

Using transcriptomics to investigate evolution and toxicology in *Gambierdiscus*.¹

Key words: *Gambierdiscus*, ciguatoxin, pan-transcriptome

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Abstract

Species of the genus *Gambierdiscus* produce Ciguatoxins (CTXs), the causative agent of ciguatera fish poisoning, a potentially debilitating seafood borne illness. Species of *Gambierdiscus* possess very large genomes, 32 - 35 Gbp, and, as with other dinoflagellates, possess unique genomic characteristics, such as highly repetitive and complex genome architecture. The exact toxins produced by species of *Gambierdiscus* remain largely unclear. It has been verified using LCMS on multiple strains that the species *Gambierdiscus polynesiensis* produces analogs of CTXs. Other species appear to produce maitotoxins, gambierol, and other uncharacterised toxins. An understanding of the evolution of *Gambierdiscus* and their toxins requires information regarding their genetics. Transcriptomic sequencing is a feasible alternative to genome sequencing. In this study, we generated de novo RNA-seq libraries for *Gambierdiscus polynesiensis*, *Gambierdiscus carpenteri*, *Gambierdiscus* cf. *silvae* and *Gambierdiscus lapillus*, compared these to a previously sequenced *Gambierdiscus australes*, to discover a set of core genes shared by all species. We present a *Gambierdiscus* core transcriptome, which might be used to investigate candidate genes related to toxin production.

Further to investigate CTX production more specifically, we compared two CTX - producing strains of *Gambierdiscus polynesiensis* to one non-CTX producing strain, verified by LC-MS/MS, to look for clues about pathways involved in ciguatoxin production.

To do:

- re-structure as per Tim's comments

Introduction

general dino stuff, genetic peculiarities

- loss of nucleosomes [27]
- high DNA content [17]
- thymine nucleotide replacement with uracil [24]
- plastid and mitochondrial genes widely incorporated into - genome [2]
- horizontal gene transfer rife
- potential unlinking of transcriptome and observed protein expression [3]
- large gene families
- trans-splicing creates pool of fully mature, useable mRNA that can then be selected from .. may be why transcriptome good approximation of genome [20, 29]
- weird as fuck transcription factors [10]
- dinoSL .. trans-spliced leader present in 2/3 of genes detected and in 12/15 of highly expressed [3, 20]
- genes of highly expressed proteins are present in tandem arrays [3]
- post-transcriptional control of protein expression and extremely long mRNA half lives [21]
- smRNA as possible post-transcriptional regulation mechanism [4]

importance of ref genome

The analysis of any genetic data relies on the reference to a known, closely related entity. Without a functional protein or genome reference database, the generation of sequencing data would be like pissing in the wind. A reference is essential in determining both the adequacy of the sequencing methodology as well as interpretation of results.

due to no ref gen, importance of transcriptome and features that allow for approximation

other pan-tran work ie koid and bacterial.. yeast?

In this study.. go through LC-MS and toxicology info

Methods

Scripts used for this project are available on Github under `hydrahamster/pan-tran`. Venn diagrams were created with InteractiVenn [12].

Transcriptome acquisition

Species of *Gambierdiscus* used in this chapter are summarized in Table 1. Toxicity and toxin profile reports are specific to the strains used due to the possibility of inter-species variation in toxin production as recently reported, unless noted otherwise [18, 26]. The *G. polynesiensis* toxin profile was elucidated by Tim Harwood at the Cawthron institute with the same methodology as for *G. lapillus* in **Chapter 1**. Seq libraries were assembled as per the transcriptome assembly subsection in the methods of **chapter 4**, without `diginorm`.

To do:

- check if Shauna submitted CAWD149 and if it was from a single RNA extraction

Table 1: *Gambierdiscus* species transcriptomes used in this study along with their toxicity, toxin profile, accession numbers and source. Where possible, information is strain specific & otherwise denoted with *

Species	<i>G. australes</i>	<i>G. carpenteri</i>	<i>G. lapillus</i>	<i>G. polyne-siensis</i>	<i>G. cf. silvae</i>
Strain	CAWD149	UTSMER9A	HG4	CG15	HG5
Transcriptome source	MMETSP	chapter 4	chapter 4	chapter 4	chapter 4
Accession ID	MMETSP0766	SRR6821720	SRR6821722	SRR6821723	SRR6821721
Isolation location	Rarotonga, Cook Islands (2007)	Merimbula, Australia (2014)	Heron Island, Australia (2014)	Rarotonga, Cook Islands (2014)	Heron Island, Australia (2014)
Toxin profile (LC-MS/MS)	CTX -ve; MTX +ve	CTX -ve; MTX -ve	CTX -ve; MTX +ve	CTX +ve; MTX +ve	CTX -ve; MTX +ve
Toxicity via bioassay	CTX +ve; MTX N/A	CTX -ve; MTX +ve	CTX +ve*; MTX +ve*	CTX N/A; MTX N/A	CTX +ve; MTX +ve
References	[15, 22, 25]	[18]	[16, 18]		[16, 18]

Spliced leader search

The spliced leader sequences reported by Zhang et al. (2007) were used to build a hmmer library. The transcriptome assemblies were searched with the dinoSL hmmer library to investigate for spliced leader presence. All cluster iterations following were matched against the contig IDs with spliced leader presence.

Homolog clustering

Cd-hit was used to cluster highly similar transcripts to reduce redundancy with the flags T 10 -M 5000 -G 0 -c 1.00 -aS 1.00 -aL 0.005 as shown by Cerveau and Jackson (2016) [5, 9]. Transdecoder was used to predict coding regions on the clustered nucleotide sequences [11]. Protein clusters were annotated with Interproscan v5.27 with local lookup server [23]. Protein clusters were processed to include the species of origin instead of the TRINITY tag and concatenated for input to get_homologues [28]. The -t 0 flag was used for get_homologues to acquire all possible clusters even with only one species representative, and -G for the OMCL algorithm. The resulting pan-, core- and softcore-clusters were matched with their interpro annotations and GO terms were queried with GOSUM against the basic Gene Ontology (GO) database [1, 6, 14]. GOSUM was run at levels 1 and 2 of GOs with the go-basic GO reference.

PKS search

The transcriptome assemblies were queried for the five conserved PKS active domains using hmmer [8] with libraries developed for this project. The contigs which were identified to contain an active domain were then searched for within the clusters to identify how the active domains grouped together between species.

To do:

- need to download ACP, ET, KR, DR, AT and TE sequences and make hmmer libs – inclusion of these extra domains is heavily dependent on time. Pretty much all studies so far just looked for the KS domain, though not sure why - all 7 are necessary to synthesize a polyketide structure.

- find conserved sequences of each KS cluster (yay for hmmer) and align clusters, run BI phylogeny to see if there is anything interesting there

Last common ancestor determination of contigs

Predicted proteins of each transcriptome were searched against the Uniprot databases SwissProt and trEMBL [7]. BASTA was used to extract the taxonomic determination from the database search for each contig and the associated last common ancestor [13].

Results

General info

The progression of clustering and annotation results per transcriptome can be found in Table 2. A total of 287,546 clusters were found across all five species.

Table 2: Progression of clusters found in each *Gambierdiscus* transcriptome during processing.

Species	<i>G. aus- trales</i>	<i>G. carpenteri</i>	<i>G. lapillus</i>	<i>G. polyne- siensis</i>	<i>G. cf. sil- vae</i>
Contigs #	102,863	263,829	148,972	270,315	191,224
dinoSL #	304	683	232	1,570	1,524
Nucleotide clusters # (cd-hit)	102,861	263,743	148,966	270,265	191,205
Predicted coding regions # (Transde- coder)	63,299	180,568	111,862	176,290	132,688
Contigs anno- tated # (In- terpro Scan)	131,970	334,737	225,324	225,324	254,844
Contigs with Uniprot hits #					
Part of core transcriptome clusters	13,750	13,750	13,750	13,750	13,750
Part of soft- core transcrip- tome clusters	2,372	16,058	16,297	16,557	16,636
Pan- transcriptome clusters	35,356	61,494	32,341	60,769	41,350

Tim: It seems kinda conspicuous that the unique clusters of *G. carpenteri* & *G. poly* are almost twice the number of *G. lapillus* and *G. silvae*, the first two were sequenced together with 150bp read length while the other two had 75bp read length during sequencing. Does this seem odd to you too?

Comparison of *Gambierdiscus* inter-species transcriptome annotations

The GOs were split up into the three functional groups defined by the consortium: 1) Molecular processes (Figs. 3 & 6) defined as biochemical or a macromolecule directly interacting with other molecules; 2) Cellular components (Figs. 1 & 5) defined by the location within the cell where a molecular process takes place; and 3) Biological process (Figs. 2 & 4) which is defined as a molecular machinery participating in the execution of the cell's genetic programming, e.g. cell division. GO basic is structured in a hierarchical manner, with parent and child terms where child terms are more specific than parent terms. For a general overview of functions present in each transcriptome, level 1 GO terms were elucidated (Figs. 2, 1 & 3). A more in depth query of the functions present in each transcriptome was conducted with a GO search of the child terms at level 2 (Figs. 4, 5 & 6).

To do:

- **Tim** I think I'm going to need to take out anything that links to Bacterial or unknown LCA and then re-run GOSUM. Thoughts?
- heatmap (evol relationship inferred from clustering, compare to phylogeny in **chapter 4**) from get_hom is throwing up errors
- describe differences in graphs once I know what needs to be taken out and re-run
- GOSUM lvl2 graphs are partially missing descriptions on x-axis. Fix when re-run

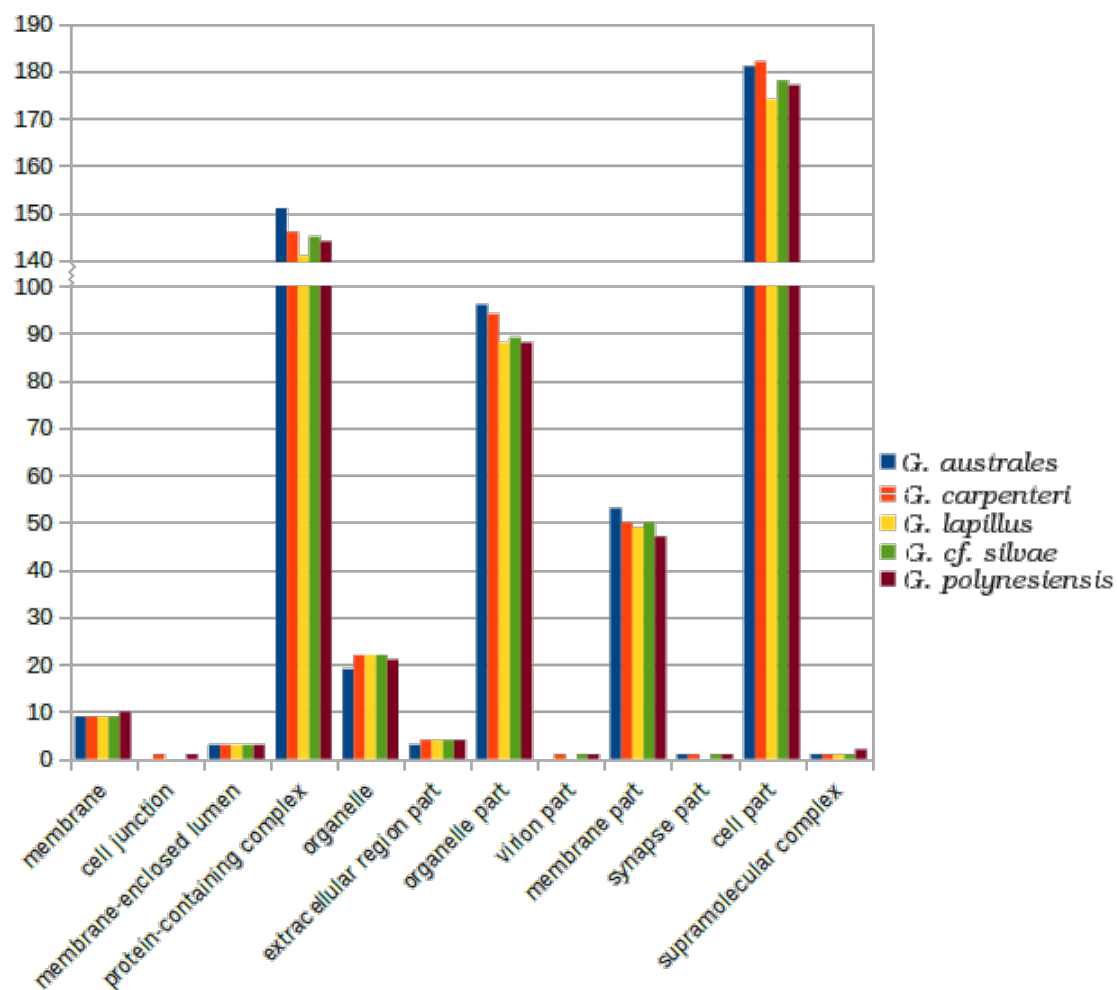


Figure 1: Summary of cellular GO annotations between *Gambierdiscus* species at GO-SUM level 1 from Suppl. table 5.

