
ESTIMATING THE **CONTAMINATION** FACTOR'S DISTRIBUTION IN UNSUPERVISED ANOMALY DETECTION

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ABSTRACT

Anomaly detection methods identify examples that do not follow the expected behaviour, typically in an unsupervised fashion, by assigning real-valued anomaly scores to the examples based on various heuristics. These scores need to be transformed into actual predictions by thresholding, so that the proportion of examples marked as anomalies equals the expected proportion of anomalies, called contamination factor. Unfortunately, there are no good methods for estimating the contamination factor itself. We address this need from a Bayesian perspective, introducing a method for estimating the posterior distribution of the contamination factor of a given unlabeled dataset. We leverage on outputs of several anomaly detectors as a representation that already captures the basic notion of anomalousness and estimate the contamination using a specific mixture formulation. Empirically on 22 datasets, we show that the estimated distribution is well-calibrated and that setting the threshold using the posterior mean improves the anomaly detectors' performance over several alternative methods. All code is publicly available for full reproducibility.

Keywords Anomaly Detection · Unsupervised Learning · Bayesian Methods · Contamination Factor

1 Introduction

Anomaly detection [Chandola et al., 2009] is the task of automatically identifying examples that do not conform to the expected behaviour. The anomalies are often indicative of critical events such as intrusions in web networks [Malaiya et al., 2018, Xin et al., 2018], failures in the petroleum extraction [Martí et al., 2015], excessive usage of water [Ver-cruyssen et al., 2018], or breakdowns in wind turbines [Zaher et al., 2009] and gas turbines [Yan and Yu, 2019]. Such events have an associated high cost and detecting the anomalies in time avoids wasting resources.

Typically, anomaly detection is tackled from an unsupervised perspective [Maxion and Tan, 2000, Goldstein and Uchida, 2016, Zong et al., 2018, Perini et al., 2020a, Han et al., 2022] because labeled examples, especially anomalies, may be expensive and difficult to acquire (e.g., you do not want to voluntarily break the equipment simply to observe anomalous behaviours), or simply rare (e.g., you may need to inspect many examples before finding an anomalous one). Unsupervised anomaly detectors exploit data-driven heuristic assumptions to assign a real-valued score to each example denoting how anomalous it is. For instance, kNNO [Angiulli and Pizzuti, 2002] assigns the example's distance to its k-th nearest neighbour, while AUTOENCODER [Chen et al., 2018] uses the example's reconstruction error. Using such anomaly scores enables ranking the examples from most to least anomalous.

Converting the anomaly scores into discrete predictions would practically allow the user to flag the anomalies. Commonly, one chooses a predictive threshold and assigns the anomaly label to the examples with higher score, while all the remaining ones are considered normal. However, setting the threshold is a challenging task as it cannot be tuned (e.g., by maximizing the model performance) due to the absence of labels [Soenen et al., 2021]. One approach

is to set the threshold such that the proportion of scores above it matches the dataset’s *contamination factor* γ , the expected proportion of anomalies. This allows perfect rankings to be transformed into perfect predictions. Usually, the contamination factor is assumed to be given by a domain expert, but in most of the real-world scenarios it is actually unknown.

Estimating the contamination factor is challenging. Existing works provide an estimate by either collecting some normal labels [Perini et al., 2020b] or by exploiting domain knowledge [Perini et al., 2022]. Moreover, one can apply statistical threshold estimators that directly set a threshold on the anomaly scores, and derive the contamination factor as proportion of scores higher than the threshold. For instance, the Modified Thompson Tau test thresholder (MTT) is based on the modified Thompson Tau test [Rengasamy et al., 2021], while the Inter-Quartile Region thresholder (IQR) sets the threshold as the third quartile plus 1.5 times the inter-quartile region [Bardet and Dimby, 2017]. We will later explain and evaluate several estimators in Section 4.

Transforming the scores using an incorrect estimate of the contamination factor (or, equivalently, an incorrect threshold) deteriorates the anomaly detector’s performance [Fourure et al., 2021, Emmott et al., 2015] and reduces the trust in the detection system. If such an estimate was coupled with a measure of uncertainty, one could take into account this uncertainty to improve decisions.

We study the estimation of the contamination factor from a Bayesian perspective and propose γ GMM, the first algorithm for estimating the contamination factor’s (posterior) distribution in unlabeled anomaly detection setups. First, we use a set of unsupervised anomaly detectors to assign anomaly scores for all examples and use these scores as a new representation for the data. Second, we fit a Bayesian Gaussian Mixture model with a Dirichlet process prior (DPGMM) [Ferguson, 1973, Rasmussen, 1999, Görür and Edward Rasmussen, 2010] in this new space. If we knew which components contain the anomalies, we could derive the contamination factor’s posterior distribution as the distribution of the sum of such components’ weights. Because we do not know this, as a third step γ GMM estimates the probability that the k most extreme components are jointly anomalous, and uses this information to construct the desired posterior. The method explained in detail in Section 3.

In summary, we make four contributions. First, we adopt a Bayesian perspective and introduce the problem of estimating the contamination factor’s posterior distribution. Second, we propose an algorithm that is able to sample from this posterior. Third, we demonstrate experimentally that the implied uncertainty-aware predictions are well calibrated and that taking the posterior mean as point estimate of γ outperforms several other algorithms in common benchmarks. Finally, we show that using the posterior mean as a threshold improves the actual anomaly detection accuracy.

2 Preliminaries

Let $(\Omega, \mathcal{F}, \mathbb{P})$ be a probability space, and $X: \Omega \rightarrow \mathbb{R}^d$ a random variable, from which a dataset D containing N i.i.d. examples $x_1, \dots, x_N \in \mathbb{R}^d$ is drawn. Assume that X has a distribution of the form $p(X) = (1-\gamma) \cdot p(X^{(1)}) + \gamma \cdot p(X^{(2)})$, where $X^{(1)}$ and $X^{(2)}$ are the random variables representing, respectively, normals and anomalies, and γ is the *contamination factor*, the proportion of anomalies. An (unsupervised) *anomaly detector* is a measurable function $f: \mathbb{R}^d \rightarrow \mathbb{R}$ that assigns real-valued anomaly scores $S = f(X)$ to the examples. Such anomaly scores follow the rule that *the higher the score, the more anomalous the example*.

A Gaussian mixture model (GMM) with K components [Roberts et al., 1998] is a generative model such that $p(x) = \sum_{k=1}^K \pi_k \mathcal{N}(x|\mu_k, \Sigma_k)$, where π_k is the mixing proportion, μ_k and Σ_k are the mean and covariance matrix of the component k . For finite mixtures we typically have a Dirichlet prior over $\pi = [\pi_1, \dots, \pi_K]$, but Dirichlet Process (DP) priors allow treating also the number of components as unknown [Görür and Edward Rasmussen, 2010]. For both cases we need approximate inference to estimate the posterior of the model parameters.

3 Methodology

We tackle the problem: **Given** an unlabeled dataset D and a set of M unsupervised anomaly detectors; **Estimate** the (posterior) distribution of the contamination factor γ .

Learning from an unlabeled dataset has three key challenges. First, the absence of labels forces us to make relatively strong assumptions. Second, the anomaly detectors rely on different heuristics that may or may not hold, and their performance can hence vary significantly across datasets. Third, we need to be careful in introducing user-specified hyperparameters, because setting them properly may be as hard as directly specifying the contamination factor.

