion agent addition to each solution (understanding the difficulty of measuring on this scale, exact values used will be recorded):

	Dithizone	Bipy	24-crown-8
25%	10.88mg	9.96mg	14.30mg
50%	21.76mg	19.91mg	28.59mg
75%	32.64mg	29.87mg	42.89mg

After adding these complexion agents, the falcon tubes will be shaken to mix the solvent and complexion agent. After  $\approx 5$  minutes of mixing, 5mL of metal solution will be added to each of the sample vials. These will, again, be mixed, though this time more gently so as to prevent the formation of an emulsion, which could jeopardize the experiment. At this point, excess organic solvent will be drawn off and disposed of properly. There should be no organic solvent remaining, so removal of some sample stock is acceptable, so long as at least 1mL of sample solution remains. This sample solution will be prepared as described in the previous section (see AES preparation technique). By the last hour of lab period,

each group member will have prepared nine samples for submission to ISF. These samples will be evaluated for heavy metal content only (in this case, cobalt, copper, chromium, cadmium, lead, and zinc). The last half-hour of class will be devoted to cleanup and effectively disposing of waste, as well as preparing for the next laboratory session.

Week Two:

Matrix: Hard Water

Solvent Assignments: Brighton-Tetrachloroethylene; Myron-Hexanes

Note that for all weeks (week 5 excepted) the in-lab procedure is invariant. That is, we are performing the same experiment, only using a matrix modifier acting on the standard heavy metal matrix prepared in the first week. During week two, we will turn our focus toward a hard water[4] matrix (one containing the carbonate salts of magnesium and calcium). These substances, being unpleasant to dissolve, will be added to a 125mL aliquot of the metal matrix prepared in week one in  $11.3(\pm 4._3)mg$  quantities (we are solubility limited, speaking on a level of ppm count, this is  $\frac{1}{5}$  the normal concentration). The salts will be added to the solution and mixed to equilibrium via stir plate. From here, the same procedure used in the last week will be implemented. The quantity of the complexing agent will remain the same, as we wish to hold the heavy metal to phase transfer agent constant so as to reduce the number of experimental variables. The 50mL falcon tubes used in this portion of the experiment will be reused (but washed thoroughly) from the previous week.

Week Three:

Matrix: Seawater

Solvent Assignments: Brighton-Hexanes; Myron-Tetrachloroethylene

This week is identical to the previous week, the only difference being that we will be using  $56.3(\pm 13)mg$  of each of the following matrix modifiers:  $NaNO_3$ ,  $KNO_3$ ,  $Fe(NO_3)_3$ . This will again bring us to a roughly 400 - 500ppm working concentration from which we can investigate complexing agent binding tendency.

Week Four:

Matrix: Acidified water (modeling acid rain)

 $\textbf{Solvent Assignments:} \ \, \textbf{Brighton-} \textit{Hexanes}; \ \, \textbf{Myron-} \textit{Tetrachloroethylene}$ 

For the fourth week we will model the effects of acid rain on the heavy metal extraction

process using sulfuric and nitric acids (note that this is different, as hydrogen will not act as a standard cation, so to speak). As adding acid will require a dilution, this experiment will utilize a different set of volumes (the exact numbers are pending notification of the acid strength available).

### Week Five:

Week five will be the only week where we do not perform physical experiments. Rather, the scope of the lab time throughout the fifth week will be to utilize computational methods (namely Q-chem software[6]) to determine the physical properties of the solvents and complexing agents so as to attempt to derive a correlation between physical properties and empirical data. Dependent upon computational time needed, this lab period will also be focused on data processing, as we will be able to determine numerous values, including coordination numbers, relative stabilities, and a whole host of other potentially useful data which we will need to carefully scrutinize. Numerical methods utilized will be based off of the guidance of former labs in conjunction with literature searches[7][8].

### Instrumentation and Specialty Equipment

Regarding instrumentation for this experiment, we require the use of only one advanced instrument: the MP-AES unit. As the crux of our experiment is dependent upon the quantification of atomic composition in aqueous solution, we plan to submit samples for this kind of processing at a rate of 18 per day (submitted on March  $30^{th}$ , April  $7^{th}$ ,  $14^{th}$ , and  $21^{st}$ ). Though this seems a large number of samples to run, the procedure has been approved by the staff at ISF[9].

### Safety Issues

Regarding the safety issues inherent to the process outlined above, the primary issues fall into two categories: solvent associated and salt associated. As we will be working with organic solvents, we must note the toxicity of their fumes, as both solvents used (tetrachloroethylene and hexanes) are volatile. The hexanes bear an NFPA health hazard rating of two and the tetrachloroethylene a safety rating of three. To mitigate the hazard associated with these solvents, we will store them in capped bottles, only opening them

to remove *small* amounts of solvent. Regarding the hazards presented by the salts we will be using, we note that we aim to recreate an aqueous matrix roughly analogous to one contaminated with heavy metals. As many heavy metals are nephro/neurotoxic and/or carcinogenic, we limit the risk of others by using only small amounts of the salts ( $\approx 0.250g$ )

## References

- [1] Doong, R.; Lee, S.; Sun, Y.; Wu, S. Marine Pollution Bulletin 2008, 57, 846-857
- [2] Chemistry Lab Development,. Quantitative Analysis of a Solution Containing Co and Cu, Part 1; University of California, Berkeley, 2016.
- [3] Sundberg, L., Crown Ethers: Applications in Inorganic Synthesis, 1978 Senior Scholar Papers. Paper 232
- [4] Silverman, L.; Trego, K. Determination Of Small Amounts Of Cadmium In Lead By A One-Colour Dithizone Method. *The Analyst* 1952, 77, 143.
- [5] Servaes, K.; De Houwer, S.; Görller-Warland, C.; Binnemans, K. Spectroscopic Properties Of Uranyl Crown Ether Complexes in Non-Aqueous Solvents. *Phys. Chem. Chem. Phys.* 2004, 6, 2946-2950
- [6] Chemistry Lab Development,. Using Computation and Experimentation to Model Aquatic Toxicity, Part 1 and 2; University of California, Berkeley, 2015.
- [7] Brubaker, G.; Johnson, D. Molecular Mechanics Calculations In Coordination Chemistry. *Coordination Chemistry Reviews* 1984, 53, 1-36.
- [8] Allinger, N.; Zhou, X.; Bergsma, J. Molecular Mechanics Parameters. *Journal of Molecular Structure: THEOCHEM* 1994, 312, 69-83.
- [9] Ly, P. 2016