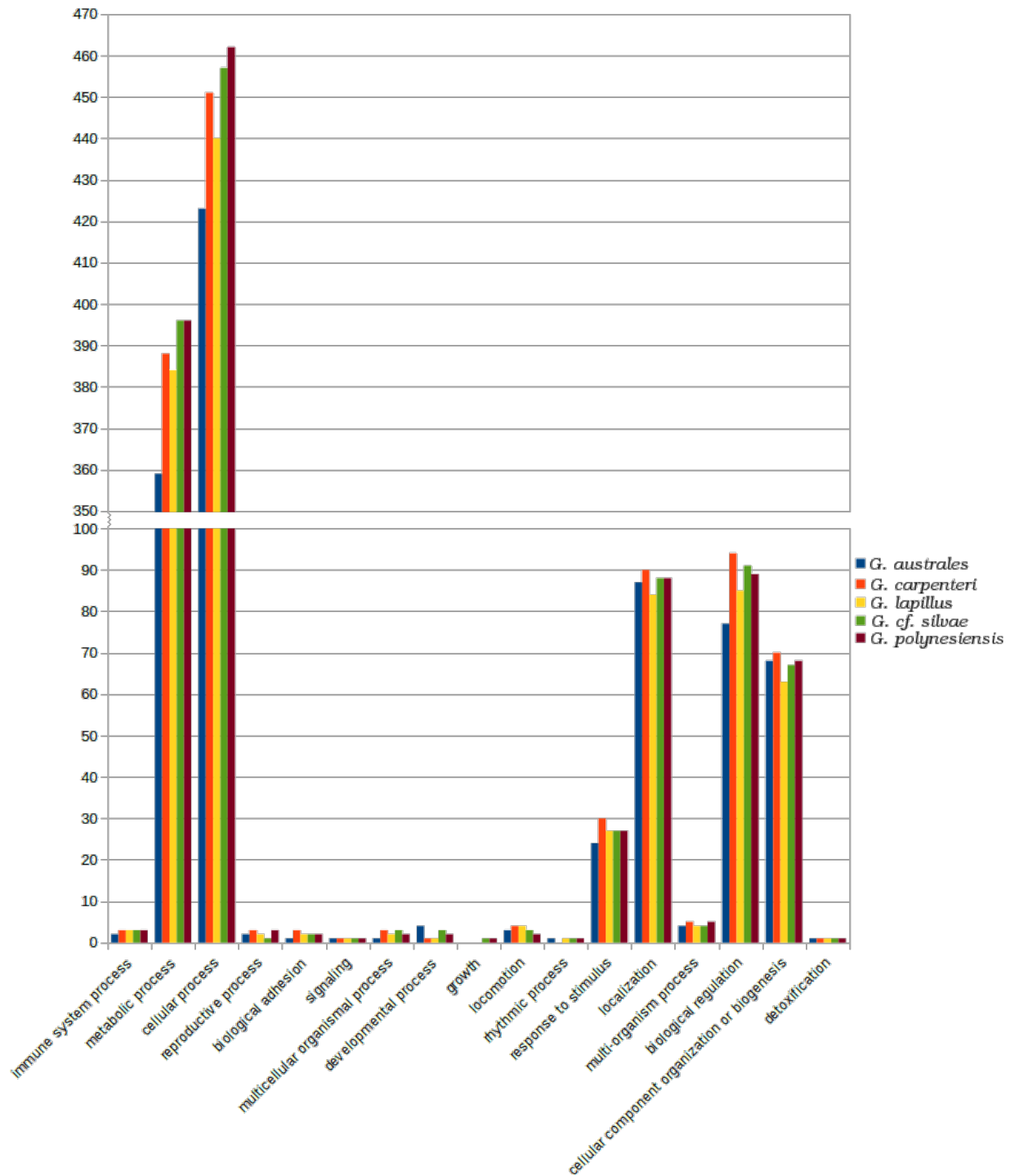


Figure 2: Summary of biological processes GO annotations between *Gambierdiscus* species at GOSUM level 1 from Suppl. table 5.

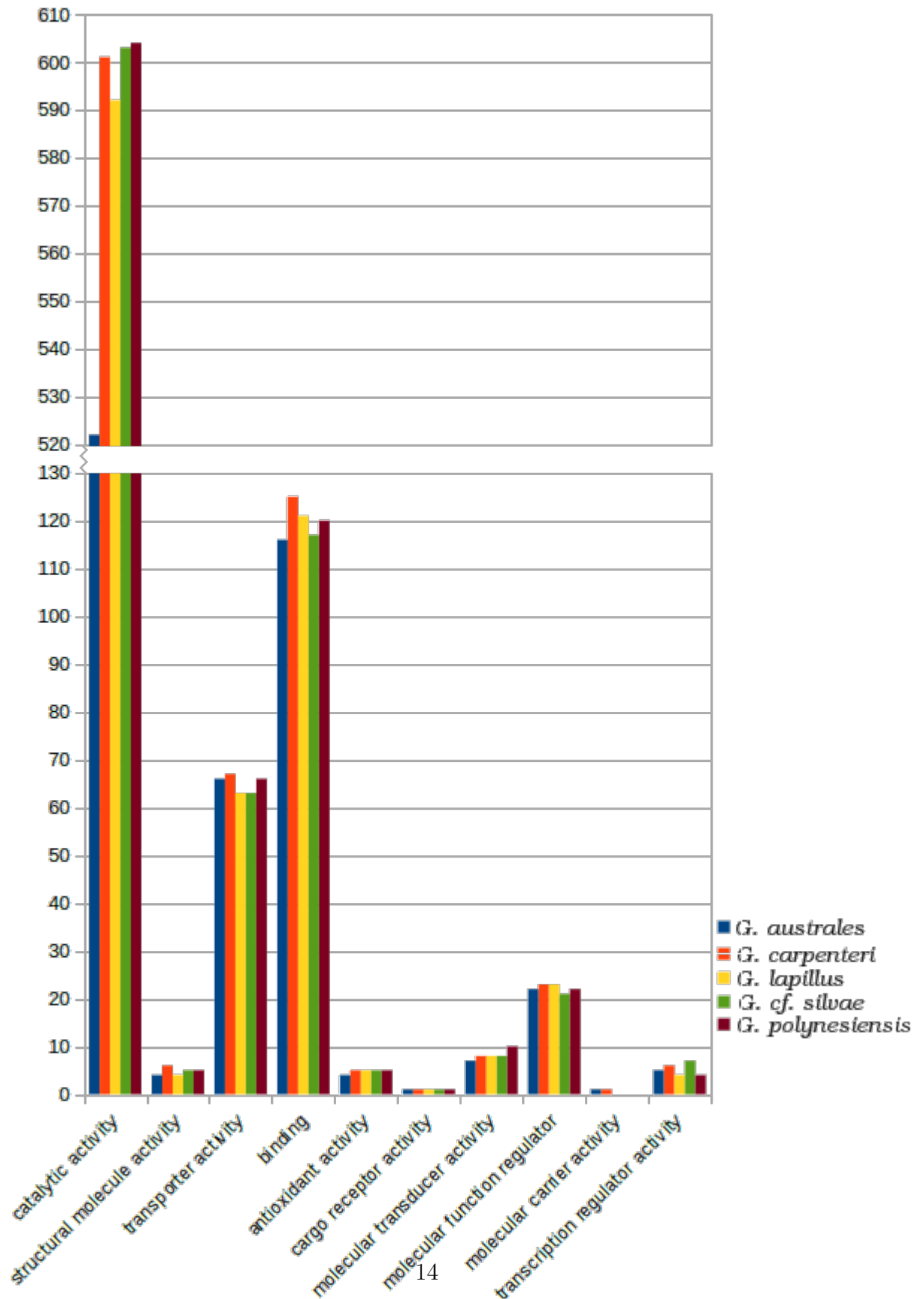


Figure 3: Summary of molecular GO annotations between *Gambierdiscus* species at GOSUM level 1 from Suppl. table 5.

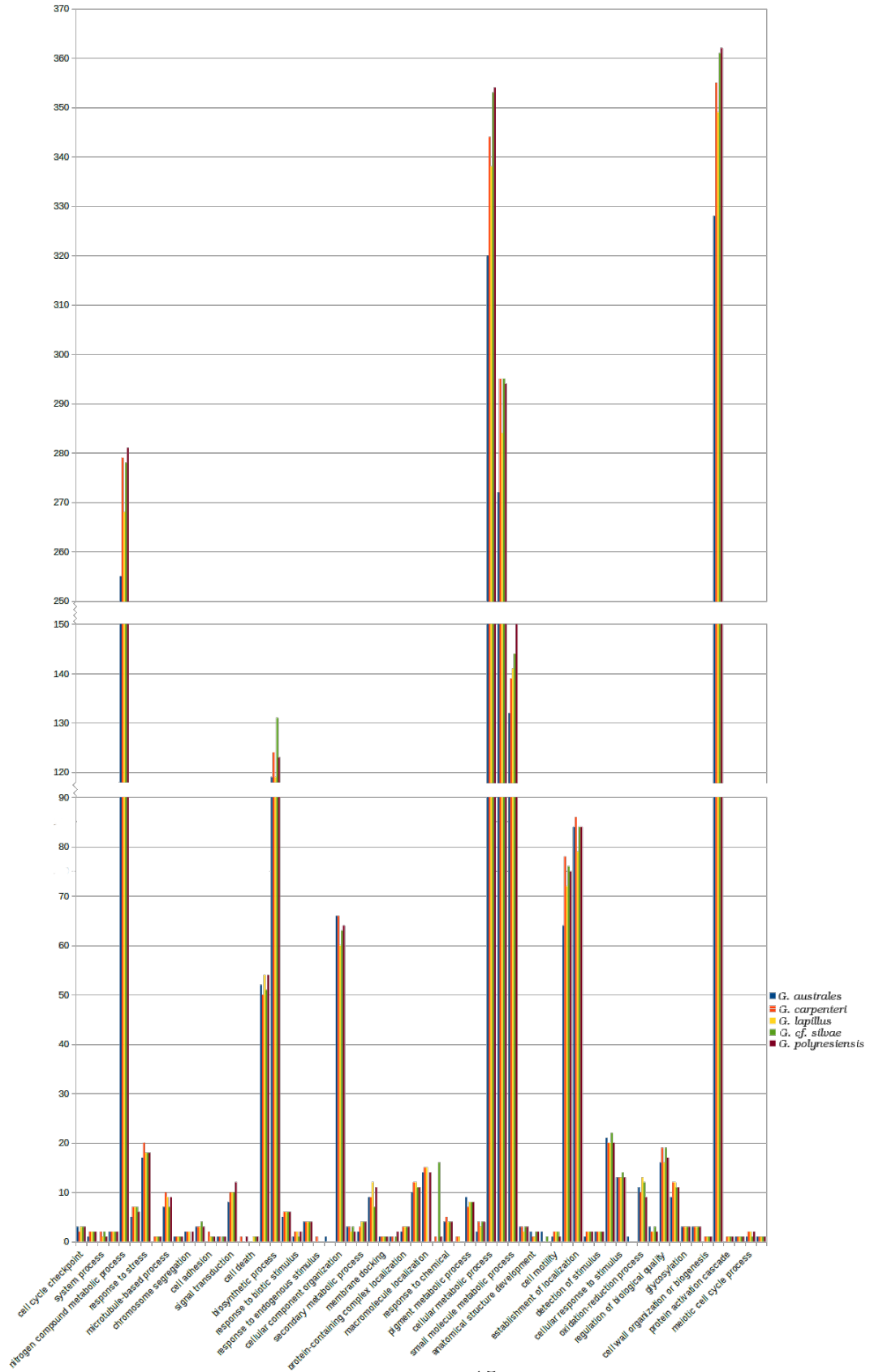


Figure 4: Summary of biological processes GO annotations between *Gambierdiscus* species at GOSUM level 2 from Suppl. table 6.

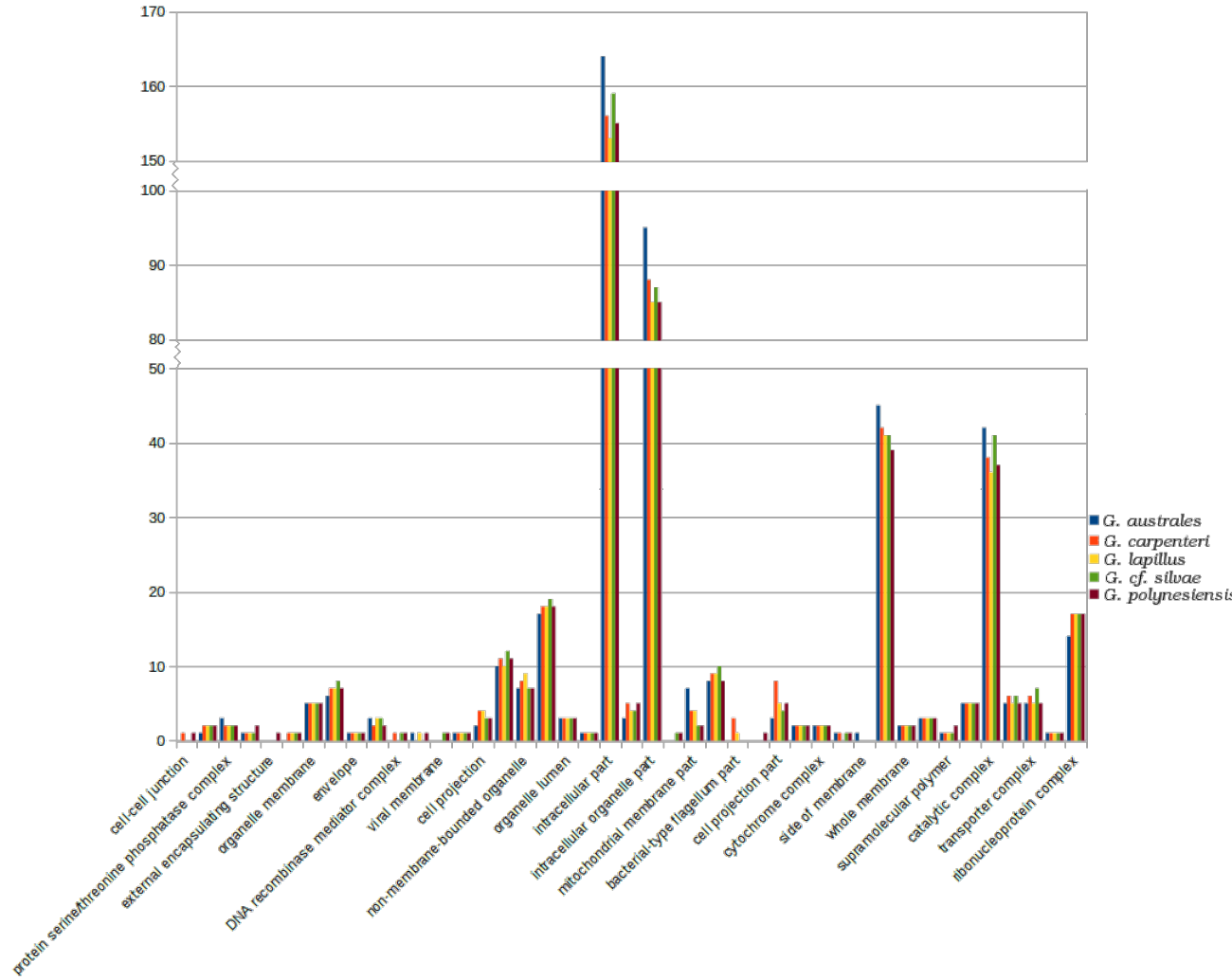


Figure 5: Summary of cellular GO annotations between *Gambierdiscus* species at GO-SUM level 2 from Suppl. table 6.