In this paper we propose γ GMM, a novel approach that estimates the contamination factor’s posterior distribution using a Bayesian approach. It consists of four steps, which we introduce on a conceptual level now and mathematically, in

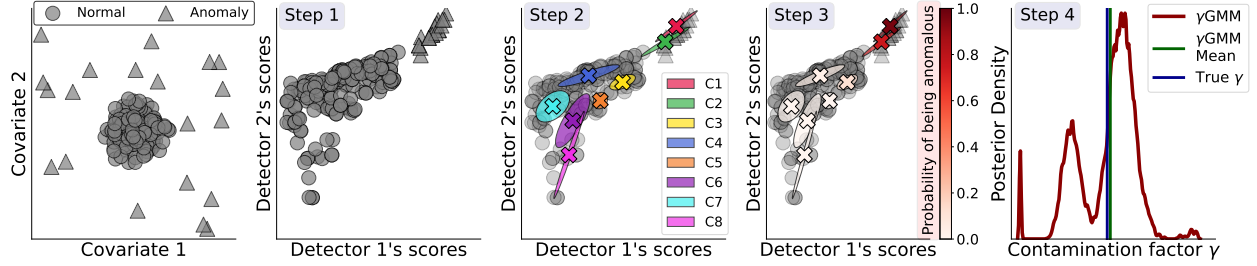


Figure 1: Illustration of the γ GMM’s four steps on a 2D toy dataset (left plot): we 1) map the 2D dataset into an $M = 2$ dimensional anomaly space, 2) fit a DPGMM model on it, 3) compute the components’ probability of being anomalous (conditional, in the plot), 4) derive $\gamma|S$ ’s posterior. γ GMM’s mean is an accurate point estimate for the true value γ^* .

more detail, afterwards. See Figure 1 for an illustration.

Step 1. Because anomalies may not follow any particular pattern, γ GMM maps the covariates X into an M dimensional anomaly space, where the dimensions correspond to the anomaly scores assigned by the M unsupervised anomaly detectors. Within each dimension of such a space, the evident pattern is that “the higher the more anomalous”.

Step 2. We model the data in this space using a Dirichlet Process Gaussian Mixture Model (DPGMM) [Neal, 1992, Rasmussen, 1999, Görür and Edward Rasmussen, 2010]. We assume that each of the (potentially many) mixture components contains either only normals or only anomalies. If we knew which components contained anomalies, we could then easily derive γ ’s posterior as the sum of the mixing proportions π of the anomalous components. However, such information is not available in our setting.

Step 3. Thus, we estimate the probability of the most extreme k components being anomalous. This poses three challenges: (a) how to represent each component in M -dimensional space by a single value to sort them from the most to the least anomalous, (b) how to compute the probability that the k th component is anomalous given that the $k - 1$ th is such, and (c) how to derive the target probability that exactly k components are jointly anomalous.

Step 4. γ GMM estimates the contamination factor’s posterior by exploiting such a joint probability and the components’ mixing proportions posterior.

In the following, we describe these steps in detail.

3.1 Representing data using anomaly scores

Learning from an unlabeled anomaly detection dataset has two major challenges. First, anomalies are rare and sparse events, which makes it hard to use common unsupervised methods like clustering [Breunig et al., 2000]. Second, making assumptions on the unlabeled data is challenging due to the absence of specific patterns in the anomalies, which makes it hard how to choose a proper anomaly detector.

Therefore, we use a set of M anomaly detectors to map the sample space into an M -dimensional score space \mathbb{R}^M , such that an example x gets a score s :

$$\mathbb{R}^d \ni x \rightarrow [f_1(x), f_2(x), \dots, f_m(x)] = s \in \mathbb{R}^M.$$

This has two main effects: (1) it introduces an interpretable space where the evident pattern is that, within each dimension, higher scores are more likely to be anomalous, and (2) it accounts for multiple inductive biases by using multiple arbitrary anomaly detectors.

To make the dimensions comparable, we (independently for each dimension) linearly map the scores to the positive reals, apply the log transform to shorten heavy right tails, and normalize them to have zero mean and unit variance.

3.2 Modeling the density with DPGMM

We use mixture models as basis for quantifying the distribution of the contamination factor, relying on their ability to model proportions of examples using the mixture weights. For flexible modelling we use a Dirichlet Process Gaussian

Mixture Model (DPGMM) [Görür and Edward Rasmussen, 2010]

$$\begin{aligned} s_i &\sim \mathcal{N}(\tilde{\mu}_i, \tilde{\Sigma}_i) \\ (\tilde{\mu}_i, \tilde{\Sigma}_i) &\sim G \\ G &\sim DP(G_0, \alpha) \\ G_0 &= \mathcal{NIW}(m, \lambda, V, u) \end{aligned}$$

where G is a random distribution of the means μ_i and covariance matrices Σ_i , drawn from a DP with base distribution G_0 . We use the explicit representation $G = \sum_{k=1}^{\infty} \pi_k \delta_{(\mu_k, \Sigma_k)}(\tilde{\mu}_i, \tilde{\Sigma}_i)$, where $\delta_{(\mu_k, \Sigma_k)}$ is the delta distribution at (μ_k, Σ_k) and π_k follow the stick-breaking distribution. We set G_0 as Normal Inverse Wishart [Nydic, 2012] with parameters m, λ, V, u common to all components.

The DP formulation allows us to refrain from specifying the number of components K . After fitting the model, we only consider the components with at least one observation assigned to them and propagate all the remaining density uniformly over the active components. Thus, for the following steps we can still proceed as if the model was a finite mixture with π following a Dirichlet distribution.

We use variational inference (VI; see Blei et al. [2017] for details) for approximating the posterior as VI is computationally efficient and sufficiently accurate for our purposes. Alternative methods (e.g., Markov Chain Monte Carlo Brooks et al. [2011]) could also be used but were not considered worth the additional computational effort here.

3.3 Estimating the components’ anomalousness

We assume each mixture component only contains anomalous or normal samples, which is a reasonable assumption when K is large. If we knew which components contain anomalies, we could directly derive the posterior of the contamination factor γ as the sum of the mixing proportions π_k of those components. This is naturally not the case, but instead we need to estimate it in an unsupervised fashion.

More formally, we estimate the probability that k (out of all K) components are anomalous such that we can derive γ ’s posterior by averaging over all the values $0 \leq k \leq K$. We do this in three steps. Initially, we sort the components by degree of anomalousness, which comes natural from the representation we made in Step 1 (Sec. 3.1). Then, our insight is that the k th component c_k can be anomalous only if the $k - 1$ th is such. This points to the estimation of conditional probabilities, i.e., the probability of c_k being anomalous given that c_{k-1} is anomalous. Finally, the probability that exactly the first k components are anomalous can be obtained using basic rules of probability theory.

Assigning an ordering to the components. As initial step for computing the joint probability, we need to design an ordering map for the components, where the “top-right” component is the most anomalous one. We do this in a manner that accounts for the uncertainty of the components’ parameters to rank high the components that can be reliably identified as anomalous: We want the means to be high but the variance low, to avoid the risk that also examples with low anomaly scores could belong to the component.

We construct the overall ranking using dimension-specific scores because our normalization cannot remove all statistical differences between the different detectors. Formally, let $r: \mathbb{R}^M \times \mathbb{R}^{M \times M} \rightarrow \mathbb{R}$ be the function of the mean μ_k and the covariance matrix Σ_k that assigns a representative value to the component k . We set r as

$$r(\mu_k^{(z)}, \Sigma_k^{(z)}) = \frac{1}{M} \sum_{j=1}^M \frac{\mu_k^{j(z)}}{1 + \sqrt{\Sigma_k^{j,j(z)}}}, \quad (1)$$

where $\mu_k^{(z)}$ and $\Sigma_k^{(z)}$ are samples from the parameters’ posterior distributions of the k th component c_k . We obtain a representative value of the whole component by taking the expected value of r , i.e. through $\mathbb{E}[r(\mu_k, \Sigma_k)]$.

