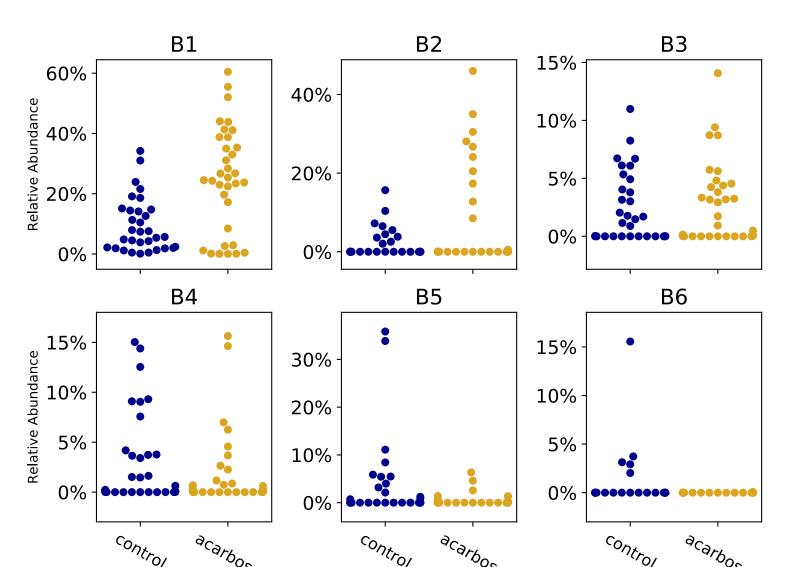
Byron J. Smith<sup>1</sup>, Thomas M. Schmidt<sup>2</sup>

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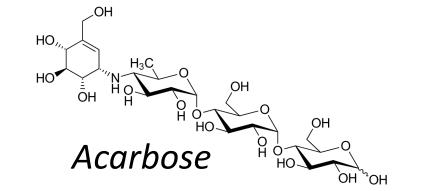
## Introduction

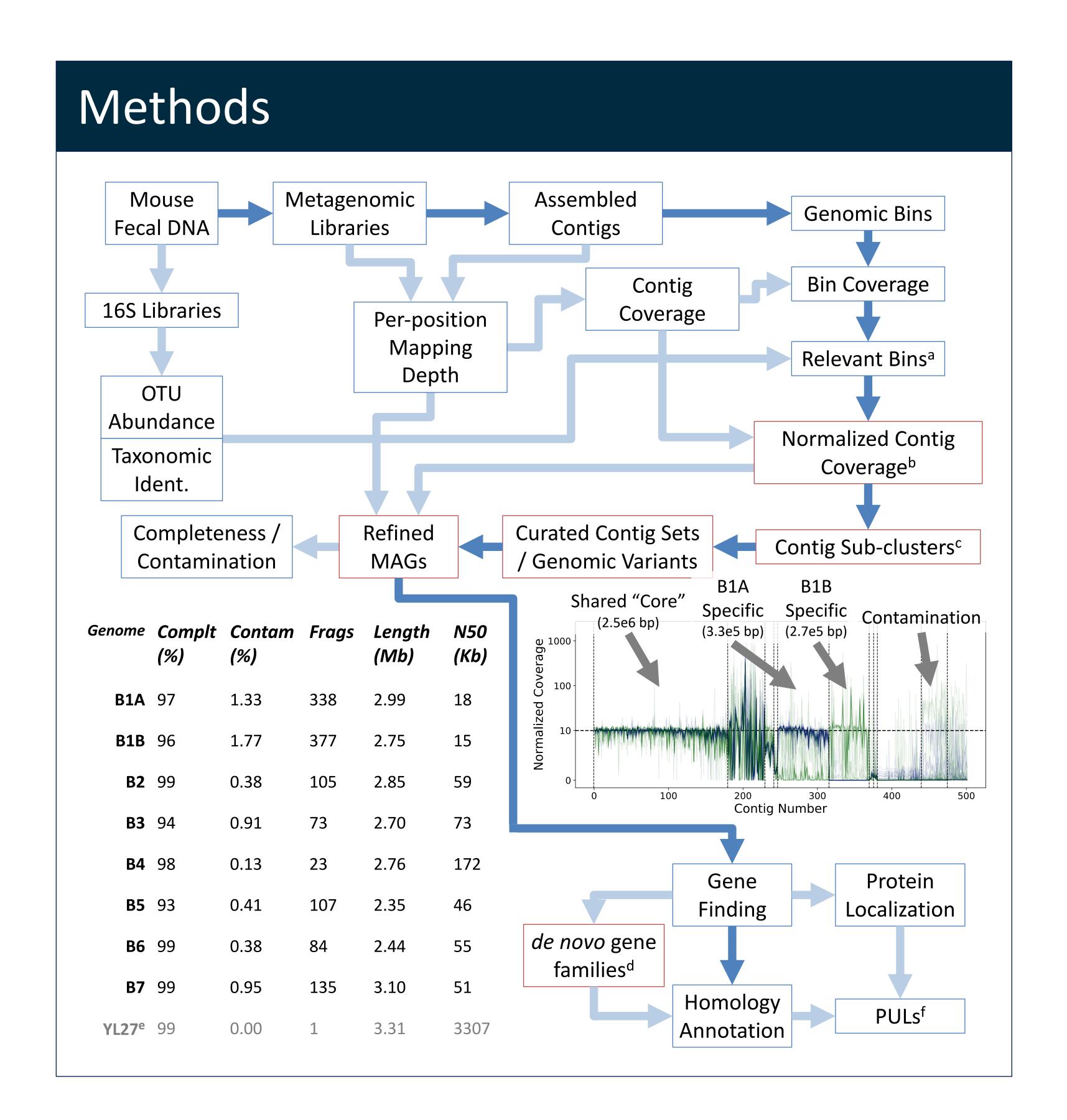
Mice treated with the  $\alpha$ -amylase and  $\alpha$ -glucosidase inhibitor acarbose live longer<sup>1</sup> and have notably different fecal microbial communities and short-chain fatty acid concentrations<sup>2</sup>, presumably due to increased starch entering the lower digestive system. The two bacterial taxa most dramatically enriched by this treatment are in the recently described family