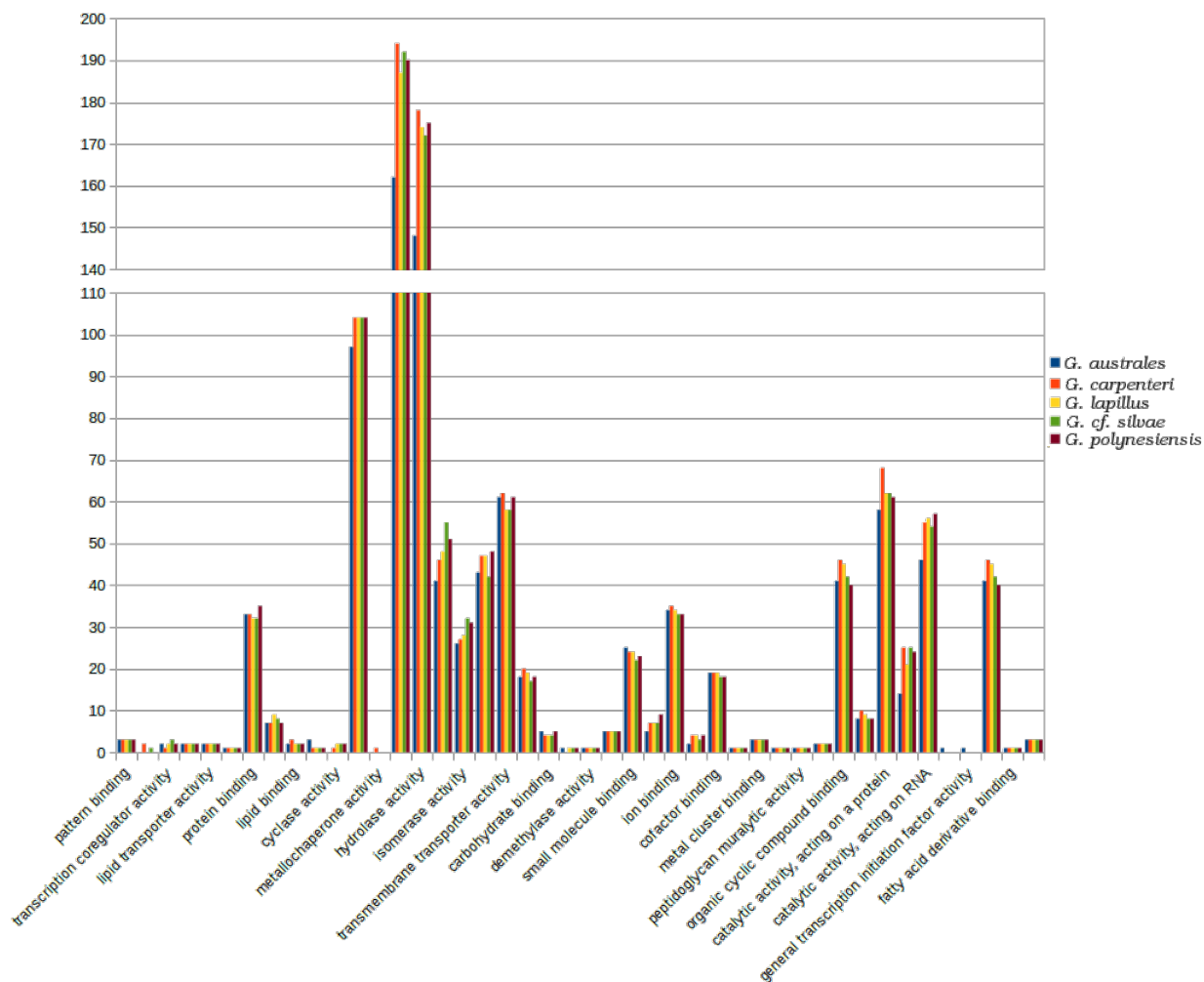


Figure 6: Summary of molecular GO annotations between *Gambierdiscus* species at GOSUM level 2 from Suppl. table 6.

Transcriptome similarity clustering

To do:

- describe differences in graphs once I know what needs to be taken out and re-run
- GOSUM lvl2 graphs are partially missing descriptions on x-axis. Fix when re-run

Potentially interesting points, if still there after bact and unknown outtakes:

- intracellular parts in pan (gosum2 cell)
- organelle memb in core and softcore, seem essential and not in unique (gosum2 cell)
- core and unique pretty evenly matched in most entries for gosum2 molec, except catalytic activity binding on DNA is much higher in unique and a little higher for binding RNA

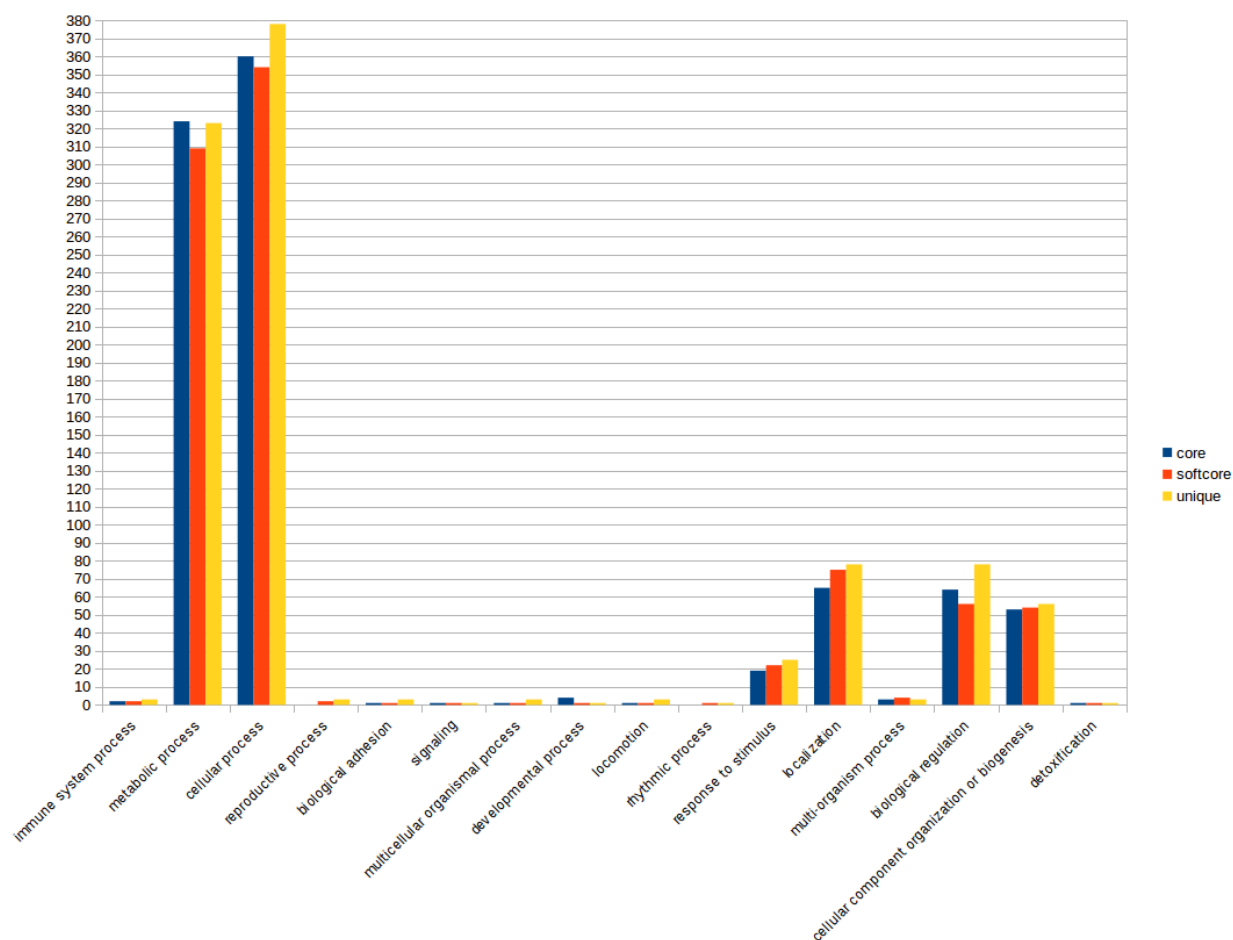


Figure 8: Summary of biological processes GO annotations between core, softcore and unique clusters at GOSUM level 1.

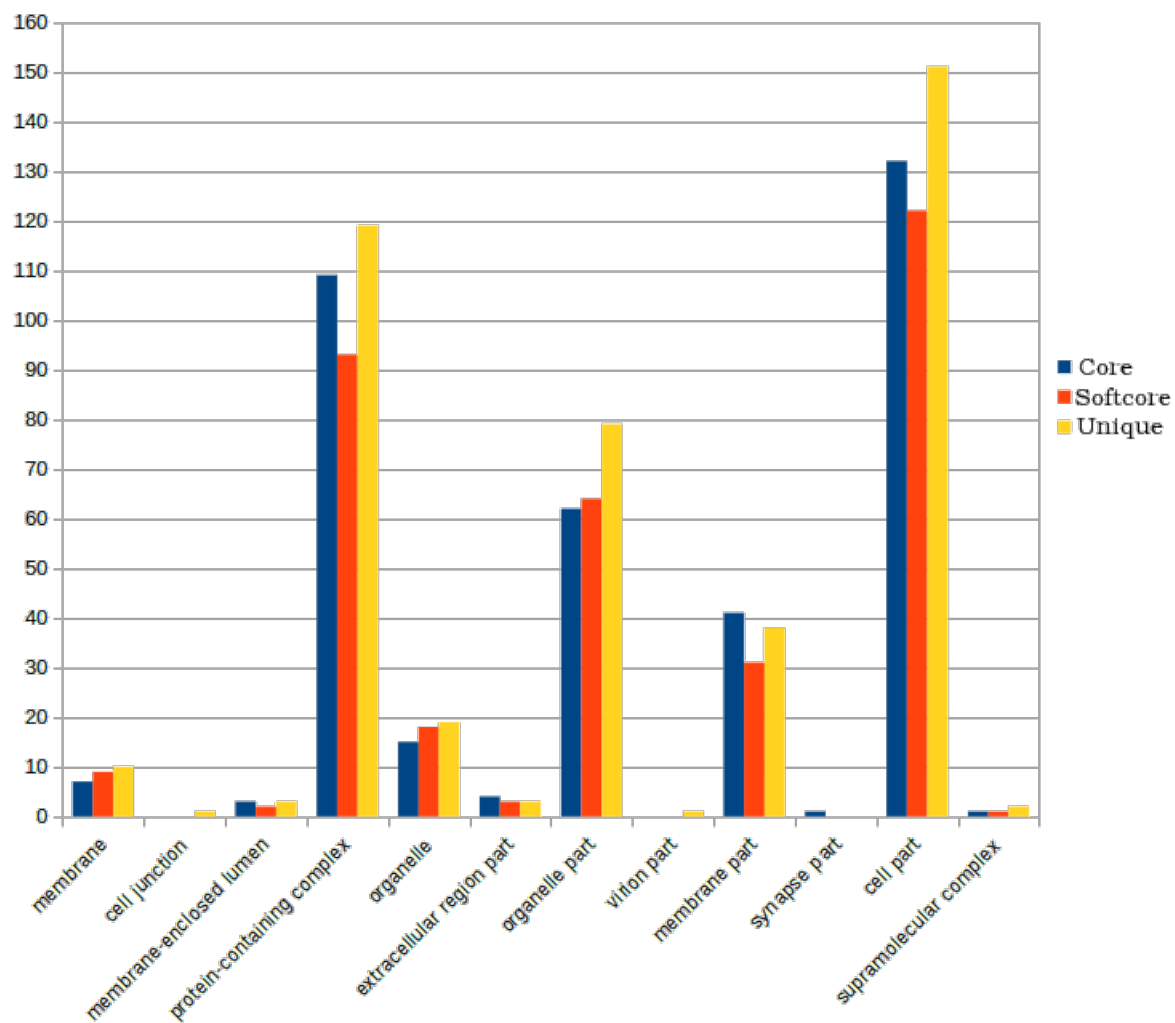


Figure 9: Summary of cellular GO annotations between core, softcore and unique clusters at GOSUM level 1.

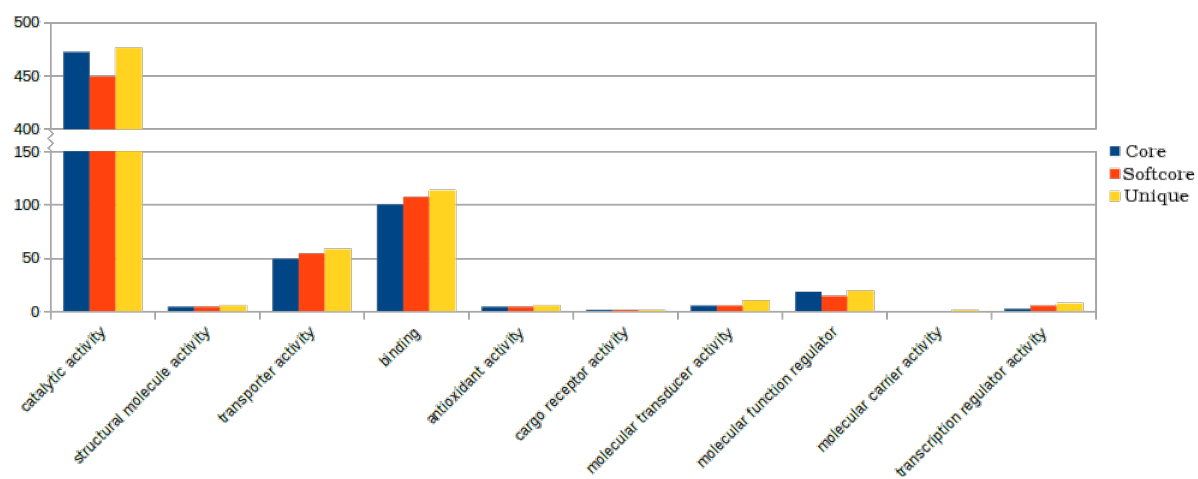


Figure 10: Summary of molecular GO annotations between core, softcore and unique clusters at GOSUM level 1.