We add 1 to the component’s standard deviation for two reasons. First, if a component contains examples with almost the same covariate values, the standard deviation would be close to 0 and the ratio would explode towards infinity, masking any effect of the mean. Second, adding 1 is reasonable because it is equal to the theoretical upper bound of the components variances, as they are normalized (Sec. 3.1).

Without loss on generality, from now on we assume that the components’ index k is ordered based on their representative value such that the k th component c_k has higher value (i.e., more anomalous) than the $k + 1$ th component.

Estimating the probability that the k th component is anomalous. Because the components are sorted by anomalousness, our key insight is that *the k th component c_k can be anomalous only if the $k - 1$ th is anomalous*. Formally,

$$\begin{aligned}\mathbb{P}(c_k \text{ is anomalous} \mid c_{k-1} \text{ is anomalous}) &> 0 \\ \mathbb{P}(c_k \text{ is anomalous} \mid c_{k-1} \text{ is not anomalous}) &= 0\end{aligned}$$

for any $1 < k \leq K$. For the sake of brevity, we use $\mathbb{P}(c_k \mid c_{k-1})$ as $\mathbb{P}(c_k \text{ is anomalous} \mid c_{k-1} \text{ is anomalous})$.

Moreover, we assume $\mathbb{P}(c_1) = \mathbb{P}(c_1 \text{ is anomalous}) \in (0, 1)$. That is, we allow for the data to not have anomalies (< 1) but exclude certain knowledge of no anomalies (> 0). This is a sensible assumption because, if one knew for sure that no anomalies are in the data, then we trivially have $\gamma = 0$, whereas we still need to allow for the data to be free of anomalies if evidence suggests so.

We estimate the conditional probability as

$$\mathbb{P}(c_k \mid c_{k-1}) = \frac{1}{1 + e^{(\tau + \delta \cdot r(\mu_k, \Sigma_k))}}, \quad (2)$$

where τ and δ are the two hyperparameters of the sigmoid function, which will be carefully discussed in the next sections. Note that the principle itself is not restricted to this particular choice of functional form. One could apply any transformation that maps to $[0, 1]$, but the detailed derivations of the parameters would naturally be different.

Deriving the components' joint probability. Given the conditional probability $\mathbb{P}(c_k \mid c_{k-1})$, the joint probability $\mathbb{P}(\text{exact } c_1, \dots, c_k)$ follows from simple steps. Taking inspiration from the sequential ordinal models [Bürkner and Vuorre, 2019], our insight is that exactly k components are jointly anomalous if and only if each of them is conditionally anomalous and the $k + 1$ th is not anomalous. Essentially,

$$\mathbb{P}(c_1, \dots, c_k) = \mathbb{P}(c_1) \prod_{t=1}^{k-1} \mathbb{P}(c_{t+1} \mid c_t) (1 - \mathbb{P}(c_{k+1} \mid c_k)) \quad (3)$$

for any $k \leq K$, where $\mathbb{P}(c_{K+1} \mid c_K) = 0$ by convention.

3.4 Estimating the contamination factor's distribution

Given the joint probability that the first k components are anomalous (for $k \leq K$), the contamination factor γ 's posterior distribution can be obtained as

$$p(\gamma \mid S) = \sum_{k=1}^K \mathbb{P}(c_1, \dots, c_k) \cdot p\left(\sum_{j=1}^k \pi_j \mid S\right), \quad (4)$$

where $\pi_j \mid S$ is the posterior marginal distribution of the mixing proportion for the j th component. For any k , we have $p(\sum_{j=1}^k \pi_j \mid S) = \text{BETA}(\sum_{j=1}^k \alpha_j, \sum_{j=k+1}^K \alpha_j)$, where $(\pi_1 \mid S, \dots, \pi_K \mid S) \sim \text{DIR}(\alpha_1, \dots, \alpha_K)$ [Lin, 2016].

Setting the sigmoid's hyperparameters τ and δ . Introducing new hyperparameters when the task is to estimate the contamination factor γ 's posterior is risky because setting their value may be as difficult as directly providing a point estimate of γ . With regard to τ and δ , our key insight is that we can obtain them by asking the user two simple questions: (a) How likely is that no anomalies are in the data? (b) How likely is that a large amount of anomalies occurred, say, more than $t = 15\%$ of the data? Both of these values are supposed to be low. Let's call p_0 the first answer and p_{high} the second one. Formally,

$$\begin{aligned}p_0 &= 1 - \mathbb{P}(c_1) = 1 - \frac{1}{1 + e^{(\tau + \delta \cdot r(\bar{\mu}_1, \bar{\Sigma}_1))}} \\ p_{\text{high}} &= \mathbb{P}(\gamma \geq t \mid S) = \sum_{k=1}^K \mathbb{P}(c_1, \dots, c_k) \mathbb{P}\left(\sum_{j=1}^k \pi_j \geq t \mid S\right)\end{aligned}$$

One can use a numerical solver for non-linear equations with linear constraints to find the values of τ and δ that satisfy such constraints. The problem has a unique solution whenever $p_{\text{high}} \geq \mathbb{P}(\pi_1 \geq t)$. This holds almost always in our experimental cases, but in the Supplement we describe a procedure that can be used in the other scenarios. In the experiments, we show that changing the p_0 and p_{high} does not have a large impact on γ 's posterior (see Q5, Sec. 4).

Sampling from γ ’s posterior distribution. Our estimate of the contamination factor’s posterior $p(\gamma|S)$ does not have a simple closed form. However, we can sample from the distribution using a simple process. The DPGMM inference determines an approximation for $p(\pi, \mu, \Sigma|S)$ and all the quantities required for Equations 2, 3, 4 can be computed based on samples from the approximation. A description of the detailed sampling process is provided in the Supplement.

Additional technical details. Because our method uses the variational inference approximation, we run it 10 times and concatenate the samples to reduce the risk of biased distributions due to local minima. Moreover, after sorting the components, we set $\mathbb{P}(c_k|c_{k-1}) = 0$ for all $k > K' = \arg \max\{k: \mathbb{E}[\sum_{j=1}^k \pi_j] < 0.25\}$. This has the effect of setting an upper bound of 0.25 to the contamination factor γ . Because anomalies must be rare, we realistically assume that it is not possible to have more than 25% of them. Note that $\mathbb{E}[\pi_1] \geq 0.25$ cannot occur, as otherwise we could not set the hyperparameters p_0 and p_{high} .

4 Experiments

We empirically evaluate two aspects of our method: (a) whether it accurately estimates the contamination factor’s posterior, and (b) how thresholding the scores using our method affects the anomaly detectors’ performance. To this end, we address the following five experimental questions:

- Q1. Is the posterior estimate sharp and well-calibrated?
- Q2. How does γ GMM compare to alternative threshold estimators?
- Q3. Does a better point estimate of γ improve the anomaly detector performance?
- Q4. What is the impact of the number of detectors M ?
- Q5. How sensitive the method is to p_0 and p_{high} ?

