

# **SNPiO: A Python API for Population Genetic File Processing, Filtering, and Encoding**

## **Getting Started with SNPiO**

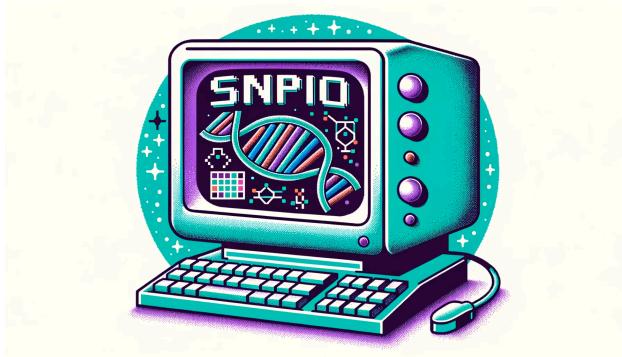
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## Getting Started with SNPiO

This guide provides an overview of how to get started with the SNPiO library. It covers the basic steps to read, manipulate, and analyze genotype data using the VCFReader, PhylipReader, StructureReader, and NRemover2 classes. SNPiO is designed to simplify the process of handling genotype data and preparing it for downstream analysis, such as population genetics, phylogenetics, and machine learning. The library supports various file formats, including VCF, PHYLIP, and STRUCTURE, and provides tools for filtering, encoding, and visualizing genotype data. This guide will help you get up and running with SNPiO quickly and efficiently.

VCFReader, PhylipReader, and StructureReader classes are used to read genotype data from VCF, PHYLIP, and STRUCTURE files, respectively. These classes load the data into a GenotypeData object that has various useful methods and properties.

The `NRemover2` class is used to filter genotype data based on various criteria, such as missing data, minor allele count, minor allele frequency, and more. The `GenotypeEncoder` class is used to encode genotype data into different formats, such as one-hot encoding, integer encoding, and 0-1-2 encoding, for downstream analysis and machine learning tasks.

The `Plotting` class provides methods to visualize genotype data, such as running principal component analysis (PCA) and generating missing data reports. The `PopGenStatistics` class is used to perform population genetic analyses on SNP datasets, such as D-statistics, Fst outliers, heterozygosity, nucleotide diversity, and Analysis of Molecular Variance (AMOVA).

The `TreeParser` class is used to load and parse phylogenetic trees in Newick and NEXUS formats. It can read and parse tree files, modify tree structures, draw trees, and save trees in different formats.

The `PopGenStatistics` class is designed to perform a suite of population genetic analyses on SNP datasets. It supports calculations such as D-statistics, Fst outliers, heterozygosity, nucleotide diversity, and Analysis of Molecular Variance (AMOVA). These analyses are essential for understanding genetic structure, diversity, and differentiation within and between populations.

Below is a step-by-step guide to using SNPiO to read, filter, encode genotype data for analysis, and calculate population genetic statistics. The guide covers the basic steps to get started with SNPiO and provides examples of how to use the main classes and methods in the library.

## Installation

Before using SNPiO, ensure it is installed in your Python environment. You can install it using pip. In the project root directory (the directory containing `pyproject.toml`), type the below command into your terminal.

### Tip

We recommend using a virtual environment to manage your Python packages. If you do not have a virtual environment set up, you can create one using the following command and then activate it and install SNPiO:

```
python3 -m venv snpio_env  
source snpio_env/bin/activate  
pip install snpio
```

This will create a virtual environment named `snpio_env` and activate it. You can then install SNPiO in this virtual environment using the pip command mentioned above.

### Note

SNPiO does not support Windows operating systems at the moment. We recommend using a Unix-based operating system such as Linux or MacOS. If you have Windows, you can use the Windows Subsystem for Linux (WSL) to run SNPiO, which runs a Linux distribution on Windows.

### Note

We aim to support anaconda environments in the future. For now, we recommend using a virtual environment with pip to install SNPio.

## Importing SNPio

To start using SNPio, import the necessary modules:

```
# Import the SNPio modules.  
from snpio import NRemover2, VCFReader, PhylipReader, StructureReader,  
Plotting, GenotypeEncoder, PopGenStatistics, TreeParser
```

Example usage:

```
# Define input filenames  
vcf = "snpio/example_data/vcf_files/phylogen_subset14K_sorted.vcf.gz"  
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"  
  
# Load the genotype data from a VCF file  
gd = VCFReader(filename=vcf, popmapfile=popmap, force_popmap=True, verbose=  
True, plot_format="png", plot_fontsize=20, plot_dpi=300, despine=True,  
prefix="snpio_example")
```

You can also include or exclude any populations from the analysis by using the `include_pops` and `exclude_pops` parameters in the reader classes. For example:

```
# Only include the populations "ON", "DS", "EA", "GU", and "TT"  
# Exclude the populations "MX", "YU", and "CH"  
gd = VCFReader(filename=vcf, popmapfile=popmap, force_popmap=True, verbose=  
True, plot_format="png", plot_fontsize=20, plot_dpi=300, despine=True,  
prefix="snpio_example", include_pops=["ON", "DS", "EA", "GU"],  
exclude_pops=["MX", "YU", "CH", "OG"])
```

The `include_pops` and `exclude_pops` parameters are optional and can be used to filter the populations included in the analysis. If both parameters are provided, the populations in `include_pops` will be included, and the populations in `exclude_pops` will be excluded. However, populations cannot overlap between lists.

### Note

If you provide both parameters, the populations in `include_pops` will take precedence.

### Note

The `VCFReader`, `PhylipReader`, `StructureReader`, `NRemover2`, `PopGenStatistics`, and `GenotypeEncoder` classes treat the following characters as missing data: “N”, “.”, “?” , “-” .

### Note

The VCFReader class can read both uncompressed and compressed VCF files (gzipped). If your input file is in PHYLIP or STRUCTURE format, it will be forced to be biallelic. To handle more than two alleles per site, use the VCF format. However, also note that many of the analyses implemented in PopGenStatistics and NRemover2 are designed for biallelic and diploid data.

## The Population Map File

To use VCFReader, PhylipReader, or StructureReader, you can optionally use a population map (popmap) file. This is a simple two-column, whitespace-delimited or comma-delimited file with SampleIDs in the first column and the corresponding PopulationIDs in the second column. It can optionally contain a header line, with the first column labeled “SampleID” and the second column labeled “PopulationID” (case-insensitive). The population IDs can be any string, such as “Population1”, “Population2”, etc, or an integer. SampleIDs must match the sample names in the alignment file.

For example:

```
Sample1,Population1  
Sample2,Population1  
Sample3,Population2  
Sample4,Population2
```

Or, with a header:

```
SampleID,PopulationID  
Sample1,Population1  
Sample2,Population1  
Sample3,Population2  
Sample4,Population2
```

The population map file is used to assign samples to populations and is useful for filtering and visualizing genotype data by population. If you do not provide a population map file, the samples will be treated as a single population.

The population map file can be provided as an argument to the reader classes. For example:

```
vcf = "snpio/example_data/vcf_files/phylogen_subset14K_sorted.vcf.gz"  
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"  
  
gd = VCFReader(filename=vcf, popmapfile=popmap, force_popmap=True, verbose=  
    True, plot_format="png", plot_fontsize=20, plot_dpi=300, despine=True,  
    prefix="snpio_example")
```

### Note

The `force_popmap` parameter in the reader classes is used to force the population map file to align with the samples in the alignment without an error. If set to `False`, the population map file must match the samples in the alignment exactly, and if they do not match, an error will be raised. If set to `True`, the population map file will be forced to align with the samples in the alignment by removing extra samples, and vice versa. This parameter is set to `False` by default.

The `verbose` parameter in the reader classes is used to print additional information about the genotype data and filtering steps. If set to `True`, the reader classes will print information about the genotype data, such as the number of samples, loci, and populations, and the filtering steps applied. This parameter is set to `False` by default.

The `plot_format`, `plot_fontsize`, `plot_dpi`, and `despine` parameters in the reader classes are used to customize the output plots generated by the reader classes. See API documentation for more details.

## Reading Genotype Data

SNPiO provides readers for different file formats. Here are examples of how to read genotype data from various file formats: VCF, PHYLIP, and STRUCTURE.