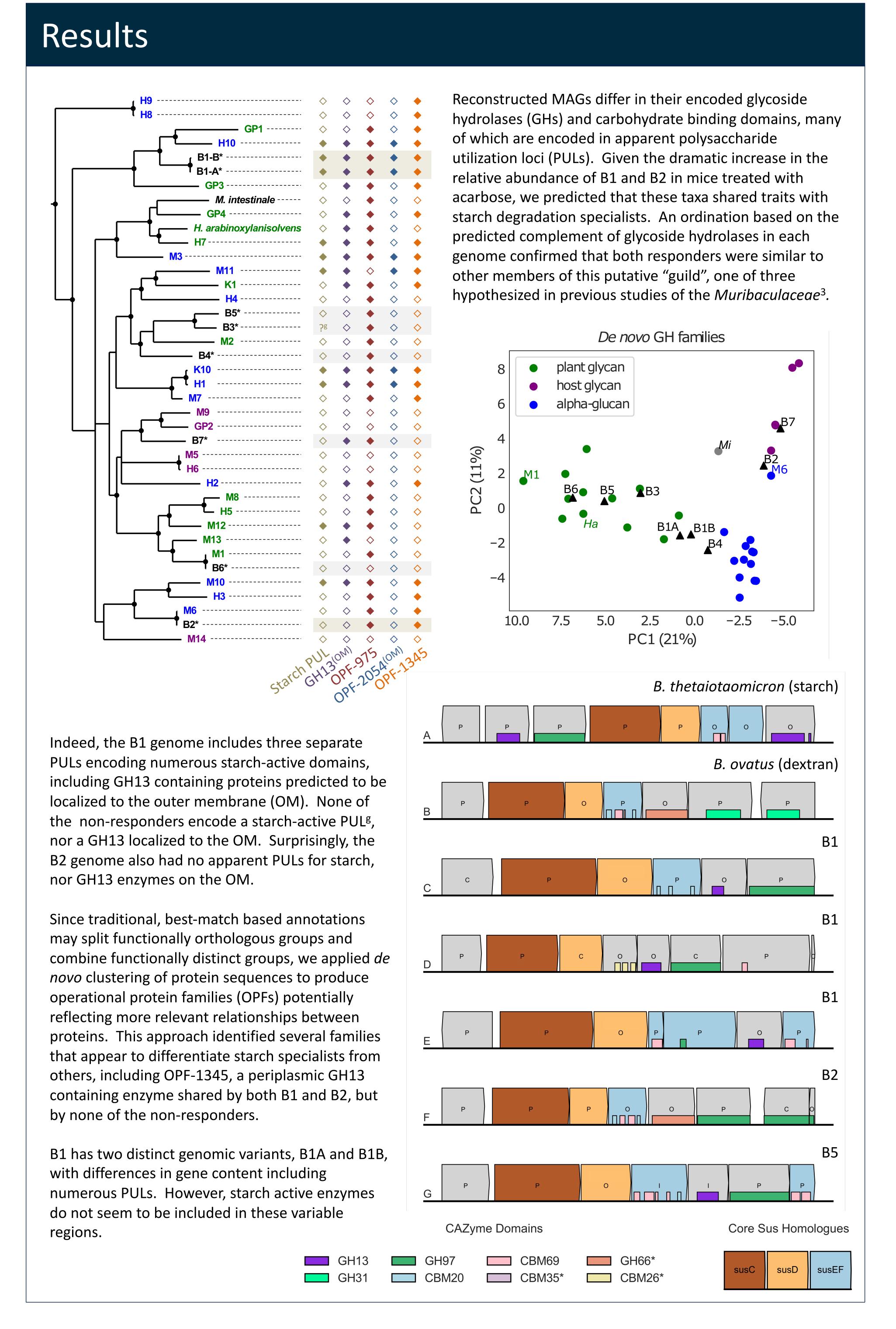


Muribaculaceae (also called: S24-7 or Homeothermaceae). However, other members of this clade decrease in abundance or do not respond. Previous studies of this bacterial family suggest that members specialize on a diversity of fibers including starch, plant polysaccharides, and glycans derived from their animal hosts<sup>3,4</sup>. However, these explorations have been limited by the availability of isolates, or have studied genomes assembled from metagenomes (MAGs) in the absence of experimental results. Here we compare 7

novel MAGs and find genomic features that may explain differences between *Muribaculaceae* that differentiate acarbose "responders", here referred to as B1 and B2, from non-responders, B3-B7.







## Conclusions

- We have reconstructed **eight high-quality Muribaculaceae genomes** from
  metagenomes.
- Members of this family are taxonomically and functionally diverse in the mouse gut microbiome, and may specialize in the degradation of complex carbohydrates.
- Differential response to acarbose treatment reflects variation in encoded polysaccharide utilization loci and glycoside hydrolases.
- The most abundant taxon, B1, is composed of two distinct genomic variants, differentiated by more than 600 protein coding genes.
- High-quality MAGs can be recovered from microbial communities currently underrepresented in strain collections, and provide a powerful platform for ecological discovery.