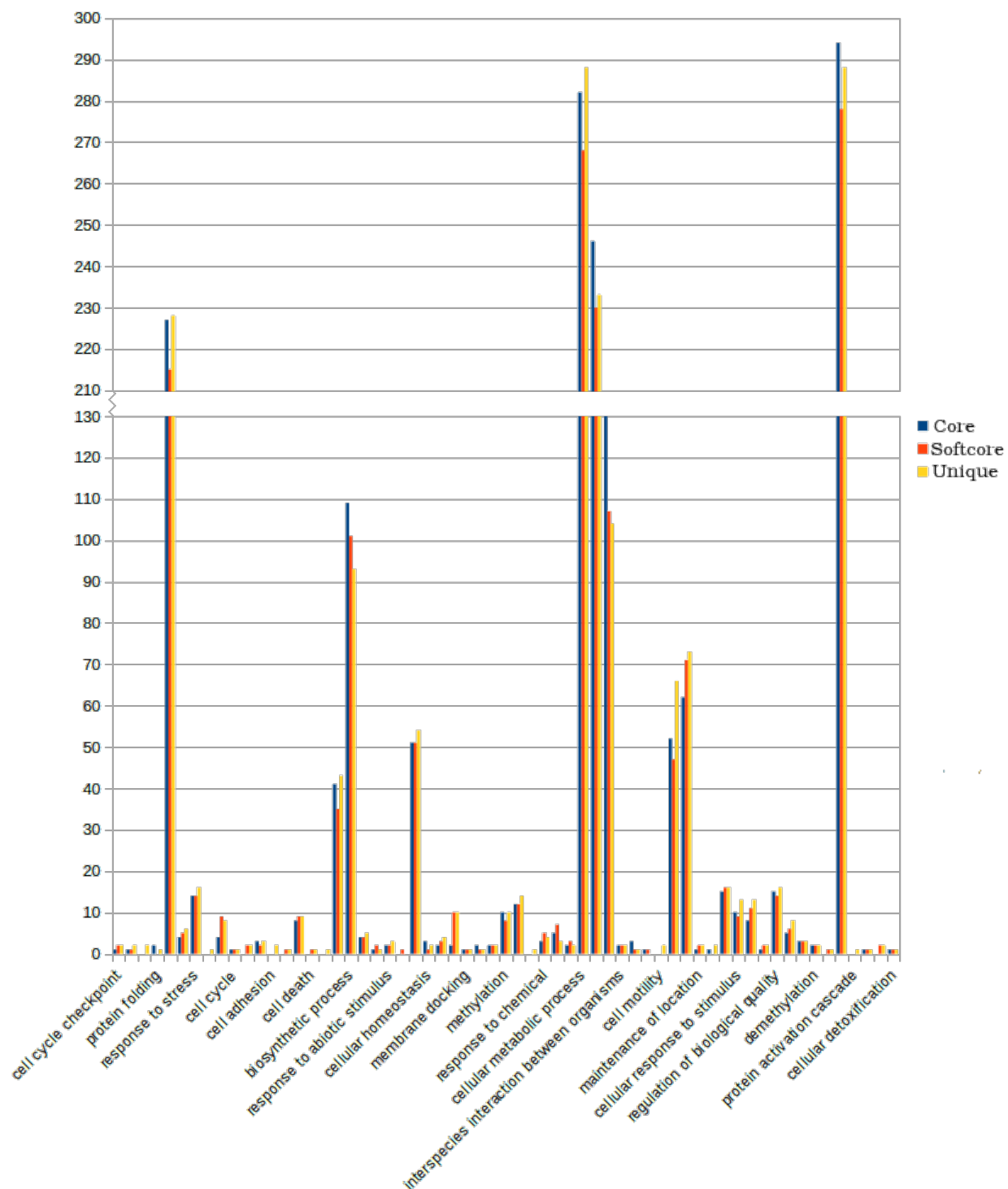


Figure 11: Summary of biological processes GO annotations between core, softcore and unique clusters at GOSUM level 2.

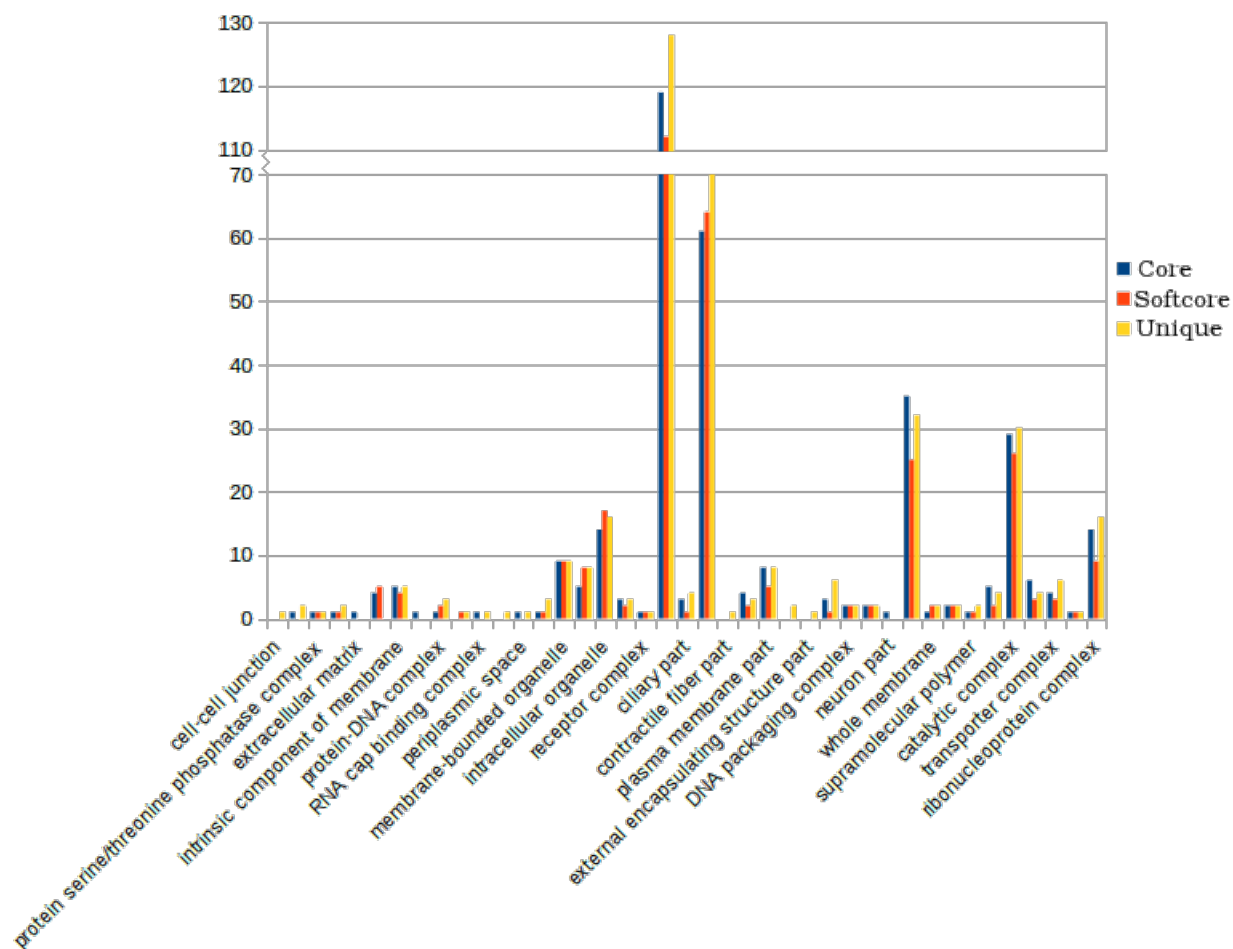


Figure 12: Summary of cellular GO annotations between core, softcore and unique clusters at GOSUM level 2.

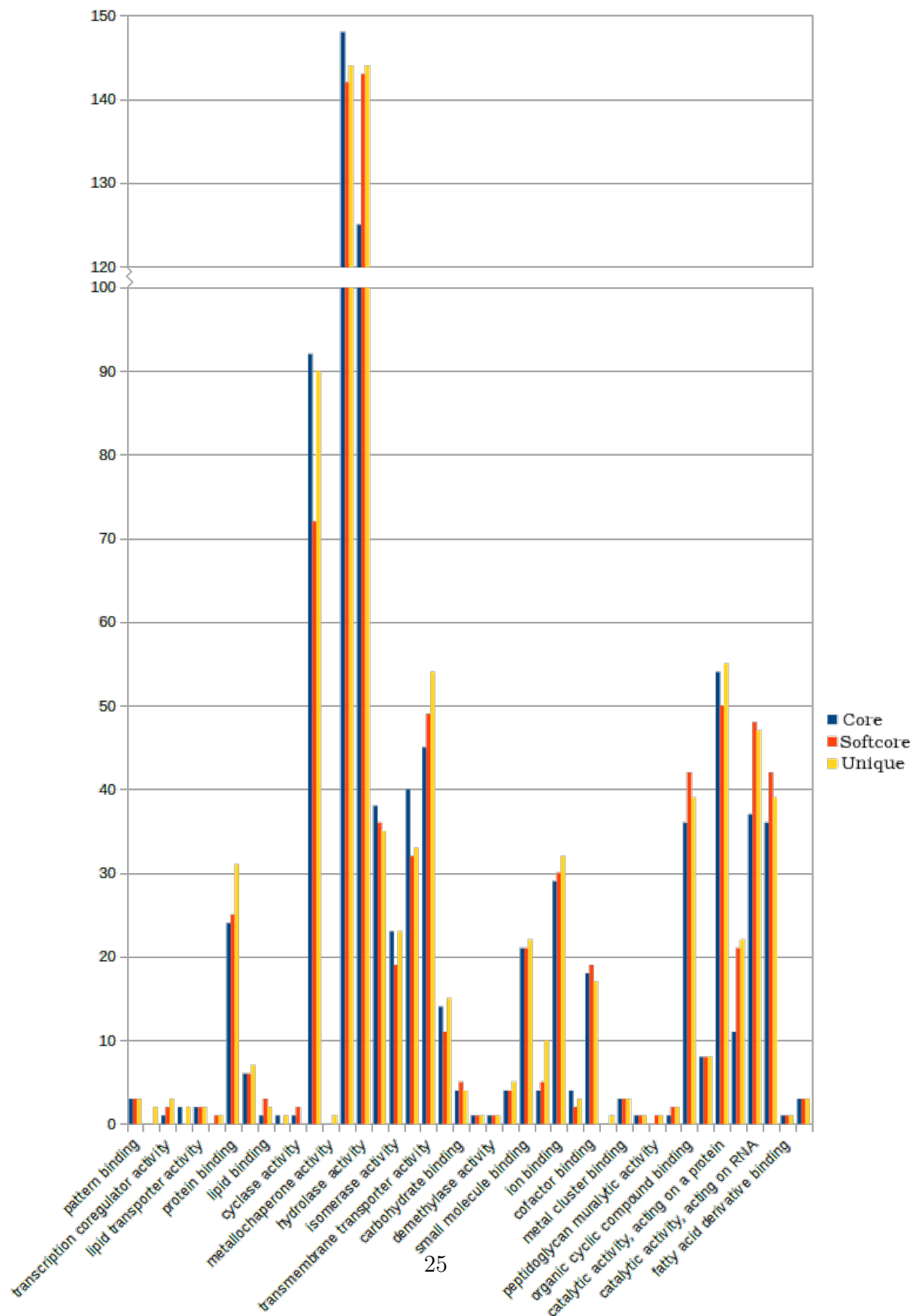


Figure 13: Summary of molecular GO annotations between core, softcore and unique clusters at GOSUM level 2.

Core transcriptome

A set of core genes common to all five species of *Gambierdiscus* were found. This set consisted of 13,750 amino acid clusters (Table 2) of which 45 % were annotated with GO terms (Suppl. table 7 & 8). The highest number of contigs in any core cluster was 180 cluster of unknown function with 23, 45, 32, 31 and 49 from *G. australes*, *G. carpenteri*, *G. lapillus*, *G. polynesiensis* and *G. cf. silvae* respectively. Twelve of the core clusters contained 100 or more contigs, of which 3 were unannotated. The predicted protein coding regions for the other nine clusters, in descending order of contig numbers: an enzyme with catalytic activity involved in metabolic process; a calcium binding transmembrane transport channel; a protein involved in calcium binding; a protein binding enzyme; a domain for unspecified protein binding; an enzyme with O-glucosyl hydrolase activity involved in carbohydrate metabolic process; membrane bound ion transporter with cation channel activity & ionotropic glutamate receptor activity; a transmembrane transporter with voltage-gated calcium channel activity; and calcium ion binding transmembrane ion transporter. A total of 3,943 core clusters contained 10 or more contigs, so 71.32 % of the total core clusters consisted of less than 10 contigs. The majority of clusters fell within metabolic processes, cellular processes and catalytic activity with %, % and % of annotated clusters respectively. **Tim - so adding up the lvl1 gosum counts for bio, cell and molec doesn't add up to the total annotated clusters.. am I correct in thinking that this is because annotations can go to other functions too?**

Softcore transcriptome

A softcore with 4 out of the five *Gambierdiscus* species examined was identified. The softcore consisted of an additional 16,980 clusters (Table 2) of which 48 % were annotated (Suppl. table 7 & 8). The most prolific cluster in the softcore contained 163 contigs with unknown function, where *G. carpenteri*, *G. lapillus*, *G. polynesiensis* and *G. cf. silvae* contained 50, 42, 41 & 30 contigs respectively. A further 5 clusters contained more than 100 contigs, four of which had GO annotations. Of the six clusters with over 100 contigs, none had representatives contigs from *G. australes*. *G. australes* was absent from 86 % of the softcore clusters. In descending order of contigs, they matched to: a protein involved in selective protein binding; a protein involved in actin binding; a

protein involved in calcium binding; and a protein with cysteine-type peptidase activity. Of the softcore, 14,035 clusters contained 10 or more contigs.

Pan-transcriptome

Clusters with single species representatives, or the pan-transcriptome to the five *Gambierdiscus* species examined, numbered 231,310 clusters. Of the unique clusters, only 15.23 % of clusters were annotated. Single species clusters from *G. australes*, *G. carpenteri*, *G. lapillus*, *G. polynesiensis* and *G. cf. silvae* numbered 35,356, 62,494, 32,341, 60,796 & 41,350 clusters respectively (Table 2). The highest number of contigs in a unique cluster were 37, found in two clusters from *G. carpenteri*. One of these was annotated for RNA and metal ion binding activity. Of the unique clusters, 83.1 % contained only one contig and 97.8 % of clusters have 5 contigs or less.

Last common ancestor identification of contigs

THIS IS WITH SWISSPROT ONLY, 20% similarity and only 100bp overlap. trEMBL still running

Table 3: basta stuff found in each *Gambierdiscus* transcriptome during processing.

Species	<i>G. aus- trales</i>	<i>G. carpenteri</i>	<i>G. lapillus</i>	<i>G. polyne- siensis</i>	<i>G. cf. sil- vae</i>
Swissprot hits on contigs	176,000				
BASTA posi- tive ID	60,811	35263			
Eukaryotic origin	10,720	35,263	22,643	32,098	24,096
Bacterial ori- gin	826	2,784	1,799	2,438	1,943
Unknown ori- gin	7,709	22,429	15,471	22,571	17,072

Unknown origin

To do:

- work out if PKS domains are within unknown
- may be bacterial origin - IF they have dinoSL, keep. If not, remove from core/pan analysis

Bacterial origin

To do:

- re-running with uniprot_trembl.fasta to see how percentage identity values differ to swissprot database
- merge trEMBL and swissprot databases and see how BASTA goes in comparison
- check if LCA is specific enough for Proteobacteria or gamma-Proteobacteria regarding Quorum sensing taxa
- make new directory with bacterial origin

- dinoSL search to see if any of bact origin are from dinos
- look if bact contigs found in unique or core clusters
- check if core bacteriome (how wanky is that word) or any species specific
- check for regional link of host association. Lapillus and silvae are from Heron Island from same collection trip, poly and australes are from Rarotonga collected 9 years apart, carp is from temperate Merimbula Merimbula)

Looking into toxin producers

Clusters that don't have *G. carpenteri* in

Rationale: This strain of carpenteri is the only one of the 5 which is a verified non-CTX producer, by LC-MS and bioassay.