### VCFReader

```
vcf = "snpio/example_data/vcf_files/phylogen_subset14K_sorted.vcf.gz"
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"

gd = VCFReader(filename=vcf, popmapfile=popmap, force_popmap=True, verbose=
    True, plot_format="png", plot_fontsize=20, plot_dpi=300, despine=True,
    prefix="snpio_example", exclude_pops=["MX", "YU", "CH"], include_pops=[
        "ON", "DS", "EA", "GU", "TT"])
```

This will read the genotype data from a VCF file and apply the population map if provided.

### PhylipReader

If you would like to read a Phylip file, you can use the `PhylipReader` class:

```
phylip = "snpio/example_data/phylip_files/phylogen_subset14K.phy"
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"

gd = PhylipReader(filename=phylip, popmapfile=popmap, force_popmap=True,
    verbose=True, plot_format="png", plot_fontsize=20, plot_dpi=300, despine=
    True, prefix="snpio_example", exclude_pops=["MX", "YU", "CH"],
    include_pops=[ "ON", "DS", "EA", "GU", "TT" ])
```

This will read the genotype data from a PHYLIP file and apply the population map (if provided).

## StructureReader

If you would like to read in a Structure file, you can use the `StructureReader` class. For example:

```
structure = "snpio/example_data/structure_files/phylogen_subset14K.str"
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"

gd = StructureReader(filename=structure, popmapfile=popmap, force_popmap=
    True, verbose=True, plot_format="png", plot_fontsize=20, plot_dpi=300,
    despine=True, prefix="snpio_example", exclude_pops=["MX", "YU", "CH"],
    include_pops=["ON", "DS", "EA", "GU", "TT"])
```

This will read the genotype data from a STRUCTURE file and apply the population map (if provided).

### Note

The `StructureReader` class will automatically detect the format of the STRUCTURE file. It can be in one-line or two-line format (see STRUCTURE documentation), and can optionally contain population information in the file as the second tab-delimited column. If the population information is not provided in the STRUCTURE file, you can provide a population map file to assign samples to populations.

## Key Methods in VCFReader, PhylipReader, and StructureReader

Function/Method	Description
VCFReader	Reads and writes genotype data from/to a VCF file and applies a population map if provided.
write_vcf	Writes the filtered or modified genotype data back to a VCF file.
PhylipReader	Reads and writes genotype data from/to a PHYLIP file and applies a population map.
write_phylip	Writes the filtered or modified genotype data back to a PHYLIP file.
StructureReader	Reads and writes genotype data from/to a STRUCTURE file and applies a population map.

Function/Method	Description
<code>write_structure</code>	Writes the filtered or modified genotype data back to a STRUCTURE file.

The `write_vcf`, `write_phylip`, and `write_structure` methods are used to write the filtered or modified genotype data back to a VCF, PHYLIP, or STRUCTURE file, respectively.

#### Note

The `write_vcf`, `write_phylip`, and `write_structure` methods can be used to write the filtered or modified genotype data back to a new file. The new file will contain the filtered or modified genotype data based on the filtering criteria applied.

## Other GenotypeData Methods

The `GenotypeData` along with the `Plotting` classes have several useful methods for working with genotype data:

1. `Plotting.run_pca()`: Runs principal component analysis (PCA) on the genotype data and plots the results. The PCA plot can help visualize the genetic structure of the populations in the dataset, with each point representing an individual. Individuals are colored by missing data proportion, and populations are represented by different shapes. A 2-dimensional PCA plot is generated by default, but you can specify three PCA axes as well. For example:
2. `GenotypeData.missingness_reports()`: Generates missing data reports and plots for the dataset. The reports include the proportion of missing data per individual, per locus, and per population. These reports can help you identify samples, loci, or populations with high levels of missing data. For example:
3. The `GenotypeData` class will automatically create a plot showing the number of individuals present in each population, if a `popmapfile` is provided. For example:

## Filtering Genotype Data with NRemover2

The `NRemover2` class provides a variety of filtering methods to clean your genotype data. Here is an example of how to apply filters to remove samples and loci with too much missing data, monomorphic sites, singletons, minor allele count (MAC), minor allele frequency (MAF), and more:

```
# Apply filters to remove samples and loci with too much missing data
gd_filt = nrm.filter_missing_sample(0.75).filter_missing(0.75) .
    filter_missing_pop(0.75).filter_mac(2).filter_monomorphic(
    exclude_heterozygous=False).filter_singletons(exclude_heterozygous=False)
    .filter_biallelic(exclude_heterozygous=False).resolve()
```

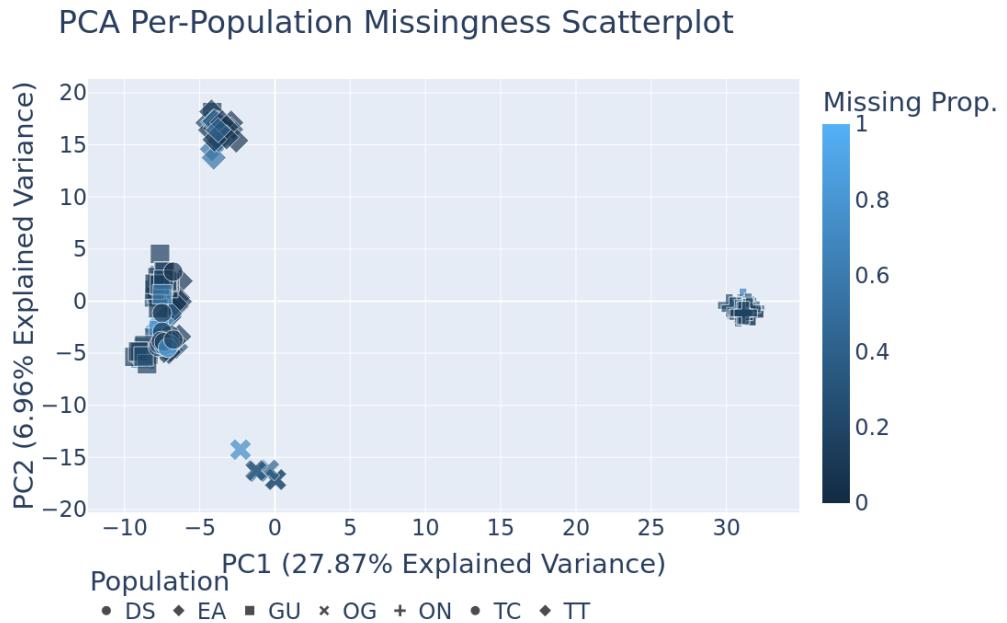


Figure 1: PCA Plot with samples colored by missing data proportion and populations represented by different shapes.

```
# Write the filtered VCF to a new file
gd_filt.write_vcf("filtered_output.vcf")
```

## Key Methods in NRemover2

Function/Method	Description
filter_missing_sample	Filters samples with missing data above the threshold.
filter_missing	Filters loci with missing data above the threshold.
filter_mac	Filters loci with a minor allele count below the threshold.
filter_maf	Filters loci with a minor allele frequency below the threshold.
filter_monomorphic	Filters monomorphic loci (sites with only one allele).

### Note

You must call `resolve()` at the end of the filtering chain to apply the filters and return the filtered GenotypeData object. The `resolve()` method is required to finalize the filtering process and return the filtered dataset.