4.1 Experimental Setup.

Methods. We compare the sample mean of γ GMM with 21 threshold estimators: *Kernel-based* (FGD [Qi et al., 2021] and AUCP [Ren et al., 2018]); *Curve-based* (EB [Friendly et al., 2013] and WIND [Jacobson et al., 2013]); *Normality-based* (ZSCORE [Bagdonavičius and Petkevičius, 2020], DSN [Amagata et al., 2021] and CHAU [Bol’shev and Ubaidullaeva, 1975]); *Regression-based* (CLF and REGR [Aggarwal, 2017]); *Filter-based* (FILTER [Hashemi et al., 2019] and HIST [Thanammal et al., 2014]); *Statistical test-based* (GESD [Alrawashdeh, 2021], MCST [Coin, 2008] and MTT [Rengasamy et al., 2021]); *Statistical moment-based* (BOOT [Martin and Roberts, 2006], KARCH [Afsari, 2011] and MAD [Archana and Pawar, 2015]); *Quantile-based* (IQR [Bardet and Dimby, 2017] and QMCD [Iouchtchenko et al., 2019]); *Transformation-based* (MOLL [Keyzer and Sonneveld, 1997] and YJ [Raymaekers and Rousseeuw, 2021]). We apply each threshold estimator to the univariate anomaly scores of each detector at a time. We then average the contamination factors over the M detectors and use this value as the final point estimate for each dataset. See the Supplement for more details.

Data. We carry out our study on 20 commonly used benchmark datasets and additionally 2 (proprietary) real tasks. The benchmark datasets contain semantically useful anomalies widely used in the literature [Campos et al., 2016]. The datasets vary in size, number of features, and true contamination factor. See Table 1 in the Supplement for details. For the real tasks, our experiments focus on preventing blade icing in wind turbines. We use two public wind turbine datasets, where sensors collect various measurements (e.g., wind speed, power energy, etc.) every 7 seconds for either 8 weeks (turbine 15) or 4 weeks (turbine 21). Following Zhang et al. [2018], we construct feature-vectors by taking the average over the time segment of one minute.

Metrics. We use three evaluation metrics to assess the performance of the methods. Contrary to all the threshold estimators, our method estimates the posterior of γ . Therefore, we measure the **probabilistic calibration** of γ GMM’s posterior using a QQ-plot with the x-axis representing the expected probabilities (i.e., assuming a perfectly calibrated distribution) and on the y-axis the empirical frequencies. That is, for $v \in [0, 0.5]$,

$$\begin{aligned} \text{Expected Prob.} &= \mathbb{P}(\gamma^* \in [q(0.5 - v), q(0.5 + v)]) = 2v \\ \text{Empirical Freq.} &= \frac{|\{\gamma \in [q(0.5 - v), q(0.5 + v)]\}|}{\# \text{experiments}}, \end{aligned}$$

where $q(u)$ is the quantile at the value u of our distribution, for $u \in [0, 1]$, and γ^* refers to the true dataset’s contamination factor. For evaluating the point estimate of the methods, we use the mean absolute error (MAE) between the method’s point estimate and the true value. Finally, we measure the impact of thresholding the scores using the methods’ point estimate through the F_1 score, as common metrics like the Area Under the ROC curve and the Average Precision are not affected by different thresholds. Specifically, we measure the *relative deterioration* of the F_1 score:

$$F_1 \text{ deterioration} = \frac{F_1(f_m, D, \gamma^*) - F_1(f_m, D, \hat{\gamma})}{F_1(f_m, D, \hat{\gamma})}$$

where we compute the F_1 score on the dataset D using the anomaly detector f_m , and either the true value γ^* or an estimate $\hat{\gamma}$ to threshold the scores. The F_1 deterioration of a method is (mostly) negative, and the higher the better.

Setup. In the experiments we assume a transductive setting [Campos et al., 2016, Scott and Blanchard, 2008, Toron et al., 2022], where a dataset D is used both for training and testing. This is the typical setting of anomaly detection [Breunig et al., 2000, Schölkopf et al., 2001, Angiulli and Pizzuti, 2002, Liu et al., 2012, Goldstein and Dengel, 2012, Chen et al., 2018] because the absence of labels and patterns (for the anomaly class) avoids overfitting issues.

For each dataset, we proceed as follows: (i) use a set of M anomaly detectors to assign the anomaly scores S to each observation in the dataset D ; (ii) map each anomaly score $s \in S$ to $\log(s - \min(S) + 0.01)$ and normalize them to have mean equal to 0 and standard deviation equal to 1; (iii) either use our method to estimate the contamination factor’s posterior and extract the posterior mean as point estimate $\hat{\gamma}$, or use one of the threshold estimators to directly obtain a point estimate $\hat{\gamma}$ of the contamination factor (see methods paragraph above); (iv) evaluate the point estimates using the mean absolute error (MAE) between such estimate and the true value γ^* ; (v) use the contamination factor’s point estimate to threshold the anomaly scores of each of the M anomaly detectors f_m (individually); (vi) finally, we measure the F_1 score and compute the relative deterioration.

Hyperparameters, anomaly detectors and priors. Our method introduces two new hyperparameters: p_0 and p_{high} that we both set to 0.01 as default value.

We use 10 anomaly detectors with different inductive bias [Soenen et al., 2021]: kNN [Angiulli and Pizzuti, 2002], IForest [Liu et al., 2012], LOF [Breunig et al., 2000], OCSVM [Green and Richardson, 2001], AE [Chen et al., 2018], VAE [Kingma and Welling, 2013], LSCP [Zhao et al., 2019a], HBOS [Goldstein and Dengel, 2012], LODA [Pevný, 2016], and COPOD [Li et al., 2020]. See the Supplement for additional details. We use the python library PyOD [Zhao et al., 2019b] for the code. The threshold estimators are implemented in the Python library PYTHRESH¹ with default hyperparameters.

For the variational inference of DPGMM we use the implementation in SKLEARN, with the Stick-breaking representation [Dunson and Park, 2008], with 100 as upper bound of the number of components K . The parameter priors are set to the default values: means are set to 0, while the covariance matrices are set to identities of appropriate dimension. We opt for such (in our context) weakly-informative priors because sensible prior knowledge of the DPGMM hyperparameters is hard to come by in practice.

4.2 Experimental Results

Q1. Does our method estimate a sharp and well-calibrated posterior of γ ? Figure 2 shows the contamination factor γ ’s posterior estimated by our method in 10 representative datasets (see Figure 8 in the Supplement for all the plots). In the top five plots, the distribution looks accurate as γ ’s true value (blue line) is close to the posterior mean (i.e., the expected value, the green line). On the contrary, the five plots below represent the five worst cases: although γ ’s true value sometimes falls on low density regions (Arrhythmia and Shuttle datasets), in many cases it would be quite likely to sample the true value from our posterior (KDDCup99, Parkinson, Glass datasets), which makes the density still quite reliable. Figure 3 shows the calibration plot. The posterior is well calibrated as it is very close to the dashed black line indicating perfectly calibrated distribution. The empirical frequencies deviate from the real probabilities by less than 5% (dark shadow grey) in more than 76% of the cases, while never deviating by more than 10% (light shadow grey).

Q2. How does γ GMM compare to the threshold estimators? We take γ GMM’s posterior mean as our best point estimate of γ and compare such value to the point estimates obtained from the threshold estimators. Figure 4 illustrates the ordered MAE (mean \pm std.) between the methods’ estimate and the true γ . On average, γ GMM obtains a MAE of 0.026 that is 20% lower than the best runner-up MTT and 27% lower than the third best method QMCD (MAE of 0.033

¹Link: <https://github.com/KulikDM/pythresh>.