To do:

- find clusters excluding carp
- look for clusters with higher number of contigs from poly and silvae as those are the two more toxic ones
- check for dinoSL and LCA of clusters

G. polynesiensis solo clusters

- number of clusters
- percentage annotated
- pathways present (another GOSUM adventure?)
- as *G. silvae* and to a much reduced extend, *G. lapillus*, also produce CTX, is the solo polynesiensis section relevant?

Polyketide synthase active domain search

Table 4: PKS active domains found in the *Gambierdiscus* species queries.

Active domain	<i>G. australes</i>	<i>G. carpenteri</i>	<i>G. lapillus</i>	<i>G. polynesiensis</i>	<i>G. cf. silvae</i>	Total contigs	# clusters
ACP							
AT							
DR							
ET							
KS	130	195	150	221	154	850	314
KR							
TE							

KS domains. A total of 850 contigs were identified with KS domains which assembled into 314 clusters (table 4). Nine clusters contained more than 10 contigs, with the highest number of 130 contigs from all species. 9 clusters contained 10 contigs or more, of which only two did not contain all the taxa examined. 57 of the 314 clusters contained contigs from multiple species, so 81.8 % of KS clusters were species specific while 78.7 % contained only a single contig (Fig. 14). The non-ciguatoxic *G. carpenteri* was absent from 73.6 % of the clusters. Of the clusters without *G. carpenteri*, none contained all four other species. However one cluster contained *G. lapillus*, *G. polynesiensis* and *G. cf. silvae* with equally represented transcript numbers. Four contigs contained *G. polynesiensis* and *G. cf. silvae* only, one of which had a higher contig representation of *G. polynesiensis* than *G. cf. silvae*. *G. polynesiensis* was the only representative species in 71 clusters, of which three clusters contained 2 contigs and one cluster contained 3 contigs. *G. cf. silvae* was representative as the only species in 23 clusters, one of which contained 3 contigs while the other clusters contained single contigs. *G. australes*, *G. carpenteri* and *G. lapillus* were the solo representatives of 81, 39 & 35 KS clusters respectively.

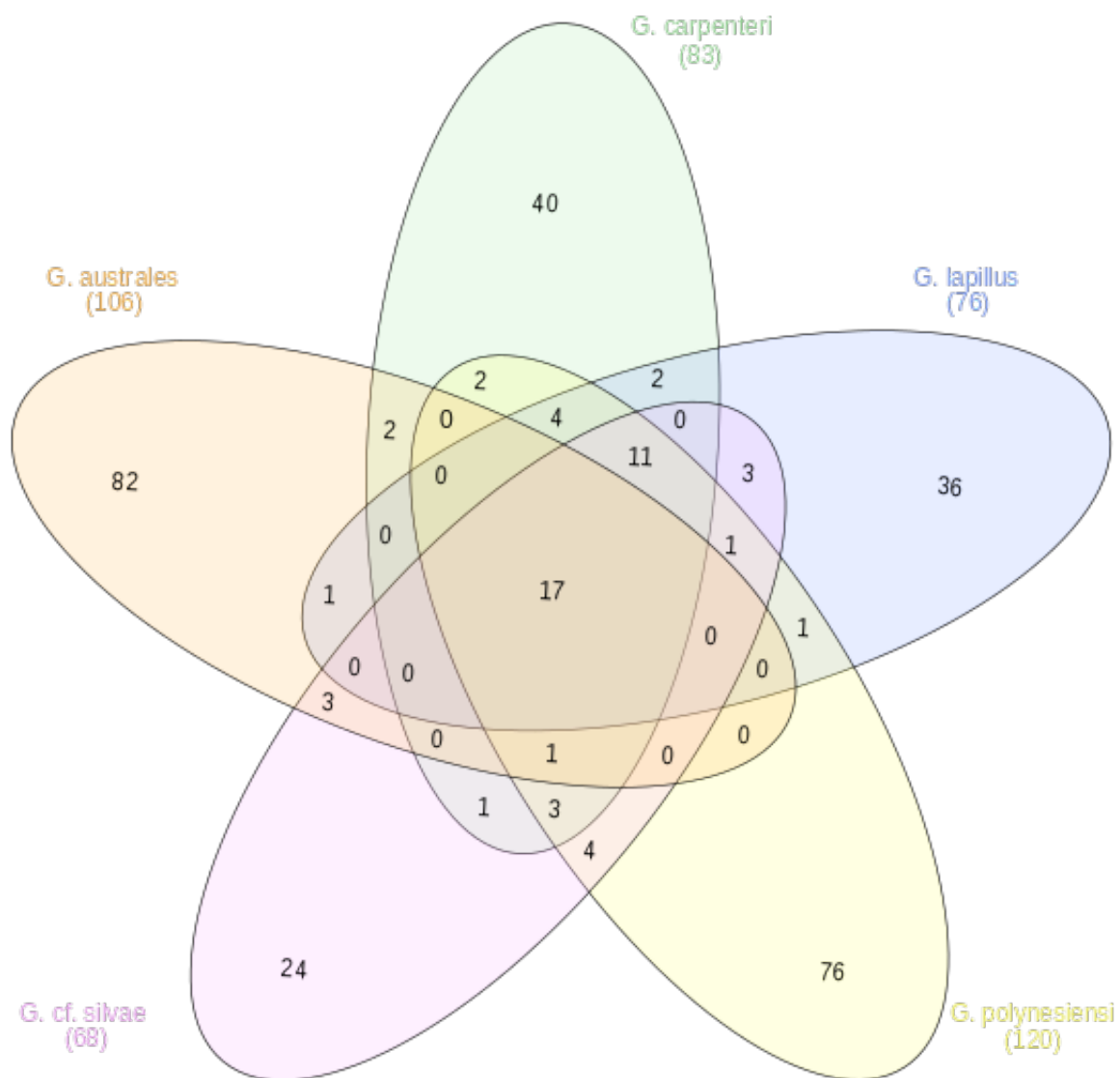


Figure 14: Venn diagram of species in KS clusters.

To do:

- are there any multi-domain transcripts?

Discussion

To go here:

- overall summary of study
- core and pan more likely to be accurate without being axenic unless same contamination
- spliced leader sites really low. potentially interesting - the two highest ones are from same phylogenetic clade, while the other three are representatives from the other two main clades. Also poly and silvae are from separate seq runs, so not an artefact from that front.
- *G. australes* seq is quite bad in comparison as can be seen in the GOSUM figs and the comparative number of contigs, predicted proteins and softcore clusters
- **Tim** not sure if I can do something like Fig 2 - only 5 isolates to put in, and I think I need to look at that again with more sleep to work out what's going on and if I could transfer the concept., there are over 200,000 pan-tran clusters, I don't think I can work out whether they are genophyletic or monophyletic for that many

core *Gambierdiscus* transcriptome

[19] comprehensive index of genes in *K. brevis* to compare to as well as functional summaries

discuss common & different functions found

Koid 14 pan-transcriptome of 4 prymnesiophyte algae. Compare functional findings (KOG vs. this) and contigs as well as predicted protein coding regions are just a fraction of the ones here. eg.30,000-56,000 contigs vs. lowest for in this study is 148,972. Other

study transcriptomes are part of MMETSP, but even australes here is over 100,000 contigs which is from the same study so more likely it's a Gambi thing rather than a seq thing. same with australes and Koid for peptides predicted, almost double. Way higher for other gambis.

Expression of genes involved in polyketide production

- discuss if different gene sets were expressed between toxic and non- toxic strains (ie. not carp)
- discuss KS containing contigs per species plus distribution and number of contigs in KS clusters
- point at Venn diagram intersections that could be of interest for further investigation for both MTX and CTX
- discuss KS conserved region phylogeny
- **Tim** I'm not sure we know enough about these pathways to do something like fig 5

Bacterial association with host

- really depends what the basta results are and if anything interesting is found

discuss usefulness for future studies

- Usefulness of core transcriptome for RNA sequencing studies
- Investigate poly only KS clusters or clusters with high number of poly reps

discuss potential short commings

- from different seq runs and methods and seq depth may vary, especially *G. australes*
- intra-species variation so one isolate per species may not be representative

- unknown if processes other than PKS play a role in toxin production

Conclusion

Supplementary

- need to add australes

Table 5: GO terms and number of contigs per species at GO ontology level 1.

GO accession	GO terms	<i>G. carpenteri</i>	<i>G. lapillus</i>	<i>G. polyne-siensis</i>	<i>G. cf. sil-vae</i>
Biological processes					
GO:0002376	immune system process	3	3	3	3
GO:0008152	metabolic process	388	384	396	396
GO:0009987	cellular process	451	440	457	462
GO:0022414	reproductive process	3	2	1	3
GO:0022610	biological adhesion	3	2	2	2
GO:0023052	signaling	1	1	1	1
GO:0032501	multicellular organismal process	3	2	3	2
GO:0032502	developmental process	1	1	3	2
GO:0040007	growth	0	0	1	1
GO:0040011	locomotion	4	4	3	2
GO:0048511	rhythmic process	0	1	1	1
GO:0050896	response to stimulus	30	27	27	27
GO:0051179	localization	90	84	88	88
GO:0051704	multi-organism process	5	4	4	5
GO:0065007	biological regulation	94	85	91	89

GO:0071840	cellular component organization/biogenesis	70	63	67	68
GO:0098754	detoxification	1	1	1	1
Cellular components					
GO:0016020	membrane	9	9	9	10
GO:0030054	cell junction	1	0	0	1
GO:0031974	membrane-enclosed lumen	3	3	3	3
GO:0032991	protein-containing complex	146	141	145	144
GO:0043226	organelle	22	22	22	21
GO:0044421	extracellular region part	4	4	4	4
GO:0044422	organelle part	94	88	89	88
GO:0044423	virion part	1	0	1	1
GO:0044425	membrane part	50	49	50	47
GO:0044456	synapse part	1	0	1	1
GO:0044464	cell part	182	174	178	177
GO:0099080	supramolecular complex	1	1	1	2
Molecular function					
GO:0003824	catalytic activity	601	592	603	604
GO:0005198	structural molecule activity	6	4	5	5
GO:0005215	transporter activity	67	63	63	66
GO:0005488	binding	125	121	117	120
GO:0016209	antioxidant activity	5	5	5	5
GO:0038024	cargo receptor activity	1	1	1	1
GO:0060089	molecular transducer activity	8	8	8	10

GO:0098772	molecular function regulator	23	23	21	22
GO:0140104	molecular carrier activity	1	0	0	0
GO:0140110	transcription regulator activity	6	4	7	4

Table 6: GO terms and number of contigs per species at GO ontology level 2, child terms of Table 5.

GO accession	GO terms	<i>G. carpen- teri</i>	<i>G. lapillus</i>	<i>G. polyne- siensis</i>	<i>G. cf. sil- vae</i>
Biological processes					
GO:0000075	cell cycle checkpoint	2	3	3	3
GO:0002252	immune effector process	2	2	2	2
GO:0003008	system process	2	1	2	1
GO:0006457	protein folding	2	2	2	2
GO:0006807	nitrogen compound metabolic process	279	268	278	281
GO:0006928	movement of cell or subcellular component	7	7	7	6
GO:0006950	response to stress	20	18	18	18
GO:0006955	immune response	1	1	1	1
GO:0007017	microtubule-based process	10	9	7	9
GO:0007049	cell cycle	1	1	1	1
GO:0007059	chromosome segregation	2	2	0	2
GO:0007154	cell communication	3	3	4	3

GO:0007155	cell adhesion	2	1	1	1
GO:0007163	establishment or maintenance of cell polarity	1	0	1	1
GO:0007165	signal transduction	10	10	10	12
GO:0008037	cell recognition	1	0	0	1
GO:0008219	cell death	0	1	1	1
GO:0009056	catabolic process	50	54	51	54
GO:0009058	biosynthetic process	124	119	131	123
GO:0009605	response to external stimulus	6	6	6	6
GO:0009607	response to biotic stimulus	2	2	1	2
GO:0009628	response to abiotic stimulus	4	4	4	4
GO:0009719	response to endoge- nous stimulus	1	0	0	0
GO:0016043	cellular component or- ganization	66	60	63	64
GO:0019725	cellular homeostasis	3	2	3	2
GO:0019748	secondary metabolic process	3	4	4	4
GO:0022402	cell cycle process	9	12	7	11
GO:0022406	membrane docking	1	1	1	1
GO:0030029	actin filament-based process	1	0	1	2
GO:0031503	protein-containing complex localization	3	3	3	3
GO:0032259	methylation	12	12	11	11
GO:0033036	macromolecule local- ization	15	15	0	14

GO:0035036	sperm-egg recognition	1	0	16	1
GO:0042221	response to chemical	5	4	4	4
GO:0042330	taxis	1	1	0	0
GO:0042440	pigment metabolic process	7	8	8	8
GO:0044085	cellular component biogenesis	4	3	4	4
GO:0044237	cellular metabolic process	344	338	353	354
GO:0044238	primary metabolic process	295	284	295	294
GO:0044281	small molecule metabolic process	139	141	144	150
GO:0044419	interspecies interaction between organisms	3	2	3	3
GO:0048856	anatomical structure development	1	1	2	2
GO:0048869	cellular developmental process	0	0	1	
GO:0048870	cell motility	2	2	2	1
GO:0050789	regulation of biological process	78	72	76	75
GO:0051234	establishment of localization	86	79	84	84
GO:0051235	maintenance of location	2	2	2	2
GO:0051606	detection of stimulus	2	2	2	2
GO:0051641	cellular localization	20	20	22	20
GO:0051716	cellular response to stimulus	13	13	14	13

GO:0055114	oxidation-reduction process	10	13	12	9
GO:0061919	process utilizing autophagic mechanism	2	2	3	2
GO:0065008	regulation of biological quality	19	16	19	17
GO:0065009	regulation of molecular function	12	12	11	11
GO:0070085	glycosylation	3	3	3	3
GO:0070988	demethylation	3	3	3	3
GO:0071554	cell wall organization or biogenesis	1	1	1	1
GO:0071704	organic substance metabolic process	355	349	361	362
GO:0072376	protein activation cascade	1	1	1	1
GO:0140029	exocytic process	1	1	1	1
GO:1903046	meiotic cell cycle process	2	2	1	2
GO:1990748	cellular detoxification	1	1	1	1
Cellular components					
GO:0005911	cell-cell junction	1	0	0	1
GO:0005929	cilium	2	2	2	2
GO:0008287	protein serine/threonine phosphatase complex	2	2	2	2
GO:0019867	outer membrane	1	1	1	2
GO:0030312	external encapsulating structure	0	0	0	1
GO:0031012	extracellular matrix	1	1	1	1
GO:0031090	organelle membrane	5	5	5	5