## Missingness Report

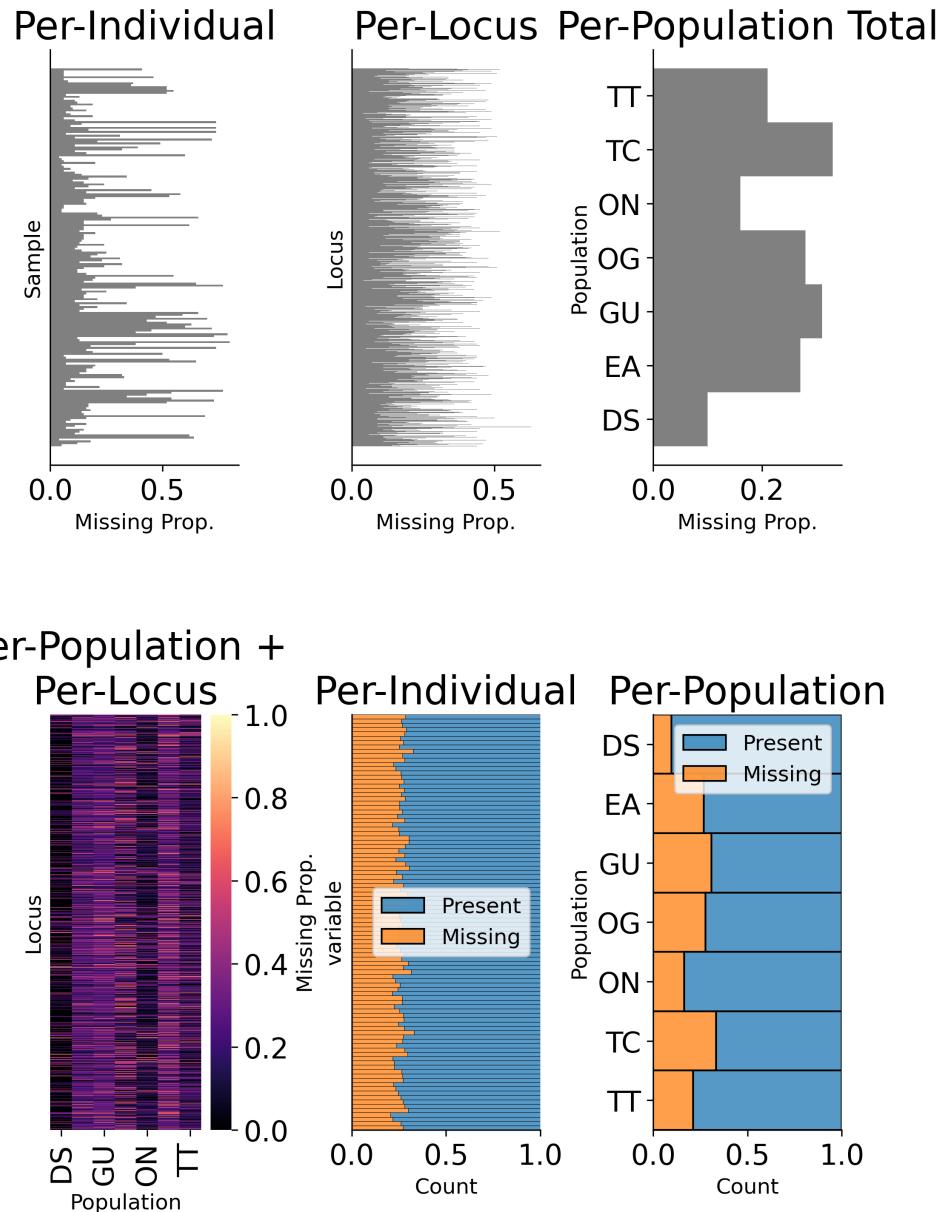


Figure 2: Missing Data Report with plots depicting missing data per sample, locus, and population.

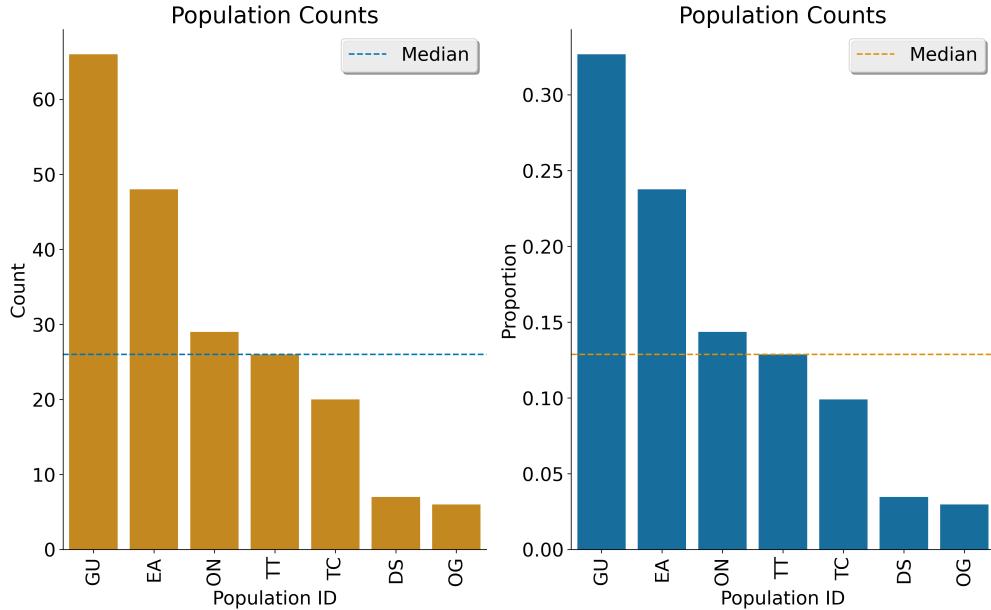


Figure 3: Population Counts and proportion bar plots with median indicated.

#### Note

The `exclude_heterozygous` parameter in `filter_monomorphic`, `filter_singletons`, and `filter_biallelic` methods allows you to exclude heterozygous genotypes from the filtering process. By default, heterozygous genotypes are included in the filtering process.

#### Note

`thin_loci` and `filter_linked` are only available for `VCFReader` and not for `PhylipReader` and `StructureReader`. These methods are used to thin loci by removing loci within a specified distance of each other on the same locus or chromosome, as defined in the VCF file. The `thin_loci` method removes loci within a specified distance of each other, while the `filter_linked` method filters loci that are linked within a specified distance.

#### Warning

The `filter_linked(size)` method might yield a limited number of loci with unlinked SNP data. It is recommended to use this method with caution and check the output carefully.

## Additional Methods in NRemover2

`search_thresholds()` searches a range of filtering thresholds for all missing data, minor allele frequency (MAF), and minor allele count (MAC) filters. This method helps you find

the optimal thresholds for your dataset. It will plot the threshold search results so you can visualize the impact of different thresholds on the dataset.

With `search_thresholds()`, you can specify the thresholds to search for and the order in which to apply the filters:

```
# Initialize NRemover2 with GenotypeData object
nrm = NRemover2(gd)

# Specify filtering thresholds and order of filters
nrm.search_thresholds(thresholds=[0.25, 0.5, 0.75, 1.0], maf_thresholds
                      =[0.01, 0.05], mac_thresholds=[2, 5], filter_order=[
                        "filter_missing_sample", "filter_missing", "filter_missing_pop", "
                        filter_mac", "filter_monomorphic", "filter_singletons", "filter_biallelic
                      ])
```

The `search_thresholds()` method will search across thresholds for missing data, MAF, MAC, and the boolean filters based on the specified thresholds and filter order. It will plot the results so you can visualize the impact of different thresholds on the dataset.

Below are example plots that are created when running the `search_thresholds()` method:

### Note

The `search_thresholds()` method is incompatible with both `thin_loci(size)` and `filter_linked()` being in the `filter_order` list.

### Warning

The `search_thresholds()` method can be called either before or after any other filtering, but note that it will reset the filtering chain to the original state. If you call `search_thresholds()` after applying other filters, it will reset the filtering chain to the original state and apply the search across the specified thresholds.

`plot_sankey_filtering_report()` generates a Sankey plot to visualize how SNPs are filtered at each step of the pipeline. For example:

```
from snpio import NRemover2, VCFReader

vcf = "snpio/example_data/vcf_files/phylogen_subset14K_sorted.vcf.gz"
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"

gd = VCFReader(filename=vcf, popmapfile=popmap, force_popmap=True, verbose=
               True, plot_format="png", plot_fontsize=20, plot_dpi=300, despine=True,
               prefix="snpio_example")

# Initialize NRemover2.
nrm = NRemover2(gd)

# Apply filters to remove samples and loci.
```