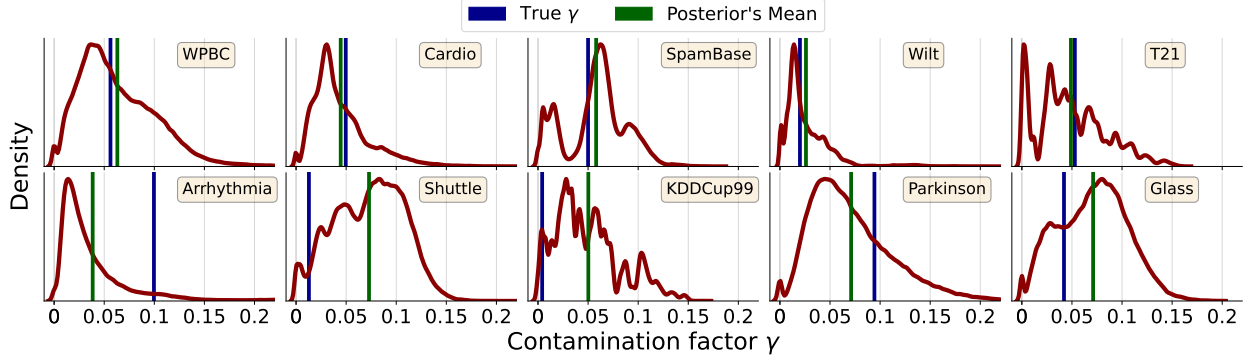


Figure 2: Illustration of how γ GMM estimates γ 's posterior on 10 representative datasets. On the top, the density has the sample mean close to the true value of γ , while on the bottom, the true value of γ falls far from our point estimate.

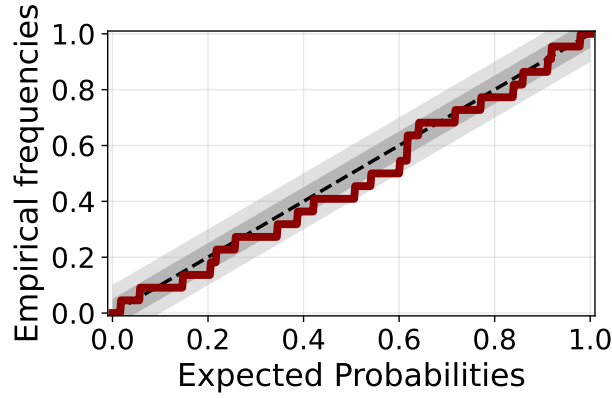


Figure 3: QQ-plot showing how calibrated γ GMM's estimate of the distribution is. The black dashed line illustrates the perfect calibration scenario. Dark and light grey shades indicate a deviation of 5% and 10% from the black line.

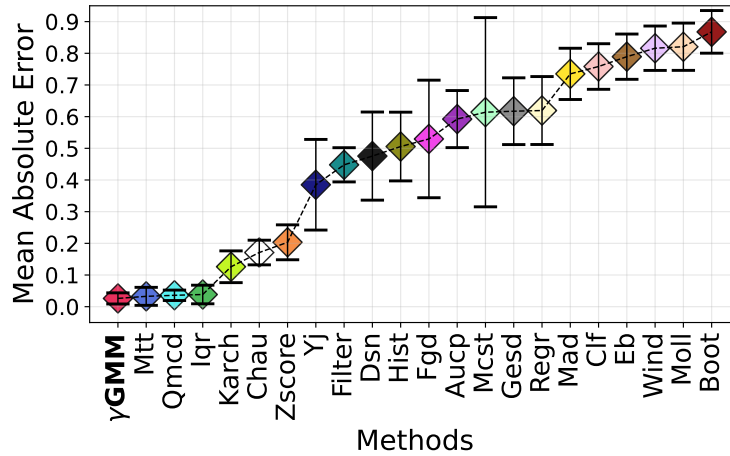


Figure 4: Average MAE (\pm std.) of γ GMM's sample mean compared to the other methods. Our method has the lowest (better) average, which is 20% lower than the runner-up.

and 0.036). For each experiment, we rank the methods from the best (position 1, lowest MAE) to the worst (position 22, greatest MAE). Our method has the best average rank (2.13 ± 1.04). Moreover, γ GMM ranks first 8 times ($\approx 36\%$ of the cases), and for 13 times ($\approx 60\%$ of the cases) it is in the top two. The next best method, MTT, ranks first in 6 cases with average rank of 2.30 ± 1.10 .

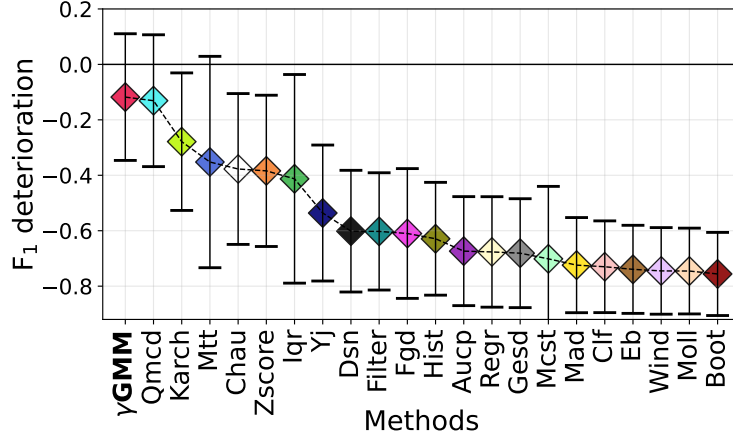


Figure 5: F_1 deterioration (mean \pm std) for each method, where the higher the better. γ GMM ranks as best method, obtaining $\approx 10\%$ higher average than QMCD.

Q3. Does a better contamination improve the anomaly detectors’ performance? We use γ GMM’s posterior mean as point estimate to measure the F_1 score of the anomaly detectors, because sampling from the distribution would not make a fair comparison against the other methods that can only provide a point estimate.

Anomaly detectors that fail to rank the samples accurately perform poorly even when using the correct contamination factor. Since our focus is studying the effect of the contamination factor, we compare F_1 scores only over the detectors that work well for each of the datasets. For each dataset D , we use as set of detectors those achieving the greatest F_1 score using the true contamination factor, i.e. $\arg \max_{f_m} \{F_1(f_m, D, \gamma^*)\}$. The Supplement contains the list of detectors used for each experiment.

Figure 5 shows the average (\pm std.) deterioration for each of the methods. On average, γ GMM has the best F_1 deterioration (-0.117 ± 0.228) that is around 10% better than the runner-up QMCD (-0.131 ± 0.238), and 58% better than the next best KARCH (-0.279 ± 0.248). For 25% of the cases we get higher F_1 score with γ GMM than when using the true γ^* . This is due to the (still incorrect) ranks made by the detectors, which achieve better performance with slightly incorrect contamination factors. Supplement provides further details on how the methods perform in terms of false alarms and false negatives.

Q4. What is the impact of the number M of detectors on the contamination factor’s posterior? In the previous experiments, we used $M = 10$ detectors. To evaluate the effect of M , we repeated the experiments for random choices of 3, 5, or 7 detectors, with 10 replicates using different detectors for each case. Figure 6 shows that the calibration suffers if using fewer detectors, but already with $M = 5$ the method works fairly well. The variance of the results (over repeated experiments) also increases for lower M .