GO:0031224	intrinsic component of membrane	7	7	8	7
GO:0031975	envelope	1	1	1	1
GO:0032993	protein-DNA complex	2	3	3	2
GO:0033061	DNA recombinase mediator complex	1	0	1	1
GO:0034518	RNA cap binding complex	0	1	0	1
GO:0036338	viral membrane	0	0	1	1
GO:0042597	periplasmic space	1	1	1	1
GO:0042995	cell projection	4	4	3	3
GO:0043227	membrane-bounded organelle	11	10	12	11
GO:0043228	non-membrane-bounded organelle	8	9	7	7
GO:0043229	intracellular organelle	18	18	19	18
GO:0043233	organelle lumen	3	3	3	3
GO:0043235	receptor complex	1	1	1	1
GO:0044424	intracellular part	156	153	159	155
GO:0044441	ciliary part	5	4	4	5
GO:0044446	intracellular organelle part	88	85	87	85
GO:0044449	contractile fiber part	0	0	1	1
GO:0044455	mitochondrial membrane part	4	4	2	2
GO:0044459	plasma membrane part	9	9	10	8
GO:0044461	bacterial-type flagellum part	3	1	0	0
GO:0044462	external encapsulating structure part	0	0	0	1

GO:0044463	cell projection part	8	5	4	5
GO:0044815	DNA packaging complex	2	2	2	2
GO:0070069	cytochrome complex	2	2	2	2
GO:0097458	neuron part	1	0	1	1
GO:0098796	membrane protein complex	42	41	41	39
GO:0098805	whole membrane	2	2	2	2
GO:0099023	tethering complex	3	3	3	3
GO:0099081	supramolecular polymer	1	1	1	2
GO:0120114	Sm-like protein family complex	5	5	5	5
GO:1902494	catalytic complex	38	36	41	37
GO:1990204	oxidoreductase complex	6	5	6	5
GO:1990351	transporter complex	6	5	7	5
GO:1990391	DNA repair complex	1	1	1	1
GO:1990904	ribonucleoprotein complex	17	17	17	17
Molecular function					
GO:0001871	pattern binding	3	3	3	3
GO:0003700	DNA-binding transcription factor activity	2	0	1	0
GO:0003712	transcription coregulator activity	1	2	3	2
GO:0004133	glycogen debranching enzyme activity	2	2	2	2
GO:0005319	lipid transporter activity	2	2	2	2

GO:0005326	neurotransmitter transporter activity	1	1	1	1
GO:0005515	protein binding	33	32	32	35
GO:0008144	drug binding	7	9	8	7
GO:0008289	lipid binding	3	2	2	2
GO:0008565	protein transporter activity	1	1	1	1
GO:0009975	cyclase activity	1	2	2	2
GO:0016491	oxidoreductase activity	104	104	104	104
GO:0016530	metallochaperone activity	1	0	0	0
GO:0016740	transferase activity	194	187	192	190
GO:0016787	hydrolase activity	178	174	172	175
GO:0016829	lyase activity	46	48	55	51
GO:0016853	isomerase activity	27	28	32	31
GO:0016874	ligase activity	47	47	42	48
GO:0022857	transmembrane transporter activity	62	58	58	61
GO:0030234	enzyme regulator activity	20	19	17	18
GO:0030246	carbohydrate binding	4	4	4	5
GO:0030545	receptor regulator activity	0	1	1	1
GO:0032451	demethylase activity	1	1	1	1
GO:0033218	amide binding	5	5	5	5
GO:0036094	small molecule binding	24	24	22	23
GO:0038023	signaling receptor activity	7	7	7	9
GO:0043167	ion binding	35	34	33	33

GO:0044877	protein-containing complex binding	4	4	3	4
GO:0048037	cofactor binding	19	19	18	18
GO:0050824	water binding	1	1	1	1
GO:0051540	metal cluster binding	3	3	3	3
GO:0060090	molecular adaptor activity	1	1	1	1
GO:0061783	peptidoglycan murelytic activity	1	1	1	1
GO:0072341	modified amino acid binding	2	2	2	2
GO:0097159	organic cyclic compound binding	46	45	42	40
GO:0097367	carbohydrate derivative binding	10	9	8	8
GO:0140096	catalytic activity, acting on a protein	68	62	62	61
GO:0140097	catalytic activity, acting on DNA	25	21	25	24
GO:0140098	catalytic activity, acting on RNA	55	56	54	57
GO:1901363	heterocyclic compound binding	46	45	42	40
GO:1901567	fatty acid derivative binding	1	1	1	1
GO:1901681	sulfur compound binding	3	3	3	3

Table 7: GO terms and number of contigs found in core, softcore and pan-transcriptome of *Gambierdiscus* at GO ontology level 1.

GO accession	GO terms	Core	Softcore	Pan
Biological processes				
GO:0002376	immune system process	2	2	3
GO:0008152	metabolic process	324	309	323
GO:0009987	cellular process	360	354	378
GO:0022414	reproductive process	0	2	3
GO:0022610	biological adhesion	1	1	3
GO:0023052	signaling	1	1	1
GO:0032501	multicellular organismal process	1	1	3
GO:0032502	developmental process	4	1	1
GO:0040011	locomotion	1	1	3
GO:0048511	rhythmic process	0	1	1
GO:0050896	response to stimulus	19	22	25
GO:0051179	localization	65	75	78
GO:0051704	multi-organism process	3	4	3
GO:0065007	biological regulation	64	56	78
GO:0071840	cellular component organization or biogenesis	53	54	56
GO:0098754	detoxification	1	1	1
Cellular components				
GO:0016020	membrane	7	9	10
GO:0030054	cell junction	0	0	1

GO:0031974	membrane-enclosed lumen	3	2	3
GO:0032991	protein-containing complex	109	93	119
GO:0043226	organelle	15	18	19
GO:0044421	extracellular region part	4	3	3
GO:0044422	organelle part	62	64	79
GO:0044423	virion part	0	0	1
GO:0044425	membrane part	41	31	38
GO:0044456	synapse part	1	0	0
GO:0044464	cell part	132	122	151
GO:0099080	supramolecular complex	1	1	2
Molecular function				
GO:0003824	catalytic activity	472	449	476
GO:0005198	structural molecule activity	4	4	5
GO:0005215	transporter activity	49	54	58
GO:0005488	binding	100	107	113
GO:0016209	antioxidant activity	4	4	5
GO:0038024	cargo receptor activity	1	1	1
GO:0060089	molecular transducer activity	5	5	10
GO:0098772	molecular function regulator	18	14	19
GO:0140104	molecular carrier activity	0	0	1
GO:0140110	transcription regulator activity	2	5	7

Table 8: GO terms and number of contigs found in core, softcore and pan-transcriptome of *Gambierdiscus* at GO ontology level 2, childer to Table 7.

GO accession	GO terms	Core	Softcore	Pan
Biological processes				
GO:0000075	cell cycle checkpoint	1	2	2
GO:0002252	immune effector process	1	1	2
GO:0003008	system process	0	0	2
GO:0006457	protein folding	2	0	1
GO:0006807	nitrogen compound metabolic process	227	215	228
GO:0006928	movement of cell or subcellular component	4	5	6
GO:0006950	response to stress	14	14	16
GO:0006955	immune response	0	0	1
GO:0007017	microtubule-based process	4	9	8
GO:0007049	cell cycle	1	1	1
GO:0007059	chromosome segregation	0	2	2
GO:0007154	cell communication	3	2	3
GO:0007155	cell adhesion	0	0	2
GO:0007163	establishment or maintenance of cell polarity	0	1	1
GO:0007165	signal transduction	8	9	9
GO:0008037	cell death	0	1	1
GO:0008219	cell death	0	0	1

GO:0009056	catabolic process	41	35	43
GO:0009058	biosynthetic process	109	101	93
GO:0009605	response to external stimulus	4	4	5
GO:0009607	response to biotic stimulus	1	2	1
GO:0009628	response to abiotic stimulus	2	2	3
GO:0009719	response to endogenous stimulus	0	1	0
GO:0016043	cellular component organization	51	51	54
GO:0019725	cellular homeostasis	3	1	2
GO:0019748	secondary metabolic process	2	3	4
GO:0022402	cell cycle process	2	10	10
GO:0022406	membrane docking	1	1	1
GO:0030029	actin filament-based process	2	1	1
GO:0031503	protein-containing complex localization	2	2	2
GO:0032259	methylation	10	8	10
GO:0033036	macromolecule localization	12	12	14
GO:0035036	sperm-egg recognition	0	0	1
GO:0042221	response to chemical	3	5	4
GO:0042440	pigment metabolic process	5	7	3
GO:0044085	cellular component biogenesis	2	3	2

GO:0044237	cellular metabolic process	282	268	288
GO:0044238	primary metabolic process	246	230	233
GO:0044281	small molecule metabolic process	130	107	104
GO:0044419	interspecies interaction between organisms	2	2	2
GO:0048856	anatomical structure development	3	1	1
GO:0048869	cellular developmental process	1	1	0
GO:0048870	cell motility	0	0	2
GO:0050789	regulation of biological process	52	47	66
GO:0051234	establishment of localization	62	71	73
GO:0051235	maintenance of location	1	2	2
GO:0051606	detection of stimulus	1	0	2
GO:0051641	cellular localization	15	16	16
GO:0051716	cellular response to stimulus	10	9	13
GO:0055114	oxidation-reduction process	8	11	13
GO:0061919	process utilizing autophagic mechanism	1	2	2
GO:0065008	regulation of biological quality	15	14	16

GO:0065009	regulation of molecular function	5	6	8
GO:0070085	glycosylation	3	3	3
GO:0070988	demethylation	2	2	2
GO:0071554	cell wall organization or biogenesis	0	1	1
GO:0071704	organic substance metabolic process	294	278	288
GO:0072376	protein activation cascade	0	0	1
GO:0140029	exocytic process	1	1	1
GO:1903046	meiotic cell cycle process	0	2	2
GO:1990748	cellular detoxification	1	1	1
Cellular components				
GO:0005911	cell-cell junction	0	0	1
GO:0005929	cilium	1	0	2
GO:0008287	protein serine/threonine phosphatase complex	1	1	1
GO:0019867	outer membrane	1	1	2
GO:0031090	extracellular matrix	1	0	0
GO:0031090	organelle membrane	4	5	0
GO:0031224	intrinsic component of membrane	5	4	5
GO:0031975	envelope	1	0	0
GO:0032993	protein-DNA complex	1	2	3
GO:0033061	DNA recombinase mediator complex	0	1	1
GO:0034518	RNA cap binding complex	1	0	1

GO:0036338	viral membrane	0	0	1
GO:0042597	periplasmic space	1	0	1
GO:0042995	cell projection	1	1	3
GO:0043227	membrane-bounded organelle	9	9	9
GO:0043228	non-membrane-bounded organelle	5	8	8
GO:0043229	intracellular organelle	14	17	16
GO:0043233	organelle lumen	3	2	3
GO:0043235	receptor complex	1	1	1
GO:0044424	intracellular part	119	112	128
GO:0044441	ciliary part	3	1	4
GO:0044446	intracellular organelle part	61	64	74
GO:0044449	contractile fiber part	0	0	1
GO:0044455	mitochondrial membrane part	4	2	3
GO:0044459	plasma membrane part	8	5	8
GO:0044461	bacterial-type flagellum part	0	0	2
GO:0044462	external encapsulating structure part	0	0	1
GO:0044463	cell projection part	3	1	6
GO:0044815	DNA packaging complex	2	2	2
GO:0070069	cytochrome complex	2	2	2
GO:0097458	neuron part	1	0	0
GO:0098796	membrane protein complex	35	25	32
GO:0098805	whole membrane	1	2	2

GO:0099023	tethering complex	2	2	2
GO:0099081	supramolecular polymer	1	1	2
GO:0120114	Sm-like protein family complex	5	2	4
GO:1902494	catalytic complex	29	26	30
GO:1990204	oxidoreductase complex	6	3	4
GO:1990351	transporter complex	4	3	6
GO:1990391	DNA repair complex	1	1	1
GO:1990904	ribonucleoprotein complex	14	9	16
Molecular function				
GO:0001871	pattern binding	3	3	3
GO:0003700	DNA-binding transcription factor activity	0	0	2
GO:0003712	transcription coregulator activity	1	2	3
GO:0004133	glycogen debranching enzyme activity	2	0	2
GO:0005319	lipid transporter activity	2	2	2
GO:0005326	neurotransmitter transporter activity	0	1	1
GO:0005515	protein binding	24	25	31
GO:0008144	drug binding	6	6	7
GO:0008289	lipid binding	1	3	2
GO:0008565	protein transporter activity	1	0	1
GO:0009975	cyclase activity	1	2	0

GO:0016491	oxidoreductase activity	92	72	90
GO:0016530	metallochaperone activity	0	0	1
GO:0016740	transferase activity	148	142	144
GO:0016787	hydrolase activity	125	143	144
GO:0016829	lyase activity	38	36	35
GO:0016853	isomerase activity	23	19	23
GO:0016874	ligase activity	40	32	33
GO:0022857	transmembrane transporter activity	45	49	54
GO:0030234	enzyme regulator activity	14	11	15
GO:0030246	carbohydrate binding	4	5	4
GO:0030545	receptor regulator activity	1	1	1
GO:0032451	demethylase activity	1	1	1
GO:0033218	amide binding	4	4	5
GO:0036094	small molecule binding	21	21	22
GO:0038023	signaling receptor activity	4	5	10
GO:0043167	ion binding	29	30	32
GO:0044877	protein-containing complex binding	4	2	3
GO:0048037	cofactor binding	18	19	17
GO:0050824	water binding	0	0	1
GO:0051540	metal cluster binding	3	3	3
GO:0060090	molecular adaptor activity	1	1	1

GO:0061783	peptidoglycan mura- lytic activity	0	1	1
GO:0072341	modified amino acid binding	1	2	2
GO:0097159	organic cyclic com- pound binding	36	42	39
GO:0097367	carbohydrate deriva- tive binding	8	8	8
GO:0140096	catalytic activity, act- ing on a protein	54	50	55
GO:0140097	catalytic activity, act- ing on DNA	11	21	22
GO:0140098	catalytic activity, act- ing on RNA	37	48	47
GO:1901363	heterocyclic com- pound binding	36	42	39
GO:1901567	fatty acid derivative binding	1	1	1
GO:1901681	sulfur compound binding	3	3	3

Table 9: KS domains found per cluster and total number of contigs present.