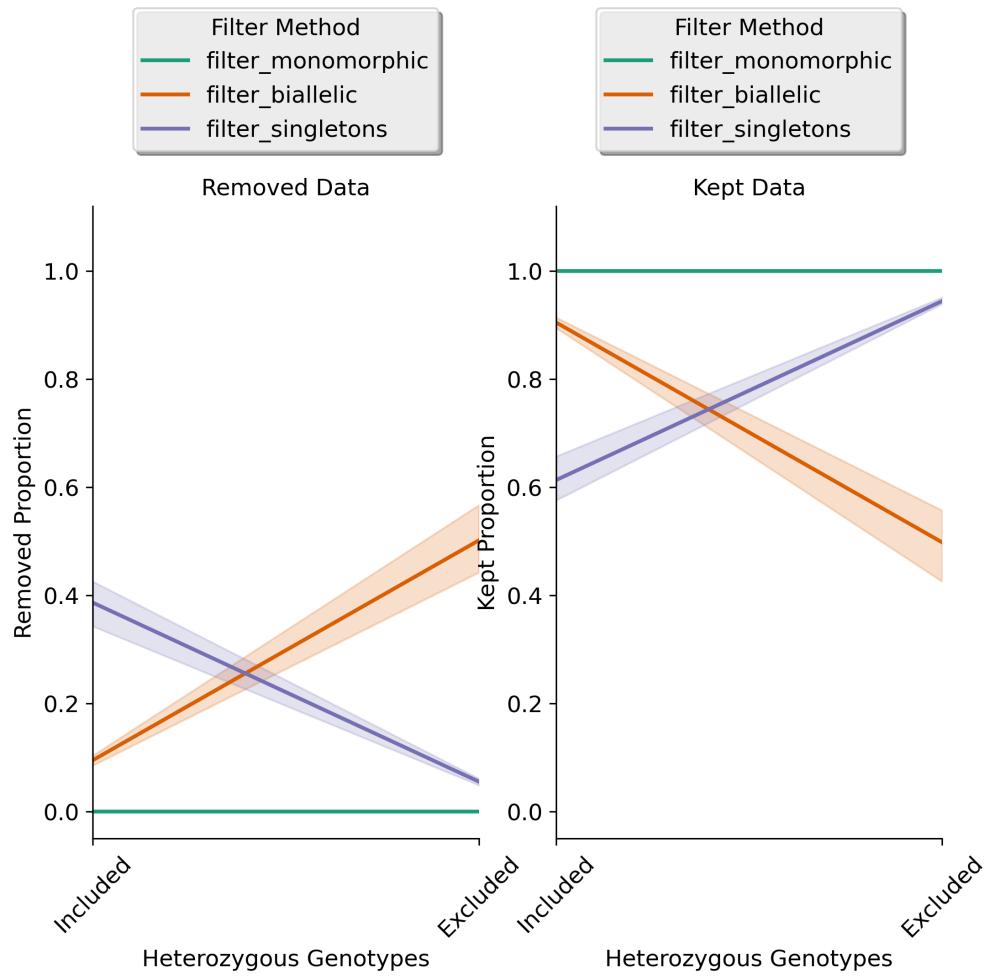


Figure 4: Filtering results for boolean filtering methods.

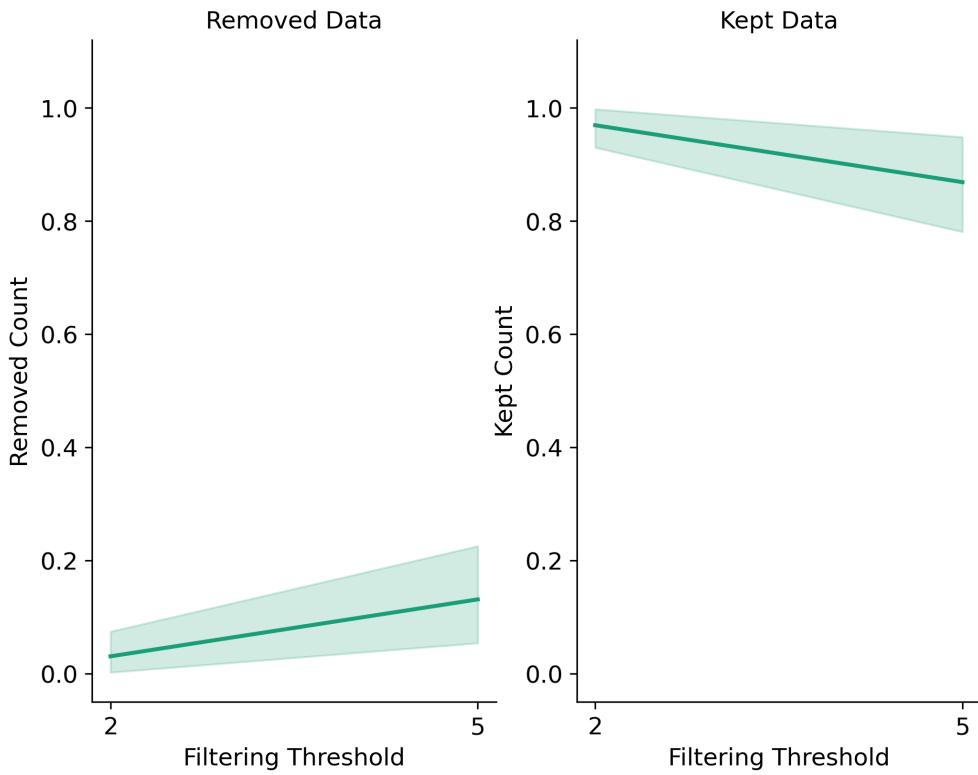


Figure 5: Filtering results for minor allele count (MAC).

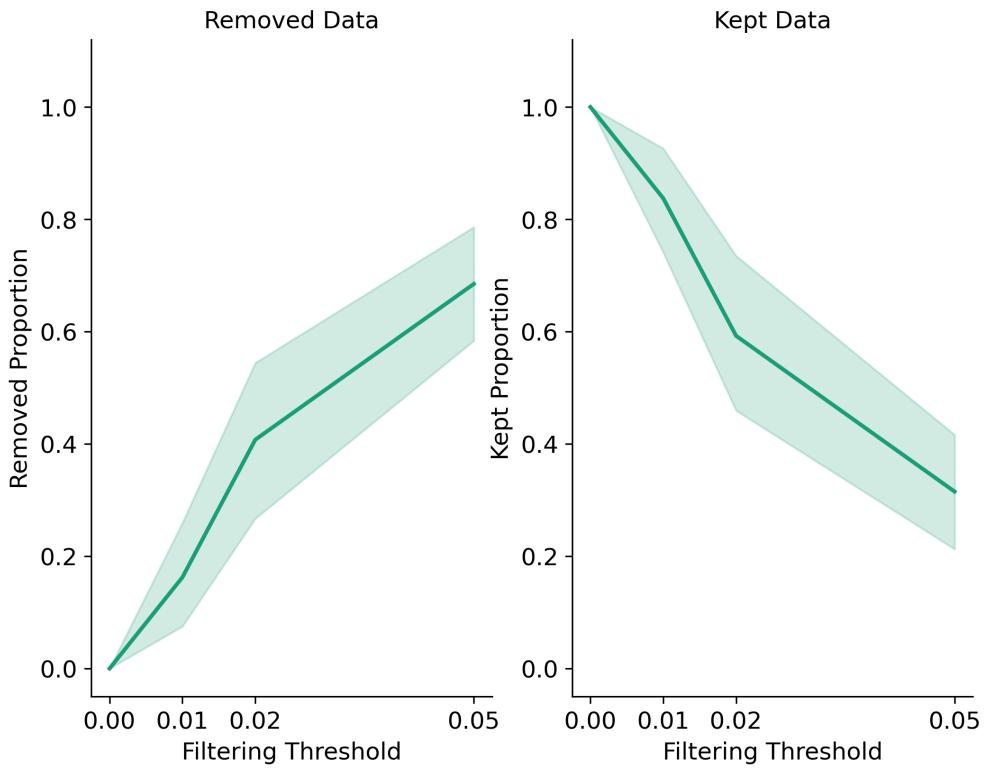


Figure 6: Filtering results for minor allele frequency (MAF).