## Footnotes and Citations

• <sup>a</sup> Bins were matched to OTUs based on a partial least squares cross-decomposition of coverage and relative abundance. • <sup>b</sup> Contig coverage was normalized to the mean coverage of contigs confidently identified as part of the core genome based on manual inspection. • <sup>c</sup> Contigs were clustered based on normalized coverage across samples, effectively partitioning between contigs in the core genome, in particular genomic variants, or in other, "contaminating" genomes. • <sup>d</sup> *De novo* gene families were assigned based on an all-by-all protein BLAST using the MCL algorithm. • <sup>e</sup> *Muribaculum intestinale* YL27 • <sup>f</sup> PULs were defined as a *susC* homolog within 5000 bases of, on the same strand as, and upstream of a *susD* homolog. • <sup>g</sup> While B3 does encode a possible starch-active PUL, the GH13 lipoprotein is predicted to localize to the inner membrane, rather than the OM. • <sup>1</sup> Harrison, D. E. *et al.* (2014). *Aging Cell, 13*(2), 273–282. doi:10.1111/acel.12170 • <sup>2</sup> Smith, B. J. *et al.* (2019). *BMC Microbiology, 19*(130), 16. doi:10.1186/s12866-019-1494-7 • <sup>3</sup> Ormerod, K. L. *et al.* (2016). *Microbiome, 4*(1), 36. doi:10.1186/s40168-016-0181-2 • <sup>4</sup> Lagkouvardos, I. *et al.* (2019). *Microbiome, 7*(1), 1–15. doi:10.1186/s40168-019-0637-2

## Acknowledgments & Contact

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