Q5. Impact of the hyperparameters p_0 and p_{high} . We evaluate the impact of the hyperparameters p_0 and p_{high} by running the experiments with much smaller and larger values than the default (0.01): we vary, one at the time, p_0 , $p_{\text{high}} \in [0.0001, 0.001, 0.05, 0.01]$ and keep the other value set as default. Figure 7 shows the QQ-plot for p_0 (left) and p_{high} (right). In both cases, smaller hyperparameters lead to slightly under-estimated expected probabilities. Overall, our method is robust to different values of p_0 , while p_{high} affects the calibration slightly more. We also compare the resulting 8 variants of γ GMM in terms of MAE. Overall, the posterior means produce similar values than our default setting, obtaining an MAE that varies from 0.252 ($p_{\text{high}} = 0.001$, the best) to 0.32 ($p_0 = 0.0001$, the worst).

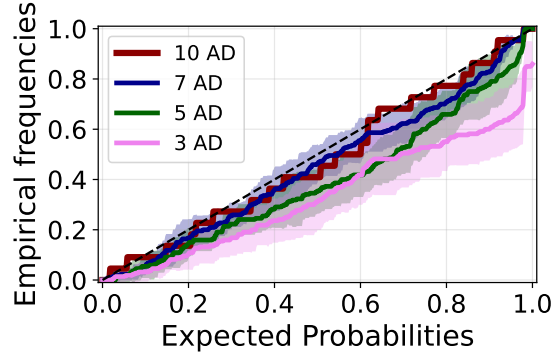


Figure 6: QQ-plot comparing the calibration curves of γ GMM when a different number M of detectors is used. The colored shades report the uncertainty obtained by sampling randomly the chosen amount of detectors in a set of 10 detectors. The plot shows that the higher the number of detectors, the more calibrated the distribution.

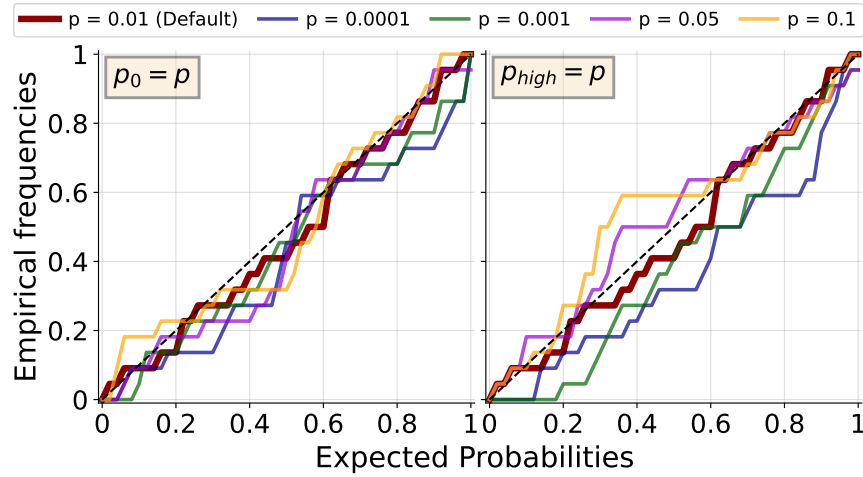


Figure 7: QQ-plot showing how calibrated γ GMM’s posterior mean would be if we varied p_0 (left) and p_{high} (right). While p_0 does not have a large impact on the method, the empirical frequencies slightly under (over) estimate the expected probabilities for low (high) values of p_{high} .

5 Conclusion

The literature on anomaly detection has, correctly, focused on unsupervised algorithms, but largely ignored practical challenges in their application. The algorithms are evaluated on performance metrics focusing on the ranking of the samples (e.g., AUC), and the ultimate choice of detecting the actual anomalies by thresholding the predictions is left for the practitioners. They lack good means for thresholding and thus often resort to using labels for such goal. This largely defeats the point of using unsupervised methods.

We presented the first practical method for estimating the posterior distribution of the contamination factor γ in a completely unsupervised manner. We empirically demonstrated on 22 datasets that our mean estimates effectively solve the question of where to threshold the predictions. We outperform all 21 comparison methods and show that the gap in detection accuracy between our estimate and the ground truth (available for these benchmark datasets) is small.

Besides solving the practical question of thresholding the predictions, we seek to transform the mindset of the anomaly detection community by presenting a fully probabilistic solution for the problem. Especially in unsupervised settings it would be completely unreasonable to expect the contamination factor could be identified exactly, but rather we need to characterize its uncertainty. However, we are not aware of any previous works even attempting this. As shown in Fig. 2,

the implied posterior distribution of γ may not only be wide but often also multi-modal. Communicating these aspects to the practitioner is critical so that they can e.g. use additional domain knowledge to interpret the possible alternatives. We showed that our estimates have near perfect calibration over the broad range of datasets used here and hence can be relied on in practical use.

On a first impression, the success of our method in solving this challenging and seemingly ill-posed problem may seem surprising. However, it can be attributed to a careful choice of strong inductive biases built into the underlying probabilistic model. We argue that all of the following elements are necessary, each substantially contributing to the overall success: (i) representing the data in the space of anomaly detector scores defines a meaning for the dimensions and allows borrowing inductive biases of arbitrary detector algorithms, (ii) the mixture model encodes a natural clustering assumption for both the normal samples and the anomalies, (iii) the ordering used for determining the final distribution incorporates both the location and shape of the mixture components in a carefully balanced manner, and (iv) the transformation from the ordering to probabilities is robustly parameterized via just two intuitive hyperparameters, enabling use of the same defaults for all cases.

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Supplementary Materials

This Supplement contains additional details about our method and the experiments.

6 Methodology

In this section we further explain (1) how to find the optimal values of our hyperparameters δ and τ , and (2) how to collect samples from our estimate of $\gamma|S$ posterior.

1. Finding the values of δ and τ . We introduce two new hyperparameters δ and τ that have the role of properly calibrating the sigmoid function. By setting the following equations:

$$p_0 = 1 - \mathbb{P}(c_1) = 1 - \frac{1}{1 + e^{(\tau + \delta \cdot r(\bar{\mu}_1, \bar{\Sigma}_1))}}$$

$$p_{\text{high}} = \mathbb{P}(\gamma \geq t|S) = \sum_{k=1}^K \mathbb{P}(c_1, \dots, c_k) \mathbb{P}\left(\sum_{j=1}^k \pi_j \geq t|S\right)$$

we aim at finding the unique (optimal) value of δ and τ . We solve the optimization problem using the least square optimizer as implemented in SKLEARN.²

However, while we can always find some values such that the first condition holds (on p_0), the second one can be satisfied only if $p_{\text{high}} \geq \mathbb{P}(\pi_1 \geq t)$. Experimentally π_1 has almost always low values, which means $\mathbb{P}(\pi_1 \geq t) = 0$ for $t = 0.15$. However, in case the such a constraint cannot be satisfied, we keep running again the variational inference method (with different starting point) for the DPGMM until the constraint on p_{high} holds. If this cannot happen or does not happen within 100 iterations, we reject the possibility of too high contamination factors and just set it to 0.

2. Sampling algorithm. After creating the M -dimensional space using the M anomaly detectors, we fit the DPGMM model and obtain an approximation for the posterior $p(\pi, \mu, \Sigma|S)$. Then we derive a sample from $p(\gamma|S)$ in four steps by repeating the next operations for all $k \leq K$. First, we draw a sample $\pi_k^{(z)}, \mu_k^{(z)}, \Sigma_k^{(z)}$ from π_k (Dirichlet), μ_k (Normal), Σ_k (Inverse Wishart). Second, we transform $\pi_k^{(z)}$ by taking the cumulative sum and obtain a sample $\sum_{j=1}^k \pi_j^{(z)}$. Third, we pass $\mu_k^{(z)}$ and $\Sigma_k^{(z)}$ through the sigmoid function (Eq.2) to get the conditional probabilities $\mathbb{P}(c_k | c_{k-1})$, and transform them into the exact joint probabilities $\mathbb{P}(c_1, \dots, c_k)$ using the equation 3. Finally, we multiply the samples following the Formula 4 and obtain a sample $\gamma^{(z)}$ from $p(\gamma|S)$.