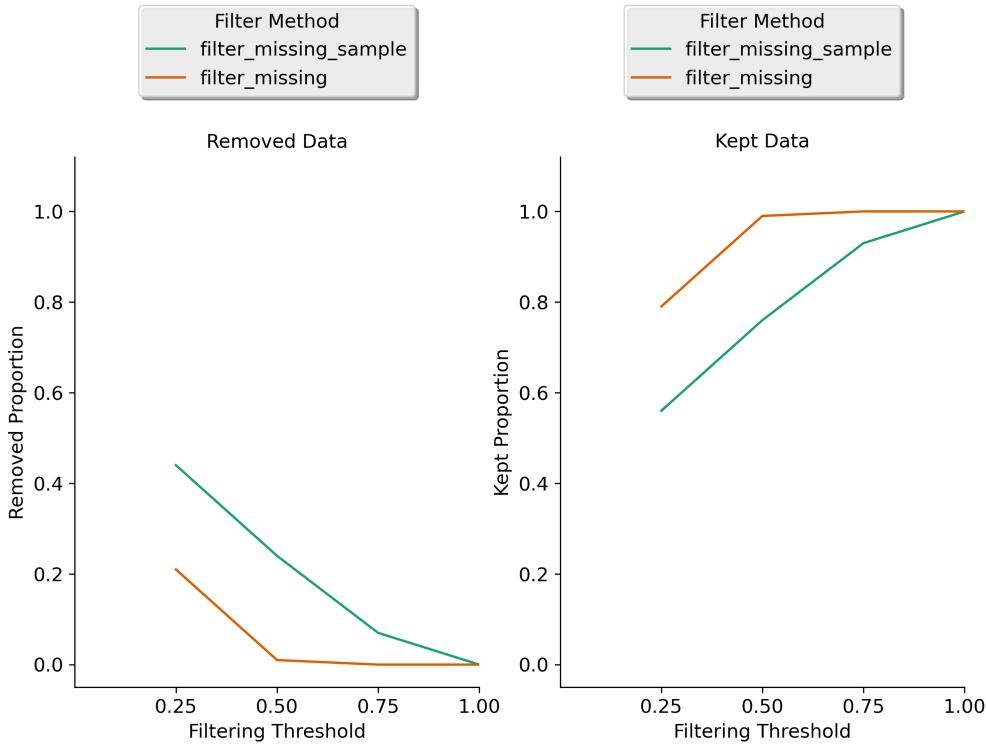


Figure 7: Missing data filtering results for loci and samples.

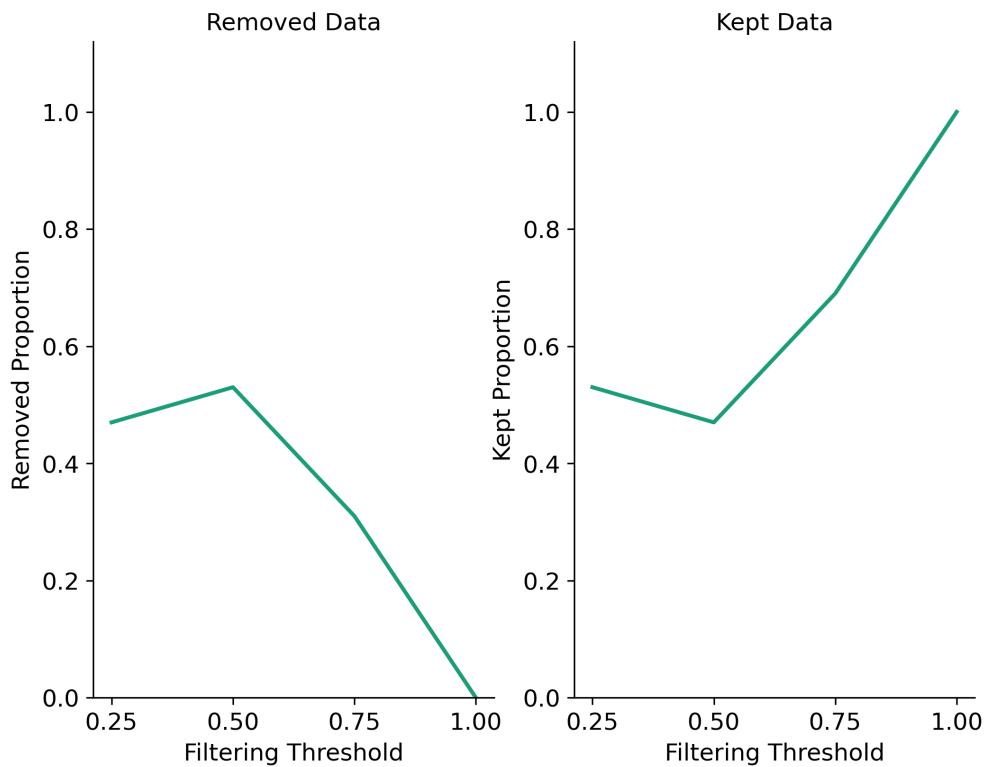


Figure 8: Missing data filtering results for populations.

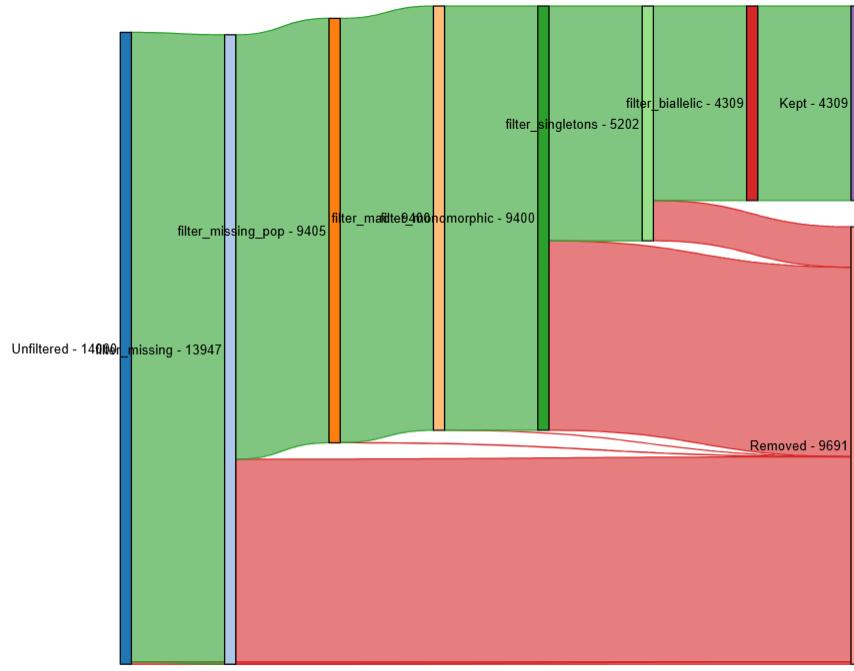


Figure 9: Sankey plot depicting loci retained and removed at each filtering step.

```

gd_filt = nrm.filter_missing_sample(0.75).filter_missing(0.75).
    filter_missing_pop(0.75).filter_mac(2).filter_monomorphic(
        exclude_heterozygous=False).filter_singletons(exclude_heterozygous=False)
    .filter_biallelic(exclude_heterozygous=False).resolve()

nrm.plot_sankey_filtering_report()

```

This will automatically track the number of loci at each filtering step and generate a Sankey plot to visualize the filtering process. The Sankey plot shows how many loci are removed at each step of the filtering process. For example:

#### Note

The `plot_sankey_filtering_report()` must be called after filtering and calling the `resolve()` method to generate the Sankey plot. It is also incompatible with `thin_loci()`, `filter_linked()`, and `random_subset_loci()` being in the `filter_order` list.

`plot_sankey_filtering_report()` also only plots loci removed at each filtering step and does not plot samples removed. It is designed to visualize the filtering process for loci only.

## GenotypeData Properties

Once genotype data is loaded using any of the readers, you can access several useful properties from the `GenotypeData` object:

Attribute	Description
<code>num_snps</code>	Number of SNPs or loci in the dataset.
<code>num_inds</code>	Number of individuals in the dataset.
<code>populations</code>	List of populations in the dataset.
<code>popmap</code>	Mapping of SampleIDs to PopulationIDs.
<code>popmap_inverse</code>	Dictionary with population IDs as keys and lists of samples as values.
<code>samples</code>	List of samples in the dataset.
<code>snpdict</code>	Dictionary with SampleIDs as keys and genotypes as values.
<code>loci_indices</code>	Numpy array with boolean values indicating the loci that passed the filtering criteria set to <code>True</code> .
<code>sample_indices</code>	Numpy array with boolean values indicating the samples that passed the filtering criteria set to <code>True</code> .
<code>snp_data</code>	2D numpy array of SNP data of shape ( <code>num_inds</code> , <code>num_snps</code> ).
<code>ref</code>	List of reference alleles for each locus.
<code>alt</code>	List of alternate alleles for each locus.
<code>inputs</code>	Dictionary of input parameters used to load the genotype data.

## Genotype Encoding with GenotypeEncoder

SNPio also includes the `GenotypeEncoder` class for encoding genotype data into formats useful for downstream analysis and commonly used for machine and deep learning tasks.