Cluster ID	<i>G. aus- trales</i>	<i>G. carpenteri</i>	<i>G. lapillus</i>	<i>G. polyne- siensis</i>	<i>G. cf. sil- vae</i>	Total contigs
988	6	40	29	24	31	130
8866	3	24	14	24	16	81
3681	7	14	16	9	12	58
1921	3	10	6	4	6	29
46550	3	4	1	8	5	21

215601	0	4	1	8	5	18
360	1	4	3	3	4	15
15645	4	2	0	4	1	11
132980	0	1	4	3	2	10
45086	1	3	1	1	3	9
78009	0	2	2	3	2	9
38915	2	2	2	1	2	9
109763	0	2	0	5	1	8
37859	2	2	1	2	1	8
24847	1	1	1	3	2	8
162333	0	2	2	2	1	7
52333	1	2	1	1	1	6
136782	0	1	2	1	2	6
301971	0	0	2	2	2	6
152898	0	3	1	1	0	5
117472	0	2	1	1	1	5
196360	0	2	1	1	1	5
145445	0	1	1	2	1	5
131919	0	1	0	1	3	5
59207	1	1	1	1	1	5
31669	1	1	1	1	1	5
55678	1	1	1	1	1	5
40462	1	1	1	1	1	5
46899	1	1	1	1	1	5
37886	1	1	1	1	1	5
475329	0	0	0	4	1	5
162320_UTSMER9A3_Gambierdiscus- carpenteri_DN15967.c2_g1_i2.p1.faa	0	1	0	1	0	4
21082_MMETSP0766_Gambierdiscus- australes_DN32692.c0_g1_i1.p1.faa	0	2	0	2	1	4

195242_UTSM0ER9A3_Gambierdiscus-	1	1	1	4
carpenteri_DN17326.c2_g5.i1.p1.faa				
83891_UTSM0ER9A3_Gambierdiscus-	1	1	1	4
carpenteri_DN13035.c1_g4.i1.p1.faa				
99486_UTSM0ER9A3_Gambierdiscus-	1	1	1	4
carpenteri_DN13588.c0_g3.i1.p1.faa				
328911_HG4_Gambierdiscus-	1	3	0	4
lapillus_DN41464.c0_g1.i1.p1.faa				
643864_HG5_Gambierdiscus-	0	0	4	4
silvae_DN47931.c1_g3.i1.p2.faa				
186957_UTSM0ER9A3_Gambierdiscus-	1	1	0	3
carpenteri_DN16979.c3_g3.i1.p1.faa				
193820_UTSM0ER9A3_Gambierdiscus-	1	1	0	3
carpenteri_DN17268.c1_g8.i4.p1.faa				
147284_UTSM0ER9A3_Gambierdiscus-	1	1	0	3
carpenteri_DN15408.c1_g3.i2.p1.faa				
116539_UTSM0ER9A3_Gambierdiscus-	2	0	0	3
carpenteri_DN14227.c2_g1.i4.p1.faa				
242595_UTSM0ER9A3_Gambierdiscus-	2	0	0	3
carpenteri_DN9176.c0_g1.i3.p1.faa				
524928_CG150Gambierdiscus-	0	3	0	3
polynesiensis_DN43543.c1_g1.i1.p1.faa				
1040_MMETSIP0766_Gambierdiscus-	0	0	2	3
australes_DN11947.c0_g1.i1.p1.faa				
38402_MMETSP0766_Gambierdiscus-	1	0	0	3
australes_DN41494.c1_g1.i3.p1.faa				
154624_UTSM0ER9A3_Gambierdiscus-	0	0	0	2
carpenteri_DN15679.c0_g6.i1.p1.faa				
63665_UTSM0ER9A3_Gambierdiscus-	0	0	0	2
carpenteri_DN10182.c0_g1.i2.p1.faa				

205876_UTSMER9A3_Gambierdiscus-carpenteri_DN17803_c0_g4_i1.p1.faa	0	0	0	2
224239_UTSMER9A3_Gambierdiscus-carpenteri_DN18618_c3_g6_i1.p1.faa	0	0	0	2
196786_UTSMER9A3_Gambierdiscus-carpenteri_DN17387_c2_g2_i1.p1.faa	0	1	0	2
131133_UTSMER9A3_Gambierdiscus-carpenteri_DN14782_c2_g4_i3.p1.faa	0	0	1	2
19133_MMETSP0766_Gambierdiscus-australes_DN30780_c0_g2_i1.p1.faa	0	0	0	2
37007_MMETSP0766_Gambierdiscus-australes_DN41205_c1_g7_i1.p1.faa	0	0	0	2
424979_CG150Gambierdiscus-polynesiensis_DN34166_c0_g9_i1.p1.faa	0	2	0	2
358554_CG150Gambierdiscus-polynesiensis_DN15070_c0_g1_i1.p2.faa	0	2	0	2
408901_CG150Gambierdiscus-polynesiensis_DN32288_c2_g1_i1.p1.faa	0	2	0	2
479997_CG150Gambierdiscus-polynesiensis_DN39607_c0_g2_i1.p1.faa	0	1	1	2
485470_CG150Gambierdiscus-polynesiensis_DN40097_c0_g1_i2.p1.faa	0	1	1	2
258909_HG4_Gambierdiscus-lapillus_DN22432_c0_g1_i2.p1.faa	0	1	1	2
263811_HG4_Gambierdiscus-lapillus_DN25138_c0_g1_i1.p1.faa	1	0	1	2
319034_HG4_Gambierdiscus-lapillus_DN40675_c3_g1_i2.p1.faa	1	0	1	2
319505_HG4_Gambierdiscus-lapillus_DN40711_c1_g8_i1.p1.faa	1	0	1	2

1041_MMETS	SP0766_Gambierdiscus-	0	0	1	2
australes_DN11947_c0_g2_i1.p1.faa					
27066_MMETS	SP0766_Gambierdiscus-	0	0	1	2
australes_DN36729_c0_g1_i1.p2.faa					
274389_HG4_C	Gambierdiscus-	2	0	0	2
lapillus_DN30113_c0_g1_i2.p1.faa					
46553_MMETS	SP0766_Gambierdiscus-	0	0	0	2
australes_DN42196_c9_g4_i1.p1.faa					
148669_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN15462_c1_g7_i1.p1.faa					
234513_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN23482_c0_g1_i1.p1.faa					
63664_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN10182_c0_g1_i1.p1.faa					
72166_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN1258_c0_g1_i1.p1.faa					
210660_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN18011_c6_g4_i1.p1.faa					
88291_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN13188_c2_g8_i2.p2.faa					
235070_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN25711_c0_g1_i1.p2.faa					
236919_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN33286_c0_g1_i1.p1.faa					
234708_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN24051_c0_g1_i1.p1.faa					
75892_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN12749_c1_g2_i3.p1.faa					
207498_UTSM	MR9A3_Gambierdiscus-	0	0	0	1
carpenteri_DN17871_c4_g9_i1.p1.faa					

234298_UTSM0ER9A3_Gambierdiscus-carpenteri_DN22896.c0_g1_i1.p1.faa	0	0	0	1
84448_UTSM0ER9A3_Gambierdiscus-carpenteri_DN13053.c3_g3_i4.p1.faa	0	0	0	1
104611_UTSM0ER9A3_Gambierdiscus-carpenteri_DN13776.c4_g7_i1.p1.faa	0	0	0	1
242597_UTSM0ER9A3_Gambierdiscus-carpenteri_DN9176.c0_g2_i2.p2.faa	0	0	0	1
233698_UTSM0ER9A3_Gambierdiscus-carpenteri_DN2009.c0_g1_i1.p1.faa	0	0	0	1
115505_UTSM0ER9A3_Gambierdiscus-carpenteri_DN14189.c2_g12_i1.p1.faa	0	0	0	1
238946_UTSM0ER9A3_Gambierdiscus-carpenteri_DN4887.c0_g1_i1.p1.faa	0	0	0	1
208524_UTSM0ER9A3_Gambierdiscus-carpenteri_DN17914.c1_g3_i4.p1.faa	0	0	0	1
131131_UTSM0ER9A3_Gambierdiscus-carpenteri_DN14782.c2_g4_i1.p1.faa	0	0	0	1
215621_UTSM0ER9A3_Gambierdiscus-carpenteri_DN18221.c2_g6_i3.p1.faa	0	0	0	1
225926_UTSM0ER9A3_Gambierdiscus-carpenteri_DN18701.c1_g3_i2.p1.faa	0	0	0	1
239297_UTSM0ER9A3_Gambierdiscus-carpenteri_DN5390.c0_g1_i1.p1.faa	0	0	0	1
233616_UTSM0ER9A3_Gambierdiscus-carpenteri_DN19857.c0_g1_i1.p1.faa	0	0	0	1
208525_UTSM0ER9A3_Gambierdiscus-carpenteri_DN17914.c1_g3_i5.p2.faa	0	0	0	1
236171_UTSM0ER9A3_Gambierdiscus-carpenteri_DN30145.c0_g1_i1.p1.faa	0	0	0	1

241217_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN7872_c0_g1_i1.p1.faa	0	0	0	1
212813_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN18098_c3_g3_i2.p1.faa	0	0	0	1
147705_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN15422_c1_g3_i1.p1.faa	0	0	0	1
242594_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN9176_c0_g1_i2.p1.faa	0	0	0	1
86631_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN13131_c1_g1_i1.p1.faa	0	0	0	1
238247_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN38343_c0_g1_i1.p1.faa	0	0	0	1
212812_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN18098_c3_g3_i1.p1.faa	0	0	0	1
211703_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN18052_c3_g5_i1.p1.faa	0	0	0	1
239230_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN5288_c0_g1_i1.p1.faa	0	0	0	1
103957_UTSM0ER9A3_Gambierdiscus-	carpenteri_DN13754_c3_g2_i4.p1.faa	0	0	0	1
462243_CG150Gambierdiscus-	polynesiensis_DN37930_c0_g1_i2.p1.faa	0	1	0	1
355979_CG150Gambierdiscus-	polynesiensis_DN10471_c0_g1_i1.p1.faa	0	1	0	1
524904_CG150Gambierdiscus-	polynesiensis_DN43540_c1_g1_i2.p1.faa	0	1	0	1
471036_CG150Gambierdiscus-	polynesiensis_DN38733_c0_g1_i1.p1.faa	0	1	0	1
527904_CG150Gambierdiscus-	polynesiensis_DN43803_c0_g1_i1.p1.faa	0	1	0	1

494332_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN40908_c1_g1_i1.p1.faa					
475327_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN39159_c1_g1_i1.p1.faa					
446377_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN36357_c3_g7_i1.p1.faa					
415511_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN33112_c0_g1_i3.p1.faa					
524930_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN43543_c1_g1_i4.p1.faa					
500254_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN41444_c1_g3_i1.p1.faa					
408903_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN32288_c3_g1_i1.p1.faa					
211708_UTSMER9A3	Gambierdiscus-	0	1	0	1
carpenteri_DN18052_c3_g5_i7.p1.faa					
524905_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN43540_c1_g1_i3.p1.faa					
528784_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN47453_c0_g1_i1.p3.faa					
528223_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN44935_c0_g1_i1.p2.faa					
362866_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN18821_c0_g1_i1.p1.faa					
408898_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN32288_c1_g1_i1.p1.faa					
473656_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN39000_c2_g2_i1.p1.faa					
505619_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN41913_c1_g3_i1.p2.faa					

357110.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN13123_c0_g1_i2.p2.faa						
529123.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN48937_c0_g1_i1.p1.faa						
419597.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN33575_c2_g1_i1.p1.faa						
486622.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN40207_c2_g2_i2.p1.faa						
518712.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN43045_c0_g2_i6.p1.faa						
505617.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN41913_c1_g2_i1.p1.faa						
419857.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN33604_c1_g1_i1.p1.faa						
319033.HG40	Gambierdiscus	0	0	1	0	1
lapillus_DN40675_c3_g1_i1.p1.faa						
505612.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN41913_c0_g1_i1.p1.faa						
505621.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN41913_c1_g5_i1.p2.faa						
368243.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN21805_c0_g1_i1.p1.faa						
531066.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN7198_c0_g1_i1.p1.faa						
411779.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN32643_c5_g2_i3.p2.faa						
529709.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN51840_c0_g1_i1.p1.faa						
424815.CG150	Gambierdiscus	0	0	1	0	1
polynesiensis_DN34144_c0_g1_i6.p1.faa						

388829_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN29147_c0_g1_i1.p1.faa					
528991_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN4849_c0_g1_i1.p2.faa					
529886_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN52795_c0_g1_i1.p1.faa					
517572_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN42942_c0_g1_i1.p1.faa					
162319_UTSMER9A3_Gambierdiscus-		0	1	0	1
carpenteri_DN15967_c2_g1_i1.p1.faa					
486374_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN40177_c0_g2_i3.p1.faa					
424977_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN34166_c0_g6_i1.p2.faa					
480000_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN39607_c0_g2_i4.p1.faa					
524933_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN43543_c1_g1_i7.p1.faa					
529340_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN50363_c0_g1_i1.p1.faa					
382787_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN27509_c0_g1_i1.p1.faa					
455767_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN37290_c0_g4_i1.p1.faa					
454667_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN37192_c1_g3_i1.p1.faa					
505616_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN41913_c1_g1_i3.p1.faa					
408904_CG150	Gambierdiscus-	0	1	0	1
polynesiensis_DN32288_c3_g2_i1.p1.faa					

519735_CG150Gambierdiscus-polynesiensis_DN43127_c3_g5_i1.p1.faa	0	1	0	1
524932_CG150Gambierdiscus-polynesiensis_DN43543_c1_g1_i6.p1.faa	0	1	0	1
419608_CG150Gambierdiscus-polynesiensis_DN33575_c2_g2_i1.p1.faa	0	1	0	1
489214_CG150Gambierdiscus-polynesiensis_DN40447_c0_g1_i2.p1.faa	0	1	0	1
407098_CG150Gambierdiscus-polynesiensis_DN32057_c0_g1_i2.p1.faa	0	1	0	1
486620_CG150Gambierdiscus-polynesiensis_DN40207_c2_g1_i2.p2.faa	0	1	0	1
529847_CG150Gambierdiscus-polynesiensis_DN52688_c0_g1_i1.p1.faa	0	1	0	1
355910_CG150Gambierdiscus-polynesiensis_DN1036_c0_g1_i1.p2.faa	0	1	0	1
419599_CG150Gambierdiscus-polynesiensis_DN33575_c2_g1_i11.p1.faa	0	1	0	1
368244_CG150Gambierdiscus-polynesiensis_DN21805_c0_g2_i1.p1.faa	0	1	0	1
528301_CG150Gambierdiscus-polynesiensis_DN45312_c0_g1_i1.p1.faa	0	1	0	1
431157_CG150Gambierdiscus-polynesiensis_DN34812_c2_g1_i1.p1.faa	0	1	0	1
429838_CG150Gambierdiscus-polynesiensis_DN3467_c0_g1_i1.p1.faa	0	1	0	1
485799_CG150Gambierdiscus-polynesiensis_DN40132_c0_g3_i1.p1.faa	0	1	0	1
449384_CG150Gambierdiscus-polynesiensis_DN36673_c0_g1_i3.p1.faa	0	1	0	1