7 Experiments

In this section we explain the threshold estimators and the ten anomaly detectors used by γ GMM to assign the anomaly scores. Moreover, we provide additional details on the datasets used, and we extend the results of Q1, Q2 and Q3.

Threshold estimators. In statistics there are several methods that, given mono-dimensional scores belonging to two distributions, set a threshold to split them. These methods can be applied in our setting to estimate the contamination factor as the proportion of examples greater than the set threshold. We cluster these statistical methods in 9 groups:

1. *Kernel-based.* FGD [Qi et al., 2021] and AUCP [Ren et al., 2018] both use the kernel density estimator to estimate the score density; FGD exploits the inflection points of the density’s first derivative, while AUCP uses the percentage of the total kernel density estimator’s AUC to set the threshold;
2. *Curve-based.* EB [Friendly et al., 2013] creates elliptical boundaries by generating pseudo-random eccentricities, while WIND [Jacobson et al., 2013] is based on the topological winding number with respect to the origin;
3. *Normality-based.* ZSCORE [Bagdonavičius and Petkevičius, 2020] exploits the Z-scores, DSN [Amagata et al., 2021] measures the distance shift from a normal distribution, and CHAU [Bol’shev and Ubaidullaeva, 1975] follows the Chauvenet’s criterion before using the Z-score;
4. *Regression-based.* CLF and REGR [Aggarwal, 2017] are two regression models that separate the anomalies based on the y-intercept value;
5. *Filter-based.* FILTER [Hashemi et al., 2019], and HIST [Thanammal et al., 2014] use, respectively, the wiener filter and the Otsu’s method to filter out the anomalous scores;

²https://docs.scipy.org/doc/scipy/reference/generated/scipy.optimize.least_squares.html

6. *Statistical test-based.* GESD [Alrawashdeh, 2021], MCST [Coin, 2008] and MTT [Rengasamy et al., 2021] are based on, respectively, the generalized extreme studentized, the Shapiro-Wilk, and the modified Thompson Tau statistical tests;

7. *Statistical moment-based.* BOOT [Martin and Roberts, 2006] derives the confidence interval through the two sided bias-corrected and accelerated bootstrap; KARCH [Afsari, 2011] and MAD [Archana and Pawar, 2015] are based on means and standard deviations, i.e., respectively, the Karcher mean plus one standard deviation, and the mean plus the median absolute deviation over the standard deviation;

8. *Quantile-based.* IQR [Bardet and Dimby, 2017] and QMCD [Iouchtchenko et al., 2019] set the threshold based on quantiles, i.e., respectively, the third quartile Q_3 plus 1.5 times the inter-quartile region $|Q_3 - Q_1|$, and the quantile of one minus the Quasi-Monte Carlo discrepancy;

9. *Transformation-based.* MOLL [Keyzer and Sonneveld, 1997] smooths the scores through the Friedrichs’ mollifier, while YJ [Raymaekers and Rousseeuw, 2021] applies the Yeo-Johnson monotonic transformations.

Anomaly Detectors. We use 10 anomaly detectors with different inductive biases: KNN [Angiulli and Pizzuti, 2002] assumes that the anomalies are far away from normals, IFOREST [Liu et al., 2012] assumes that the anomalies are easier to isolate, LOF [Breunig et al., 2000] exploits the examples’ density, OCSVM [Green and Richardson, 2001] encapsulates the data into a multi-dimensional hypersphere, AE [Chen et al., 2018] and VAE [Kingma and Welling, 2013] use the reconstruction error as anomaly score function in a, respectively, deterministic and probabilistic perspective, LSCP [Zhao et al., 2019a] is an ensemble method that selects competent detectors locally, HBOS [Goldstein and Dengel, 2012] calculates the degree of anomalousness by building histograms, LODA [Pevný, 2016] is an ensemble of weak detectors that build histograms on randomly generated projected spaces, and COPOD [Li et al., 2020] is a copula based method. All these methods are implemented in the python library PyOD [Zhao et al., 2019b].

Table 1: Properties of the 22 datasets used. For each dataset, we report the number examples, the number of original covariates, and the ground-truth contamination factor.

Dataset	# Examples (N)	# Covariates	True γ^*
ALOI	12384	27	0.0304
Anthyroid	7129	21	0.0749
Arrhythmia	271	259	0.0996
Cardiotocography	1734	21	0.0496
Glass	214	7	0.0421
InternetAds	1682	1555	0.0499
KDDCup99	48113	40	0.0042
Lymphography	148	47	0.0405
PageBlocks	5473	10	0.1023
Parkinson	53	22	0.0943
PenDigits	9868	16	0.0020
Pima	526	8	0.0494
Shuttle	1013	9	0.0128
SpamBase	2661	57	0.0500
Stamps	340	9	0.0912
T15	42125	10	0.0668
T21	18509	10	0.0529
WBC	223	9	0.0448
WDBC	367	30	0.0272
WPBC	160	33	0.0562
Waveform	3443	21	0.0290
Wilt	4655	5	0.0200

Data. Table 1 shows the details of the used datasets. The datasets vary in terms of number of examples (from around 50 to more than 48000), number of covariates (from 5 to more than 1500) and the contamination factor (from around 0.004 to more than 0.10). Note that even the highest contamination factor is around 0.10, confirming the general assumption of anomalies being rare.

Q1-Q2. γ GMM’s estimated distribution. Figure 8 shows our estimate of $\gamma|S$ ’s posterior on the 22 used datasets. Moreover, Table 2 shows the MAE between γ GMM’s sample mean and the true value γ^* on a per-dataset basis. With respect to the true value γ^* and out of 22 experiments, the sample mean is:

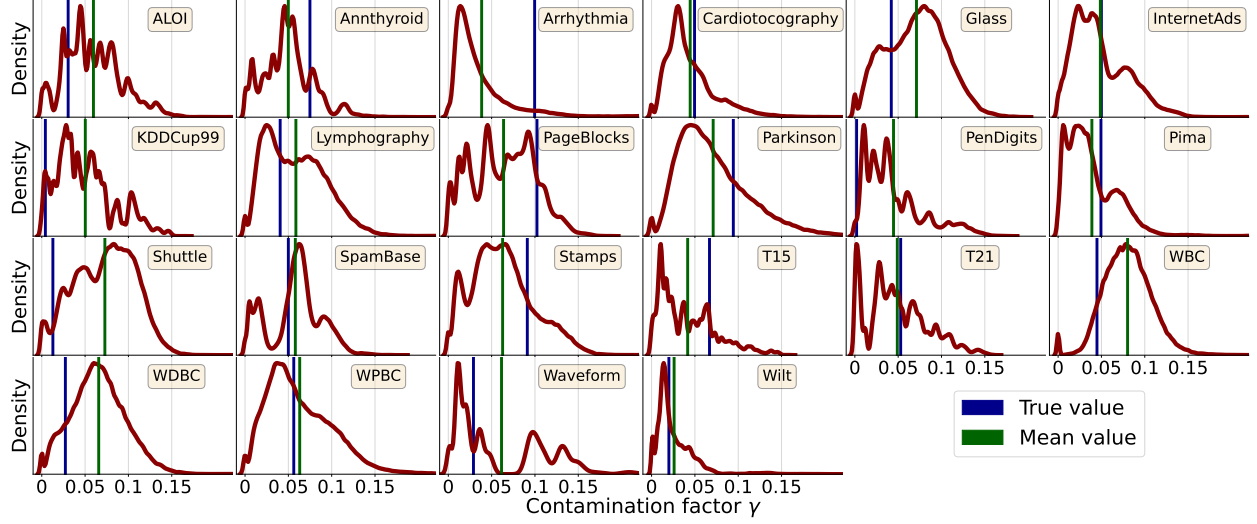

 Figure 8: Illustration of how γ GMM estimates γ 's posterior on the 22 datasets.