The `GenotypeEncoder` class provides three encoding properties:

`genotypes_onehot`: Encodes genotype data into one-hot encoding, where each possible biallelic IUPAC genotype is represented by a one-hot vector. Heterozygotes are represented as multi-label vectors as follows:

```
onehot_dict = {
    "A": [1.0, 0.0, 0.0, 0.0],
```

```

    "T": [0.0, 1.0, 0.0, 0.0],
    "G": [0.0, 0.0, 1.0, 0.0],
    "C": [0.0, 0.0, 0.0, 1.0],
    "N": [0.0, 0.0, 0.0, 0.0],
    "W": [1.0, 1.0, 0.0, 0.0],
    "R": [1.0, 0.0, 1.0, 0.0],
    "M": [1.0, 0.0, 0.0, 1.0],
    "K": [0.0, 1.0, 1.0, 0.0],
    "Y": [0.0, 1.0, 0.0, 1.0],
    "S": [0.0, 0.0, 1.0, 1.0],
    "N": [0.0, 0.0, 0.0, 0.0],
}

```

`genotypes_int`: Encodes genotype data into integer encoding, where each possible biallelic IUPAC genotype is represented by an integer as follows: as follows: A=0, T=1, G=2, C=3, W=4, R=5, M=6, K=7, Y=8, S=9, N=-9. Missing values are represented as -9.

`genotypes_012`: Encodes genotype data into 0-1-2 encoding, where 0 represents the homozygous reference genotype, 1 represents the heterozygous genotype, and 2 represents the homozygous alternate genotype. Missing values are represented as -9.

Example Usage:

```

from snpio import VCFReader, GenotypeEncoder

vcf = "snpio/example_data/vcf_files/phylogen_subset14K_sorted.vcf.gz"
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"

gd = VCFReader(filename=vcf, popmapfile=popmap, force_popmap=True, verbose=
    True, plot_format="png", plot_fontsize=20, plot_dpi=300, despine=True,
    prefix="snpio_example")

encoder = GenotypeEncoder(gd)

# Convert genotype data to one-hot encoding
gt_ohe = encoder.genotypes_onehot

# Convert genotype data to integer encoding
gt_int = encoder.genotypes_int

# Convert genotype data to 0-1-2 encoding.
gt_012 = encoder.genotypes_012

```

The `GenotypeEncoder` allows you to seamlessly convert genotype data into formats often used by machine and deep learning workflows.

You can also inversely convert the encoded data back to the original genotypes by just setting the `GenotypeEncoder` properties to a new value. For example:

```

# Convert one-hot encoded data back to genotypes
encoder.genotypes_onehot = gt_ohe

# Convert integer encoded data back to genotypes

```

```

encoder.genotypes_int = gt_int

# Convert 0-1-2 encoded data back to genotypes
encoder.genotypes_012 = gt_012

```

This will automatically update the original genotype data in the GenotypeData object and convert it to the original format stored in the `snp_data` property of the GenotypeData object.

## Population Genetics Analysis with PopGenStatistics

The `PopGenStatistics` class is designed to perform a suite of population genetic analyses on SNP datasets. It supports calculations such as D-statistics, Fst outliers, heterozygosity, nucleotide diversity, and Analysis of Molecular Variance (AMOVA). These analyses facilitate understanding of the genetic structure, diversity, and differentiation within and between populations.

The `PopGenStatistics` class provides several methods for calculating population genetic statistics and performing analyses on genotype data:

Class Method	Description	Supported Algorithm(s)
<code>calculate_d_statistics</code>	Calculates D-statistics and saves as CSV.	Patterson's, partitioned, and D-foil D-statistics.
<code>detect_fst_outliers</code>	Identifies Fst outliers. Supports P-values.	DBSCAN clustering, Traditional bootstrapping.
<code>summary_statistics</code>	Calculates genetic summary statistics.	Observed heterozygosity (Ho), Expected heterozygosity (He), Nucleotide diversity (Pi), Weir and Cockerham's Fst.

## PopGenStatistics Core Methods

Here is an example of how to use the `PopGenStatistics` class to perform population genetic analyses:

```

from snpio import VCFReader, PopGenStatistics

vcf = "snpio/example_data/vcf_files/phylogen_subset14K_sorted.vcf.gz"
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"

gd = VCFReader(filename=vcf, popmapfile=popmap, force_popmap=True, verbose=True,
               plot_format="png", plot_fontsize=20, plot_dpi=300, despine=True,
               prefix="snpio_example")

pgs = PopGenStatistics(gd)

# Calculate summary statistics.

```

```

summary_stats = pgs.summary_statistics(n_bootstraps=1000, n_jobs=-1,
                                         save_plots=True)

# Calculate D-statistics.
dstats_df, overall_results = pgs.calculate_d_statistics(
    method="patterson",
    population1="EA",
    population2="GU",
    population3="TT",
    outgroup="ON",
    num_bootstraps=10,
    n_jobs=1,
    max_individuals_per_pop=6,
)

# NOTE: Takes a while to run.
amova_results = pgs.amova(
    regionmap={
        "EA": "Eastern",
        "GU": "Eastern",
        "TT": "Eastern",
        "TC": "Eastern",
        "DS": "Ornate",
    },
    n_bootstraps=10,
    n_jobs=1,
    random_seed=42,
)

nei_dist_df, nei_pvals_df = pgs.neis_genetic_distance(n_bootstraps=1000)

summary_stats = pgs.summary_statistics(save_plots=True)

df_fst_outliers_boot, df_fst_outlier_pvalues_boot = pgs.detect_fst_outliers(
    correction_method="bonf",
    use_bootstrap=True,
    n_bootstraps=1000,
    n_jobs=1,
    tail_direction="upper",
)

df_fst_outliers_dbSCAN, df_fst_outlier_pvalues_dbSCAN = pgs.
    detect_fst_outliers(
        correction_method="bonf", use_bootstrap=False, n_jobs=1
)

```

The `PopGenStatistics` class provides a comprehensive suite of methods for calculating population genetic statistics and performing analyses on genotype data. These methods can help you understand the genetic structure, diversity, and differentiation within and between populations, and identify outliers and patterns in the data.

Below is an example of the output from the `neis_genetic_distance` method:

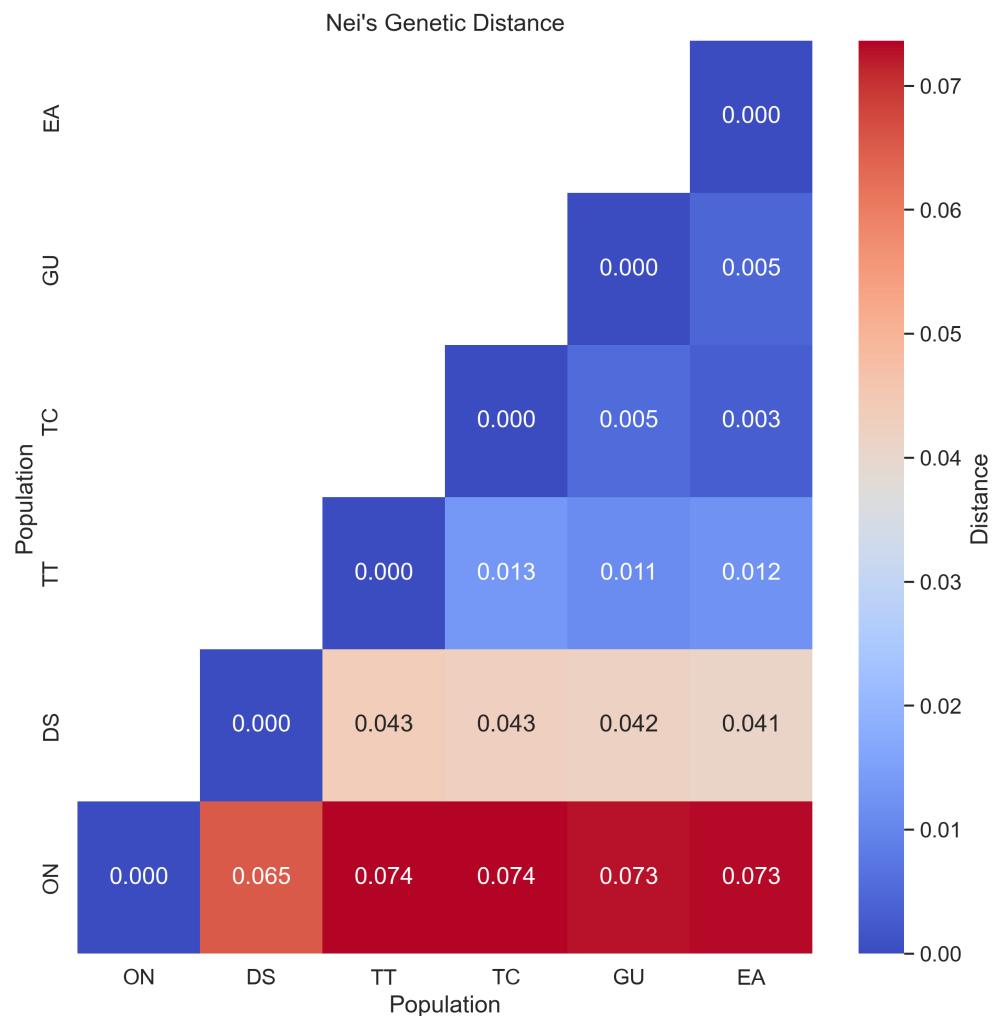


Figure 10: Nei's Genetic Distance Matrix.