530384.CG150	Gambierdiscus- polynesiensis_DN55090_c0_g1_i1.p1.faa	0	1	0	1
357109.CG150	Gambierdiscus- polynesiensis_DN13123_c0_g1_i1.p2.faa	0	1	0	1
466543.CG150	Gambierdiscus- polynesiensis_DN38313_c1_g3_i1.p1.faa	0	1	0	1
367731.CG150	Gambierdiscus- polynesiensis_DN21547_c0_g1_i1.p1.faa	0	1	0	1
438506.CG150	Gambierdiscus- polynesiensis_DN35575_c0_g1_i7.p1.faa	0	1	0	1
491823.CG150	Gambierdiscus- polynesiensis_DN40690_c4_g5_i2.p1.faa	0	1	0	1
530249.CG150	Gambierdiscus- polynesiensis_DN54681_c0_g1_i1.p1.faa	0	1	0	1
661643.HG5	Gambierdiscus- silvae_DN57114_c0_g1_i1.p1.faa	0	0	1	1
601478.HG5	Gambierdiscus- silvae_DN43780_c7_g8_i1.p1.faa	0	0	1	1
567939.HG5	Gambierdiscus- silvae_DN35530_c0_g3_i1.p1.faa	0	0	1	1
593688.HG5	Gambierdiscus- silvae_DN42661_c0_g1_i1.p1.faa	0	0	1	1
540524.HG5	Gambierdiscus- silvae_DN20879_c0_g2_i1.p1.faa	0	0	1	1
649671.HG5	Gambierdiscus- silvae_DN48408_c0_g1_i4.p1.faa	0	0	1	1
620146.HG5	Gambierdiscus- silvae_DN45801_c1_g1_i1.p2.faa	0	0	1	1
589550.HG5	Gambierdiscus- silvae_DN41996_c3_g12_i1.p1.faa	0	0	1	1

643868_HG5_ Cambierdiscu silvae_DN47931_c1_g3_i5.p1.faa	0	0	1	1
657026_HG5_ Cambierdiscu silvae_DN48988_c0_g3_i1.p1.faa	0	0	1	1
589562_HG5_ Cambierdiscu silvae_DN41996_c3_g5_i1.p2.faa	0	0	1	1
608846_HG5_ Cambierdiscu silvae_DN44648_c2_g1_i1.p1.faa	0	0	1	1
593690_HG5_ Cambierdiscu silvae_DN42661_c0_g2_i3.p1.faa	0	0	1	1
550256_HG5_ Cambierdiscu silvae_DN27602_c0_g2_i1.p1.faa	0	0	1	1
608853_HG5_ Cambierdiscu silvae_DN44648_c2_g6_i1.p1.faa	0	0	1	1
559711_HG5_ Cambierdiscu silvae_DN32102_c0_g1_i2.p1.faa	0	0	1	1
575231_HG5_ Cambierdiscu silvae_DN38322_c1_g2_i1.p1.faa	0	0	1	1
591087_HG5_ Cambierdiscu silvae_DN42232_c1_g4_i1.p2.faa	0	0	1	1
657027_HG5_ Cambierdiscu silvae_DN48988_c0_g3_i2.p1.faa	0	0	1	1
540525_HG5_ Cambierdiscu silvae_DN20879_c0_g3_i1.p1.faa	0	0	1	1
601479_HG5_ Cambierdiscu silvae_DN43780_c7_g9_i1.p1.faa	0	0	1	1
589619_HG5_ Cambierdiscu silvae_DN42009_c0_g1_i3.p1.faa	0	0	1	1
596728_HG5_ Cambierdiscu silvae_DN43120_c1_g4_i4.p1.faa	0	0	1	1

254977_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN19871_c0.g1.i1.p1.faa						
244474_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN10661_c0.g1.i1.p1.faa						
354441_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN7536_c0.g1.i1.p2.faa						
277633_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN31491_c0.g2.i1.p1.faa						
312699_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN40082_c0.g1.i1.p1.faa						
319501_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN40711_c1.g5.i1.p1.faa						
244476_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN10661_c0.g2.i1.p1.faa						
355588_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN9793_c0.g1.i1.p1.faa						
351360_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN46619_c0.g1.i1.p2.faa						
319490_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN40711_c1.g10.i1.p1.faa						
249529_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN15767_c0.g4.i1.p1.faa						
350445_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN4403_c0.g2.i1.p1.faa						
249527_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN15767_c0.g2.i1.p1.faa						
247959_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN14263_c0.g2.i1.p1.faa						
354628_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN8017_c0.g2.i1.p1.faa						

327310_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN41349_c0.g1.i2.p1.faa						
245201_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN11411_c0.g1.i1.p1.faa						
328839_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN41459_c1.g5.i1.p1.faa						
332373_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN41718_c2.g1.i1.p1.faa						
310068_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN39797_c2.g1.i1.p1.faa						
355491_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN9601_c0.g2.i1.p1.faa						
264742_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN25642_c0.g1.i1.p1.faa						
310329_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN39821_c0.g4.i1.p1.faa						
351275_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN46352_c0.g1.i1.p1.faa						
298246_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN38038_c1.g5.i1.p1.faa						
312700_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN40082_c0.g2.i1.p1.faa						
245202_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN11411_c0.g1.i2.p1.faa						
270811_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN28598_c0.g3.i1.p1.faa						
311957_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN40004_c3.g1.i2.p1.faa						
270397_HG4_	Cambierdiscus		1	0	0	1
lapillus.DN2840_c0.g1.i2.p1.faa						

351199_HG4_Cambierdiscus- lapillus_DN46156_c0_g1_i1.p1.faa	0	1	0	0	1
308197_HG4_Cambierdiscus- lapillus_DN39584_c0_g1_i1.p1.faa	0	1	0	0	1
354586_HG4_Cambierdiscus- lapillus_DN792_c0_g1_i1.p1.faa	0	1	0	0	1
350059_HG4_Cambierdiscus- lapillus_DN43172_c0_g1_i1.p1.faa	0	1	0	0	1
264744_HG4_Cambierdiscus- lapillus_DN25642_c0_g2_i1.p1.faa	0	1	0	0	1
27277_MMETSP0766_Gambierdiscus- australes_DN36847_c0_g3_i1.p1.faa	0	0	0	0	1
38401_MMETSP0766_Gambierdiscus- australes_DN41494_c1_g1_i2.p1.faa	0	0	0	0	1
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33030_MMETSP0766_Gambierdiscus- australes_DN39895_c0_g4_i1.p1.faa	0	0	0	0	1
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40251_MMETSP0766_Gambierdiscus- australes_DN41766_c4_g24_i2.p1.faa	0	0	0	0	1
18687_MMETSP0766_Gambierdiscus- australes_DN30415_c0_g4_i1.p1.faa	0	0	0	0	1
18486_MMETSP0766_Gambierdiscus- australes_DN30257_c0_g2_i1.p1.faa	0	0	0	0	1

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16842_MMETSP0766_Gambierdiscus-australes_DN28731_c0_g3_i1.p2.faa	0	0	0	1

1 References

- [1] ASHBURNER, M., BALL, C. A., BLAKE, J. A., BOTSTEIN, D., BUTLER, H., CHERRY, J. M., DAVIS, A. P., DOLINSKI, K., DWIGHT, S. S., EPPIG, J. T., ET AL. Gene Ontology: tool for the unification of biology. *Nature genetics* 25, 1 (2000), 25.
- [2] BACHVAROFF, T. R., CONCEPCION, G. T., ROGERS, C. R., HERMAN, E. M., AND DELWICHE, C. F. Dinoflagellate expressed sequence tag data indicate massive transfer of chloroplast genes to the nuclear genome. *Protist* 155, 1 (2004), 65–78.
- [3] BACHVAROFF, T. R., AND PLACE, A. R. From stop to start: tandem gene arrangement, copy number and trans-splicing sites in the dinoflagellate *Amphidinium carterae*. *PLoS One* 3, 8 (2008), e2929.
- [4] BAUMGARTEN, S., BAYER, T., ARANDA, M., LIEW, Y. J., CARR, A., MICKLEM, G., AND VOOLSTRA, C. R. Integrating microRNA and mRNA expression profiling in *Symbiodinium microadriaticum*, a dinoflagellate symbiont of reef-building corals. *BMC genomics* 14, 1 (2013), 704.
- [5] CERVEAU, N., AND JACKSON, D. J. Combining independent de novo assemblies optimizes the coding transcriptome for nonconventional model eukaryotic organisms. *BMC bioinformatics* 17, 1 (2016), 525.
- [6] CONSORTIUM, G. O. Expansion of the Gene Ontology knowledgebase and resources. *Nucleic acids research* 45, D1 (2016), D331–D338.
- [7] CONSORTIUM, U. Ongoing and future developments at the Universal Protein Resource. *Nucleic acids research* 39, suppl.1 (2010), D214–D219.
- [8] EDDY, S., AND WHEELER, T. HMMER: biosequence analysis using profile hidden Markov models, 2015. hmmerr.org/.
- [9] FU, L., NIU, B., ZHU, Z., WU, S., AND LI, W. CD-HIT: accelerated for clustering the next-generation sequencing data. *Bioinformatics* 28, 23 (2012), 3150–3152.

- [10] GUILLEBAULT, D., SASORITH, S., DERELLE, E., WURTZ, J.-M., LOZANO, J.-C., BINGHAM, S., TORA, L., AND MOREAU, H. A new class of transcription initiation factors, intermediate between TATA box-binding proteins (TBPs) and TBP-like factors (TLFs), is present in the marine unicellular organism, the dinoflagellate *Cryptothecodinium cohnii*. *Journal of Biological Chemistry* 277, 43 (2002), 40881–40886.
- [11] HAAS, B., AND PAPANICOLAOU, A. TransDecoder (find coding regions within transcripts), 2016.
- [12] HEBERLE, H., MEIRELLES, G., DA SILVA, F., TELLES, G., AND MINGHIM, R. Interactivenn: a web-based tool for the analysis of sets through venn diagrams. *BMC bioinformatics* 16 (2015), 169.
- [13] KAHLKE, T. Basta 1.2 - Basic Sequence Taxonomy Annotation (Version 1.2), 2018. <https://doi.org/10.5281/zenodo.1137870>.
- [14] KAHLKE, T. GOSUM: Gene Ontology Summarizer version 0.1, 2018. <http://doi.org/10.5281/zenodo.1344306>.
- [15] KEELING, P. J., BURKI, F., WILCOX, H. M., ALLAM, B., ALLEN, E. E., AMARAL-ZETTLER, L. A., ARMBRUST, E. V., ARCHIBALD, J. M., BHARTI, A. K., BELL, C. J., ET AL. The Marine Microbial Eukaryote Transcriptome Sequencing Project (MMETSP): illuminating the functional diversity of eukaryotic life in the oceans through transcriptome sequencing. *PloS one* (2014).
- [16] KRETZSCHMAR, A. L., VERMA, A., HARWOOD, T., HOPPENRATH, M., AND MURRAY, S. Characterization of gambierdiscus lapillus sp. nov.(gonyaulacales, dinophyceae): A new toxic dinoflagellate from the great barrier reef (australia). *Journal of phycology* 53, 2 (2017), 283–297.
- [17] LAJEUNESSE, T. C., LAMBERT, G., ANDERSEN, R. A., COFFROTH, M. A., AND GALBRAITH, D. W. *Symbiodinium* (Pyrrophyta) genome sizes (DNA content) are smallest among dinoflagellates. *Journal of Phycology* 41, 4 (2005), 880–886.
- [18] LARSSON, M. E., LACZKA, O. F., HARWOOD, D. T., LEWIS, R. J., HIMAYA, S., MURRAY, S. A., AND DOBLIN, M. A. Toxicology of *Gambierdiscus*

- spp.(Dinophyceae) from tropical and temperate Australian waters. *Marine drugs* 16, 1 (2018), 7.
- [19] LIDIE, K. B., RYAN, J. C., BARBIER, M., AND VAN DOLAH, F. M. Gene expression in Florida red tide dinoflagellate *Karenia brevis*: analysis of an expressed sequence tag library and development of DNA microarray. *Marine Biotechnology* 7, 5 (2005), 481–493.
 - [20] LIDIE, K. B., AND VAN DOLAH, F. M. Spliced leader rna-mediated trans-splicing in a dinoflagellate, *Karenia brevis*. *Journal of Eukaryotic Microbiology* 54, 5 (2007), 427–435.
 - [21] MOREY, J. S., AND VAN DOLAH, F. M. Global analysis of mrna half-lives and de novo transcription in a dinoflagellate, *Karenia brevis*. *PLoS One* 8, 6 (2013), e66347.
 - [22] MUNDAY, R., MURRAY, S., RHODES, L. L., LARSSON, M. E., AND HARWOOD, D. T. Ciguatoxins and maitotoxins in extracts of sixteen gambierdiscus isolates and one fukuyoa isolate from the south pacific and their toxicity to mice by intraperitoneal and oral administration. *Marine drugs* 15, 7 (2017), 208.
 - [23] QUEVILLON, E., SILVENTOINEN, V., PILLAI, S., HARTE, N., MULDER, N., APWEILER, R., AND LOPEZ, R. InterProScan: protein domains identifier. *Nucleic acids research* 33, suppl_2 (2005), W116–W120.
 - [24] RAE, P. Hydroxymethyluracil in eukaryote DNA: a natural feature of the pyrophyta (dinoflagellates). *Science* 194, 4269 (1976), 1062–1064.
 - [25] RHODES, L. L., SMITH, K. F., MUNDAY, R., SELWOOD, A. I., McNABB, P. S., HOLLAND, P. T., AND BOTTEIN, M.-Y. Toxic dinoflagellates (Dinophyceae) from Rarotonga, Cook Islands. *Toxicon* 56, 5 (2010), 751–758.
 - [26] RHODES, L. L., SMITH, K. F., MURRAY, S., HARWOOD, D. T., TRNSKI, T., AND MUNDAY, R. The epiphytic genus *Gambierdiscus* (Dinophyceae) in the Kermadec Islands and Zealandia regions of the southwestern Pacific and the associated risk of ciguatera fish poisoning. *Marine drugs* 15, 7 (2017), 219.

- [27] RIZZO, P. J., AND NOODÉN, L. D. Chromosomal proteins in the dinoflagellate alga *Gyrodinium cohnii*. *Science* 176, 4036 (1972), 796–797.
- [28] VINUESA, P., AND CONTRERAS-MOREIRA, B. Robust identification of orthologues and paralogues for microbial pan-genomics using GET_HOMOLOGUES: a case study of pIncA/C plasmids. In *Bacterial Pangenomics*. Springer, 2015, pp. 203–232.
- [29] ZHANG, H., HOU, Y., MIRANDA, L., CAMPBELL, D. A., STURM, N. R., GAASTERLAND, T., AND LIN, S. Spliced leader RNA trans-splicing in dinoflagellates. *Proceedings of the National Academy of Sciences* 104, 11 (2007), 4618–4623.