 Table 2: Mean Absolute Error (MAE) between the true contamination factor and γ GMM's sample mean for the 22 datasets.

Dataset	γ GMM's sample mean	True γ^*	MAE
ALOI	0.0596	0.0304	0.0292
Annthyroid	0.0499	0.0749	0.0250
Arrhythmia	0.0385	0.0996	0.0611
Cardiotocography	0.0446	0.0496	0.0050
Glass	0.0711	0.0421	0.0290
InternetAds	0.0487	0.0499	0.0012
KDDCup99	0.0502	0.0042	0.0460
Lymphography	0.0587	0.0405	0.0182
PageBlocks	0.0638	0.1023	0.0385
Parkinson	0.0711	0.0943	0.0232
PenDigits	0.0446	0.0020	0.0426
Pima	0.0390	0.0494	0.0104
Shuttle	0.0728	0.0128	0.0600
SpamBase	0.0580	0.0500	0.0080
Stamps	0.0627	0.0912	0.0285
T15	0.0417	0.0668	0.0251
T21	0.0490	0.0529	0.0039
WBC	0.0802	0.0448	0.0354
WDBC	0.0657	0.0272	0.0385
WPBC	0.0631	0.0562	0.0069
Waveform	0.0614	0.0290	0.0324
Wilt	0.0260	0.0200	0.0060

- a *good estimate* (i.e., $MAE \leq 0.01$) for 7 datasets (Cardiotocography, InternetAds, Pima, SpamBase, T21, WPBC, Wilt);
- a *slightly imprecise estimate* (i.e., $0.01 < MAE \leq 0.03$) for 7 datasets (ALOI, Annthyroid, Glass, Lymphography, Parkinson, Stamps and T15);
- a *not-optimal estimate* (i.e., $0.03 < MAE \leq 0.05$) for 6 datasets (KDDCup99, PageBlocks, PenDigits, WBC, WDBC, and Waveform);
- a *bad estimate* ($MAE > 0.05$) for just two datasets (Arrhythmia, and Shuttle).

This shows, again, that the estimated distribution is well-calibrated.

Table 3: List of detectors with the greatest F_1 score when using the true contamination factor to set the threshold. For each dataset, we use such subset of detectors to compute the deterioration.

Dataset	Anomaly Detectors
ALOI	KNN
Annthyroid	HBOS
Arrhythmia	IForest-HBOS-COPOD
Cardiotocography	KNN
Glass	LOF
InternetAds	LSCP
KDDCup99	COPOD
Lymphography	KNN-LOF-OCSVM-HBOS
PageBlocks	LOF
Parkinson	LSCP-HBOS-COPOD-LSCP-HBOS-COPOD
PenDigits	KNN-IForest-LOF-OCSVM-LSCP-Ae-VAE-HBOS-LODA-COPOD
Pima	IForest
Shuttle	KNN-OCSVM-Ae-VAE-HBOS-KNN-OCSVM-Ae-VAE-HBOS
SpamBase	LSCP
Stamps	LSCP
T15	OCSVM
T21	OCSVM
WBC	KNN-LOF-OCSVM-LODA-COPOD
WDBC	KNN-LOF-OCSVM-LSCP-Ae-VAE-LODA-COPOD
WPBC	OCSVM
Waveform	OCSVM
Wilt	LOF

Q3. Selecting the anomaly detectors to compute the F_1 score. Because we aim at studying the effect of the contamination factor on the detectors’ performance, we compare the F_1 scores only over the detectors that work well for each of the dataset. For each dataset D , we use as set of detectors those achieving the greatest F_1 score using the true contamination factor, i.e. $\arg \max_{f_m} \{F_1(f_m, D, \gamma^*)\}$. This means that, for each dataset, we (1) use each detector separately to make predictions using the true contamination factor γ^* , (2) measure their F_1 score, (3) keep those detectors that obtains the greatest F_1 , and (4) use them to compute the F_1 deterioration using the point-estimates of the contamination factor. Table 3 lists the detectors used for each dataset to compute the F_1 deterioration. Observe that sometimes only a single detector obtains the greatest F_1 score, while sometimes several detectors get the same F_1 score.

Q3. False alarms and false negatives. Finally, Table 4 shows the false alarm (false positive) rate and the false negative rate. The majority of the threshold estimators provide extremely high estimates for the contamination factor, shown here as extremely low false negative rates, but they would be useless in practice because of their high false alarm rate. In fact, this metric is important as false alarms result in real costs for the company (e.g., turning the wind turbine off to wait until the ice on the blades melts down), while reducing trust in the detection system. Our method reduces the false alarm rate compared to most of the baselines, including QMCD and KARCH that achieve the second and third best F_1 scores on average. On the other hand, IQR and MTT have the lowest false positive rates, due to the fact that they often underestimate the contamination factor as supported by the false negative table.

Table 4: Mean and standard deviation of the false alarm rate (left) and false negative rate (right) obtained by using each method's γ estimate to set the threshold (the lower the better). Regarding the false alarms, γ GMM has the third best mean and outperforms QMCD and KARCH, which are the second and third best baseline when measuring the F_1 score. On the other hand, γ GMM obtains higher false negative rates than almost all the competitors, due to the fact that the threshold estimators overestimate the true contamination factor.

False Alarm Rate		False Negative Rate	
Method	Mean \pm std.	Method	Mean \pm std
IQR	0.009 ± 0.008	BOOT	0.001 ± 0.002
MTT	0.027 ± 0.024	WIND	0.001 ± 0.002
γGMM	0.042 ± 0.015	MOLL	0.001 ± 0.002
QMCD	0.059 ± 0.018	EB	0.001 ± 0.003
KARCH	0.147 ± 0.047	MAD	0.002 ± 0.003
CHAU	0.190 ± 0.035	CLF	0.002 ± 0.003
ZSCORE	0.221 ± 0.050	GESD	0.003 ± 0.005
YJ	0.390 ± 0.139	REGR	0.003 ± 0.005
FILTER	0.454 ± 0.054	AUCP	0.004 ± 0.006
DSN	0.477 ± 0.134	HIST	0.006 ± 0.008
HIST	0.513 ± 0.100	FGD	0.007 ± 0.011
FGD	0.533 ± 0.183	DSN	0.007 ± 0.009
AUCP	0.591 ± 0.088	FILTER	0.007 ± 0.009
MCST	0.611 ± 0.287	MCST	0.007 ± 0.012
GESD	0.616 ± 0.106	YJ	0.009 ± 0.010
REGR	0.643 ± 0.105	ZSCORE	0.017 ± 0.015
MAD	0.731 ± 0.083	CHAU	0.019 ± 0.016
CLF	0.757 ± 0.077	KARCH	0.021 ± 0.018
EB	0.785 ± 0.077	QMCD	0.034 ± 0.024
WIND	0.809 ± 0.076	γGMM	0.036 ± 0.025
MOLL	0.816 ± 0.082	MTT	0.042 ± 0.028
BOOT	0.862 ± 0.079	IQR	0.044 ± 0.029