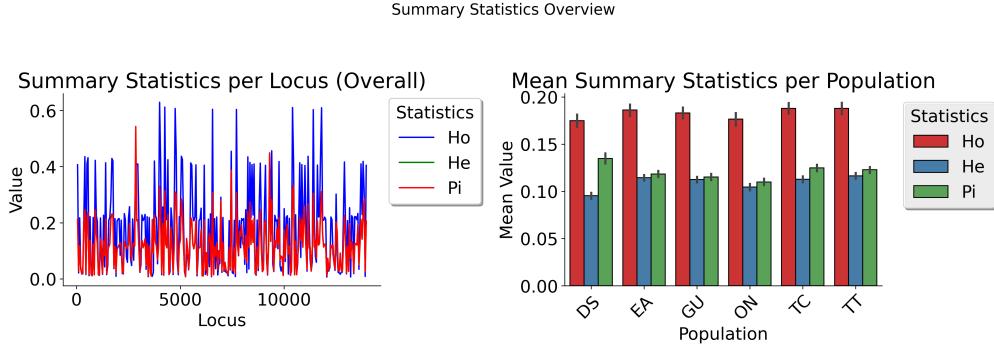


Figure 11: Summary statistics report with heterozygosity and nucleotide diversity.

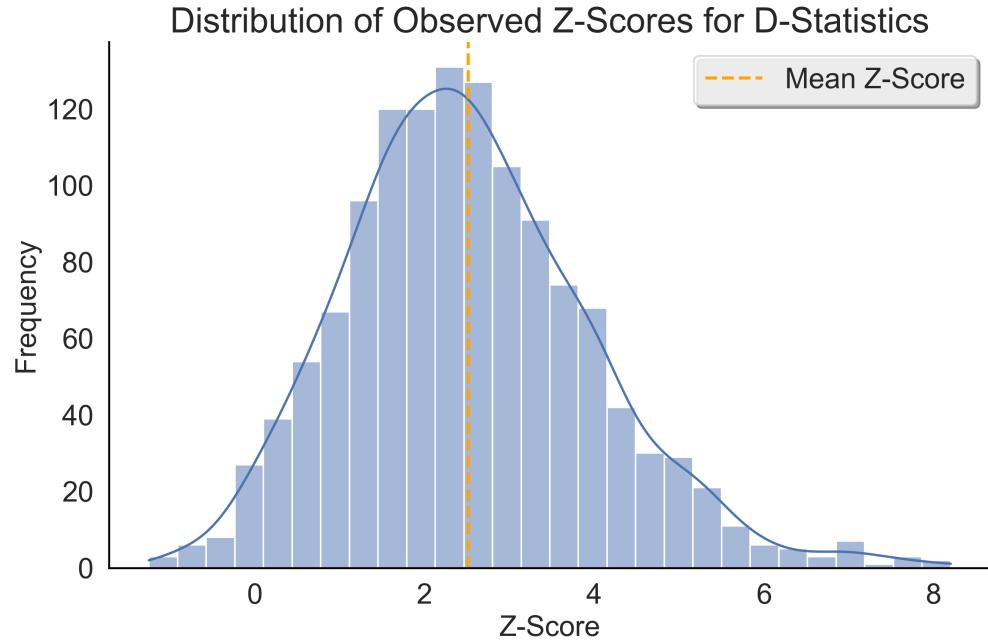


Figure 12: D-statistics distribution histogram plot.

The summary statistics method generates a summary report with observed heterozygosity (Ho), expected heterozygosity (He), nucleotide diversity (Pi), and Weir and Cockerham's Fst values for each population. The report includes plots of the summary statistics for each population, which can help you visualize the genetic diversity and differentiation within and between populations. Below is an example figure generated by the `summary_statistics` method:

The D-statistics method calculates Patterson's D-statistics, partitioned D-statistics, and D-foil D-statistics for the specified population groups. The method returns a DataFrame with the D-statistics values and overall results for the analysis. Below are three example visualizations made by the `calculate_d_statistics` method:

Below is an example of the plot made by the `detect_fst_outliers` method:

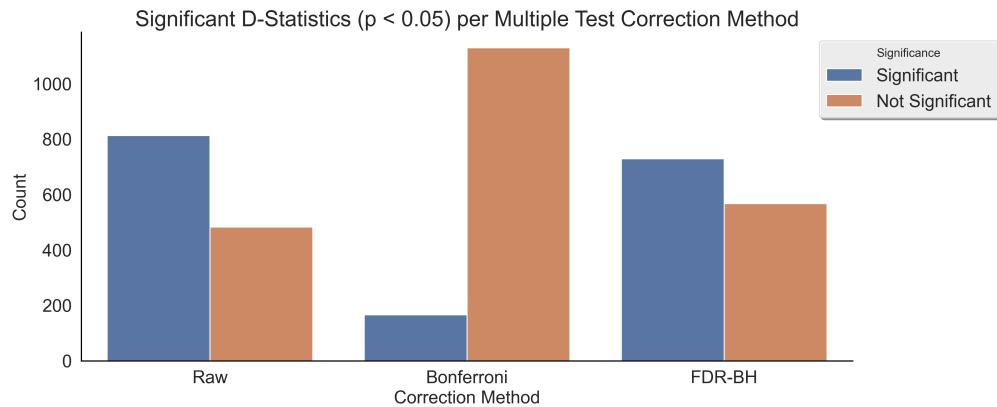


Figure 13: D-statistics significance counts bar plot.

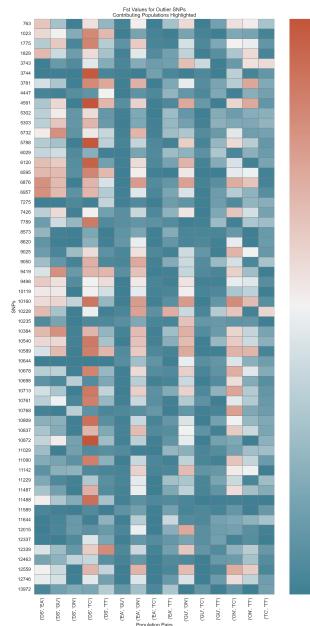


Figure 14: Fst outlier SNPs heatmap.

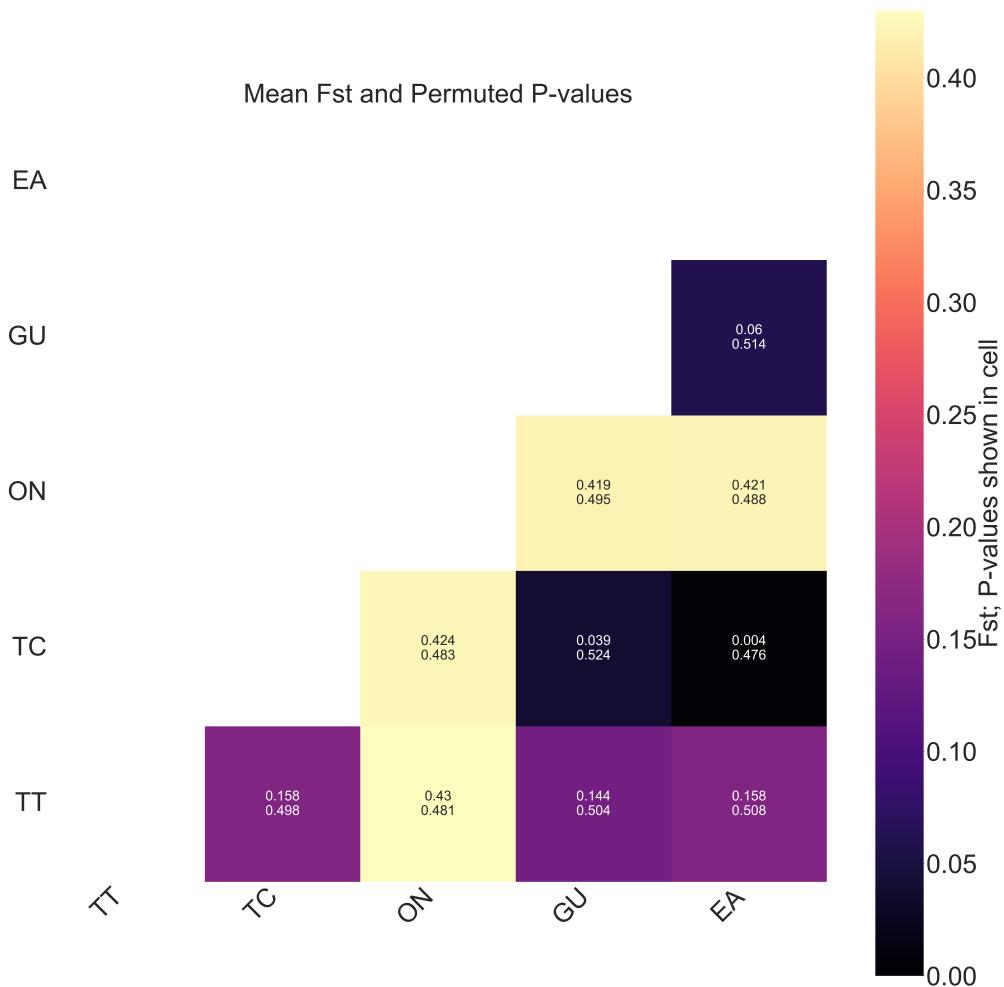


Figure 15: Pairwise Fst heatmap.

Finally, below is a plot depicting the results of the per-population pairwise Fst analysis:

## Loading and Parsing Phylogenetic TreeParser

SNPio also provides a `TreeParser` class to load and parse phylogenetic trees in Newick and NEXUS formats. The `TreeParser` class can read and parse tree files, modify tree structures, draw trees, and save trees in different formats. You can use the `TreeParser` class to analyze and visualize phylogenetic trees and extract relevant information for downstream analysis.

Here are some examples of how to load and parse a phylogenetic tree using the `TreeParser` class:

```
from snpio import TreeParser, VCFReader

vcf = "snpio/example_data/vcf_files/phylogen_subset14K_sorted.vcf.gz"
popmap = "snpio/example_data/popmaps/phylogen_nomx.popmap"

gd = VCFReader(filename=vcf, popmapfile=popmap, force_popmap=True, verbose=
```

```

        True, plot_format="pdf", plot_fontsize=20, plot_dpi=300, despine=True,
        prefix="snpio_example")

# Load a phylogenetic tree from a Newick file
tp = TreeParser(genotype_data=gd, treefile="snpio/example_data/trees/test.
tre", siterates="snpio/example_data/trees/test14K.rates", qmatrix="snpio/
example_data/trees/test.iqtree", verbose=True)

tree = tp.read_tree()

tree.draw(); # Draw the tree

# Save the tree in Newick format
tp.write_tree(tree, save_path="snpio/example_data/trees/test_newick.tre")

# Save the tree in NEXUS format
tp.write_tree(tree, save_path="snpio/example_data/trees/test_nexus.nex",
nexus=True)

# Returns the tree in Newick format as a string
tp.write_tree(tree, save_path=None)

# Get the tree stats. Returns a dictionary of tree stats.
print(tp.tree_stats())

# Reroot the tree at any nodes containing the string 'EA' in the sampleID.
# Use the '~' character to specify a regular expression pattern to match.
tp.reroot_tree("~EA")

# Get a distance matrix between all nodes in the tree.
print(tp.get_distance_matrix())

# Get the Rate Matrix Q from the Qmatrix file.
print(tp.qmat)

# Get the Site Rates from the Site Rates file.
print(tp.site_rates)

# Get a subtree with only the samples containing 'EA' in the sampleID.
# Use the '~' character to specify a regular expression pattern to select
# all
# tips containing the pattern.
subtree = tp.get_subtree("~EA")

# Prune the tree to remove samples containing 'ON' in the sampleID.
pruned_tree = tp.prune_tree("~ON")

# Write the subtree and pruned tree. Returns a Newick string if 'save_path'
# is None. Otherwise saves it to 'save_path'.
print(tp.write_tree(subtree, save_path=None))
print(tp.write_tree(pruned_tree, save_path=None))

```

The TreeParser class provides several methods for working with phylogenetic trees, in-

cluding reading, writing, and modifying trees. You can use these methods to analyze and manipulate phylogenetic trees for your research and analysis tasks.

The `TreeParser` class also provides methods for calculating tree statistics, rerooting trees, getting distance matrices, and extracting subtrees based on sample IDs. These methods can help you analyze and visualize phylogenetic trees and extract relevant information for downstream analysis.

The `Rate` matrix  $Q$  and site Rates can be accessed from the `Qmatrix` and `Site Rates` files, respectively. These matrices can be used to calculate evolutionary distances and rates between samples in the phylogenetic tree. The `siterates` file can be output by IQ-TREE or specified as a one-column file with the rates for each site in the alignment (header optional). The `qmatrix` file can be obtained from the IQ-TREE standard output ('.iqtree' file) or from a stand-alone `Qmatrix` file with the rate matrix  $Q$ . In the latter case, the file should be a tab-delimited or comma-delimited file with the rate matrix  $Q$  with substitution rates in the order: "A", "C", "G", "T". A header line is optional.

The rate matrix and site rates objects can be accessed by their corresponding properties:

- `tp.qmat`: Rate matrix  $Q$ .
- `tp.site_rates`: Site rates.

The `TreeParser` class is designed to simplify the process of working with phylogenetic trees and extracting relevant information for downstream analysis. You can use the `TreeParser` class to load, parse, and manipulate phylogenetic trees in Newick and NEXUS formats, and extract tree statistics, distance matrices, and subtrees based on sample IDs. For more information on the `TreeParser` class and its methods, please refer to the API documentation.

## Benchmarking the Performance

You can benchmark the filtering performance using the `Benchmark` class to visualize how thresholds affect the dataset, if you have installed the `snpio dev` requirements:

```
pip install snpio[dev]
```

Then, you can use the `Benchmark` class to plot performance metrics for your filtered genotype data after the `resolve()` method is called. For example:

```
from snpio.utils.benchmarking import Benchmark  
  
Benchmark.plot_performance(nrm.genotype_data, nrm.genotype_data.  
    resource_data)
```

This function will plot performance metrics for your filtered genotype data and for the `vcFReader` class, giving insights into data quality changes.

The `Benchmark` class is designed to help you evaluate the performance of your filtering process and visualize the impact of different thresholds on the dataset. For more information on the `Benchmark` class and how to use it, see the API documentation.

## Conclusion

This guide provides an overview of how to get started with the SNPio library. It covers the basic steps to read, manipulate, and analyze genotype data using the VCFReader, PhylipReader, StructureReader, and NRemover2 classes. SNPio is designed to simplify the process of handling genotype data and preparing it for downstream analysis, such as population genetics, phylogenetics, and machine learning. The library supports various file formats, including VCF, PHYLIP, and STRUCTURE, and provides tools for filtering, encoding, and visualizing genotype data. This guide will help you get up and running with SNPio quickly and efficiently.

For more information on the SNPio library, please refer to this API documentation and examples provided in the repository. If you have any questions or feedback, please feel free to reach out to the developers. We hope you find SNPio useful for your bioinformatic analyses!

### Note

The SNPio library is under active development, and we welcome contributions from the community. If you would like to contribute to the project, please check the GitHub repository for open issues and submit a pull request. We appreciate your support and feedback!

If you encounter any issues or have any questions about the SNPio library, please feel free to reach out to the developers or open an issue on the GitHub repository. We are here to help and improve the library based on your feedback.

The SNPio library is licensed under the GPL3 License, and we encourage you to use it for your research and analysis tasks. If you find the library useful, please cite it in your publications. We appreciate your support and feedback! We hope you find SNPio useful for your research and analysis tasks! Thank you for using SNPio!